

Timo Vesikari
Pierre Van Damme
Editors



Pediatric Vaccines and Vaccinations

A European Textbook

 Springer

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Timo Vesikari

Vaccine Research Centre
University of Tampere Medical School
Tampere
Finland

Pierre Van Damme

Vaccine & Infectious Disease Institute
Centre for the Evaluation of Vaccination
University of Antwerp
Antwerpen
Belgium

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Preface

A European textbook of pediatric vaccines and vaccinations may be due and timely. Europe has contributed significantly to the development, testing, and production of many key vaccines. Europe also has advanced childhood immunization programs that include the latest vaccines and reach high coverage. There is no European unity on childhood immunization calendars – despite abortive attempts at “harmonization” – but the programs in most European countries have been very successful, showing that excellent results can be achieved using more than one approach. In terms of elimination and eradication, some European countries have been the first in the world or the most enduringly successful or both.

Historically, the important vaccine inventions were made in Europe. Edward Jenner’s smallpox vaccine was based on cross protection between animal and human pathogens at a time when the virus and indeed any concept of microbial pathogenesis were still unknown. With the global eradication of smallpox, this may be regarded as the most successful vaccine to date. Louis Pasteur’s rabies vaccine in 1885 was based on systematic attenuation of a pathogen and paved the way for many others. Diphtheria antitoxin and later vaccine were based on the works of von Behring and Ehrlich in Germany. Tuberculosis vaccine, bacille Calmette-Guérin or BCG, was developed in France in the 1920s and is still today in use in most parts of the world.

More recently, a well-known example of European development is hepatitis B vaccine (Engerix[®], GSK), the first licensed vaccine, in 1986, based on genetic engineering. A more recent high technology example is the first vaccine against group B meningococcus (Bexsero[®], Novartis), licensed in 2013, and developed using “reverse vaccinology.”

Other vaccinology landmarks are more geographically diverse. Polysaccharide protein conjugation technology was first developed in the USA, but many subsequent applications have taken place in Europe, including some *Haemophilus influenzae* type b (Hib) vaccines, one pneumococcal conjugate vaccine, and meningococcal C and ACWY conjugate vaccines.

Many live attenuated viral vaccines have been American developments, starting with the propagation of poliovirus in monkey kidney cells by Enders, Weller, and Robbins in 1948 that was followed by the development of inactivated (Salk) and live (Sabin) oral poliovirus vaccines (IPV, OPV). During the classical era of virology in the USA in the 1950s and 1960s, attenuated vaccines were developed against measles, mumps, and rubella, leading to the MMR combination vaccine. Live attenuated varicella vaccine was developed using the same technology, but eventually, the successful vaccine was developed in Japan.

Although the development of poliovirus vaccines was definitively an American success story, it should be noted that the major field study demonstrating the efficacy and safety of the Salk IPV had a significant European contribution (Finland). Moreover, the most widely used IPV today was developed in the Netherlands by van Wezel and coworkers in 1985. Europe also played an important part in the implementation of OPV. The vaccine was tested in the former Soviet republics of Estonia and Latvia resulting in the eradication of wild-type poliovirus in these countries, and this observation formed the basis of worldwide application of OPV for global eradication. Israel was the first country globally to introduce universal immunization plan against hepatitis A virus (HAV), an enteric virus closely related to poliovirus.

The political system in Eastern Europe made it possible to apply mandatory vaccination, with 99% coverage of measles vaccination, for example, in the former German Democratic Republic, and consequent total elimination of measles, in contrast to the much lower uptake and less effective disease control in the former Federal Republic of Germany. Mandatory vaccinations are, in general, not a favoured approach in Europe, but publicly available national medical care and immunization programs in many European countries have made it possible to reach high vaccine coverage without compulsion. MMR vaccine had been implemented in the USA for more than a decade when it was adopted using a two-dose schedule in Finland and Sweden in 1982, reaching over 95% coverage, and Finland became the first country to eliminate indigenous measles, mumps, and rubella by the early 1990s.

However, measles control is also a “dark area” for European vaccination programs. While the Americas have been able to eliminate measles, Europe has become a reservoir and a continuing source of measles virus to the Americas and elsewhere. Europe has been plagued by outbreaks of measles in many countries, because of failure to reach sufficient, sustained vaccine coverage to eliminate or eradicate measles virus. The reasons for this may vary from country to country, related to a multitude of factors allowing sufficient numbers of nonimmune individuals to sustain viral transmission following introductions.

On the bacterial side, Europe has a considerable track record of achievements in vaccination programs. Finland was the first country to eliminate invasive Hib disease and also carriage in the 1990s. The UK reacted quickly to increasing levels of meningococcal group C (MenC) disease and, in 1999, introduced an extensive vaccination program with newly developed MenC conjugate vaccines that resulted in drastic reduction and sustained control of the disease. The UK is also the first country

to have launched universal MenB vaccination of infants with the aim of controlling much of the remaining meningococcal disease.

The extensive trials in Sweden, Germany, and Italy have been crucial in demonstrating the efficacy of acellular pertussis vaccines.

The use of pentavalent and hexavalent pediatric combination vaccines in public health immunization programs is characteristic of Europe, in contrast to the USA where adoption of such multivalent formulations has been much slower. These vaccines have greatly facilitated the introduction of new antigens into infant programs (e.g., hepatitis B) and control of all diseases covered by the combinations by facilitating overall high levels of coverage of “routine” infant immunizations in Europe. Still, universal hepatitis B vaccination immunization has yet to be achieved in some countries in the north of Europe that use pentavalent (without hepatitis B) combination vaccines rather than hexavalent.

Immunization calendars vary between European countries, but the European experience largely shows that childhood diseases covered by the combination vaccines can be controlled regardless of variation between schedules; thus, there is no absolute need for “harmonization.” Clear recommendations on the minimal number of vaccines and vaccine antigens needed by certain ages might be more helpful to European countries, particularly those with less well-developed programs, than attempts to implement a harmonized schedule. Some countries use the same 2-, 4-, and 6-month schedule as the USA, while others have adopted an “accelerated” schedule of 2, 3, and 4 months followed, in some cases and for some antigens, by booster doses at or around 12 months of age. A 2 + 1 immunization schedule was first introduced in Italy and Scandinavia as a 3-, 5-, and 11-month calendar and is now used also elsewhere, including Israel

(2, 4, and 12 months). This shortened schedule is more convenient and cheaper and seems to be sufficient to control the diseases included in the hexavalent vaccine provided that the immunization coverage is sufficiently high. This variation between immunization schedules across Europe may be regarded as a richness. Scientifically, it confers a wealth of evidence on the impact of numbers of vaccine doses and flexibility of schedules. They show that there is no single correct way of preventing childhood infectious diseases by vaccination and that this goal can be reached in more than one way.

Given the diversity of cultures and opinions that exist in Europe, it is perhaps unsurprising that anti-vaccine groups have achieved some prominence in many countries. In general, anti-vaccine misinformation and “alternative truth” has not had much impact on the coverage of routine programs for infants. In contrast, MMR vaccination has suffered on occasion as, for example, in the late 1990s in the UK, where vaccine coverage was severely affected for several years leading to subsequent outbreaks of measles in unvaccinated children. Overall, newer vaccines tend to be less well accepted than established ones, and this may in part be due to anti-vaccine group activity. Conversely, new platforms for vaccination, such as during pregnancy, may be surprisingly well adopted both by the general public and by healthcare work-

ers, especially where the rationale for vaccination is comprehensive and clear.

Parents and the public in general actively seek information about vaccines on the Internet. Unfortunately, much of the most prominent and accessible information flow is alarmingly negative and critical about vaccination. Against this background, it is particularly important that European students and practitioners of medicine and other healthcare professionals become better taught and are provided with accurate, accessible, and up-to-date information about vaccines and their use.

This book aims to provide just such essential information on the current vaccines that are used in childhood immunization programs in Europe and the principles which underlie them. The book is written by leading European experts on the topics that they know best. This ensures that the subject matter is covered comprehensively even though the text is compact. We believe that we have been able to produce a book that is both readable and thorough. We hope the textbook will find a readership not only among “vaccinologists” but also among European and other pediatricians and practitioners who deal with vaccines and vaccinations. We also hope that the book will be used in the curriculum of future doctors, pharmacists, midwives, and nurses for their vaccinology classes in Europe and perhaps even more widely.

Timo Vesikari

Tampere, Finland

Pierre van Damme

Antwerpen, Belgium

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Contributors

R.H. Behrens

London School of Hygiene
and Tropical Medicine
London, UK
Ron.Behrens@lshtm.ac.uk

Shalom Ben-Shimol

Faculty of Health Sciences, Ben-Gurion
University of the Negev
Beer-Sheva, Israel
shalomb2@clalit.org.il

Paolo Bonanni

Department of Health Sciences,
University of Florence
Florence, Italy
paolo.bonanni@unifi.it

Robb Butler

WHO Regional Office for Europe
Copenhagen, Denmark
RBU@euro.who.int

Ron Dagan

Faculty of Health Sciences, Ben-Gurion
University of the Negev
Beer-Sheva, Israel
rdagan@bgu.ac.il

Ener Cagri Dinleyici

Eskisehir Osmangazi
University Faculty of Medicine,
Department of Pediatrics TR-26480
Eskisehir, Turkey
timboothtr@yahoo.com

Susanna Esposito

Pediatric Clinic, Università degli Studi di
Perugia, Perugia, Italy
susanna.esposito@unimi.it

Adam Finn

UHB Education Centre
Bristol Children's Vaccine Center
Bristol, UK
Adam.Finn@bristol.ac.uk, cdahrf@bristol.ac.uk

Nathalie Garçon, PharmD, PhD

Bioaster, Lyon, France
Nathalie.GARCON@bioaster.org

Carlo Giaquinto

Department of Women and Child Health
University of Padova
Padua, Italy
carlo.giaquinto@unipd.it

Paul T. Heath

Paediatric Infectious Diseases Research
Group & Vaccine Institute
St Georges, University of London
London, UK
pheath@sgul.ac.uk

Ulrich Heininger

Pediatric Infectious Diseases
and Vaccinology University of
Basel Children's Hospital
Basel, Switzerland
ulrich.heininger@ukbb.ch

Tapani Hovi

National Institute for Health and Welfare
Helsinki, Finland
tapani.hovi@thl.fi

Chrissie E. Jones

Faculty of Medicine and
Institute for Life Sciences
University of Southampton and
University Hospital Southampton NHS
Foundation Trust
Southampton, UK
cjones@sgul.ac.uk

Herwig Kollaritsch

Institute for Specific Prophylaxis and Tropical
Medicine, Medical University of Vienna
Vienna, Austria
herwig.kollaritsch@meduni.wien.ac.at

Kirsty Le Doare

Centre for International Child Health
Imperial College
London, UK
k.mehring-le-doare@imperial.ac.uk

Johannes Liese

Department of Pediatrics
University of Würzburg
Würzburg, Germany
Liese_J@ukw.de

Carlos Martin, MD, PhD

Microbiology at the Faculty of Medicine
at University of Zaragoza and member
of the Advisory Committee of
Tuberculosis Vaccine Initiative (TBVI)
Zaragoza, Spain

CIBER Enfermedades Respiratorias
Instituto de Salud Carlos III
Madrid, Spain

Servicio de Microbiología, Hospital
Universitario Miguel Servet, ISS Aragón
Zaragoza, Spain
carlos@unizar.es

Federico Martinon-Torres, MD, PhD

Pediatrics, Hospital Clínico
Universitario de Santiago
Santiago de Compostela, Spain

Genetics, Vaccines and Infectious Diseases
Research Group (GENVIP)
Healthcare Research Institute
of Santiago (IDIS), University of Santiago
Santiago de Compostela, Spain
Federico.Martinon.Torres@sergas.es

Andrew J. Pollard

Oxford Vaccine Group, Department of
Paediatrics, University of Oxford and NIHR
Biomedical Research Centre, Oxford University
Hospitals NHS Trust
Level 2, Children's Hospital
Oxford, UK
andrew.pollard@paediatrics.ox.ac.uk

N. Prevatt

London School of Hygiene
and Tropical Medicine
London, UK
drnatprevatt@hotmail.com

Francesca Rocchi

University-Hospital Paediatric Department
Bambino Gesù Children's Hospital
Rome, Italy
Francesca.Rocchi@pentafoundation.org

Manish Sadarangani

Oxford Vaccine Group, Department of
Paediatrics, University of Oxford and NIHR
Biomedical Research Centre
Oxford University Hospitals NHS Trust
Oxford, UK

Vaccine Evaluation Center, BC Children's
Hospital, University of British Columbia
Vancouver, BC, Canada
manish.sadarangani@paediatrics.ox.ac.uk

Mary P.E. Slack, MA, MB BChir, FRCPath

School of Medicine, Griffith University
Nathan, QLD, Australia

Haemophilus Reference Unit, Respiratory &
Vaccine Preventable Bacteria Reference Unit
Public Health England
Colindale, London, UK

WHO Collaborating
Centre for Haemophilus influenzae
London, UK
mpe.slack@gmail.com

Matthew D. Snape

Oxford Vaccine Group, Department of
Paediatrics, University of Oxford and NIHR
Biomedical Research Centre
Oxford University Hospitals NHS Trust
Oxford, UK
matthew.snape@paediatrics.ox.ac.uk

Vana Spoulou

Department of Infectious Diseases,
University of Athens
Athens, Greece
vspoulou@med.uoa.gr

Vytautas Usonis

Vilnius University Faculty of Medicine
Clinic of Children Diseases
Vilnius, Lithuania
vytautas.usonis@mf.vu.lt

Pierre Van Damme

Centre for the Evaluation of Vaccination
Vaccine & Infectious Disease Institute
University of Antwerp
Antwerp, Belgium
pierre.vandamme@uantwerpen.be

Timo Vesikari

Vaccine Research Center
University of Tampere
Tampere, Finland
Timo.Vesikari@staff.uta.fi

General Vaccinology

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Expected and Unexpected Effects of Vaccination

Federico Martinon-Torres

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1.1 Introduction

Vaccination is widely considered to be one of the greatest medical achievements of civilization and one of the top major breakthroughs of humanity.

From an almost empirical origin of vaccinology to the present vaccinomics, our knowledge has evolved substantially and we have learned important lessons. Although the main target of a vaccine is direct protection against a particular microorganism or disease, the scope of vaccination has expanded with the discovery that vaccines can also protect unvaccinated people through herd protection, or even that certain vaccines can protect against additional diseases different from those that they were designed to prevent, through so-called heterologous effects.

1.2 Effectiveness and Impact of Vaccination

Disease prevention through vaccination is the most cost-effective health care intervention available. The World Health Organization (WHO) estimates that every year immunization saves between two and three million lives across the

world. One hundred years ago, infectious diseases were the main cause of death worldwide, even in the most developed countries. Today, common childhood diseases of previous generations are becoming increasingly rare thanks to vaccines, and there are new vaccines on the horizon with the potential to prevent even more.

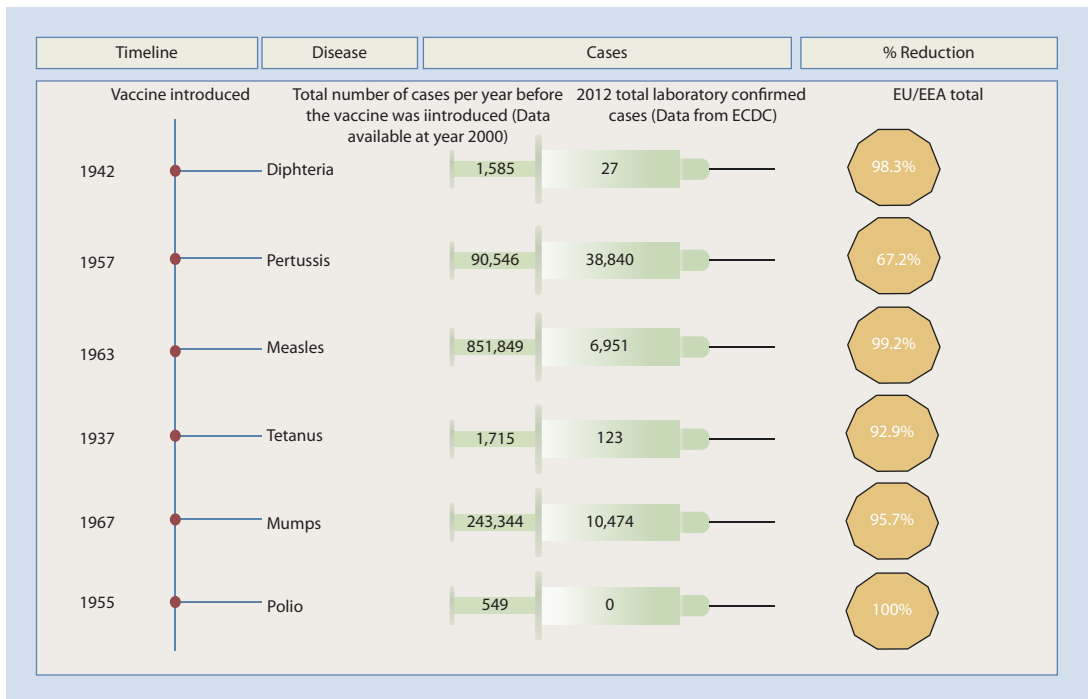
Mass immunization programs have proven successful at controlling or even eliminating disease (■ Fig. 1.1).

1.2.1 SmallPox

Before a vaccination campaign eliminated all natural occurrences of smallpox in 1980, the disease threatened 60% of the world's population and killed 1 in 4 patients. Approximately 350 million people are estimated to have been spared from smallpox infection, and 40 million from dying, since the disease was eradicated.

1.2.2 Measles

Between 2000 and 2014, deaths from measles dropped by 79% worldwide, preventing an



■ Fig. 1.1 Effectiveness and impact of the introduction of various vaccines in Europe

estimated 17.1 million deaths and making the measles vaccine one of the best buys in public health. Since 1974, the number of reported measles deaths has dropped from 2 million to 150,000 per year, although the fight to eradicate the disease is still under way for reasons other than vaccine effectiveness. Measles eradication is in sight if we are able to deal with hesitancy regarding vaccination and anti-vaccine lobbies, and to maintain vaccination coverage at an adequate level.

1.2.3 Polio

Total eradication of polio is within our reach. Since the creation of the Global Polio Eradication Initiative in 1988 by the WHO and its partners, reported cases of polio have fallen by 99%, with paralysis being prevented in an estimated ten million people.

1.2.4 Haemophilus

The conjugate vaccines are effective tools for preventing Hib infections, which were the most common severe invasive childhood infections in industrialized countries. Several prospective studies have shown an efficacy exceeding 90% from the first months of life. The impact of vaccination in different European countries is summarized in [Table 1.1](#).

1.2.5 Diphtheria

Before vaccination against diphtheria became readily available in the 1980s, it is estimated that approximately one million cases occurred in the countries of Eastern Europe each year. Although diphtheria is still present in some European countries and epidemics broke out in Eastern Europe during the 1990s, it is now drastically reduced thanks to vaccination.

1.2.6 Invasive Pneumococcal Disease

Several European countries have reported a significant decline in rates of invasive pneumococcal infection and mucosal forms of pneumococcal

disease (mainly otitis and pneumonia) as a result of pneumococcal conjugate vaccination. This benefit also seems to have spread to unvaccinated populations through indirect protection.

1.2.7 Invasive Meningococcal Disease

Mass vaccination of children and adolescents with group A + C meningococcal conjugate vaccine, together with routine childhood immunization, have yielded reductions in hospitalization and mortality in Africa. In Europe, meningococcal group C (MenC) infections and deaths decreased by more than 90% after the deployment in 1999 of a vaccination campaign with a MenC conjugate vaccine in the UK. A similar result was found in other countries that included the MenC vaccine in their schedules, such as the Netherlands or Spain.

1.2.8 Rotavirus

Within 8 years of their initial introduction into Europe, rotavirus vaccines have been shown to be highly effective, with a substantial impact on the rotavirus gastroenteritis-related health care burden, including hospitalizations, nosocomial infections, and outpatient visits. These findings are consistent in several studies and countries across Europe, and comparable with observations from Australia and the USA. Some examples show a >95% effectiveness in the reduction of hospital admissions for rotavirus gastroenteritis in several European countries (Finland, Spain, France, and the UK) and a >60% reduction in the number of hospital admissions and emergency-department visits in countries with universal rotavirus vaccination (e.g., Austria, UK, Finland, and Belgium).

1.3 Expanded and Unexpected Effects

The main expected benefit from vaccination is protection against the pathogen for which it is designed. This is a direct effect on a particular target infection. For many years, however, epidemiological data indicated some unexpected, beneficial effects brought about indirectly by some vaccines. These expanded and somewhat

Table 1.1 Annual <i>Haemophilus</i> cases prevented by conjugate vaccines in children aged 0–4 years in various European countries									
Country and year of comparison	Number of children 0–4 years old	Incidence before vaccination		Cases/year before vaccination		Incidence after vaccination		Cases prevented by vaccination/year	
		Meningitis	All entities	Meningitis	All entities	Meningitis	All entities	Meningitis	All entities
Scandinavia 1970s vs 1995	1,581,000	31	51	490	810	<1	1	470	770
Austria, Vienna, 1991 vs 1993–1996	485,000	11		55		<1		50	
France – Val-de-Marne, 1980s vs 1992–1993 Whole country	45,000		21–25	>500	18		4		15
	3,777,000		23		870		4	420	720
Germany – Rhein–Main area, 1989 vs 1993–1995 Whole country	100,000		33	950	33	0.8	1	900	1800
	4,115,000	23	46		1900	0.9	1.3		
Ireland, 1991–1993 vs 1995	260,000		25		65		2.6		60
The Netherlands, 1970s vs 1993–1994	981,000	22–40	80	390	78	0.3	1	385	770
Spain, Basque region, 1993–1995 vs 1997	80,000	14	21	13	18	0	2	13	16
Switzerland, 1976–1990 vs 1991–1993	447,000	26	84	115	375	8	10	80	330
United Kingdom – England and Wales, 1991–1992 vs 1993–1994 Whole country	3,434,000	15	31	515	1060	0.6	2	500	990
	3,831,000	24	36	920	1380	0.6	1	895	1340

unexpected effects have broadened the benefits of vaccines. Using these mechanisms, it is possible to generate direct protection against antigens different from the immunogen contained in the vaccine (cross-protection), protect or even eradicate a disease without having to vaccinate the entire population (indirect protection), or even protect against pathogens different from those targeted by the vaccine (heterologous protection).

1.3.1 Cross-Protection and Heterologous Immunity

The concept of cross-protection denotes the ability of the immune system to recognize various antigens that differ from the immunogen, through certain flexibility in peptide recognition (*cross-immunity*). For this reason, different antigens appear similar to the immune system, thereby challenging the theoretical specificity postulated by the clonal selection theory. To understand this issue, it is useful to distinguish between cross-neutralization and cross-protection. In *cross-neutralization*, antibodies elicited by vaccination with a certain serotype neutralize other serotypes in vitro. *Cross-protection* means that immunization with a certain vaccine type provides clinically significant protection against infection or disease (or both) owing to another serotype, i.e., that the cross-neutralizing response has a functional impact.

One example is the HPV vaccine. Immunity to HPV is type-specific. However, if we look at the phylogenetic tree that includes the various HPV types, we observe that some degree of cross-protection is possible, given the high level of homology of some viral types with vaccine types. This is the case, for instance, for HPV-31 and -35 (strictly related to HPV-16), and for HPV-45 (strictly related to HPV-18). Another example can be seen with rotavirus vaccines. The antibodies elicited by these vaccines not only protect against those circulating strains sharing the same G or P variant as that contained in the vaccine strain, but also other non-matching G and P strains (heterotypic protection). According to this, type-specific antibodies targeted at neutralizing VP7 or VP4 epitopes are not solely responsible for their protective effect. The comparable effectiveness of RV1 and RV5 reinforces this conclusion: neutralizing antibody titers induced by RV1 or

RV5 consistently underestimates the protection conferred by the vaccine. Other examples of this cross-reactivity have been confirmed in humans, involving influenza virus-specific immunity, or pneumococcal conjugate vaccines, among others.

Cross-protection was described five decades ago and later termed *heterologous immunity*. The initial observation was that CD8⁺ - T cells are able to cross-recognize peptides from two distinct viruses and may play roles not only in protective immunity, but also in immunopathology (autoimmunity). According to this phenomenon, memory T cells that are specific to one pathogen can become activated during infection with an unrelated heterologous pathogen. As such, previous host exposure to unrelated infectious agents can greatly alter immune response to an infection. T cells recognize processed peptides that are presented at the cell surface in antigen-binding grooves of class I major histocompatibility class (MHC) proteins. At the same time, the T-cell receptor (TCR) binds to the peptide-MHC complex. Thus, a TCR that recognizes a given MHC-presented peptide may also recognize other peptides that fit into the appropriate MHC groove, and has amino acid chains that are able to bind to TCR. This degeneration of the T-cell recognition is called *molecular mimicry* when the cross-reactive peptide has similar determinants and interacts with TCR in the same manner as the original peptide. It is called *alternative recognition* when different determinants of the TCR are involved in recognition. A third explanation for cross-reactivity is when a given T cell expresses two different TCRs as a result of an incomplete allelic exclusion of a second TCR chain; in this way, the two distinct TCRs formed may recognize different antigens.

When the term cross-protection is applied to *vaccination*, it typically refers to *heterosubtypic immunity* defined as protection by virus (influenza is the best-known case) of one strain, against a challenge infection with other strains differing in subtype. However, very recently, cross-protective immunity has also been highlighted as one of the mechanisms for the unexpected beneficial effects of BCG vaccination on infections other than tuberculosis. Researchers showed that BCG vaccination induces a long-lasting, nonspecific potentiation effect of heterologous T-helper responses, Th1 (IFN-gamma) and Th17 (IL-17 and IL-22), to non-mycobacterial stimulation. Previously,

other authors had demonstrated that both effector and memory CD8+ cells had the potential to secrete IFN-gamma in the absence of related antigens. According to these findings, vaccination can provide not only a heterosubtypic protection, but also heterologous protection through a cross-immunity mechanism.

1.3.2 Indirect Protection

The term “herd immunity” was coined a century ago, but its use has become widespread in recent decades to describe the reduced risk of infection among susceptible individuals in a population, induced by the presence and proximity of vaccinated individuals. Herd immunity makes it possible to protect a whole community from infectious disease by immunizing a critical percentage of the population. Just as a herd of sheep uses its sheer number to protect individual members from predators, herd immunity protects a community from infectious disease by virtue of the number of immune individuals. The more members of a human herd are immunized, the better protected the whole population will be from an outbreak of disease.

The terms *herd immunity* and *herd effect* are frequently used indistinctly, but they do not reflect the same concept. *Herd immunity* refers only to the proportion of subjects immunized in a given population, while the *herd effect* is used to describe the indirect protection observed in the non-immunized segment of the population. Furthermore, herd immunity applies to immunization or infection, human to human transmission. Conversely, the herd effect applies exclusively to immunization achieved by vaccination or other health intervention that reduces the probability of transmission.

Vaccination has been revealed as an indirect way of protecting members of the community who cannot be vaccinated. Vaccinated individuals protect themselves from disease, but also, moreover, they prevent the spread of the infectious agent and limit potential disease outbreaks. The herd effect achieved through vaccination for a given disease depends on the efficacy and coverage of the vaccine in addition to the transmissibility of the infection.

There are numerous examples of herd immunity, illustrating the importance of indirect pro-

tection for predicting the impact of vaccination programs. The basis for the herd effect is that individuals who are immune to a disease act as a barrier in the spread of disease, slowing or preventing the transmission of disease to others. When a given proportion of the population – known as the herd immunity threshold – becomes immunized, the disease may no longer persist in this population. This threshold is defined based on the “basic reproduction number” (R_0), which represents the number of people in an unprotected population that could receive the disease from one infected individual. The more contagious the disease, the higher this number, and thus the higher the threshold to be reached to protect the community. For example, measles, an extremely contagious disease, has a threshold of 95% to ensure community protection. On the other hand, mumps, which is not as contagious, needs a threshold of 80% (■ Fig. 1.2, ■ Table 1.2).

A clear example of herd protection is the case of the meningococcal serogroup C conjugate vaccine in the UK, the Netherlands, and subsequently in other countries. The impact of this vaccine on the prevalence of the disease was higher than expected according to the population covered with the vaccine, also reducing the number of cases in a nonvaccinated population. This indirect protection was due to the high efficacy of the vaccination at preventing nasopharyngeal carriage and thus, spreading of the pathogen to the rest of the population.

Mass vaccination is the best way to rapidly increase herd immunity either for accelerating disease control and to rapidly increase coverage with a new vaccine or in the setting of an existing or potential outbreak, thereby limiting the morbidity and mortality that might result.

Even if the increase in population immunity is not sufficient to achieve infection elimination owing to low vaccine efficacy or insufficient coverage, the risk of infection among unvaccinated persons may still be reduced. This may be particularly important for those for whom vaccination is contraindicated. The paradox is that for an individual, with regard to vaccination in a population, the best option is that everybody else is vaccinated and the individual is not. This way the individual is protected from infection because of the herd effect, but suffers none of the potential adverse effects of vaccination. Finally, these indirect effects may eventually be deleterious, if

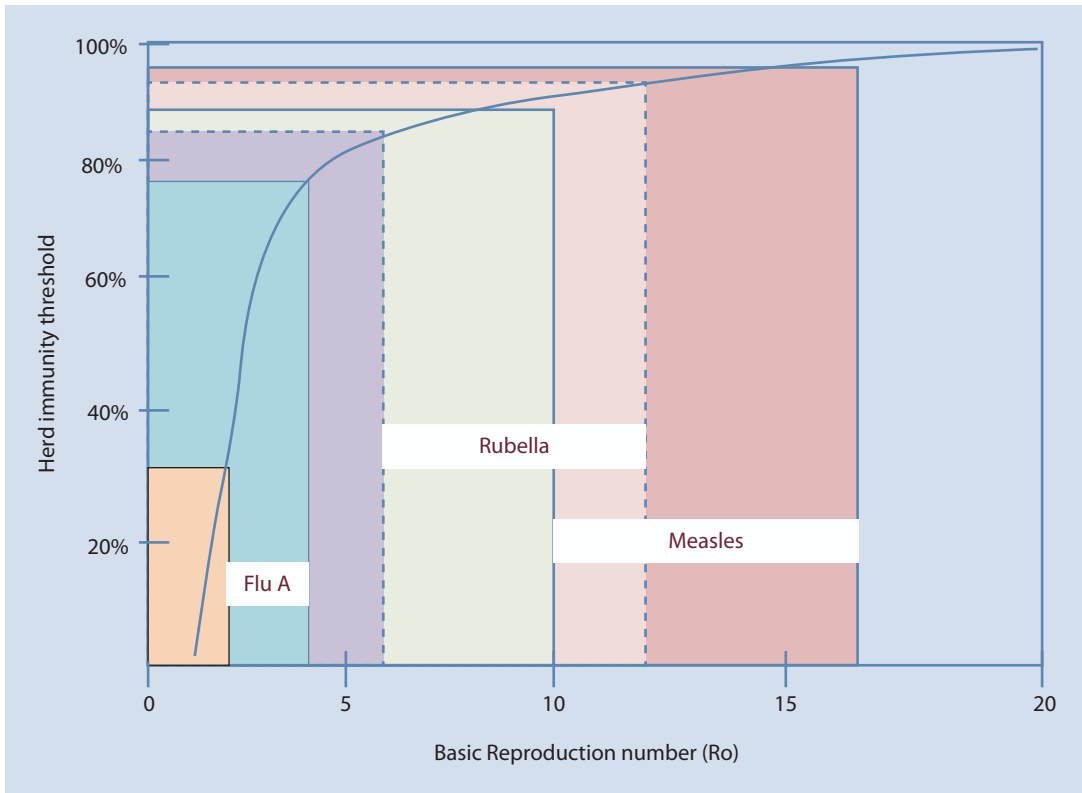


Fig. 1.2 Simple threshold concept of herd immunity. Relationship between the herd immunity threshold, $(R_0 - 1)/R_0 = 1 - 1/R_0$, and basic reproduction number, R_0 , in a randomly mixing homogeneous population. Note the

implications of ranges of R_0 , which can vary considerably between populations, for ranges of immunity coverage required to exceed the threshold

Table 1.2 R_0 values for well-known infectious diseases

Infectious disease	Transmission	R_0
Measles	Air transmission	12–18
Whooping cough	Airborne droplets	12–17
Diphtheria	Saliva	6–7
Smallpox	Social contact	5–7
Polio	Fecal–oral	5–7
Rubella	Airborne droplets	5–7
Mumps	Airborne droplets	4–7
HIV	Sexual contact	2–5
SARS	Airborne droplets	2–5
Influenza	Airborne droplets	2–3
Ebola	Contact with body tissues or fluids	2–3

as a consequence of reducing the risk of infection among those susceptible, there is a displacement of the risk of infection to other age groups and/or to a more vulnerable population, as has been suggested for varicella or hepatitis A in certain scenarios.

1.3.3 Heterologous (Nonpecific) Effects of Vaccination

Some vaccines can broadly enhance immune responses to a range of distinct pathogens or even to other vaccines, indicating that immune protection may be influenced by previous exposure to unrelated microorganisms or microbial components. First described for BCG vaccine, epidemiologists showed a reduction in mortality or hospitalization rates in the BCG-vaccinated population versus the nonvaccinated that could not be explained by the reduction in deaths due to the prevented pathogen. In recent years, a plethora

of scientific papers have documented this unexpected effect of vaccination, and explained it as resulting from an indirect action of vaccines on the immune system, other than their specific expected effect. These so-called heterologous effects of vaccines are now being explored not only for BCG – the most frequently studied in this regard – but also for polio, measles, influenza, rotavirus, and others. Scientific data reveal a dual mechanism for these heterologous properties of vaccines: cross-protective immunity (an old and well-known phenomenon described above) and immune training, a new and revolutionary concept referring to the innate immunological memory and its ability to be trained through vaccination.

Immunological memory, or the ability to remember the encounter with a pathogen, used to be considered an exclusive virtue of the adaptive immune system. For some years now, this concept is changing and immunological memory is recognized too as an ability of the innate host defense. *Immune training* is the term applied to this recently described feature of innate immunity, and its demonstration in humans was first documented with BCG vaccination by Kleinnijenhuis et al.: they showed a BCG-induced trained immunity mechanism of nonspecific protection from infections through epigenetic reprogramming of innate immune cells as monocytes. This revolutionary concept represents a plausible explanation for the rapid protective effects observed after BCG vaccination, unexplained by the cross-protective effect of the adaptive system – the latter, with long-term effects but slow to develop.

According to this concept, vaccination would induce an enhanced innate immunity state mediated by natural killer or monocytes/macrophages, which would provide nonspecific protection against nonrelated infections. As a consequence of vaccination, innate immune cells become more efficient cells, and better responders against microbial aggressions. Epigenetic and metabolic modifications during innate cell development in the bone marrow would be responsible for the maintenance of these enhanced features to influence the functions of innate cells for longer periods. Epigenetic reprogramming of cells through tri-methylation of histones leads to a stronger gene transcription upon re-stimulation through the NOD2 receptor, an intracellular pattern recognition receptor (PRR). Metabolic processes would also be affected, with a cell metabolic shift toward

an aerobic glycolytic (transformation of pyruvate to lactate) pathway, as opposed to the classic and less efficient aerobic oxidative phosphorylation of pyruvate. This shift of glucose metabolism is also known as the “Warburg effect,” and allows the rapid production of energy for the proliferation of cancer cells or activated lymphocytes.

This epigenetic and metabolic reprogramming is not the only mechanism involved in the immune training of innate cells. Other mechanisms include an increased expression of recognition receptor pathogens (PRRs) on the cell surface following vaccination, and enhanced cytokine release, particularly inflammatory signals for a protective function.

Future research should seek a better understanding of innate immune training mechanisms induced by vaccines, including the impact of age, host genetics, geographical location, and sociological factors. It is also important to explore the timing and the combination of vaccines to avoid negative side effects and fully exploit their potential benefits. This will help us to improve the beneficial heterologous effects of vaccination. In addition, vaccines that were removed from the immunization schedule could now be re-considered in view of these beneficial nonspecific effects.

Positive Heterologous Effects

The paradigmatic case of vaccines providing heterologous benefits is that of *bacillus Calmette-Guérin* (BCG). Several randomized controlled trials have indicated that BCG, a vaccine introduced in 1921 to fight against tuberculosis, has beneficial, heterologous, nonspecific effects in children from developing countries, reducing morbidity and mortality caused by unrelated pathogens. Old epidemiological data had already pointed toward a protective nonspecific effect, without a mechanism that could explain it. More recently, it has been demonstrated that this beneficial effect was not restricted to developing countries, with reduced early childhood hospitalization and mortality rates also observed in high-income settings.

Apart from this heterologous effect on mortality and hospitalization of children, BCG has been revealed in recent years to be a potent immunomodulator, with potential applications in the treatment of immune-based disorders (type 1 diabetes and multiple sclerosis) and as immunotherapy for treating early-stage bladder cancer.

There are, however, reports describing heterologous effects for other vaccines, either live or attenuated. Similar to the BCG vaccine, *measles-containing vaccines* have been demonstrated to reduce mortality and hospital admissions from causes other than measles infection, in both low- and high-income countries. Incidence of infectious diseases other than measles has been found to correlate strongly with incidence of measles in different countries, in both pre- and post-vaccine periods. It has been recently described that the prevention of immunosuppressive effects of measles infection through vaccination might explain these long-term benefits of measles vaccination.

The effect of *oral polio vaccine* (OPV) on mortality has only been assessed in a few studies, which concluded that OPV is associated to lower infant mortality and morbidity through these non-specific effects. The observations of this beneficial effect of OPV have generated a controversial debate on the substitution of oral polio vaccine for the inactivated polio vaccine, and the possible consequences of this decision on the mortality increment.

Negative Heterologous Effects

Negative heterologous effects are also possible. An association between the AS03-adjuvanted influenza pandemic vaccine and the development of narcolepsy has been described in some children and infants due to cross-reactivity to host antigens. In this case, molecular mimicry between a fragment of one of the influenza antigens (nucleoprotein) and a portion of the human brain receptor that promotes wakefulness (hypocretin receptor 2) has been reported as an explanation for this heterologous effect.

Unlike BCG, measles vaccine or OPV, the diphtheria–tetanus–pertussis (DTP) vaccine has not shown the same beneficial effect, and in fact some studies have suggested detrimental effects on children's survival. In 2013, a strategic advisory group of experts commissioned by the WHO reviewed all evidence concerning possible nonspecific effects of DTP-containing vaccines on survival and all-cause mortality in children under 5 years of age, concluding that findings on DTP vaccines were inconsistent. Further research into the potential nonspecific effects of DTP vaccines is warranted. Based on current knowledge, it is suggested that the order in the administration of DTP vaccines with other scheduled vaccines

(especially BCG) is important in the generation of these nonspecific effects, as DTP seems to oppose the positive heterologous effects of live vaccines.

In summary, vaccine effectiveness and impact have exceeded expectations, often ahead of our actual understanding of all the mechanisms behind this success. We are now beginning to understand these mechanisms for the oldest vaccines, and are now applying this knowledge to the design of the next generation of vaccines.

Further Reading

- Aaby P, Roth A, Ravn H, et al. Randomized trial of BCG vaccination at birth to low-birth-weight children: beneficial non-specific effects in the neonatal period? *J Infect Dis*. 2011;204:245–52.
- Benn CS, Jacobsen LH, Fisker AB, Rodrigues A, Sartono E, Lund N, Whittle HC, Aaby P. Campaigns with oral polio vaccine may lower mortality and create unexpected results. *Vaccine*. 2017;35(8):1113–6. <https://doi.org/10.1016/j.vaccine.2016.11.006>.
- Byberg S, Thysen SM, Rodrigues A, Martins C, Cabral C, Careme M, Aaby P, Benn CS, Fisker AB. A general measles vaccination campaign in urban Guinea-Bissau: comparing child mortality among participants and non-participants. *Vaccine*. 2017;35(1):33–9. <https://doi.org/10.1016/j.vaccine.2016.11.049>.
- Contreras G. Effect of the administration of oral poliovirus vaccine on infantile diarrhoea mortality. *Vaccine*. 1989;7:211–2.
- De Castro MJ, Pardo-Seco J, Martinon-Torres F. Non-specific (heterologous) protection of neonatal BCG vaccination against hospitalization due to respiratory infection and sepsis. *Clin Infect Dis*. 2015;60:1611–9.
- Do VA, Biering-Sørensen S, Fisker AB, Balé C, Rasmussen SM, Christensen LD, Jensen KJ, Martins C, Aaby P, Benn CS. Effect of an early dose of measles vaccine on morbidity between 18 weeks and 9 months of age: a randomized, controlled trial in Guinea-Bissau. *J Infect Dis*. 2017. pii:jiw512. <https://doi.org/10.1093/infdis/jiw512>.
- Faustman DL, Wang L, Okubo Y, et al. Proof-of-concept, randomized, controlled clinical trial of Bacillus-Calmette-Guerin for treatment of long-term type 1 diabetes. *PLoS One*. 2012;7:e41756.
- Fine PE. Herd immunity: history, theory, practice. *Epidemiol Rev*. 1993;15:265–302.
- Goodridge HS, Ahmed SS, Curtis N, Kollmann TR, Levy O, Netea MG, Pollard AJ, van Crevel R, Wilson CB. Harnessing the beneficial heterologous effects of vaccination. *Nat Rev Immunol*. 2016;16(6):392–400. <https://doi.org/10.1038/nri.2016.43>.
- Jensen KJ, Karkov HS, Lund N, et al. The immunological effects of oral polio vaccine provided with BCG vaccine at birth: a randomised trial. *Vaccine*. 2014;32:5949–56.
- Jensen KJ, Benn CS, van Crevel R. Unravelling the nature of non-specific effects of vaccines—A challenge for innate

- immunologists. *Semin Immunol.* 2016;28(4):377–83. <https://doi.org/10.1016/j.smim.2016.05.005>.
- John TJ, Samuel R. Herd immunity and herd effect: new insights and definitions. *Eur J Epidemiol.* 2000;16:601–6.
- Kandasamy R, Voysey M, McQuaid F, de Nie K, Ryan R, Orr O, Uhlig U, Sande C, O'Connor D, Pollard AJ. Non-specific immunological effects of selected routine childhood immunisations: systematic review. *BMJ.* 2016;355:i5225. <https://doi.org/10.1136/bmj.i5225>.
- Kleinnijenhuis J, Quintin J, Preijers F, et al. Bacille Calmette-Guerin induces NOD2-dependent non-specific protection from reinfection via epigenetic reprogramming of monocytes. *Proc Natl Acad Sci U S A.* 2012;109:17537–42.
- Lund N, Andersen A, Hansen AS, et al. The effect of oral polio vaccine at birth on infant mortality: a randomized trial. *Clin Infect Dis.* 2015;61:1504–11.
- Mina MJ, Metcalf CJ, de Swart RL, Osterhaus AD, Grenfell BT. Long-term measles-induced immunomodulation increases overall childhood infectious disease mortality. *Science.* 2015;348:694–9.
- Netea MG. Training innate immunity: the changing concept of immunological memory in innate host defence. *Eur J Clin Investig.* 2013;43:881–4.
- Pollard AJ, Finn A, Curtis N. Non-specific effects of vaccines: plausible and potentially important, but implications uncertain. *Arch Dis Child.* 2017 pii: archdischild-2015-310282. <https://doi.org/10.1136/archdischild-2015-310282>. [Epub ahead of print] Review
- Rivero-Calle I, Gomez-Rial J, Martinon-Torres F. Systemic features of rotavirus infection. *J Infect.* 2016;72(Suppl):S98–S105.
- Shann F. The heterologous (non-specific) effects of vaccines: implications for policy in high-mortality countries. *Trans R Soc Trop Med Hyg.* 2015;109:5–8.
- Trotter CL, Maiden MC. Meningococcal vaccines and herd immunity: lessons learned from serogroup C conjugate vaccination programs. *Expert Rev Vaccines.* 2009;8:851–61.

How Vaccinating People Can Also Protect Others

Adam Finn

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2.1 Smallpox Was Not Eradicated by Immunising Everyone

Edward Jenner demonstrated direct protection against smallpox in a human challenge study in a single subject conducted a century before the pioneering work of Pasteur and Koch laid the foundations of our current understanding of the microbial causes of infection. His paper “On the origin of the vaccine inoculation” published in 1801 concludes with the words: “..and it now becomes too manifest to admit of controversy, that the annihilation of the Small Pox, the most dreadful scourge of the human species, must be the final result of this practice”. So Jenner accurately predicted the eradication of smallpox some 175 years later using the technique he had discovered. There are no words with which adequately to do justice to his remarkable foresight. However, Jenner must have taken his observation in James Phipps, the boy he vaccinated with material from a cowpox lesion and then repeatedly challenged with material from smallpox lesions and multiplied it in his head by the number of people living on the planet. Even he could not have known then, what we know now, namely, that his vaccine and nearly all the others developed and widely used since, can do much more than protect recipients against target infections. Setting aside the possibility of non-specific effects, which are beyond the scope of this chapter (see ► Chap. 1), vaccines can break the train of transmission of their target infections between humans, and so vaccinating just some people can be enough to protect everyone. In the cases of smallpox and more recently polio virus type 2, mass vaccination has led to eradication and thus protection for everyone who will ever live. No other advance in medicine comes anywhere close to this extraordinary power of vaccines.

The strategy adopted in the final phase of the eradication of smallpox in the 1970s reflects the growing understanding at the time that vaccinating people can also protect others. The very visible clinical features of smallpox made it relatively easy to recognise each individual case and then immunise around it, generating a ring of human immunity that the virus could not escape from. In fact many countries stopped universal smallpox vaccination long before global eradication had been achieved. Thus vaccine supplies could be used exclusively in areas where the infection was

still circulating. Ring vaccination reappeared recently in the context of the Ebola epidemic in West Africa.

2.2 Why Direct Protection and Indirect Protection Are Not the Same Thing

It is easy to fall into the trap of thinking that direct and indirect protection afforded by vaccines are both one and the same thing. Immunise James Phipps and he will not get smallpox. That he will also therefore not infect his brother seems simply an inevitable consequence of the protection he got from the vaccine himself. To demonstrate that it is not as simple as that, it is worth considering the example of developmental “transmission-blocking” malaria vaccines. Given to humans, these consist of antigens expressed by the malaria parasite only during the stages of its life cycle when it is resident in the mosquito. When the insect takes a blood meal from an immunised human, it ingests not only malaria parasites but also vaccine-induced antibodies, which bind to them as they develop inside the insect reducing their viability and thus affording protection to the human providing the mosquito’s next blood meal. The altruism inherent in such vaccines creates problems for their licensure as regulation of medicines is driven by considerations of safety and of benefit to the recipient, not others. For these vaccines in particular, but actually for most other vaccines as well, we need a new developmental paradigm that recognises that they really work for the common good and need to be deployed towards that end to achieve maximum impact and cost benefit.

Of course the concept of indirect protection – previously often referred to as “herd immunity” – is not novel. Implicit in long-standing advice to attain and then maintain 95% (and not 100%) coverage with measles containing vaccine was the recognition that while there would always be some who would not receive or make protective responses to the vaccine, disease control for all could nevertheless be achieved, even for an infection as contagious as measles. However the ubiquitous nature of such effects among the vaccines used in universal programmes (tetanus – acquired from soil bacteria, not other people, being the one unequivocal exception) and the dominance of

such effects in ensuring the effectiveness of many programmes have only become evident more recently. Indirect effects are no longer considered a “bonus extra” but are understood to be at the core of how vaccines impact on disease and, to an increasing extent, they drive the design of the programmes used – the numbers of doses of vaccines given and the ages of the recipients.

2.3 Immunising Teenagers and Protecting Everyone Against Meningococci

The recent history of the deployment of conjugate meningococcal vaccines in the UK is a particularly informative example of this. In the 1990s, in the wake of the successful introduction of conjugate vaccines against *Haemophilus influenzae* type b, several protein-polysaccharide conjugate vaccines against *Neisseria meningitidis* capsular group C were developed. A rapid rise in the number of severe and fatal cases of meningitis and septicaemia had been occurring in the UK during that decade due to spread of a hyperinvasive strain (clonal complex (cc) 11) bearing this capsule, both in young children and teenagers. The target of the rolling programme introduced in late 1999 was infants who received three doses of vaccine, while a one-off “catch up” programme offering vaccine to all children up to the age of 20 years was also rapidly implemented with the aim of preventing cases in older age cohorts. The licensure of the vaccines was based upon their ability to induce serum bactericidal antibody. From this, it was inferred that they would protect recipients against invasive disease. Between 2009 and 2015, a very similar epidemic of hyperinvasive cc11 meningococcal disease was detected, this time expressing group W capsule. Once again the UK authorities acted rapidly and decisively to attempt to control the epidemic using conjugate vaccines. However the strategy used was entirely different. This time infants and young children were not immunised at all – despite the availability of licenced vaccines and the fact that severe cases were being seen in this age group. Instead vaccination has been targeted exclusively at teenagers – the age group among whom upper respiratory tract carriage of meningococcus is most prevalent. In the 15 years between the two interventions, it had become clear that conjugate

meningococcal vaccines actually work at the population level by eliminating the circulation of hypervirulent strains among the target capsular group(s) of the vaccine(s) used. Infants and young children may have the highest risk of disease, but carriage is comparatively rare in this age group. By immunising adolescents, (in whom the vaccines also induce larger and more long-lasting responses than in young children), all age groups are indirectly protected (see ► Chap. 22).

2.4 Maternal Immunisation

Problems with control of pertussis by childhood immunisation have been an emerging concern since early in the twenty-first century. Development, licensure and adoption of acellular pertussis vaccines for infants and young children in many wealthier countries, alongside continued use of whole cell vaccines in others, both combined with diphtheria, tetanus and other antigens for infants, led initially to effective pertussis control (see ► Chap. 18). However the first decade of the twenty-first century saw new resurgences of disease in several acellular vaccine-using countries. Several lines of evidence suggest that pertussis vaccines in general and acellular vaccines in particular induce protection that is shorter lived and incomplete against onward transmission. Pertussis presenting as chronic cough in adolescents and young adults has become more widely recognised, and transmission from these individuals to their newborn unimmunised infants can result in severe cases and deaths. The UK authorities responded to just such a resurgence in 2012 by offering vaccine to pregnant mothers. Subsequent case-screening and case-control evaluations of effectiveness have provided convincing evidence that this approach works and many other countries have now followed suit (see ► Chap. 6). Protecting infants by immunising their mothers is, again, nothing new, having been used to prevent neonatal tetanus for many years in poorer settings where this is a significant public health problem. It has also been an observed benefit of maternal influenza immunisation programmes implemented to protect pregnant women at high risk from flu. Its success and widespread acceptance as a means to prevent pertussis is likely to accelerate development of similar programmes using developmental maternal vac-

cines against other severe neonatal infections including group B *Streptococcus* and respiratory syncytial virus (see Part IV).

2.5 Indirect Effects of Influenza Vaccines in Healthcare Workers and Children

Over the many years they have been available, the vast majority of seasonal influenza vaccine use in most countries has aimed at direct protection of recipients. Every autumn, large numbers of doses are earmarked for elderly people and patients with a range of chronic disorders, all deemed to be at high risk of severe or fatal flu infection. Even if high coverage rates are achieved, which is unusual, this approach cannot be expected to impact significantly upon flu circulation in the population at large as transmission occurs in all age groups and particularly in childhood. However, one aspect of traditional flu vaccine use does aim higher than simple prevention of morbidity in recipients and that is immunisation of healthcare workers (HCWs). Hospitals are subject to major seasonal fluctuations in workload due to wintertime epidemics of respiratory and gastrointestinal viruses. Staff are continuously exposed and often infected. Two serious adverse consequences are that they then infect other vulnerable patients and that they may be absent from work during their illnesses reducing the capacity to deliver healthcare at times of peak demand. Immunisation of HCWs against flu has the potential to reduce these problems and is actively promoted in many settings. Given that there is good quality evidence that – at least when there is a good match between the vaccine and circulating strains – flu vaccines prevent flu in healthy adults and can also prevent onward transmission in some settings, this policy, designed to protect the function of the health service and to reduce flu morbidity and mortality among its patients as well as its employees, makes good sense. However, evidence that this approach actually delivers on these endpoints in the healthcare setting is surprisingly weak. This undermines the argument that such immunisation should be mandatory as, for example, it commonly is for hepatitis B vaccine. In addition, studies that suggest that repeated annual doses of

inactivated flu vaccine may result in progressive falls in immunogenicity and effectiveness suggest that other strategies may be needed to tackle this problem.

There is emerging evidence that annual universal childhood immunisation against influenza using the live attenuated intranasal vaccine (LAIV) could be an effective approach to population-wide influenza control. The UK started offering universal one-dose LAIV for 2- and 3-year-old children in 2014 and has progressively raised the upper age limit in successive years. Ecological data support the idea that preventing flu in young children reduces the incidence of influenza-like illness in other age groups, and, most recently, 2015–2016 data from Scotland and Northern Ireland, where the programme was implemented more effectively with higher coverage and across a wider age range, show that the incidence never crossed the epidemic threshold, unlike England and Wales where fewer children were immunised. This apparent success is bolstered by early supportive data from Canada and Finland with the nasal vaccine in children. However, recent data from the USA has failed to demonstrate effectiveness, particularly against the H1N1 strain, which has led to removal of the recommendation to use LAIV vaccine there in 2016 (see ► Chap. 14).

However, the indirect effects of childhood flu vaccination may extend further than prevention of flu in the wider population. Serious bacterial infections, including those caused by pneumococcus and meningococcus, have been associated epidemiologically with influenza, and potential pathogenic mechanisms are well described. These observations hint that preventing bacterial infections may turn out to be as effectively done using vaccines against viruses, including flu, as by using vaccines targeted at the bacteria.

2.6 The Future of Indirect Effects of Vaccines

The need to study, understand and ultimately accurately predict indirect effects of vaccines has become obvious, but the best ways to achieve this are still far from clear.

With regard to colonisation and infectiousness, molecular microbiological techniques which

accurately quantify mucosal microbes are already replacing culture techniques which tended to see colonisation as a binary endpoint – either present or absent. That bacteria may be present in the noses of toddlers across a range of up to six orders of magnitude gives the lie to the traditional approach. Development of transcriptomic signatures able to predict whether microbes are in “transmission” or “stable colonisation” mode may also be a way forward.

With regard to immunological mechanisms and markers that reliably predict that an individual is unlikely to become infected or to be infectious to others, these have, to date, been remarkably hard to define. Mucosal immune responses to both mucosally administered and injected vaccines are commonly observed, but quantifying them accurately in complex mucosal secretions and showing what they predict have proved extremely difficult. The discovery of the importance of CD4-positive T helper cells expressing IL17 for mucosal immunity to bacteria provides a new potential approach to understanding naturally acquired and vaccine-induced protection against mucosal acquisition or promotion of mucosal clearance. But again, to date, this has turned out to be hard to define accurately in humans.

Finally, there is the emerging field of vaccine non-specific effects, described in (see ► Chap. 1). Genuinely non-specific protective effects if they exist, perhaps due to innate immune activation and/or polyclonal T and B cell stimulation in early life, are really something quite different to population-wide effects on the target infection of a widely used vaccine. But infections can affect one another in numerous ways. Association between influenza and serious bacterial infections have been mentioned above. The wider impact of preventing measles and thus averting its long-known immunoparetic effects and consequent mortality has recently been highlighted. Vaccines that change human mucosal ecology, whether in the gastrointestinal, urogenital or respiratory tracts, will have knock-on effects on other microbes and therefore other infectious diseases.

This is an area full of important questions and, as yet, rather few clear answers. It is likely that this will change as recognition of the importance of the latter and emergence of new ways to find them become available.

Further Reading

- Campbell H, Saliba V, Borrow R, Ramsay M, Ladhani SN. Targeted vaccination of teenagers following continued rapid endemic expansion of a single meningococcal group W clone (sequence type 11 clonal complex), United Kingdom 2015. *Euro Surveill* : bulletin European sur les maladies transmissibles = European communicable disease bulletin. 2015;20(28).
- Cartwright KA, Jones DM, Smith AJ, Stuart JM, Kaczmarek EB, Palmer SR. Influenza A and meningococcal disease. *Lancet*. 1991;338(8766):554–7.
- Dabrera G, Amirthalingam G, Andrews N, Campbell H, Ribeiro S, Kara E, et al. A case-control study to estimate the effectiveness of maternal pertussis vaccination in protecting newborn infants in England and Wales, 2012–2013. *Clin Infect Dis*. 2015;60(3):333–7.
- Didierlaurent A, Goulding J, Patel S, Snelgrove R, Low L, Bebiën M, et al. Sustained desensitization to bacterial Toll-like receptor ligands after resolution of respiratory influenza infection. *J Exp Med*. 2008;205(2):323–9.
- Febre M, McLay K, Caccamo M, Twomey KB, Ryan RP. Advances in bacterial transcriptome and transposon insertion-site profiling using second-generation sequencing. *Trends Biotechnol*. 2011;29(11):586–94.
- Henao-Restrepo AM, Camacho A, Longini IM, Watson CH, Edmunds WJ, Egger M, et al. Efficacy and effectiveness of an rVSV-vectored vaccine in preventing Ebola virus disease: final results from the Guinea ring vaccination, open-label, cluster-randomised trial (Ebola ca Suffit!). *Lancet*. 2017;389(10068):505–18.
- Henderson DA. Principles and lessons from the smallpox eradication programme. *Bull World Health Organ*. 1987;65(4):535–46.
- Hodgson D, Baguelin M, van Leeuwen E, Panovska-Griffiths J, Ramsay M, Pebody R, et al. Effect of mass paediatric influenza vaccination on existing influenza vaccination programmes in England and Wales: a modelling and cost-effectiveness analysis. *Lancet Public Health*. 2017;2(2):e74–81.
- Jenner E. On the origin of the vaccine inoculation. London: D N Shury; 1801.
- Klugman KP, Chien YW, Madhi SA. Pneumococcal pneumonia and influenza: a deadly combination. *Vaccine*. 2009;27(Suppl 3):C9–C14.
- Ladhani SN, Beebeejaun K, Lucidarme J, Campbell H, Gray S, Kaczmarek E, et al. Increase in endemic *Neisseria meningitidis* capsular group W sequence type 11 complex associated with severe invasive disease in England and Wales. *Clin Infect Dis*. 2015;60(4):578–85.
- Loeb M, Russell ML, Moss L, Fonseca K, Fox J, Earn DJ, et al. Effect of influenza vaccination of children on infection rates in Hutterite communities: a randomized trial. *JAMA*. 2010;303(10):943–50.
- Lu YJ, Gross J, Bogaert D, Finn A, Bagrade L, Zhang Q, et al. Interleukin-17A mediates acquired immunity to pneumococcal colonization. *PLoS Pathog*. 2008;4(9):e1000159.
- McLean HQ, Thompson MG, Sundaram ME, Meece JK, McClure DL, Friedrich TC, et al. Impact of repeated vaccination on vaccine effectiveness against influenza

A(H3N2) and B during 8 seasons. *Clin Infect Dis*. 2014;59(10):1375–85.

Mina MJ, Metcalf CJ, de Swart RL, Osterhaus AD, Grenfell BT. Long-term measles-induced immunomodulation increases overall childhood infectious disease mortality. *Science*. 2015;348(6235):694–9.

Nunes JK, Woods C, Carter T, Raphael T, Morin MJ, Diallo D, et al. Development of a transmission-blocking malaria vaccine: progress, challenges, and the path forward. *Vaccine*. 2014;32(43):5531–9.

Pebody RG, Green HK, Andrews N, Boddington NL, Zhao H, Yonova I, et al. Uptake and impact of vaccinating school age children against influenza during a season with circulation of drifted influenza A and B strains, 4th ed. England, 2014/15. *Euro Surveill*. 2015;20(39): Article 4.

Thomas RE, Jefferson T, Demicheli V, Rivetti D. Influenza vaccination for healthcare workers who work with the elderly. *Cochrane Database Syst Rev*. 2006;(3):CD005187.

Thorrington D, Jit M, Eames K. Targeted vaccination in healthy school children – can primary school vaccination alone control influenza? *Vaccine*. 2015;33(41):5415–24.

Thors V, Morales-Aza B, Pidwill G, Vipond I, Muir P, Finn A. Population density profiles of nasopharyngeal carriage of 5 bacterial species in pre-school children measured using quantitative PCR offer potential insights into the dynamics of transmission. *Hum Vaccin Immunother*. 2016;12(2):375–82.

Childhood and Adolescent Immunization Programs in Europe

Pierre Van Damme

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3.1 Introduction

In 2005, the World Health Assembly adopted resolution WHA58.15 on global immunization strategy. It “urged Member States to meet immunization targets expressed in the United Nations General Assembly special session on children; to adopt the Strategy as the framework for strengthening of national immunization programmes, with the goal of achieving greater coverage and equity in access to immunizations, of improving access to existing and future vaccines, and of extending the benefits of vaccination linked with other health interventions to age groups beyond infancy; to ensure that immunization remains a priority on the national health agenda, ...”

The diversity of the European Region is reflected not only in the cultures and languages, but also by economies and health systems. The economic, cultural, and historical differences have all contributed to the resulting diversity seen in the health systems and health governance among them, differences that have contributed to the wide variation of immunization programs currently in place.

All Member States of the European Union and a large number of the non-EU countries in the WHO European Region have a national immunization technical advisory group (NITAG) on immunization, and most of these NITAGs have a legislative basis for making recommendations to the government (i.e., the Ministry of Health). The effect of the recommendations varies according to how immunization programs are organized (centralized or decentralized) and the balance between public and private sector provision of services. In countries such as Belgium, Germany, and Spain, the communities (Belgium), the Länder (Germany) or the “autonomous regions” (Spain) have the responsibility for prevention and protection of public health. Although each country has a NITAG, its recommendations can be modified at the local level, and the vaccines actually provided depend on the choice of private practitioners and reimbursement arrangements with insurance companies.

Immunization policy or practice has not been subject to European legislation for harmonization, although many relevant processes such as batch release are controlled through EU legislation.

The vaccines and immunization schedules used in the 53 countries of the WHO European Region are undergoing continuous change, with the introduction of new antigens and the increasing use of combined antigen vaccines and simplified schedules with a lower number of vaccine doses. Annual information is collected from WHO Member States on immunization programs and vaccine-preventable diseases using the WHO/UNICEF joint reporting form. This information can easily be consulted through the WHO website at: ► http://apps.who.int/immunization_monitoring/globalsummary/schedules. ECDC offers a *Vaccine Scheduler* tool, it is an interactive platform of *vaccination schedules* for individual European countries and specific age groups (► <http://vaccine-schedule.ecdc.europa.eu/Pages/Scheduler.aspx>).

Country immunization schedules can be consulted by vaccine or target disease, or compared with each other.

3.2 Childhood Vaccination

In Europe, childhood vaccination is offered through routine immunization programs at “well-baby” clinics, or through the private sector (general practitioners or pediatricians), or a combination of both public and private sector.

The current childhood immunization schedules for vaccination below 24 months of age in the EU countries can be divided into four major groups for the infant vaccination schedule:

Group 1 Early-onset 3 plus 1 schedule with vaccination at 2, 3, and 4 months of age (Bulgaria, Czech Republic, Germany, Hungary, Luxemburg, Malta, the Netherlands, and Belgium using timings of 8, 12, and 16 weeks) or the schedule similar to that of the USA of 2, 4, and 6 months of age (Croatia, Cyprus, Greece, Latvia, Liechtenstein, Lithuania, Poland, and Portugal), followed by a fourth dose in the 2nd year of life.

Group 2 Early onset according to a 2 plus 1 schedule, with vaccination at 2 and 4 months, followed by a third dose at 11 months (France, Romania, and Spain).

Group 3 Late onset 2 plus 1 schedule with vaccination at 3 and 5 months of age followed by a

third dose at 12 months of age (Denmark, Finland, Iceland, Norway, Slovakia, Sweden, and Italy).

Group 4 Late onset 3 plus 1, starting at the age of 3 months, (Estonia and Slovenia), with a fourth dose in the 2nd year of life.

Only one or two countries use only a three-dose primary immunization schedule with no penta- or hexavalent booster in the 2nd year of life. In the remaining WHO European Region countries, the Extended Program of Immunization (EPI) schedule is often implemented together with primary infant immunization offered at 6, 10, and 14 weeks – in some countries followed by infant booster immunization.

The various childhood immunization schedules in Europe evolved historically, taking into consideration the local vaccine-preventable infection epidemiology, and were based on the experiences gained from immunization with whole-cell pertussis-containing diphtheria–tetanus–pertussis (DTP) vaccines (2-, 3-, 4- and 2-, 4-, 6-month schedules), where the need for three doses was shown. The 3- and 5-month schedule, on the other hand, evolved from the vaccination priming schedule for the diphtheria–tetanus (DT) vaccine, which was introduced first in Italy in 1981 and in Sweden in 1986. That schedule was maintained in a number of countries when a pertussis vaccine was added to DT.

The four different schedules used in Europe have been shown to accomplish their primary goal, i.e., to induce rapid protection and immunological memory against the vaccine-preventable infections targeted by the immunization, in close to 100% of vaccinated infants. By starting at 2 months of age (or 8 weeks, which offers a smaller range than 2 months) protection will be achieved 1 month earlier than with a 3-, 4-, and 5-month schedule or 3- and 5-month schedule.

A measurable antibody response does not develop in all children after the priming doses and the level of the antibody responses may be low. The booster dose will induce measurable antibody responses in almost 100% of children, and result in much higher antibody levels than after the priming doses.

European vaccination schedules all call for at least one or two booster doses between the ages of 2 and 18 years, but with quite a variation in local schedules. Such a variation creates problems in

migration, as parents and physicians have to face difficult decisions on how to adapt or complete vaccination schedules when families move from one European country to another.

3.3 Adolescent Vaccination

Vaccinating adolescents offers three types of immunization opportunities: catch-up on missed vaccinations, boosting waning immunity (derived from previous childhood vaccinations such as for pertussis), and the achievement of primary immunization through administration of new vaccines best delivered during adolescence (e.g., meningococcal and human papillomavirus vaccines; ■ Table 3.1). In the future, adolescence may also be the target age range for administration of some vaccines currently in development.

Adolescent vaccination can prevent considerable morbidity in adolescent and adult age groups, and limit the spread of infectious diseases in the population. In Europe, adolescent vaccination can be provided through routine immunization programs or campaigns, run with the support and participation of either the private sector or the public sector, or both. Vaccines can be administered through clinic-based schemes (e.g., in health centers), in the community or in schools. Mixed systems of school health and private sector can offer benefits, but require coherence, coordination, and good communication between all parties.

However, because of the age of the target group – the WHO definition of an adolescent being aged between 10 and 19 – legal issues arise: parental consent, minors' consent (assent) and legality thereof, the concept of “capacity to understand” and “competence,” and action in case of parental opposition. Another feature that emerges is the disconnect between the practice of immunization and other medical procedures (“treatment”), including the role of school health services in dealing with other health problems, such as drug use, alcohol use, and violence.

Furthermore, medical issues in this age range also complicate the matter of immunization; a substantial proportion (about 10%) of young people suffer from chronic illnesses (e.g., diabetes, whose incidence in young people is increasing) that need to be considered before vaccination is given. Other temporal, coincidental associations in adolescents, e.g., asthma,

Table 3.1 Examples, advantages, and disadvantages of adolescent vaccination strategies (Brabin et al. 2008)

Vaccine implementation			
Strategy	Example vaccine	Advantages for adolescent programs	Disadvantages for adolescent programs
Universal	Meningococcal conjugate (MCV4)	Increased likelihood of achieving herd immunity	The ability to achieve herd immunity is undermined if low vaccination rates occur
		Decreased likelihood of inducing stigma around certain diseases such as sexually transmitted infections	Higher costs to society
Targeted	Hepatitis B virus (HBV)	Reduced costs if every adolescent does not require vaccination	Target groups can be difficult to identify
		Reduced risk of adverse events in the whole population	Adolescents may not perceive themselves to be high risk
			Adolescents may be unwilling to seek care if fear of judgment or lack of confidentiality exists, especially for sexually transmitted infections
Increased risk of stigmatization, particularly for sexually transmitted infections			
School-based	Rubella (MMR, MR, or R)	In countries with school-based programs, success has been mediated by the requirement to attend school and by a lack of private sector health care	School attendance by adolescents is low in many countries
			School-based health care infrastructure is generally directed at younger children; therefore, retention and/or creation of appropriate infrastructures in many countries need to be developed to create an adolescent program
			Future adolescent vaccines targeted at sexually transmitted diseases necessitate integration with health promotion; in particular, sexual health issues associated with absenteeism require development of catch-up programs
Catch-up	Pertussis (Tdap)	Maintain immunity to prevent infection and subsequent infection of un-immunized individuals	Timing of catch-up programs need to coincide with other preventive services to increase the likelihood of vaccination uptake
		Reduced healthcare costs associated with decreased disease burden	
Mass vaccination	Typhoid fever (Ty21a, Vi)	Large number of individuals can be vaccinated within a rapid timeframe	Suitable for single-dose vaccinations; however, less effective for multi-dose vaccines, as the likelihood of individuals returning for subsequent vaccination decreases with each additional dose
		Excellent for outbreak situations	
		Limited amount of resources can be mobilized	

auto-immune thyroiditis, and Guillain–Barré syndrome may raise safety concerns.

In Europe, as for the implementation of the childhood immunization program, the adolescent program differs by country and sometimes by state, region or canton, and involves the public and/or private sectors.

In general, in Europe, adolescent immunizations lag behind childhood uptake figures, in particular for the second dose of measles, mumps, and rubella vaccine, the booster dose of the pertussis vaccine, or the uptake of human papillomavirus vaccines. Waning immunity or absence of immunity in adolescents makes them reservoirs of infection, with transmission possibilities to other age groups in the population. In many countries, adolescents are an underserved group that is hard to reach because of their good health and sparse preventive medicine visits.

Studies among adolescents have identified risk factors associated with suboptimal immunization, which may include financial and logistic constraints, in addition to parental and adolescent knowledge and beliefs: e.g., socioeconomic status, lack of medical insurance, large family size, divorced parents, foreign nationality, and language barriers.

School health services have been identified as playing a specific role in the prevention and response to adolescent health problems. Where there were no strong school health facilities or vaccine programs, such as in France, Germany, and Italy, rates of adolescent vaccination have been low. With school attendance mandatory for high proportions of adolescents in Europe, the presence of a captive audience makes vaccination at school feasible. Benefits of school health programs (besides high coverage rates) include easy access to vaccination for parents (no effort required from them) and easy monitoring of coverage and side effects. On the down side, school immunization programs form only one part of a school medicine system, and cannot manage common adolescent problems including smoking, alcohol and drug use, sexual behavior, and violence, unless it is fully embedded in a comprehensive program. In addition, communication with parents is indirect, which can raise some legal issues.

The introduction of a centralized immunization information system (enabling recording, recall and informing health care workers and parents), the organization of a school health pro-

gram, offering the vaccine free of charge, and the implementation of school-entry mandates have been recognized as factors that could contribute to improved vaccination coverage in adolescents. In addition, advocacy and educational initiatives for parents, adolescents, and vaccinators should help to support these programs and safeguard the health of adolescents.

The concept of promoting health in schools seems to be successfully taking off, but health care providers alone cannot meet adolescents' needs: there has to be a partnership and networking of vaccinators, teachers, parents, and young people all playing a role. Vaccination should be integrated into other interventions in health systems (e.g., sexual health education and sports medical examinations). Various approaches are currently being successfully used by different countries to reach adolescents.

3.4 Vaccination of Refugees and Immigrants

Since 2011, Europe has been facing one of the greatest migration inflows in its history: during 2011, there were an estimated 1.7 million immigrants into the EU from countries outside the EU. According to Eurostat, after the Northern African turmoil, in 2012, EU countries received 300,000 asylum applications, which peaked at 1,300,000 in 2015, after the Syrian conflict; almost double the previous great migration inflow recorded in 1992, after the crisis in the former Yugoslavia. The UNHCR estimated that, in 2015, more than one million migrants arrived in Europe after crossing the Mediterranean Sea. Refugees and immigrants often come from countries in which poverty-related diseases are endemic, with disrupted health care systems, and consequently a fall in vaccination coverage. This explains why they are at a high risk of vaccine-preventable diseases, not to mention the risky conditions they endure during the journey to Europe (unsanitary conditions, overcrowding).

Overall, migrants and refugees have lower immunization rates than European-born individuals, with children being at a higher risk of being unvaccinated against measles, mumps, and rubella (MMR; ■ Table 3.2). The coverage for the oral polio vaccine has been estimated to be less than 15% among Syrian children refugees in Germany.

Table 3.2 Immunization coverage for 2014, according to the estimates of the WHO and UNICEF for six of the most frequent countries of origin of migrants arriving in Europe (2012), compared with five EU countries (Mipatrini et al. 2017)

Vaccine	Code	Syria	Iraq	Afghanistan	Albania	Pakistan	Eritrea	Italy	Greece	Germany	Denmark	Sweden
Bacillus Calmette–Guerin	BCG	81	95	86	99	85	97	–	–	–	–	–
Diphtheria–tetanus–pertussis first dose	DTP1	65	77	82	99	79	97	98	99	98	96	99
Diphtheria–tetanus–pertussis third dose	DTP3	43	64	75	98	72	94	94	99	96	94	98
HBV third dose	HepB3	71	62	75	98	72	94	94	96	87	–	53
HBV birth dose	HepB_BD	78	43	4	99	–	–	–	–	–	–	–
<i>Haemophilus influenzae</i> third dose	Hb3	43	64	75	98	72	94	94	99	94	94	98
Measles-containing vaccine first dose	MCV1	54	57	66	98	61	96	86	97	97	90	98
Measles-containing vaccine second dose	MCV2	49	57	39	98	52	–	–	83	92	84	95
Maternal immunization with ≥2 doses of tetanus toxoid	PAB	92	72	70	92	75	94	–	–	–	–	–
Pneumococcal conjugate vaccine	PCV3	–	–	40	99	72	–	55	96	69	93	97
Polio vaccine 3rd dose	Pol3	52	67	75	98	72	94	94	99	95	94	98
Rotavirus	RotaC	–	29	–	–	–	25	–	–	–	–	–

In 2016, the WHO, UNICEF, and UNHCR officially stated that migrants, asylum seekers, and refugees should have nondiscriminatory and equitable access to vaccinations. They recommended vaccinating these populations, avoiding delays, in accordance with the immunization schedule of the host country, and offering documentation of administered vaccines to avoid duplications.

However, access to complete vaccination is difficult to ensure: migrants are moving throughout Europe, whereas vaccines must often be given in consecutive doses; information on the immunization status of the migrants is often lacking; recommended immunization schedules differ among EU countries complicating the catch-up programs; a number of the host countries face severe economic crises, challenging migrants' access to the local health care services; migrants may refuse registration by medical authorities for the fear of legal consequences; a lack of coordination among EU public health authorities may cause either a lack of vaccine administration or duplication.

Although migrants have the right to health care under legal settlements issued by the EU, there is no standard European approach for offering health care to migrants. Each country has its own policy.

To overcome many of these issues at the EU or country level, the WHO proposes tailoring immunization services to the specific needs of the target population, to strengthen social mobilization, advocacy, and communication toward these specific populations, to develop electronic vaccination registries, and to introduce coordination among public health authorities of EU countries.

In general, the vaccination status of migrants and refugees arriving in Europe should first be assessed through documentation; when this is lacking, they should be regarded as unvaccinated, and should then be vaccinated according to the local recommended schedule. Catch-up immunization programs should prioritize MMR and inactivated poliovirus vaccines, followed by the DTP vaccines, and hepatitis B vaccine (depending on the age after first screening). Vaccination against polio should be considered a high priority for migrants coming from countries in which polio is endemic. In 2016, some countries or regions (e.g., Flanders) started to offer asylum seek-

ers polio (when indicated), MMR and diphtheria, tetanus, and acellular pertussis vaccination (for pregnant women) immediately on entry into the country, with further follow-up of the immunization in the respective centers for asylum seekers.

Clearly, under-immunization and therefore susceptibility to vaccine-preventable infections pose a risk to the health of migrants and refugees, and, in turn, can result in epidemics in the host country.

Further Reading

- Bearinger LH, Sieving RE, Ferguson J, Sharma V. Global perspectives on the sexual and reproductive health of adolescents: patterns, prevention, and potential. *Lancet*. 2007;369(9568):1220–31.
- Brabin L, Greenberg DP, Hessel L, Hyer R, Ivanoff B, Van Damme P. Current issues in adolescent immunization. *Vaccine*. 2008;26(33):4120–34.
- ECDC. Scientific panel on childhood immunisation schedule: diphtheria-tetanus-pertussis (DTP) vaccination – report. European centre for disease prevention and control, Stockholm. 2009, 40p.
- FitzSimons D, Vorsters A, Hoppenbrouwers K, Van Damme P, Viral Hepatitis Prevention Board (VHPB), European Union for School and University Health and Medicine (EUSUHM). Prevention and control of viral hepatitis through adolescent health programmes in Europe. *Vaccine*. 2007;25(52):8651–9.
- Keane MT, Walter MV, Patel BI, et al. Confidence in vaccination: a parent model. *Vaccine*. 2005;23(19):2486–93.
- Kleinert S. Adolescent health: an opportunity not to be missed. *Lancet*. 2007;369(9567):1057–8.
- Kpomezou E, Heywood AE, Kay M, Smith M, Paudel P, Sheikh M, MacIntyre CR. Improving access to immunisation for migrants and refugees: recommendations from a stakeholder workshop. *Aust N Z J Public Health*. 2016;41(2):118–20.
- Mipatrini D, Stefanelli P, Severoni S, Rezza G. Vaccinations in migrants and refugees: a challenge for European health systems. A systematic review of current scientific evidence. *Pathog Glob Health*. 2017;111(2):59–68.
- Reyes-Uruena JM, Noori T, Pharris A, Jansà JM. New times for migrants' health in Europe. *Rev Esp Sanid Penit*. 2014;16(2):48–58.
- Rosenthal SL, Kottenhahn RK, Biro FM, Succop PA. Hepatitis B vaccine acceptance among adolescents and their parents. *J Adolesc Health*. 1995;17(4):248–54.
- Sakou I-I, Tsitsika A, Papaevangelou V, Tzavela E, et al. Vaccination coverage among adolescents and risk factors associated with incomplete immunization. *Eur J Pediatr*. 2011;170:1419–26.
- Vandermeulen C, Roelens M, Theeten H, et al. Vaccination coverage and sociodemographic determinants of

measles-mumps-rubella vaccination in three different age groups. *Eur J Pediatr.* 2008;167:1161–8.

Wallace LA, Young D, Brown A, et al. Costs of running a universal adolescent hepatitis B vaccination programme. *Vaccine.* 2005;23:5624–31.

Williams GA, Bacci S, Shadwick R, Tillmann T, Rechel B, Noori T, Suk JE, Odone A, Ingleby JD, Mladovsky P, Mckee M. Measles among migrants in the European

Union and the European economic area. *Scand J Public Health.* 2016;44(1):6–13.

Wilson TR, Fishbein DB, Ellis PA, Edlavitch SA. The impact of a school entry law on adolescent immunization rates. *J Adolesc Health.* 2005;37(6):511–6.

Zimet GD, Liddon N, Rosenthal SL, Lazcano-Ponce E, Allen B. Psychosocial aspects of vaccine acceptability. *Vaccine.* 2006;24(Suppl 3):S201–9.

Vaccine Hesitancy, Acceptance, and Demand

Robb Butler

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4.1 Background

The World Health Organization defines vaccine hesitancy as “...a delay in acceptance or refusal of vaccines despite the availability of vaccination services. Vaccine hesitancy is complex and context specific varying across time, place and type of vaccine.” The hesitancy continuum extends from those that accept all vaccines, but are unsure about their decisions for some or all vaccines, through to those who refuse all vaccines, but are unsure about these decisions (■ Fig. 4.1). In that sense, hesitancy affects demand and is most closely associated with negative demand. Addressing vaccine hesitancy requires an understanding of the magnitude and setting of the problem, diagnosis of the root causes, tailoring strategies based on local evidence to address the causes, evaluation to gauge if the intervention has been successful in improving vaccine acceptance, and monitoring.

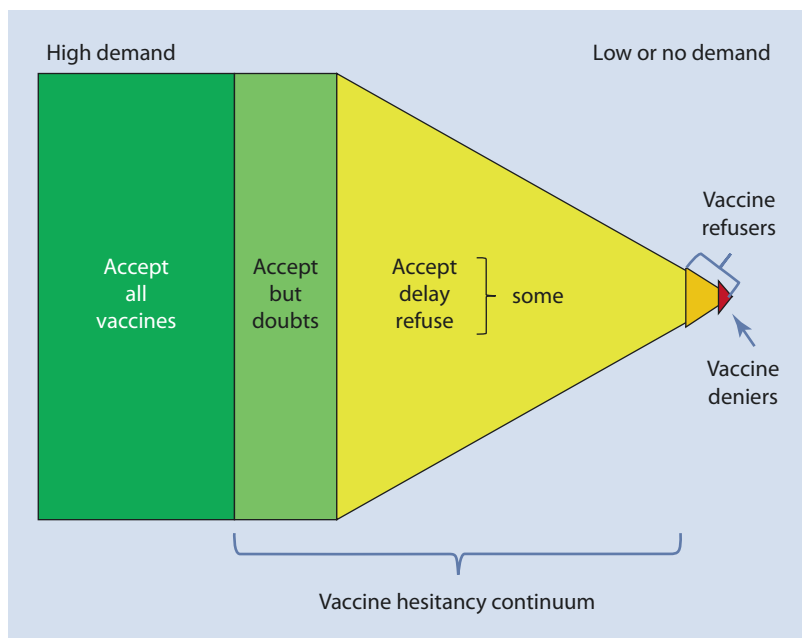
In March 2012, the Strategic Advisory Group of Experts (SAGE) on immunization established a working group to define vaccine hesitancy and its scope, provide advice on how to address vaccine hesitancy, including a landscape analysis of stakeholders working on the issue and identifying promising practices. It presented its work to SAGE at the WHO premises in Geneva, October

2014 (► http://www.who.int/immunization/sage/meetings/2014/october/1_Report_WORKING_GROUP_vaccine_hesitancy_final.pdf) and shortly thereafter published a supplement in *Vaccine* in August 2015. Later that same year, an informal working group was established to develop an understanding of “demand” (definition, components, actors, and determinants) and to explore the means of measuring progress on improving demand. The informal working group has been instrumental in building consensus and understanding around the term demand and its determinants, sharing promising practices from around the globe and considering the best approaches and methods to measuring demand and the impact of demand-generating initiatives. For the purposes of this chapter, we align with the hesitancy and demand working groups’ definitions and understanding of demand – considering hesitancy and acceptance as factors of demand. We focus primarily on hesitancy, its scope and expression in the European Region and strategies to address it from a program planning and an individual (provider–parent/patient) perspective.

■ Figure 4.1 demonstrates the spectrum of demand and the effect of vaccine hesitancy.

In Europe, program organizers have become acutely aware of the potential damage and threat that vaccine hesitancy, public mistrust of vaccines

■ Fig. 4.1 History of vaccine acceptance in Europe. Noni Mac Donald, ► www.sabin.org/sites/sabin.org/files/1-vaccine_hesitancy_final_draft_7_jan26_2017.pdf



and immunization services, and the rejection of vaccines pose. It is unclear whether vaccine hesitancy and associated risks have increased within the European public over recent years (as some observers suggest) or whether, instead, vaccination programs have become more sensitive and aware of the phenomena as they attempt to reach remaining under-immunized populations and meet ambitious coverage targets and disease control goals.

Vaccine hesitancy is not a new phenomenon. Following the introduction of small pox immunization, as early as the mid-1800s, hesitancy and vaccine objection have been documented in Europe. In the UK, the smallpox vaccination induced fear and protest; some believing that the practice of inoculation was un-Christian, others skeptical of Edward Jenner's ideas or objecting on the grounds that the practice violated their personal liberty (mandatory vaccination for infants up to 3 months of age was introduced in 1853). At that time, anti-vaccination lobbies or "leagues" were established with their own journals and communication materials.

A resurgence and lingering of vaccine-preventable diseases such as measles, rubella, diphtheria, and pertussis, resulting in hospitalization and deaths of infants, children, and adults over the past decade, has prompted renewed interest in understanding why Europe, a region rich in resources and capacity, has been unable to close the immunity gaps and meet regional disease control and elimination goals. Immunization service managers and administrators are, in turn, eager to better understand parent/patient hesitancy and health-seeking behaviors to appropriately motivate them to vaccinate and remove factors limiting their ability or opportunity to utilize immunization services. Member States of the European Region restated their commitment to immunization by adopting the European Vaccine Action Plan (EVAP) 2015–2020 in 2014, the first regional plan to openly acknowledge the extent of vaccine hesitancy, vaccine skepticism, and sub-optimal parent/patient demand for immunization services and need for vaccine trust. The EVAP second strategic objective calls for "individuals [to] understand the value of immunization services and vaccines and demand vaccination" and the third calls for "the benefits of vaccination (to be) equitably extended to all people through tailored, innovative strategies."

4.2 Shortcomings of Terminology

As a term, "hesitancy" has often been used synonymously and interchangeably with "lack of confidence" or "confidence-gap" by some academics and practitioners alike. However, in Europe its expression is multi-faceted, including but not limited to trust in vaccines and/or the authorities that provide them. Attributing recent disease incidence and outbreaks in Europe to parental or provider confidence is arguable and may deflect attention from systemic and service delivery shortcomings by placing responsibility solely on the "hesitant" parent/patient. In this sense, the term should be used with caution. In Europe, other system side factors have contributed to disease burden. Even when demand is evident, there are factors that prevent action, despite an intention to vaccinate by a parent/patient. Demand for immunization services does not equate to immunization service utilization. Vaccine supply disruptions, economic/financial/societal crises, program delivery disruption or weaknesses (e.g., delayed introduction of a second dose of measles, or a period of health worker shortages), poor-quality service delivery, including poor communication, for example, have all resulted in sub-optimal coverage and under-utilization of vaccination services in Europe. Some of these factors continue to affect program reach, coverage, and utilization, particularly in countries challenged by high vaccine prices, lack of long-term secured domestic funding for their programs, and unstable vaccine supply. Some countries, particularly those with weak infrastructures, have had to face the additional burden of addressing the migrant influx into Europe, many of whom also require immunization in addition to having other support needs.

4.3 Characteristics of Measles Outbreaks in Europe

Owing to the nature of the disease and the availability of safe and effective vaccines, measles vaccination coverage, disease incidence and outbreaks, are often considered good indicators of the immunization program performance of a country. Most of the 2012–2016 measles patients in Europe had not been fully vaccinated. This finding helped to sharpen the focus of attention given to identifying the population characteristics and the reasons

why these children and adults are un- and under-immunized. The affected were quite disparate. Since 2010, outbreaks have been reported in marginalized, hard-to-reach, and under-served populations in the region, where there is a lower probability of being vaccinated; in anthroposophic communities (Germany, Netherlands, Switzerland, Sweden), in Jewish ultra-orthodox communities (Belgium, Israel, UK), in the Roma and Sinti populations (Bulgaria, central and south-eastern European countries), in migrant communities such as Somali communities in Sweden, and in orthodox Protestant communities (“Bible Belt” residents in the Netherlands). However, large outbreaks, such as those reported in France (2011), United Kingdom (2014) Germany (2015) did not occur within a single community or group, but rather across the general population, indicating issues related to more widespread acceptance in the general population and/or supply barriers to vaccination. Several outbreaks also had their foci in health care facilities including physician waiting areas, infectious disease wards and pediatric wards (Czech Republic, Denmark, France, Greece, Latvia, Italy, Netherlands, and Spain). From aggregated annual measles data analysis, it is also apparent that over 50% of the measles cases reported in 2014–2016 were in persons over the age of 10 years (in some countries that figure is significantly higher, 78% in the UK), indicating that an increasing proportion of cases is being reported among individuals beyond childhood and emphasizing the need to reach out with vaccination services to this group. The immunity gaps leading to the measles outbreaks in Europe are attributed by the existence of these un- and under-immunized subgroups of population. It is generally agreed that the previously emphasized association between vaccination uptake and background characteristics, such as socio-economic and educational status of parents, is diminishing in Europe.

4.4 Vaccination Complacency, Convenience, and Confidence in Europe

Vaccine hesitancy includes factors such as complacency, convenience, and confidence, each of which is exhibited at parent/patient, provider, and decision-making levels in Europe today.

In terms of convenience, parent/patients are not presented with opportunities to access immunization services outside traditional working hours and in locations other than health facilities. Very few countries have considered pharmacies as an option for immunization service delivery (Ireland and Portugal are the exceptions to this), despite strong evidence from the USA and Canada that influenza vaccine rates have been boosted by the use of pharmacies, mini-marts, and other nontraditional outlets, for many years now.

Immunizations can be unnecessarily stressful and anxious events for many children and adults who fear needles and the pain of immunization. This can lead to long-term non-adherence with recommended schedules, missed immunizations, and even a shunning of health care services in general. Very few programs have considered the negative impact of pain of immunization. Few have made efforts to improve provider and parent/patient knowledge and skills to mitigate stress and anxiety during immunization. There are evidence-based strategies, including non-invasive methods such as liquid-jet injection or even distraction techniques with better positioning that can address this problem. New technologies such as micro-needles also promise to not only minimize pain, but potentially enable the delivery of services through nontraditional outlets using nonmedical personnel.

Many parents/patients in Europe have grown complacent about diseases that most communities have not seen in decades. Complacent individuals thus consider the risks of the vaccine to outweigh the risk of contracting the disease. In that sense, vaccines have become a victim of their own success. This even extends to health care providers where many have not seen, first hand, diseases such as measles, rubella, diphtheria, and pertussis in their practice. Complacency is also evident in political decision-making, with many countries unable to secure domestic resources for their programs against competing health, economic, and security priorities. This is particularly apparent in countries that have not experienced outbreaks recently. The decision-making environment in these countries faces an additional dilemma as the direct and indirect costs of outbreaks have not been calculated and appropriately understood thereby hampering adequate planning.

The overall confidence and trust in vaccine effectiveness and safety, and in the authorities that deliver them, is positive, but does vary across Europe. The proliferation of conflicting information, from multiple sources within and outside of the region, has challenged decision-making regarding parent/patient vaccine acceptance and eroded the value of and trust in provider-delivered advice and recommendations. The ability of a single anti-vaccine individual to influence the health seeking behavior of others, including the intention to vaccinate, is greater now than ever before. Indeed, such individuals who understand how new media platforms are leveraged effectively is often more influential and may even be perceived as being more trustworthy than a trained medical or public health professional. This phenomenon has damaged vaccine acceptance and trust in many European countries. In some extreme cases, a single vaccine opponent has been responsible for the suspension of a vaccine program (Ukraine [2008], Bosnia and Herzegovina [2010]) or severely undermined vaccine acceptance and uptake (human papilloma virus, Denmark, 2014). At the extreme end of the demand/hesitancy spectrum are vaccine deniers who oppose vaccines for diverse reasons, but are not open to a change of mind. In Europe, these very small groups are not organized into a cohesive, financed, coordinated body and therefore cannot be considered a “movement” or “lobby,” as is more commonplace in the USA or in Australia, for example. Recent work to mitigate the negative influence of “vocal” vaccine deniers has been undertaken by the WHO in Europe with a guidance document and training program based on psychological research into persuasion, on research into public health, on communication studies, and on WHO risk communication guidelines.

Many immunization programs in the region have relied over the years on communication campaigns solely focused on addressing misconceptions and misinformation. These fail to decrease hesitancy and in some cases, backfire entirely. To some degree, this can be attributed to a lack of understanding by the program organizers that informed individuals are not necessarily behaviorally responsive ones, that knowledge does not predict action, and as such, closing the information gaps through awareness campaigns

does not address hesitancy, ensure demand or guarantee utilization. Social copying and behavioral imitation is also manifest among parent/patients, which is largely seen to be beneficial in increasing and maintaining vaccination coverage, but is also evidently having a negative impact by amplifying nonvaccination behavior and antivaccination sentiment.

4.5 Strategies to Address Hesitancy

4.5.1 Understanding the Target Population: Diagnosing Hesitancy

As demand, hesitancy, and acceptance are context-specific, and program and community resilience variable across Europe, it should be considered a pre-requisite for a program to locally gauge and diagnose the factors influencing vaccination intentions, decisions, and behaviors, with participation of affected (under-immunized) communities. General public and subgroup attitudes, knowledge, and behaviors must be regularly monitored and assessed frequently, to be able to inform and tailor program delivery and response to match the needs of the target subgroups. Success in countering anti-vaccination sentiment and safety concerns depend on this in particular. By tracking patient/parent sentiment and behavior with the use of operational research (such as surveys or rapid assessments) the immunization program ensures that people and communities, not only diseases, are at the centre of immunization systems, and empowers people to take a more active role in their own health. Using WHO tools, behavioral insight studies have uncovered the reasons for lower vaccination uptake in Roma, migrant, Jewish ultra-orthodox, and anthroposophic communities and found that both vaccine hesitancy (individual) and inappropriate or insufficient service delivery (program) affect uptake in each of these communities. The application of such “insight” and social science techniques and methods in some European contexts clearly demonstrates how programs can adopt approaches to tailoring the extension of service delivery according to the needs of communities.

Alongside the importance of diagnosing vaccine hesitancy and demand determinants in any population group, in addition to a consideration of the factors and determinants previously noted in this chapter, we should consider evidence-informed strategies for addressing vaccine hesitancy and improving vaccine uptake from the program perspective and from the individual provider–parent/patient perspective. Some of the strategies covered in this section are adapted from *MacDonald, Dube and Butler* (2016) and are considered appropriate options in the European Region.

4.5.2 Communications Planning

The primary demand indicator of EVAP measures the presence of a communications plan as a proxy for resilience and a signal of communications and advocacy capacity. Crisis (outbreak and vaccine safety related “events”) and risk communication plans should be developed and tested by programs. The communication plans should adhere to best practice and the key principles of risk communications, and be proactive in nature. Clear roles and responsibilities of vaccination programs and emergency communication tasks should be accounted for, including the costing and resourcing of immunization communication activities. Audiences should be clearly identified and multiple channels of communication and messages envisioned. Communication plans must be bidirectional with the immunization programs being sensitive to the values and incorporates the concerns of the target audience. The drafted messages should be tailored to fit the target audience and strengthen or reinforce individuals’ understanding of the benefits and risks of vaccination and the diseases it prevents, enabling them to make evidence-based informed choices, encouraging them to seek immunization services and overcome barriers to vaccination. National vaccination programs should also acknowledge that by developing effective communications plans and capacity, the public’s perception of the credibility, trustworthiness, and competence of the program is enhanced.

4.5.3 Optimizing the Provider’s Role

Healthcare providers, pediatricians included, remain the most trusted source of information and health advice; however, there is a significant minority of providers in Europe today that do not actively promote vaccination, are hesitant or outright anti-vaccination. These providers influence their patients and parents. Therefore, national immunization programs need to ensure that the concept of vaccinology and immunology features on medical curricula in medical and nursing colleges and that opportunities for in-service training of healthcare providers are continuously provided and kept up-to-date. Such education and training should include interpersonal communication techniques and skills to tackle hesitancy.

National vaccination programs should consider reinforcing the learning about vaccine hesitancy and demand determinants with facts sheets and job aids that assist healthcare providers in explaining the risks and benefits of vaccination in a clear and concise way to the parents and patients without the use of jargon or medical terminology. Parents and patients behave more rationally when they receive information in such formats from their credible and trusted health care provider. Inconsistent messaging and contradictory information amongst healthcare providers can confuse patients and parents, prompting mistrust and inaction.

Those health care providers that actively advocate and champion vaccination should be identified and supported to share their opinions and engage a broader audience (than the parent/patient and clients they see on a daily basis). These same gatekeepers and influencers also have a role to play in communicating the value and full benefit of vaccines to other providers who themselves are hesitant and those being educated/trained to become health care professionals. Professional societies and associations should be considered here as partners in addition to prominent scientists and renowned healthcare luminaries. There is also substantial evidence that vaccine acceptance can be increased by engaging local religious and community leaders and this should be considered.

4.5.4 Interpersonal Risk Communication

People are hesitant for various reasons and their levels of concern range from very high to very low. Providers should avoid confrontation and adversarial situations. Rarely do such encounters end with a positive outcome. Providers should adopt an easy-to-understand approach and use frameworks for facing hesitancy; those based on the principles of good risk communication practices. 4-step Framework for Communicating Science: Making the CASE for Vaccines presents such an approach from the University at Albany's School of Public Health.

■ 4-step Framework for Communicating Science: Making the CASE for Vaccines

Corroborate: - acknowledge the parents' concern and find some point on which you can agree. Set the tone for a respectful, successful talk.

About me: - describe what you have done to build your knowledge base and expertise.

Science: - describe what the science says.

Explain/advise: - give your advice to the patient, based on the science.

Example: - "I want to spread out the shots so they won't overwhelm my child's immune system."

Corroborate: - children today certainly have more shots than years ago.

About me: - our practice follows the national schedule because it is carefully designed to protect children at the time they are most vulnerable to disease. I recently returned from a meeting, or I served on a committee, that reviewed the schedule...

Science: - although children undergo more shots today, they actually receive fewer antigens than when they had fewer shots, because technology has enabled us to make vaccines that have only the part of the cell that induces immune response. Plus, the immunological challenge from a vaccine is nothing compared with what kids fight off every day. An ear infection is a greater immunological challenge ("Drop in the ocean").

Explain: - we want all the kids in our practice to be immunized so that they have the greatest chance of a long, healthy life. My own children are fully vaccinated.

Providers are advised to communicate the roles and responsibility that the hesitant parent/patient needs to take on if they choose not to vaccinate and to convey that as a health professional they

are uncomfortable with the parent/patient's decision, emphasizing that it is against the overwhelming scientific consensus. How the health care provider introduces immunization at a visit also matters. Taking a presumptive approach, e.g., "Tom is due his vaccinations today," as opposed to a participatory one, e.g., "what do you want to do about vaccinating Tom today?" may also affect the likelihood of immunization acceptance; however, more research is required on this approach. For a very worried hesitant parent/patient, the provider should consider how to find and present extra evidence, information, and narratives, and how to dedicate more time, possibly through follow-up appointments. Consider using images and other ways of explaining risks, avoiding jargon and sticking to the facts. At all costs, the provider must maintain the relationship. Parent/patients who are dismissed or feel alienated ultimately find a source, possibly a provider, who supports and agrees with their decision not to vaccinate.

4.5.5 Role of the School

Reaching parents of today and tomorrow by educating pupils (and their parents) in school settings may significantly boost immunization acceptance and resilience of communities. Although little evidence has been generated from vaccination education in school settings, there is evidence that in other areas such as alcohol and substance abuse, sexual and reproductive health, nutrition, and bullying, curricula have shaped beliefs, including the successful development of "Health Promotion schools" under the WHO's Global School Health Initiative. In general, schools provide an important setting for health promotion, with the potential to reach over 1 billion children worldwide and through them, school staff, families, and whole communities. Providing education on vaccines and immunization in school settings can help children to develop informed critical thinking and decision-making skills, provide knowledge about vaccinations, promote positive attitudes toward immunization, and help to prepare them to make informed choices as parents/patients in the future and be more resilient in the face of anti-vaccine misinformation, includ-

ing influencing health-related behaviors of the teachers. Pupils around the age of 10 years might be selected as a starting point as they have the cognitive maturity and ability to understand the complexity of the immune system and think beyond the concrete concepts. There are few immunization examples to share, but inclusion of digital learning material, “edutainment,” and “gaming,” through which teachers and/or parents can guide students to make their own scientific discoveries, witness and understand the history of vaccines, could be adapted from methods used for delivery of other health and social development curricula. Just as education on the environment and ecology has shaped a generation’s perception of climate change, so can immunization perceptions be shaped.

4.5.6 Role of the Internet

For active seekers of information, the internet is an important channel that is growing in terms of its reach and influence on vaccination decisions. In Europe, reliable, trustworthy, easy to understand web-based information on vaccine-preventable diseases and the benefits of vaccines is often not available, is difficult to find or is not in a language that is helpful. Programs have a responsibility to address this and to offer parent/patients and providers a website that is well managed, well resourced, reviewed (format and content), and regularly updated with qualified and well-referenced information. Preferably, these sites should include a mechanism where user feedback and interaction is accommodated – such as a question–answer function. The WHO Global Advisory Committee on Vaccine Safety (GACVS) has compiled a list of websites that provide information on vaccine safety and follow good information practices. GACVS developed four categories of criteria for good information practices – regarding credibility, content, accessibility, and design to which sites providing information on vaccine safety should adhere. Programs are recommended to consider the VSN project when establishing their website and to become a member by meeting the criteria.

4.6 Pain Management

Immunizations are the most commonly recurring health-related procedure undertaken in childhood and the one most associated with needles. For many children, these procedures can cause unnecessary stress and anxiety, which if not mitigated, can lead to long-term non-adherence with recommended health care interventions and missed immunizations. For parents, vaccination sessions can be stressful and involve strong emotional reactions from both the infant/child and the parent. Providers are recommended to familiarize themselves with the WHO position paper: *Reducing pain at the time of vaccination (September 2015)* and consider some of the practices proven to reduce pain and anxiety. These include, but are not limited to, techniques to position the child differently or to distract the child. In addition, topical local anesthetic is very effective; however, it was not included in the guideline as it was not readily accessible in low income countries, but is recommended in Canada’s guideline.

4.7 Conclusion

It is evident that the immunization end-user’s experiences and perceptions have been undervalued and consequently under-researched. Without understanding these, in addition to the practical and structural barriers to vaccination that people face, immunization programs continue to struggle to equitably extend the benefits of vaccination to protect populations throughout the course of life and across all sectors of society.

There is no strong evidence to recommend any specific intervention for addressing vaccine hesitancy/refusal. Multi-pronged programs, community- and individual-level strategies, including innovative new methods, should be considered. Interventions should be based upon a degree of audience insight and take into consideration both supply-side modification and parent/patient behavior change, addressing more than a knowledge deficit in addressing hesitancy or sub-optimal demand. Interventions should be tested according to the target population, the context

within which the intervention is to take place, and the degree to which interventions can be tailored. At best, we can be moderately confident in the strategies presented in this chapter, as little research has been conducted into strategies and very few have been evaluated, suggesting that immunization programs might still require focus.

The attention to demand side factors, themselves at least the counterbalance to supply side issues, and acknowledgement of the value of behavioral and community insight to direct and inform policy and strategy, are necessary developments in Europe. However, it is apparent that immunization program delivery in Europe has some way to go before it becomes people-centric: designed to meet the needs of the end-users and responsive to evolving parent/patient and provider expectations of immunization service delivery.

Further Reading

- Butler R, MacDonald NE, SAGE Working Group on Vaccine Hesitancy. Diagnosing the determinants of vaccine hesitancy in specific subgroups: the guide to tailoring immunization programmes (TIP). *Vaccine*. 2015;33(34):4176–9.
- Dubé E, Gagnon D, MacDonald NE, SAGE Working Group on Vaccine Hesitancy. Strategies intended to address vaccine hesitancy: review of published reviews. *Vaccine*. 2015;33(34):4191–203.
- Fu LY, Zook K, Gingold JA, Gillespie CW, Briccetti C, Cora-Bramble D, Joseph JG, Haimowitz R, Moon RY. Strategies for improving vaccine delivery: a cluster-randomized trial. *Pediatrics*. 2016;137(6). pii: e20154603.
- Gesser-Edelsburg A, Walter N, Shir-Raz Y, Sassoni Bar-Lev O, Rosenblat S. The behind-the-scenes activity of parental decision-making discourse regarding childhood vaccination. *Am J Infect Control*. 2016. pii: S0196-6553(16)30962-2. doi:10.1016/j.ajic.2016.10.009.
- Goldstein S, MacDonald NE, Guirguis S, SAGE Working Group on Vaccine. Health communication and vaccine hesitancy. *Vaccine*. 2015;33(34):4212–4.
- Harvey H, Reissland N, Mason J. Parental reminder, recall and educational interventions to improve early childhood immunisation uptake: a systematic review and meta-analysis. *Vaccine*. 2015;33(25): 2862–80.
- Jarrett C, Wilson R, O'Leary M, Eckersberger E, Larson HJ, SAGE Working Group on Vaccine. Hesitancy strategies for addressing vaccine hesitancy - a systematic review. *Vaccine*. 2015;33(34):4180–90.
- Larson HJ, Smith DM, Paterson P, Cumming M, Eckersberger E, Freifeld CC, et al. Measuring vaccine confidence: analysis of data obtained by a media surveillance system used to analyse public concerns about vaccines. *Lancet Infect Dis*. 2013;13(7):606–13.
- Leask J, Kinnersley P, Jackson C, Cheater F, Bedford H, Rowles G. Communicating with parents about vaccination: a framework for health professionals. *BMC Pediatr*. 2012;12:154.
- MacDonald NE, SAGE Working Group on Vaccine Hesitancy. Vaccine hesitancy: definition, scope and determinants. *Vaccine*. 2015;33(34):4161–4.
- MacDonald NE, Finlay JC, Canadian Paediatric Society, Infectious Diseases and Immunization Committee. Working with vaccine-hesitant parents. *Paediatr Child Health*. 2013;18(5):265–7.
- Ndeffo Mbah ML, Liu J, Bauch CT, Tekel YI, Medlock J, Meyers LA, Galvani AP. The impact of imitation on vaccination behavior in social contact networks. 2012. <http://dx.doi.org/10.1371/journal.pcbi.1002469>.
- Nowak GJ, Gellin BG, MacDonald NE, Butler R, SAGE Working Group on Vaccine Hesitancy. Addressing vaccine hesitancy: the potential value of commercial and social marketing principles and practices. *Vaccine*. 2015;33(34):4204–11.
- Nyhan B, Reifler J, Richey S, Freed GL. Effective messages in vaccine promotion: a randomized trial. *Pediatrics*. 2014;133(4):e835–42.
- Odone A, Ferrari A, Spagnoli F, Visciarelli S, Shefer A, Pasquarella C, Signorelli C. Effectiveness of interventions that apply new media to improve vaccine uptake and vaccine coverage. *Hum Vaccin Immunother*. 2015;11(1):72–82.
- Schmidt P, MacDonald NE, Habersaat K, Butler R. Commentary to: how to respond to vocal vaccine deniers in public. *Vaccine*. 2016. pii: S0264-410X(16)30914-8 ahead of print.
- Shelby A, Ernst K. Story and science: how providers and parents can utilize storytelling to combat anti-vaccine misinformation. *Hum Vaccin Immunother*. 2013;9(8):1795–801.
- World Health Organization Regional Office for Europe. 2012. Information for parents. If you choose not to vaccinate your child, understand the risks and responsibilities. http://www.euro.who.int/__data/assets/pdf_file/0004/160753/If-You-Choose-Not-to-Vaccinate.pdf?ua=1.
- World Health Organization. Report of the SAGE Working Group on Vaccine Hesitancy. http://www.who.int/immunization/sage/meetings/2014/october/SAGE_working_group_revised_report_vaccine_hesitancy.pdf?ua=1.
- World Health Organization. Reducing pain at the time of vaccination: WHO position paper – September 2015. *Wkly Epidemiol Rec*. 2015;90:505–10.

Adjuvants in Pediatric Vaccines

Nathalie Garçon

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5.1 Introduction

The exponential evolution of scientific knowledge during the first half of the twentieth century led to the emergence of new and improved ways of producing vaccines. Vaccines were produced from cultivating the pathogens, but this has not always been possible in sufficient quantities. The rise of molecular biology and a better understanding of the key components of immune protection have allowed the development and production of what is known as recombinant antigens. Most, if not all purified and recombinant antigens, require to be effective the addition of what is known today as adjuvants. They are an important part of the development of improved or new vaccines against infectious diseases, alongside DNA or vector-based vaccines.

5.2 Definition of Adjuvants

An adjuvant, from the Latin word *adjuvare* meaning to help or aid, is a substance used to improve a vaccine's immune response by accelerating, prolonging, or enhancing the immune responses specific to the vaccine antigen(s), in particular by increasing mean antibody (Ab) titers of the population being immunized.

It is clearly accepted today that all current whole, attenuated, subunit, purified recombinant protein and peptide vaccines are adjuvanted (endogenously: part of the pathogen) or exogenously (added to the antigen formulation).

Indeed, during this evolution moving from whole killed or attenuated pathogens, to particulate vaccines, combined, with the tools of modern biotechnology, vaccines have not only seen an increased safety and lowered reactogenicity profile, but also the loss of many of the later immunological stimuli needed to trigger an effective immune response. For these vaccines, adjuvants became an important tool to ensure efficient and lasting immune response.

Until the early 1980s, adjuvantation science was limited to the use of aluminium salts. Following the emergence of HIV and the following attempts to develop HIV, it appeared that

aluminum salts were not enough to induce a protective immune response when combined with recombinant antigens. This revived the interest in adjuvants, and over the past 30 years, there has been an exponential growth of information regarding pattern recognition receptors (PRRs) that can activate leukocytes and thereby enhance immune responses.

When properly designed, selected, and combined with the relevant antigen(s), adjuvants can enable the appropriate and long-term immune response required to protect against the disease, with a safety profile acceptable in the targeted population. To date, no combination of recombinant antigen and adjuvant has demonstrated the ability to induce a CD8 immune response in naive human subjects, and adjuvants that enhance CD4 T cell responses are critical for durable vaccine immunity.

The understanding of the mode of action of adjuvants has greatly benefited from the discovery of pathogen associated molecular patterns (PAMPs) and their associated receptors (Toll-like receptors [TLR], nucleotide-binding oligomerization domain [NOD]-like receptors [NLR]), inflammasome components, and has been critical to the understanding of the link between innate and adaptive immunity and the associated pivotal role of dendritic cells. Despite these advances, a rational design approach would clearly benefit from a better understanding of the roles of innate and adaptive immunity and their impact on vaccine safety and immunogenicity.

5.3 Adjuvants in Vaccines

New vaccines based on recombinant antigens and adjuvants have put vaccine formulation at the center of vaccine development. Chemical structure, physicochemical characteristics, stability, the nature of the induced immune response, the impact on innate immune response, and the mode of action are key for their evaluation and use.

To date, there are nine different adjuvants present in licensed adjuvanted vaccines. Amongst those, seven are licensed for use in pediatric populations (■ Table 5.1).

Table 5.1 Adjuvants in vaccines licensed for pediatric populations

Aluminum salts	Phosphate or hydroxide	D, T, Pa, Hib, HBV, HAV, IPV, pneumococcus, HPV
Emulsion	MF59	Seasonal influenza
	AS03	Pandemic influenza H1N1 and H5N1
	AF03	Pandemic Influenza H1N1
Liposomes	Virosomes	Seasonal influenza
Combination	Aluminum + MPL	HPV

5.4 Aluminum Salts

The evaluation and use of aluminum salts in vaccines emerged in 1921 when a diphtheria vaccine based on inactivated diphtheria toxin (toxoid) was shown to be protective against diphtheria toxin. In 1926, aluminum precipitation was shown to enhance antibody response to diphtheria toxoid in guinea pigs, and in 1932 it was shown that alum enhances response to diphtheria toxoid immunization in humans. In 1939, Al-hydrogel became commercially

available, and since then, several billions of aluminum-containing vaccine doses have been used around the world. Several types of aluminum salts have been developed. They are particulate in nature, are different with regard to their surface charge, allowing effective adsorption of the antigen depending on its point of zero charge (pH at which the antigen has a neutral charge). The antigen adsorption increases the specific immune response and the antigen stability. Aluminum adjuvants are present in most of the currently licensed vaccines (Table 5.2). Although aluminum-containing vaccines are licensed across the world, the amount of Al present in a vaccine can vary depending on the country considered (Table 5.3).

The mode and mechanism of action by which aluminum salts have an impact on the human immune system is not fully deciphered and appears to be both direct and indirect. Through the transformation of antigens into a particulate through their adsorption on aluminum salts, antigen interaction with antigen-presenting cells (APCs) and macrophages is optimized compared to a soluble antigen formulation. To date, various possible mechanisms of action have been described (Table 5.4).

Aluminum salts have the longest and largest safety track record of all adjuvanted vaccines, with more than 3 billion vaccine doses used during the

Table 5.2 Aluminum-containing vaccines licensed for pediatric vaccines

Adjuvant	Vaccine
Alum: aluminum potassium sulphate Alhydrogel: aluminum hydroxide Adju Phos: aluminum phosphate Proprietary aluminum hydroxide and phosphate	DTaP (pediatric diphtheria, tetanus and acellular pertussis)
	DTaP, polio and <i>Haemophilus influenzae</i> type b
	DTaP, polio, <i>Haemophilus influenzae</i> type b and hepatitis B
	Hepatitis A
	Hepatitis B
	Hepatitis A/B
	Human papillomavirus-6/11/16/18
	Influenza (H5N1)
	Pneumococcus (conjugated)
This is not an exhaustive list; it focuses on the USA and Europe	

Table 5.3 Limits of elemental aluminum (Al³⁺), reported per human dose

Region	Reference/product	Limit (Al ³⁺) mg/dose
USA	21CFR Part 610 "General Biological Products Standards"	0.85
EU	European pharmacopoeia "Vaccines for Human Use"	1.25
WHO	WHO technical report series	1.25
China	DTPa	0.17–0.26
	Diphtheria vaccine adsorbed	0.52
	Tetanus vaccine adsorbed	0.52
	Diphtheria and tetanus combined vaccine, adsorbed	0.43
	HAV	0.60
	HBV	0.18–0.31
Japan	Adsorbed purified pertussis	0.15
	Adsorbed diphtheria-purified pertussis-tetanus	0.15
	HPV	0.42–0.58
	Recombinant adsorbed hepatitis B vaccine	0.325
India	HBV	1.25
	DTP	1.25

past 80 years and a positive risk–benefit ratio. Focal histological lesions were observed in vaccinees with diffuse muscular symptoms that included persistent myalgias, arthralgias, and persistent fatigue. In the approximately 130 cases observed, these lesions were identified as macrophagic myofasciitis (MMF). Intracytoplasmic inclusions in the infiltrating macrophages have been identified as containing aluminum by electron microscopy, microanalysis, and atomic adsorption spectroscopy. There is no established relationship between the presence of aluminum and MMF and the clinical symptoms, however. The Vaccine Safety Advisory Committee of the World Health Organization (WHO) reviewed MMF during a

Table 5.4 Mode of action of aluminum

Crystalline alum binds lipids on the surface of DCs	Cellular activation cascade triggering an immune response
Directly or indirectly triggers innate immunity through activation of inflammasome complexes	Likely nucleotide-binding oligomerization (NOD)-like receptor (NLR)-mediated effect is still present in MyD88 and TRIF in knockout mice
Induces cell death, which modulates the environment towards an enhanced adaptive immune response	Damage-associated molecular pattern release, such as uric acid and dsDNA, act as autologously derived adjuvants

meeting in 1999 and found no basis for recommending a change in vaccination practices (vaccine selection, schedule, delivery practices, or information on aluminum-containing vaccines). Studies have been undertaken since then, to evaluate the clinical, epidemiological, and basic science aspects of MMF. Although it is recognized that aluminum salts may be found months or years later at the intramuscular injection site after vaccination, to date, no link has been clearly established with the MMF syndrome.

5.5 Emulsions

Since the development of Freund's adjuvant, numerous emulsions have been evaluated in human. Water-in-oil emulsions (emulsified water droplets in a continuous oil phase) have been removed from testing following unacceptable reactogenicity (cysts at the injection site) and a lack of formulation reproducibility. The development of alternative emulsions (oil-in-water where oil droplets are in a continuous aqueous phase) was then undertaken. They represent the class of emulsion currently licensed in pediatric vaccines. They are made of particles of less than 200 μm (allowing for sterile filtration), are made of metabolizable naturally occurring oils such as squalene and stabilized by non-ionic surfactants such as Tween 80 and Span 85. They have been shown to enhance antibody responses and allow for antigen dose sparing particularly seasonal and pandemic

influenza vaccines, using MF59 (Fluid, Focetria), AS03 (Pandemrix), and AF03 (Humenza) as adjuvants. Oil-in-water emulsion can have a deleterious effect on antigen stability depending on the nature of the antigen, and has not yet been shown to improve antigen stability. Their mechanism of action may vary depending on the emulsion considered. Post H1N1pdm09 vaccination, reports of narcolepsy caused great concern and to date, no correlation or mechanism explaining these events has been yet established (see ► Chap. 14).

5.6 Virosomes

Virosomes are liposome-based formulations that can incorporate hydrophobic components within their membrane and hydrophilic ones as a cargo within the particle internal volume. They can act both as antigen carrier and adjuvant through the incorporation of immunomodulatory molecules.

In the case of Inflexal (seasonal influenza vaccine) the virosomes are made up of empty influenza virus envelopes that present the HA antigen within their membranes.

The mode of action of virosomes is not yet understood. It is, however, hypothesized that it relies on binding to macrophages and APC membranes, leading to the engagement of the innate and adaptive immune mechanisms.

5.7 TLR4 Agonists and Adjuvant Systems

At the forefront of PRRs are detoxified congeners of endotoxin that stimulate TLR4. Present in Cervarix, one of the human papilloma virus vaccines, it is derived from lipopolysaccharide the *Salmonella minnesota* lipopolysaccharide through a specific process that allows for a very significant reduction of its pyrogenicity (2–3 log), whilst retaining its adjuvant effect. In this vaccine, monophosphoryl lipid A (MPL) is combined with aluminum hydroxide and is known as AS04 adjuvant. Its mode and mechanism of action have been thoroughly evaluated. The efficacy and safety report in the target population has allowed for vaccine registration worldwide, making AS04 the first adjuvant, other than aluminum salts, to be present in a licensed vaccine in the US.

5.8 Additional Adjuvants in Development

Building on the successful results obtained with MPL, and a better understanding of the mechanisms of action of the current immunomodulators, a number of additional adjuvants are being evaluated in the context of various vaccines.

5.8.1 Defined Agonists of PRRs

Numerous PRR agonists targeting TLRs, NOD-like receptors or retinoic acid-inducible gene (RIG)-like receptors have been evaluated in adult human clinical trials. Several TLR agonists such as double-stranded ribonucleic acid (dsRNA), flagellin, single-stranded RNA or CpG have demonstrated different levels of activity. Several have also been shown to be capable of inducing an effective immune response in animal models, including mucosal adjuvants. Those capable of targeting the endosomal compartment have demonstrated the most robust impact on cellular immunity so far.

5.8.2 Saponins

As most of the adjuvants used or developed for human vaccines haven shown strong local reactogenicity, efforts have been undertaken to purify out from the mixture a specific molecule (QS21) that presents the optimum ratio between adjuvant effect and low local reactogenicity. This, however, was not sufficient to fully abrogate the lytic activity observed, and improvement through formulation was developed. The ability of Quil-A saponins to interact strongly with cholesterol was the cornerstone of the two formulations that were developed: one, known as ISCOMs/ISCOMATRIX, uses specific fractions of Quil-A, the other uses specific cholesterol-containing liposomes that are able to completely quench the lytic activity, while retaining the adjuvant activity. This later adjuvant combined with MPL, is known as AS01, and is present in the malaria candidate vaccine RTS,S, as well as the recombinant zoster vaccine. AS01 acts through the TLR4 activation capability of MPL, increases APC recruitment and activation, leading to a stronger and more persistent immune response.

5.8.3 Particulates

The use of particulates in vaccines goes back to the early 1920s when G. Ramon, then at the Pasteur institute, developed a method of increasing the production of hyperimmune sera, while avoiding the frequent abscesses observed in horses after toxoid administration. It is the adsorption of antigens on those particles that increases the immune response (the principle used for aluminum salts), and decreases or prevents abscesses by the concomitant adsorption of endotoxins. Bio-degradable polymers (such as polylactic, polyglycolic) have been extensively explored with the hope of designing nano- or micro-particles, where the antigens could be entrapped within or adsorbed on their surface. This should allow for a slow release of the antigens, leading to a single-shoot vaccine approach. Those polymers, however, due to their sensitivity to hydrolysis, need to be lyophilized and kept in a humidity-controlled environment until use.

Recent advances in polymer synthesis and particles engineering have allowed for the development of delivery systems with defined size, shape, and components, allowing for an approach tailored to the antigen to be delivered and, cell or cell compartment to be targeted. This has the potential for a rational design approach to the field of vaccine delivery systems.

5.9 Specific Needs for the Pediatric Population

Today, pediatric populations are the primary beneficiary of vaccination, whereas most adjuvant research and development is done for vaccines to be used in older populations. As many of the adjuvants described above can have a varying of impact on immunogenicity and reactogenicity when applied to younger populations, a better understanding of the immune status and its evolution across ages, in addition to the impact of adjuvants in those settings, is critical to understand how adjuvants may be best used in children.

5.9.1 Immunogenicity

The emergence and development of new tools first applied to drug discovery, such as medicinal chemistry for the design and synthesis of molecules tailored to the need for early life immunity, their evaluation in high throughput models based on infants' leukocytes, and their optimization through modern computational algorithms, can reasonably be seen as the next step toward the rational design of adjuvants for all target populations, including pediatrics.

The evaluation of the vaccine's immunogenicity and efficacy in animal models predictive of infant human populations can be expensive and unpredictable. In vitro approaches, which have the potential to accurately reflect the in vivo activity of those adjuvants in the target population, would allow for a rational design and selection of the adjuvant to be used, and a focused preclinical evaluation. Given the leaps that are being made today, both in fundamental science and in technology development such as organ on a chip, these approaches may be a reality in the near future.

5.9.2 Reactogenicity

A key concern regarding adjuvanted vaccine development is reactogenicity, i.e., the ability of a formulation to cause acute inflammatory events locally or systemically (such as fever). Their optimization may require adaptations such as modifying their pharmacokinetic properties to affect their biodistribution or tailoring the formulation to ensure co-delivery of the antigen(s) and adjuvant to the same APC. The discovery of biomarkers as surrogate markers of in vivo reactogenicity would allow for the rational screening of potential candidates and accelerate the selection of the optimal candidate for a specific vaccine.

5.10 Conclusion

The emergence of new diseases that can affect populations of all ages worldwide, in addition to the re-emergence of childhood diseases, needs to

be tackled using new or improved technologies. Adjuvants have been, for the past decades, one of the most promising advances in the development of new or improved vaccines. They have been developed and tested to a large extent for and in the adult population. Little has been done to specifically design adjuvants that are best suited to pediatric populations, in part because of the less advanced understanding of the pediatric immune system and the challenges posed by the small size of infants.

Given the evolution of knowledge and technologies observed during the last few decades, it is possible today to envision the identification of biomarkers predictive of better safety and immunogenicity that allow for their targeted use in pediatric populations when and where needed.

Further Reading

- Awate S, et al. Mechanisms of action of adjuvants. *Front Immunol*. 2013;4:114.
- Bachmann MF, Jennings GT. Vaccine delivery: a matter of size, geometry, kinetics and molecular patterns. *Nat Rev Immunol*. 2010;10:787–96.
- Brenner A. Macrophagic myofasciitis: a summary of Dr. Gherardi's presentations. *Vaccine*. 2002;20(Suppl 3):S5–6.
- Broadbent A, Subbarao K. Influenza virus vaccines: lessons from the 2009 H1N1 pandemic. *Curr Opin Virol*. 2011;1(4):254–62.
- Didierlaurent A, et al. Adjuvant system AS01: helping to overcome the challenges of modern vaccines. *Expert Rev Vaccines*. 2017;16:55–63.
- EMA. Assessment report immunological differences of pandemic vaccines (review of hypothesis on Pandemrix and development of narcolepsy). 2012. http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Assessment_Report_-_Variation/human/000832/WC500118056.pdf.
- Garçon N. Development of an AS04-adjuvanted HPV vaccine with the adjuvant system approach. *Biodrugs*. 2011;25(4):217–26.
- Glenny AT, Sudmersen HJ. Notes on the production of immunity to diphtheria toxin. *J Hyg*. 1921;20:176.
- Gomes AC, Mohsen M, Bachmann MF. Harnessing nanoparticles for immunomodulation and vaccines. *Vaccines (Basel)*. 2017;5
- Park WH, Schroder MC. Diphtheria toxin-antitoxin and toxoid: a comparison. *Am J Public Health Nations Health*. 1932;22:7–16.
- Plotkin SA, Plotkin SL. The development of vaccines: how the past led to the future. *Nat Rev Microbiol*. 2011;9:889–93.
- PrabhuDas M, et al. Challenges in infant immunity: implications for responses to infection and vaccines. *Nat Immunol*. 2011;12(3):189–94.
- Riedel S. Edward Jenner and the history of smallpox and vaccination. *Proc (Bayl Univ Med Cent)*. 2005;18(1): 21–5.
- Rowley DA, Fitch FW. The road to the discovery of dendritic cells, a tribute to Ralph Steinman. *Cell Immunol*. 2012;273(2):95–8.
- Schwendener R. Liposomes as vaccine delivery systems: a review of the recent advances. *Ther Adv Vaccines*. 2014;2(6):159–82.

Maternal Immunization

Timo Vesikari and Adam Finn

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Vaccination during pregnancy has been viewed with caution until recently. For example, in clinical trials of vaccines, pregnancy has been a contraindication, and stringent measures have been taken to avoid inadvertent inclusion of pregnant subjects. For this reason, safety information about specific vaccines for pregnant women has not been available and is still missing for many. However, in recent years influenza and pertussis vaccination in pregnancy have become recommended practice in many countries, reversing the previous situation.

6.1 Live Viral Vaccines

Live attenuated rubella vaccine virus can cross the placenta but is not known to cause congenital rubella nor, in fact, any symptoms in the fetus or newborn. Nevertheless, rubella vaccine is contraindicated in pregnancy. Single rubella vaccine is no longer available, but the same applies for MMR (measles-mumps-rubella) vaccine although the measles and mumps components are not known to pass transplacentally. If MMR vaccine is indicated for women of childbearing age, pregnancy should be excluded before vaccine administration, and contraceptive precautions should be advised for 1 month following vaccination.

However, if MMR vaccine is given inadvertently, no specific measures need to be taken. The vast clinical experience of inadvertent administration of rubella and MMR vaccination suggests that these vaccines will not cause any harm to the fetus.

Live attenuated varicella and MMRV vaccines should be treated like MMR, i.e. not given in pregnancy but, if given accidentally, no specific measures taken.

6.2 Tetanus Immunization

Tetanus vaccine for pregnant women has been an integral part of WHO's EPI program since its conception in 1977. At that time, neonatal tetanus caused almost one million annual deaths globally. The recommendation has been to give two doses of tetanus vaccine at any time during pregnancy. The program has been a great success: in 2001, it was estimated that the mortality had fallen to

180,000 deaths annually and in 2015 to only 34,000.

The vast experience accumulated in the global effort to eliminate neonatal tetanus by vaccination during pregnancy provides valuable evidence of general safety of non-live vaccines in pregnant women and in this way has been part of the foundation for current immunizations in pregnant women with Tdap vaccines in Europe and elsewhere.

6.3 Pertussis Immunization

Infant immunization program in Europe start at 2 or 3 months of age, and protection is insufficient after one dose. Thus, infants remain susceptible to pertussis for several months at the age when pertussis is most dangerous. With the introduction of acellular pertussis vaccines, the immunity level in young people surrounding the newborn, and indeed in young mothers, will be lower than before and the risk to newborn infants of severe pertussis even greater.

In the UK, a resurgence of pertussis in newborns with an increase of deaths was observed in 2012, and the authorities responded quickly by offering vaccine to pregnant women. The program has been highly successful and had reached around 80% coverage. The program has successfully prevented pertussis deaths in neonates, and the only two reported pertussis deaths where vaccine was used were in infants of mothers immunized only shortly before delivery (■ Fig. 6.1).

The mechanism of protection is likely neonatal IgG antibodies acquired transplacentally from the mother although IgA antibody in breast milk may also have a role. Although maternal antibody in the infant has been shown to reduce seroresponses to primary schedule vaccination, the clinical significance of such effects for strength and duration of protection are uncertain and, if they do make a difference, would do so at a time of lower risk of severe disease. Nevertheless, this raises questions about the optimal primary schedule for infants of immunized mothers. Maternal immunization seems to have a negative impact on immune response of the infants to routine immunization, including pertussis, diphtheria, and tetanus antibodies and antibody response to pneumococcal conjugate vaccine.

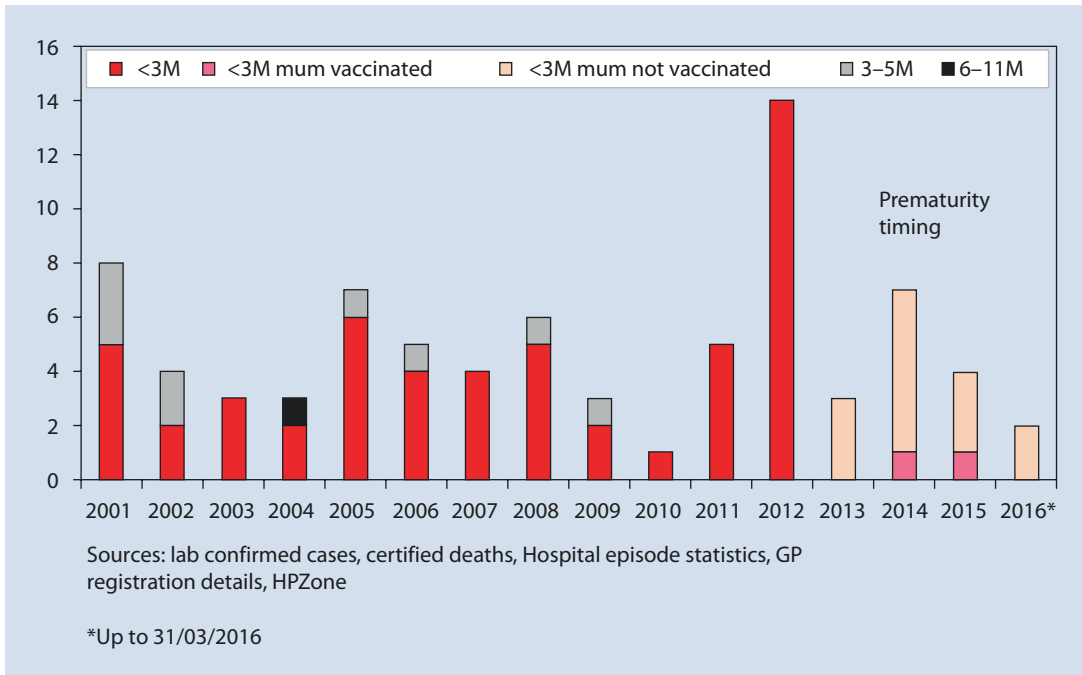


Fig. 6.1 Reconciled deaths from pertussis in infants, England 2001–2015 (From: [https://www.gov.uk/government/publications/vaccination-against-](https://www.gov.uk/government/publications/vaccination-against-pertussis-whooping-cough-for-pregnant-women)

[pertussis-whooping-cough-for-pregnant-women](https://www.gov.uk/government/publications/vaccination-against-pertussis-whooping-cough-for-pregnant-women). Vaccination against pertussis (whooping cough) for pregnant women: an update for healthcare professionals)

While the UK program aims to prevent infant pertussis, in practice a Tdap-polio combination vaccine, such as Boostrix-IPV, is given to pregnant women. Other countries, notably Belgium in 2013, have followed the UK model and started vaccination of pregnant women with combination vaccine. However, Europe is divided in this regard, and the majority of countries do not (yet) recommend pertussis vaccination of pregnant women.

6.4 Influenza Vaccination

With the emergence of H1N1pdm09 pandemic, it was soon recognized that swine flu was serious and more often fatal in pregnant women. When monovalent H1N1pdm09 vaccines became available in late 2009, they were recommended and given to pregnant women to protect them against severe pandemic influenza. Several studies were conducted on safety and efficacy of this practice, and it was confirmed that the vaccine protected pregnant women and was not only safe for the fetus but actually decreased fetal complications.

Meanwhile, in 1998, Neuzil and coworkers had already shown that influenza vaccination reduces the risk of severe complications of seasonal influenza in pregnancy. This had already led to consideration of influenza vaccination of pregnant women, and the good experience of H1N1pdm09 vaccination formed another stimulus for the US ACIP in 2010 to reinforce recommendations for influenza vaccination for all women who are pregnant during influenza season. In 2012 WHO stated that influenza vaccination of pregnant women is a “highest priority.” Several European countries have adapted the recommendation, and others may follow as there is no clear opposition to this recommendation in contrast to pertussis vaccine.

While influenza vaccination for pregnant women was introduced to protect the women, it has also been documented that immunization of mothers will also protect infants against influenza up to 6 months of age (Fig. 6.2). This is of particular importance, because young infants are a high-risk group for influenza deaths and there is no influenza vaccination policy in sight for direct protection of infants younger than 6 months of age.

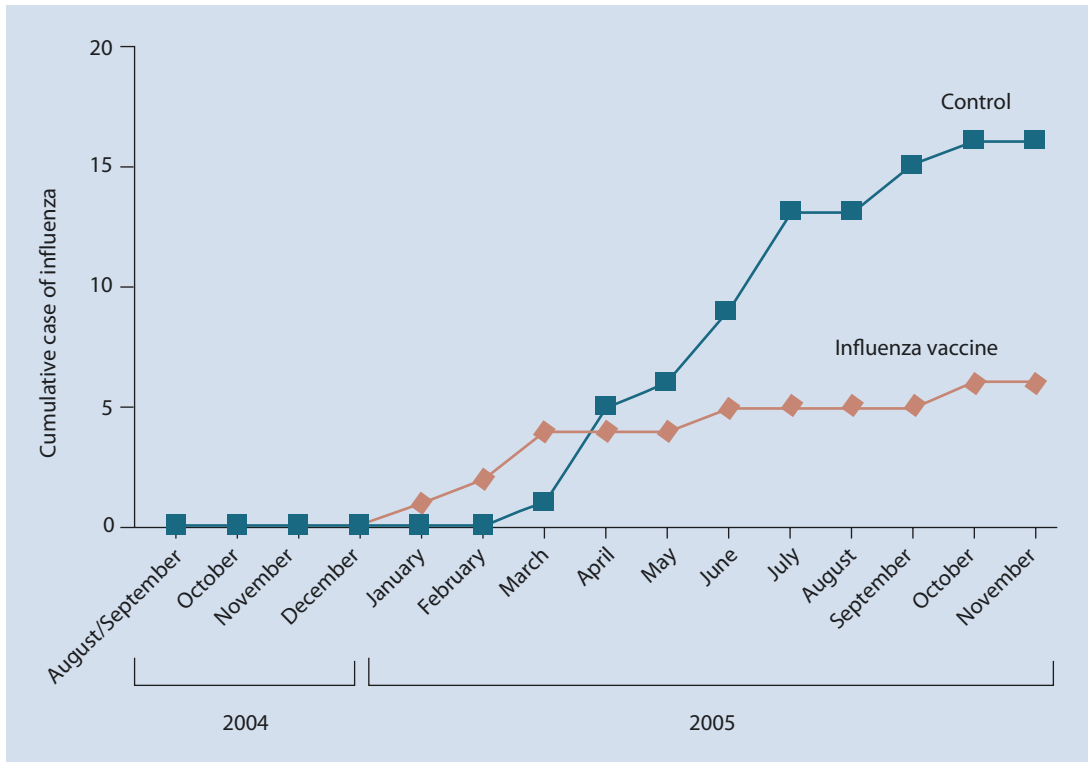


Fig. 6.2 Prevention of influenza up to 6 months of age in infants whose mothers received influenza vaccine during pregnancy (From: Zaman et al. (2008))

6.5 Future Prospects

Vaccines against respiratory influenza virus (RSV) are being developed, with non-live vaccines based on the viral F-protein being the strongest candidates. Such vaccines could conceivably be given either to young infants, even newborns, or to pregnant women to induce protection by trans-placental antibody. Both options are currently being investigated.

Other vaccines are in development with a potential to be given during pregnancy to prevent severe neonatal bacterial infections including group B streptococcal vaccine (see related chapters in Part IV).

Further Reading

Tetanus

Demicheli V, Barale A, Rivetti A. Vaccines for women for preventing neonatal tetanus. *Cochrane Database Syst Rev.* 2015;(7):CD002959.

Pertussis

Dabrera G, Amirthalingam G, Andrews N, et al. A case-control study to estimate the effectiveness of maternal pertussis vaccination in protecting newborn infants in England and Wales, 2012–2013. *Clin Infect Dis.* 2015;60(3):333–7.

Ladhani SN, Andrews NJ, Southern J, et al. Antibody responses after primary immunization in infants born to women receiving a pertussis-containing vaccine during pregnancy: single arm observational study with a historical comparator. *Clin Infect Dis Off Publ Infect Dis Soc Am.* 2015;61:1637–44.

Maertens K, Caboré RN, Huygen K, Vermeiren S, Hens N, Van Damme P, Leuridan E. Pertussis vaccination during pregnancy in Belgium: follow-up of infants until 1 month after the fourth infant pertussis vaccination at 15 months of age. *Vaccine.* 2016;34(31):3613–9.

Maertens K, Burbidge P, Van Damme P, Goldblatt D, Leuridan E. Pneumococcal immune response in infants whose mothers received Tdap vaccination during pregnancy. *Pediatr Infect Dis J.* 2017. <https://doi.org/10.1097/INF.0000000000001601>. [Epub ahead of print].

Munoz FM, Bond NH, Maccato M, Pinell P, Hammill HA, Swamy GK, et al. Safety and immunogenicity of tetanus diphtheria and acellular pertussis (Tdap) immunization during pregnancy in mothers and infants: a randomized clinical trial. *JAMA.* 2014;311:1760–9.

Influenza

- Fell DB, Platt RW, Lanes A, et al. Fetal death and preterm birth associated with maternal influenza vaccination: systematic review. *BJOG*. 2015;122:17–26.
- Håberg SE, Trogstad L, Gunnes N, et al. Risk of fetal death after pandemic influenza virus infection or vaccination. *N Engl J Med*. 2013;368:333–40.
- Moro PL, Broder K, Zheteyeva Y, et al. Adverse events following administration to pregnant women of influenza A (H1N1) 2009 monovalent vaccine reported to the vaccine adverse event reporting system. *J Obstet Gynecol*. 2011;205:473.e1–9.
- Neuzil KM, Reed GW, Mitchel EF, Simonsen L, Griffin MR. Impact of influenza on acute cardiopulmonary hospitalizations in pregnant women. *Am J Epidemiol*. 1998;148:1094–102.
- Sakala IG, Honda-Okubo Y, Fung J, Petrovsky N. Influenza immunization during pregnancy: benefits for mother and infant. *Hum Vaccin Immunother*. 2016;12(12):3065–71.
- Savulescu C, Jiménez-Jorge S, de Mateo S, et al. Using surveillance data to estimate pandemic vaccine effectiveness against laboratory confirmed influenza A(H1N1)2009 infection: two case-control studies, Spain, season 2009–2010. *BMC Public Health*. 2011;11:899.
- Zaman K, Roy E, Arifeen SE, et al. Effectiveness of maternal influenza immunization in mothers and infants. *N Engl J Med*. 2008;359:1555–64.

RSV

- Madhi SA, Dangor Z. Prospects for preventing infant invasive GBS disease through maternal vaccination. *Vaccine*. 2017;35(35Pt A):4457–60.
- Munoz FM. Respiratory syncytial virus in infants: is maternal vaccination a realistic strategy? *Curr Opin Infect Dis*. 2015;28(3):221–4.

Neonatal Immunization

Ener Cagri Dinleyici

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Neonatal immunization refers to the immunization of newborns during the first 28 postnatal days; however, neonatal immunization may also include vaccines used in the first 2 months of life and immunization practices among high-risk neonates, including preterm newborns.

Infections are more common and generally more severe in neonates and young infants than in older children and adults, particularly because immune defenses are functionally impaired in early life. There are two main approaches to preventing early life infections: maternal (mainly during pregnancy) and neonatal immunization. Considerable public health benefit can potentially be derived from the vaccination of women in pregnancy to protect newborns against specific infections (see ► Chap. 6). Neonatal immunizations have not shown the same progress as maternal immunizations, as there are some barriers and threats concerning this issue. The immunization studies have focused on the immune system of newborns, the potential use of existing vaccines during the neonatal period, immunization practices in premature babies, and new vaccines and adjuvants.

In the neonatal period, the different maturity of the immune system, particularly in premature infants, is associated with distinctive clinical, physical, and outcome characteristics with regard to infections compared with other age groups. Different etiological agents may often present with similar clinical features, and localized infections may present with systemic signs making clinical diagnosis difficult. In addition, respiratory distress syndrome, inborn errors of metabolism, and congenital heart disease have initial clinical presentations similar to severe infections. During the newborn period, diagnostic tools are limited and may lack sensitivity.

The same immune deficiencies that render newborns susceptible to infection also reduce their memory responses to most antigens, thereby potentially frustrating efforts to protect this high-risk population. As birth is the most reliable point of healthcare contact worldwide and effective vaccination at birth would provide early protection for newborns and infants, expanding and improving the available means of neonatal vaccination is a global health priority. At present, there are two good vaccines, the bacillus Calmette–Guérin (BCG) and the hepatitis B vaccines, which are routinely and widely used in neonates.

7.1 Immunity in the Neonatal Period

The neonatal immune system is no longer considered immature, but rather specifically adapted for early postnatal life, developing over time through a regulatory process that has not yet been well defined. Mohr and Siegrist described the neonatal immune system as characterized by anti-inflammatory, rather than pro-inflammatory, responses to danger signals and antigens, resulting in the preferential differentiation of CD4+ helper T cells (Th) toward Th2 cells antagonizing Th1 cells and cytotoxic responses against intracellular pathogens, based on the propensity to differentiate into immuno-regulatory cells over effector/memory cells, limited plasma cell and germinal center B cell responses, and occasionally the presence of maternal antibodies with immunomodulatory properties.

During the intrauterine period, reflecting low exposure to foreign antigens, the newborn adaptive immune system is primarily composed of naïve lymphocytes. The paucity of antigen-experienced cells confers vulnerability to serious pathogens and leaves newborns reliant on their innate immune system. This system is biased against the induction of the T helper 1 (Th1) cell polarization of cytokines, which is necessary to avoid alloimmune reactions between the mother and fetus or excess anti-inflammatory reactions, but increases susceptibility to many viral and bacterial pathogens. The neonatal immunological milieu is skewed toward T helper 2 (Th2) immunity to prevent the recognition of the developing fetus as an allograft by the maternal immune system, representing an important obstacle for vaccination during the neonatal period. In neonates, responses to two major danger pathways, the toll-like receptor signaling and interleukin (IL)-1/inflammasome pathways, are dampened and fail to induce potent pro-inflammatory responses, including that of IL-12p70, the master cytokine for Th1, and cytotoxic responses. The low responsiveness of neonatal T cells to toll-like receptor and IL-1/inflammasome pathways has an impact on the intrinsic ability of T cells to respond to vaccines and pathogens.

Early-life humoral responses are affected by B-cell intrinsic and extrinsic features/limitations, but they are primarily controlled by extrinsic fac-

tors. Follicular dendritic cells develop slowly after birth, delaying germinal cell formation, and bone marrow stromal cells provide insufficient survival factors, such as a proliferation-inducing ligand. The expansion of T follicular helper cells is a key limiting factor for the development of early life germinal complex responses.

Many factors determine the quality and quantity of the early infant antibody response, including the stage of development of the infant immune system, the type of vaccine and its intrinsic immunogenicity, the number of doses and intervals between doses, and the influence of maternal antibodies. Neonates and infants have a limited antibody repertoire, may produce suboptimal antibody responses to some polysaccharide and protein antigens, and show the limited persistence of these antibodies. Neonatal B-cell differentiation pathway is skewed toward memory B cells rather than plasma cells.

In addition to the challenge posed by the different maturity of the neonatal leukocyte compartment, effective neonatal vaccines must also overcome the potential inhibitory effect of maternal antibodies. Increasing the placental transfer of maternal antibodies effectively protects newborns and infants against some diseases, e.g., tetanus, influenza pertussis. The amount of antibody transferred is dependent on several factors, including gestational age, maternal antibody level, type of IgG subclass, and placental characteristics. Maternal antibodies may interfere with infant vaccine responses and also breast milk antibodies may affect the efficacy of vaccines. Concerns about the use of vaccines during the newborn period include the limited capacity of neonates to respond to many antigens and the potential effects of vaccinations on the immune system polarization during prenatal and early periods after birth.

7.2 BCG Vaccine

The *bacillus Calmette–Guérin* (BCG) vaccine (see also ► Chap. 17) is a live attenuated *Mycobacterium bovis* vaccine that is usually administered within the first few days of life in most low- and middle-income countries to prevent tuberculous meningitis and miliary tuberculosis. More than 120 million doses of BCG vaccine are administered

each year worldwide. Most infants receive BCG at birth in accordance with WHO recommendations. With more than 3 billion people having received this vaccine, BCG is one of the most widely used vaccines worldwide, and in general, the BCG vaccine exhibits an excellent safety profile. The protective efficacy of the neonatal BCG vaccine is 64–73% against meningitis and 77–78% against miliary tuberculosis. The efficacy varies between countries, particularly against miliary tuberculosis and tuberculosis meningitis, reflecting the disparate exposure to environmental mycobacteria, strain variations in BCG preparations, genetic or nutritional differences, and environmental factors, such as sunlight exposure and poor cold-chain maintenance. The greatest benefit of BCG immunization has been observed in regions where both the risk of tuberculosis and the rates of vaccine coverage are highest.

The efficacy of neonatal BCG administration has been linked to its ability to effectively induce anti-mycobacterial CD4⁺ T-cell Th1-polarized neonatal immune responses. BCG vaccination at birth results in neonatal IFN- γ production against mycobacterial antigens, and the levels of secreted IFN- γ are comparable with adult levels. Notably, BCG also effects the immune response to unrelated antigens in early life, boosting both Th1- and Th2-type responses to other antigens (e.g., HBV and oral polio vaccines), likely through its influence on dendritic cells (DC) maturation. Th1 responses are characterized by CD4⁺ T-cell interferon (IFN)- γ production. Enhanced neonatal Th1-polarized immune responses would be beneficial for combating infections with intracellular pathogens and toxin-producing organisms. Neonatal BCG vaccination has also been reported to reduce neonatal and infant mortalities resulting from diseases other than tuberculosis. The nonspecific beneficial effects may also include the reduction of atopic diseases (see ► Chap. 1).

Ritz et al. (2016) evaluated the CD4 and CD8 T-cell responses and cytokine release at birth and BCG immunization of infants born in Australia after the 2nd month. Cellular immunity measured at 10 weeks after BCG immunization was similar in infants administered BCG at birth and in those administered BCG at 2 months of age. These results suggest that delaying BCG immunization might not confer any immunological advantage in cellular immunity.

A major concern about the use of BCG vaccine at birth is the disseminated BCG infection. Disseminated BCG infection is a rare complication, occurring in less than one per million individuals, mainly with congenital immune disorders. BCG vaccination at birth is no longer recommended in HIV-positive infants because of the risk of disseminated BCG disease, in approximately 1%, and the limited vaccine efficacy in HIV-infected infants.

In Bulgaria, Hungary, Ireland, Latvia, Lithuania, and Portugal, the BCG vaccine is routinely recommended at 48 h following birth without tests. In Poland, the BCG vaccine is administered within 24 h of birth. In Croatia, vaccination is preferably administered at the time of delivery in the hospital, otherwise it should be administered before 1 year of age. In Cyprus and Luxembourg, vaccines are only administered upon specific indications at birth. In the Czech Republic, the BCG vaccine is administered from the 4th day until 6 weeks after birth to babies in at-risk groups. In Estonia, BCG administration is recommended in 1–5 days after birth. In Finland, France, Greece, Malta, the BCG vaccine is administered to specific at-risk groups only. In Liechtenstein, vaccination is recommended for newborns and infants under 12 months, if their parents are from countries with a high prevalence of tuberculosis. In Romania, BCG vaccination is recommended at 2–7 days after delivery. In Slovenia, vaccination is recommended for newborn infants of immigrant families who moved to Slovenia from countries with a high prevalence of tuberculosis in the last 5 years. In the UK, vaccination is recommended for babies and children who have a high chance of coming in contact with tuberculosis (see also ► Chap. 17).

7.3 Hepatitis B Vaccine

Primary prevention through immunization remains the most effective strategy for controlling the spread of hepatitis B virus. In healthy infants, one dose provides ~30–50% protection, two doses provide 50–75% protection, and three doses provide >90% protection against HBV infection. Immunity elicited by neonatal/infant HBV immunization persists life-long in the absence of antigen exposure/booster immunization (see ► Chap. 13). The WHO recommends, independent from endemicity, to offer hepatitis B vaccine univer-

sally within 24 h of birth, followed by two or three additional hepatitis vaccine doses.

In Europe, in Bulgaria, Estonia, Iceland, Lithuania, Liechtenstein, Luxembourg, Malta, the Netherlands, Poland, Portugal, Romania, UK, and Turkey, the first dose of the hepatitis B vaccine is recommended at 12–24 h after birth. In Austria, Belgium, Croatia, Cyprus, Czech Republic, Denmark, France, Germany, Greece, Ireland, Italy, Slovakia, Slovenia, Sweden, and Spain, hepatitis B vaccination is generally recommended at birth for babies born to a mother infected with hepatitis B, and an initial vaccination is offered at birth simultaneously with hepatitis B immunoglobulin. In Hungary and Latvia, the recommendation for hepatitis B vaccine at birth includes babies born to mothers with unknown immune status (see ► Chap. 13).

In 2017, the Advisory Committee on the Immunization Practices (ACIP) of the USA added monovalent hepatitis B vaccinations to all newborns within 24 h of birth. For infants born to hepatitis B surface antigen (HBsAg)-positive mothers, the ACIP recommends hepatitis B vaccine and hepatitis B immune globulin within 12 h of birth. ACIP recommendations include the administration of the hepatitis B vaccine, regardless of birth weight, when the HBsAg status of the mother is unknown.

7.4 Immunization of Premature Infants

It is generally recommended that premature infants should follow the same vaccination schedule as that generally used for full-term infants, without correcting for prematurity and regardless of birth weight. However, the routine immunization of premature infants is often delayed because many pediatricians think that the impaired immune systems of these infants could significantly suppress responses to vaccine antigens and reduce the protective effects of vaccination.

Numerous differences in vaccine responses between premature and full-term newborns have been observed. Skin, lung, and epithelial cells secrete less than adequate amounts of peptides, such as defensins, which could alter host gene expression, act as chemokines, and/or induce chemokine production, inhibit lipopolysaccharide-

induced pro-inflammatory cytokine production, and modulate the responses of dendritic cells and the cells of the adaptive immune response. For premature newborns, an impaired innate system is another important factor for immunization via antigen-presenting cell dysfunction resulting from suboptimal vaccine responses. Adaptive cellular and humoral immunity are also less efficient in premature newborns, including a suboptimal function of Th1 and a Th2 polarized response with the relative impairment of Th1 activity, significantly reduced T-cell repertoire limiting the recognition of the peptides, less IL-2 production, decreased cytolytic activity, and abnormal cytokine production associated with reduced/delayed T lymphocyte-related causes. Premature infants predominantly respond with IgM, and there is a slow or no switch to IgG. In premature infants, maternal antibody is lower than in term infants, which may actually improve vaccine responses.

Premature infants are capable of mounting overall systemic and local immune responses to inactivated polio vaccines comparable with those of full-term infants. Also, prematurity does not influence the immediate protection conferred by tetanus and diphtheria vaccines given according to calendar age. For acellular pertussis vaccines, even in extremely pre-term infants, pertussis antigens induce significant immune memory capable of providing protection for a long time. Clinical studies have shown that premature infants seroconvert in response to the hepatitis B vaccine by 30 days of age, regardless of gestational age and birth weight, suggesting that prematurity per se rather than gestational age or birth weight might be more predictive of a decreased antibody response.

The immunogenicity of the meningococcal C-conjugated vaccine in premature infants is not different from that of full-term infants. Most studies on the *Haemophilus influenzae* type b vaccine reported only marginal differences between premature and full-term infants. This finding clearly indicates that most premature infants, particularly those at a gestational age > 32 weeks, remain protected, even after the primary series. Premature infants are at an increased risk for invasive pneumococcal disease compared with term infants and are more likely to have lower vaccine responses compared with term infants. A recent clinical study that included 210 premature

newborns showed that after primary PCV13 vaccination, 75%, 88%, and 97% of participants had protective antibody concentrations for at least one-half of the PCV13 serotypes for the reduced, accelerated, and extended schedules respectively. After the booster vaccination, nearly all participants, regardless of schedule or serotype, had seroprotective IgG concentrations. A reduced priming schedule for PCV13 resulted in higher post-booster IgG concentrations, but lower post-primary concentrations.

Preterm infants are vulnerable to severe rotavirus infection resulting in hospitalization. Rotavirus vaccines are immunogenic and safe and have been demonstrated to have similar effects in preterm infants to term infants when given according to calendar age. However, preterm newborns are usually not given rotavirus vaccine at birth but only at a calendar age of 6–8 weeks.

Overall, premature infants should follow the same vaccination schedule as that generally used for full-term infants, without correcting for prematurity and regardless of birth weight. Even though an impaired immune response can reduce antibody production and cell-mediated immunity, antibody production is high enough to ensure short- and long-term protection in most premature infants.

7.5 The Need for Novel Approaches to Enhancing Neonatal Vaccination

The medical advantages inherent to neonatal vaccination at birth include the following: (1) early protection to close the window of vulnerability inherent to vaccination schedules that start later in life (e.g., 2 months), (2) the practicality of birth being a global point of contact with health care systems, and (3) potential advantages of novel vaccines that may require fewer doses to achieve efficacy.

Experimental studies have suggested that a key requirement for the induction of an effective neonatal adaptive response is the entrance of antigen into the cytoplasm of antigen-presenting cells (APC). Cytoplasmic delivery of antigens may enhance neonatal immune responses. A novel approach to neonatal vaccination has employed an attenuated strain of the intracellular pathogenic

bacterium *Listeria monocytogenes* to deliver antigen to the cytoplasm of APC.

Developing an effective Th1-inducing adjuvant for the neonatal period would be another step toward the goal of improving vaccinations against toxins (e.g., diphtheria/pertussis/tetanus) and viruses (e.g., polioviruses, measles virus) at birth. An effective neonatal Th1-inducing adjuvant should comprise a TLR2 agonist, type I IFN, and type II IFN. Novel adjuvants constitute another new promising field in neonatal immunizations. Toll-like receptor agonists might be potential antigens that experimentally induce interferon production and enhance the primary anti-tetanus toxoid immune responses. TLR8 agonists, including certain synthetic imidazoquinolines and single-stranded viral RNA, are particularly effective at activating human neonatal APC in vitro. Endosomal TLR ligand binding TLR-2, -7/8, and -9 trigger better responses in neonatal individuals and may offer a strategy for enhancing Th1 responses. C-lectins and/or neonatal plasmacytoid dendritic cells represent potential targets to elicit Th1 antiviral protection in newborns. All novel neonatal vaccines need to undergo rigorous safety evaluations and clinical studies in humans.

7.6 Conclusion

Newborns have a specific immune system that renders these individuals at a high risk for infection, while simultaneously reducing responses to most vaccines, thereby posing challenges in protecting this vulnerable population. The development of early life vaccination, including vaccines effective when administered at birth, is an important target of global health care contact. Neonatal immunization against tuberculosis and hepatitis B are widely practiced. Considering the potentially significant benefit of vaccinating at birth, the availability of a broader range of more effective neonatal vaccines is an unmet medical need and a public health priority.

Further Reading

Andersen P, Doherty TM. The success and failure of BCG – implications for a novel tuberculosis vaccine. *Nat Rev Microbiol.* 2005;3:656–62.

Berrington JE, Barge D, Fenton AC, Cant AJ, Spickett GP. Lymphocyte subsets in term and significantly preterm UK infants in the first year of life analysed by

single platform flow cytometry. *Clin Exp Immunol.* 2005;140(2):289–92.

Bonhoeffer J, Siegrist CA, Heath PT. Immunisation of premature infants. *Arch Dis Child.* 2006;91(11):929–35.

Cuenca AG, Wynn JL, Moldawer LL, Levy O. Role of innate immunity in neonatal infection. *Am J Perinatol.* 2013;30(2):105–12.

Demirjian A, Levy O. Neonatal vaccination: a once in a lifetime opportunity. *Pediatr Infect Dis J.* 2009a;28(9):833–5.

Demirjian A, Levy O. Safety and efficacy of neonatal vaccination. *Eur J Immunol.* 2009b;39(1):36–46.

Esposito S, Fumagalli M, Principi N. Immunogenicity, safety and tolerability of vaccinations in premature infants. *Expert Rev Vaccines.* 2012;11(10):1199–209.

Fine PE. Variation in protection by BCG: implications of and for heterologous immunity. *Lancet.* 1995;346:1339–45.

Gagneur A, Pinquier D, Quach C. Immunization of pre-term infants. *Hum Vaccin Immunother.* 2015;11(11):2556–63.

<http://vaccine-schedule.ecdc.europa.eu/Pages/Scheduler.aspx>

<https://www.cdc.gov/vaccines/schedules/downloads/child/0-18yrs-child-combined-schedule.pdf>

Kilich E, Sadarangani M. Use of rotavirus vaccines in pre-term babies on the neonatal unit. *Expert Rev Vaccines.* 2016;15(12):1463–5.

Levy O. Innate immunity of the newborn: basic mechanisms and clinical correlates. *Nat Rev Immunol.* 2007;7:379–90.

Libraty DH, Zhang L, Woda M, Acosta LP, Obcena A, Brion JD, Capeding RZ. Neonatal BCG vaccination is associated with enhanced T-helper 1 immune responses to heterologous infant vaccines. *Trials Vaccinol.* 2014;3:1–5.

Linterman MA, Hill DL. Can follicular helper T cells be targeted to improve vaccine efficacy? *F1000Res.* 2016;5

Mohr E, Siegrist CA. Vaccination in early life: standing up to the challenges. *Curr Opin Immunol.* 2016;41:1–8.

Morris MC, Surendran N. Neonatal vaccination: challenges and intervention strategies. *Neonatology.* 2016;109(3):161–9.

Ritz N, Casalaz D, Donath S, Tebruegge M, Dutta B, Connell TG, Robins-Browne R, Britton WJ, Hanekom WA, Curtis N. Comparable CD4 and CD8 T cell responses and cytokine release after at-birth and delayed BCG immunisation in infants born in Australia. *Vaccine.* 2016;34(35):4132–9.

Siegrist CA. The challenges of vaccine responses in early life: selected examples. *J Comp Pathol.* 2007;137(Suppl 1):S4–9.

Simon AK, Hollander GA, McMichael A. Evolution of the immune system in humans from infancy to old age. *Proc Biol Sci.* 2015;282(1821):20143085.

Stensballe LG, Sørup S, Aaby P, Benn CS, Greisen G, Jeppesen DL, Birk NM, Kjærgaard J, Nissen TN, Pihi GT, Thøstesen LM, Kofoed PE, Pryds O, Ravn H. BCG vaccination at birth and early childhood hospitalisation: a randomised clinical multicentre trial. *Arch Dis Child.* 2017;102(3):224–31.

Swaminathan S, Rekha B. Pediatric tuberculosis: global overview and challenges. *Clin Infect Dis.* 2010;50(Suppl 3):S184–94.



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Poliovirus Vaccines

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8.1 The Disease

Acute anterior poliomyelitis was recognized as a clinical entity in the late nineteenth century, and was shown to be caused by a virus in the early twentieth century. Initially, it was considered to be mainly a disease of young children, hence the old name “infantile paralysis”. Frequent large outbreaks through the Western world during the first half of the twentieth century – together with individual adult victims of the disease among persons with powerful positions in the USA – increased the interest in research and facilitated its funding. There were about 35,000 cases of paralytic polio annually in the USA before the introduction of vaccination in the mid-1950s.

Development of the cell culture techniques and propagation of polioviruses in the late 1940s enabled detailed studies of the disease, confirmation of the diagnosis by virological laboratory tests, and eventually, development of vaccines. Polioviruses are small, non-enveloped RNA viruses belonging to the family *Picornaviridae*, genus *Enterovirus*. Polioviruses infect only cells and tissues of humans or other primates, and humans are the only natural hosts of the virus. Polioviruses are divided into three distinct serotypes, referred to as poliovirus types 1, 2, and 3. Two types of poliovirus vaccines have been available since the late 1950s and the virus has been eliminated from circulation in human populations in most parts of the world. The last cases in Europe were reported in 1996 in Albania, Greece, and Kosovo and 1998 in Turkey. However, even with this rarity of new cases, the maintenance of immunity to poliovirus will still be important for years to come, as discussed in detail below. For the whole of 2016, a total of 37 cases worldwide caused by wild type 1 poliovirus (WPV1) have been reported to WHO.

8.1.1 Pathogenesis and Symptoms

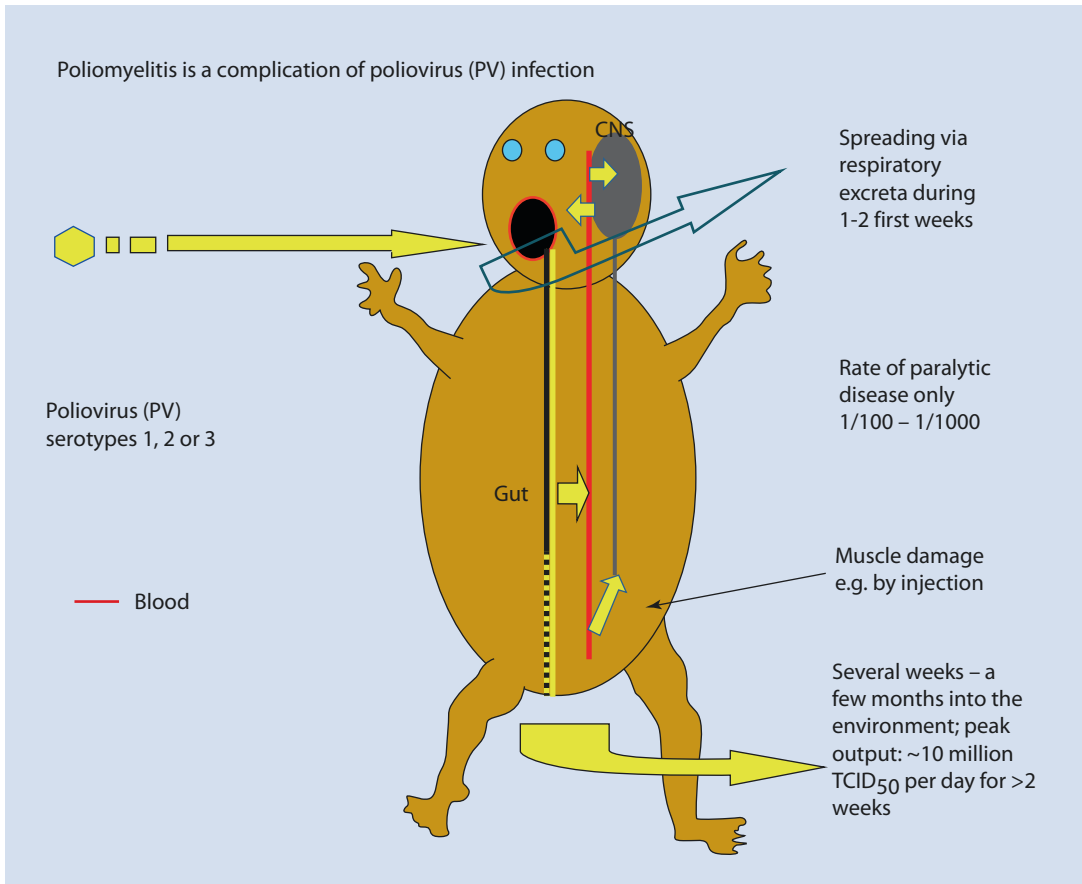
The virus enters the body in contaminated food or via close physical contacts to infected persons or their excreta. Primary virus replication takes place in the oropharyngeal or intestinal mucosa, and the virus then spreads to submucosal lymphatic tissues. This phase of the infection may present with nonspecific symptoms of acute infection. The virus is shed in the excreta of the oropharynx during the first 2 weeks of infection, and in the stools for sev-

eral weeks, up to a couple of months (■ Fig. 8.1). From the lymphatic tissues, the virus may enter the blood circulation, and thereby reach secondary replication sites, including the oropharynx and the central nervous system (CNS). In the CNS, the most common, but not exclusive target tissue is the medullary anterior horn (hence the full name, *acute anterior poliomyelitis*). Apart from crossing the blood–brain barrier, a viral route into the CNS can be initiated through mechanical damage of the axons of the motor neurons, for instance, by intramuscular injections and subsequent retrograde transport of the virus into the soma of the neuron. Lytic infection of the upper motor neurons results in rapid paralysis of the corresponding muscular fibers in the skeletal muscles. In the more severe forms of poliomyelitis, the bulbar nuclei are involved, and destruction of those regulating respiration and circulation may result in death.

Only 0.2–1% of immunologically naive individuals who are infected develop paralytic symptoms. The “typical” paralytic presentation of the disease could thus be considered a complication of the infection that is largely asymptomatic or associated with mild nonspecific symptoms of common acute infection. Acute mortality of paralytic patients is about 10%. Of the survivors, about one third recover to become symptom-free within a few months; another third have lifelong sequelae complicating mobility and skeletal development; and the rest live with milder, persisting symptoms. No specific treatment is available.

8.1.2 Immunity

An immune response follows the natural course of poliovirus infection irrespective of associated clinical symptoms. Both virus-specific class IgM, IgA, and IgG antibodies appear in the circulation and class IgA antibodies are excreted in oropharyngeal and gastrointestinal mucosa. Intestinal IgA antibodies are crucial for protection against reinfection of individuals and for the limitation of virus transmission in the population (herd immunity). Levels of IgM and IgA decay within months whereas the neutralizing class IgG response gives lifelong protection from paralytic disease. A cellular immune response can be demonstrated, but its potential role in the recovery, initial virus elimination, and later protective immunity is not well understood. The neutralization activity of the



■ Fig. 8.1 Schematic picture of poliovirus infection (T. Hovi, unpublished)

antibodies is type-specific and a person surviving paralytic poliomyelitis caused by, say, type 1 poliovirus, remains susceptible to type 2 and type 3 poliovirus infection, and in principle, could fall ill with a second episode of poliomyelitis.

8.2 Inactivated Poliovirus Vaccine

The inactivated poliovirus vaccine (IPV), also referred to as the killed poliovirus vaccine (KPV), was developed in the 1950s by Jonas Salk and his colleagues in the USA. The original Salk vaccine contained representatives of all three poliovirus strains, wild neurovirulent strains PV1/Mahoney, PV2/MEF₁, and PV3/Saukett, inactivated by a low concentration of formalin. Protective efficacy of the vaccine was demonstrated in large field studies (USA, Canada, Finland), and from 1957, several European countries started to use the vaccine for the immunization of children

and older age cohorts in the population. A few countries (Sweden, Finland, Norway, Iceland, The Netherlands) capable of reaching high vaccination coverage succeeded in eliminating poliomyelitis using this vaccine, and continued to use IPV exclusively. With sub-optimal coverage levels, however, poliovirus transmission and outbreaks continued, although at highly reduced levels. Most European countries subsequently switched to the use of the oral, attenuated poliovirus vaccine when it became available.

New techniques for virus propagation, virion purification, and vaccine manufacturing were worked out in the early 1980s at the Dutch Institute for Public Health and Environment (RIVM) by Anton van Wezel and coworkers. Together with Jonas Salk, they demonstrated that two or three doses of the new “enhanced potency” IPV suffice to induce long lasting immunity in all three poliovirus serotypes. All currently available IPV preparations are based on these principles.

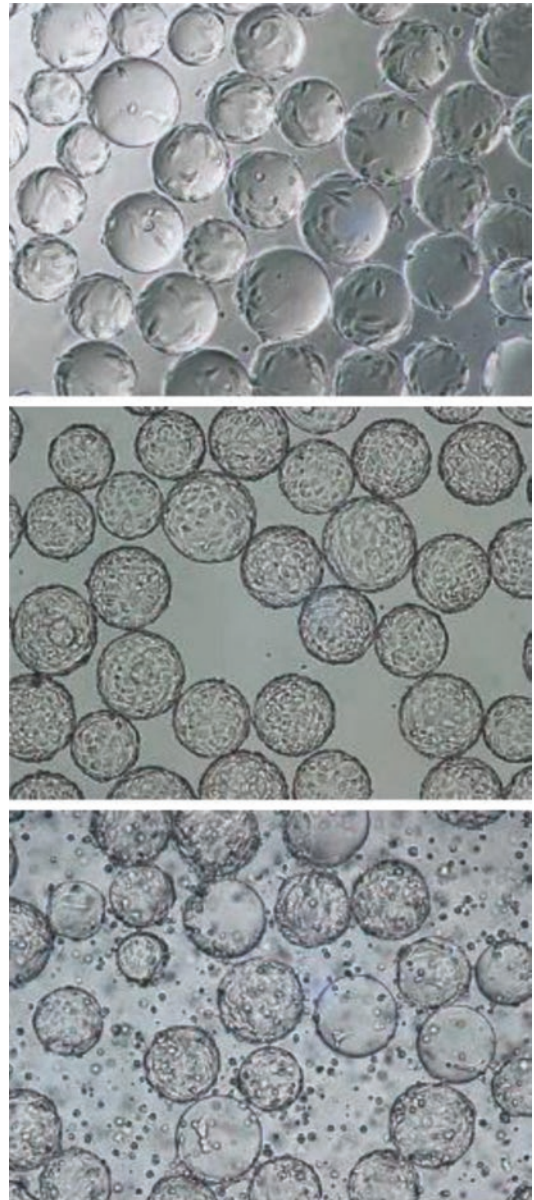
Antigenic stability of the vaccine is guaranteed by using only a standard, limited number of virus passages to create the infectious seed for individual vaccine bulk production. The cell substrate for virus propagation is a well-characterized Vero cell subline or the diploid human embryonic lung cell line MRC-5. Growth of the cells on microcarriers enables large-scale fermenter-based manufacturing of the vaccine (■ Fig. 8.2). After infection, the virus is purified from the supernate by gel filtration and ion exchange chromatography, and is inactivated by careful incubation in 3 mM formaldehyde. Standardized procedures are important because it is known that the antigenic phenotype of purified poliovirus may be changed from the neutralizing antibody-inducing D-type to the non-inducing C-type. Antigenicity of IPV is expressed in so-called D-units (DU). Typically, trivalent IPV preparations contain uneven amounts of the three serotypes, the original van Wezel version 40:8:32 DU of poliovirus types 1, 2, and 3. These serotype ratios were selected for their optimally balanced immune response toward all three serotypes. IPV-only vaccine preparations do not contain any adjuvant, but all the currently used pediatric combination vaccines contain aluminum adjuvant.

Seronegative infants seroconvert rapidly to all three poliovirus serotypes after two injections of IPV with an interval of 1 month or more between the doses. Additional doses further increase the antibody concentration in circulation. The formalin inactivation of polioviruses is known to destroy some of the several antigenic determinants involved in the induction of neutralizing antibodies. Yet, IPV-induced immunity gives full protection against paralytic poliomyelitis. Inactivated poliovirus vaccine alone does not induce significant intestinal IgA response and thus is considered inferior to the oral poliovirus vaccine in inducing protection against intestinal reinfection and in creating herd immunity in human populations. IPV injections to previously OPV-immunized individuals strongly boost the intestinal immunity.

Inactivated poliovirus-induced circulating antibodies can also prevent the post-viremic secondary replication of the virus in the oropharynx, and thus interfere with the further spread of infections in the population. Oropharyngeal shedding of poliovirus is likely to play a major role in poliovirus transmission under the Western-style hygienic conditions where the classical feco-oral

transmission route is partly blocked by well-organized sanitary systems.

The IPV-only vaccine preparations can be administered using intramuscular or subcutaneous injections, whereas the pediatric combination vaccines containing IPV are recommended for intramuscular use only. Adverse effects due to



■ Fig. 8.2 Growth of Vero cells on microcarrier particles and the effect of poliovirus infection on cells. *Upper panel:* cells soon after; *middle panel:* cells grown to confluence a couple of days before virus inoculation; *lower panel:* cytopathic effect of poliovirus before harvest (captured from a GE brochure of cell culture equipment)

IPV administration are rare and, if they do occur, are limited to common inconveniences and local reactions at the injection site.

After elimination of wild-type polioviruses in Europe, OPV-using countries have returned one by one to the IPV-only immunization programs using the new IPV. France made this switch during the 1990s and the UK in 2004. At present, all Western European countries, except Portugal and Malta, use IPV only in primary immunizations.

Although several European vaccine manufacturers make IPV, most of the IPV used is given as the pediatric combination vaccines, and the availability of IPV-only preparations needed for optional boosters for individuals exposed to poliovirus has occasionally been limited. The explosive importation of refugees to Europe in 2015 and the global switch from oral poliovirus vaccine (OPV) to IPV caused further availability problems.

► https://www.vaccineshoppecanada.com/document.cfm?file=IMOVAX_Polio_E.pdf

8.3 Oral Poliovirus Vaccine

Selected isolates from each of the three serotypes of poliovirus were serially passaged in monkeys or in cell cultures, and the desired attenuation was monitored by designated neurovirulence tests in monkeys *in vivo*. Out of the few candidates, a set of strains developed by Albert Sabin and colleagues was finally chosen for wider studies on efficacy and safety in infants. The strains are pragmatically referred to as PV1/Sabin, PV2/Sabin, and PV3/Sabin, or Sabin 1, Sabin 2, and Sabin 3 respectively. The largest field study was carried out in the Soviet Union (including Estonia, Latvia, and Lithuania) during the late 1950s and early 1960s. The trivalent OPV was shown to be highly effective and the frequency of harmful effects (vaccine-associated polio) was considered acceptable compared with the threat of the devastating disease. The vaccine was relatively inexpensive and oral administration did not require specially trained health care personnel. Hence, most national immunization programs rapidly adopted OPV for use in all infants.

The relative proportions of poliovirus serotypes in the vaccine formulation were found to be important to guarantee seroconversion to all three serotypes. Typically, a dose of OPV included 10^6 cell culture infectious units (CCU₅₀) of PV1/Sabin, 10^5 CCU₅₀ PV2/Sabin, and 3×10^6 CCU₅₀ of PV3/

Sabin (10:1:3 ratio). Replication of the attenuated polioviruses in the epithelia and submucosa of the intestines results in shedding of the virus into stools and induction of both circulating neutralizing antibodies and local (intestinal) IgA antibodies. OPV-derived polioviruses are known to spread from the primary vaccinees to close contacts, a feature initially considered beneficial to improving the nominal coverage of the vaccination. The local immune response is considered to be crucial for resistance to intestinal reinfection and for the herd immunity in immunized populations.

Wider use of OPV in the early 1960s soon revealed that vaccine-associated paralytic poliomyelitis (VAPP) may occur in the vaccinee or in a contact. Furthermore, at a low frequency, a persistent infection may be established by one or more of the OPV components in persons with a defect in the humoral immune system. In such persons, paralytic symptoms often emerge only years after receiving the vaccine. The frequency of VAPP is about one case in 700,000 primary vaccinations. Earlier vaccine doses, either OPV or IPV, decrease the risk. In Denmark, this observation was exploited by establishing a safe combination schedule for polio immunizations, starting with three doses of IPV and followed by three doses of OPV.

Most VAPP patients shed either type 2 or type 3 poliovirus related to the corresponding OPV component. Sabin 2 virus is genetically close to the wild parental virus. Sabin 3 shows only ten point mutations, differentiating it from the parental PV3/Leon strain, whereas Sabin 1 has 57 single nucleotide differences compared with the parental PV1/Mahoney strain. During a few days of replication of the OPV-derived viruses in the human body, the viruses are readapted to human tissues, lose many of the attenuating mutations, and revert to neurovirulence. Although VAPP was initially accepted as the price of an inexpensive and effective immunization program, it later became intolerable in the absence of wild poliovirus transmission and with decreasing risks of importation of wild polioviruses. Thus, one after another, European countries stopped using OPV in routine immunizations and switched to programs using various pediatric combination vaccines including IPV components.

The shift of the millennium marked a significant change in the safety consideration of OPV. An outbreak of paralytic poliomyelitis emerged on the Caribbean island of Hispaniola in 2000 and

the patients turned out to be shedding type 1 poliovirus closely related to, but significantly different from the OPV component Sabin 1. The capsid protein VP1 coding sequences differed by 2–3% from those of Sabin 1. Epidemiological analysis of the outbreak indicated that it had started in Haiti, where the routine immunization coverage had been suboptimal for years, and subsequently spread to the neighboring country of the Dominican Republic. The outbreak was contained by an aggressive immunization campaign with trivalent OPV.

The Hispaniola episode, and several similar ones subsequently discovered in different parts of the world where OPV is routinely used have led to the following updated view on the safety of OPV: in populations with low vaccine coverage, OPV-derived polioviruses of any serotype may circulate, and during circulation accumulate point mutations throughout the genome, including sites responsible for attenuation. Circulating vaccine-derived polioviruses (cVDPV) revert into neurovirulence and may cause outbreaks of paralytic poliomyelitis. Thus, they behave like wild polioviruses. By 17 August 2016, a total of 809 cases of cVDPV-induced paralytic poliomyelitis had been reported to the World Health Organization (WHO). A need to rapidly cease use of OPV has emerged.

Wild-type 2 poliovirus has not been isolated from human specimens anywhere in the world since 1999 and was declared eradicated in 2015. Because of the risk of cVDPV caused by the Sabin 2 strain in the traditional trivalent OPV, a WHO-coordinated global switch in polio immunizations took place in April 2016: all countries should have incorporated at least one dose of IPV in the routine immunization program of infants and those countries still using OPV must switch from the trivalent to a bivalent Sabin 1 + Sabin 3 vaccine.

► http://www.epid.gov.lk/web/images/pdf/Polio/switch_plan_sri%20lanka_updated_nov%202015.pdf.

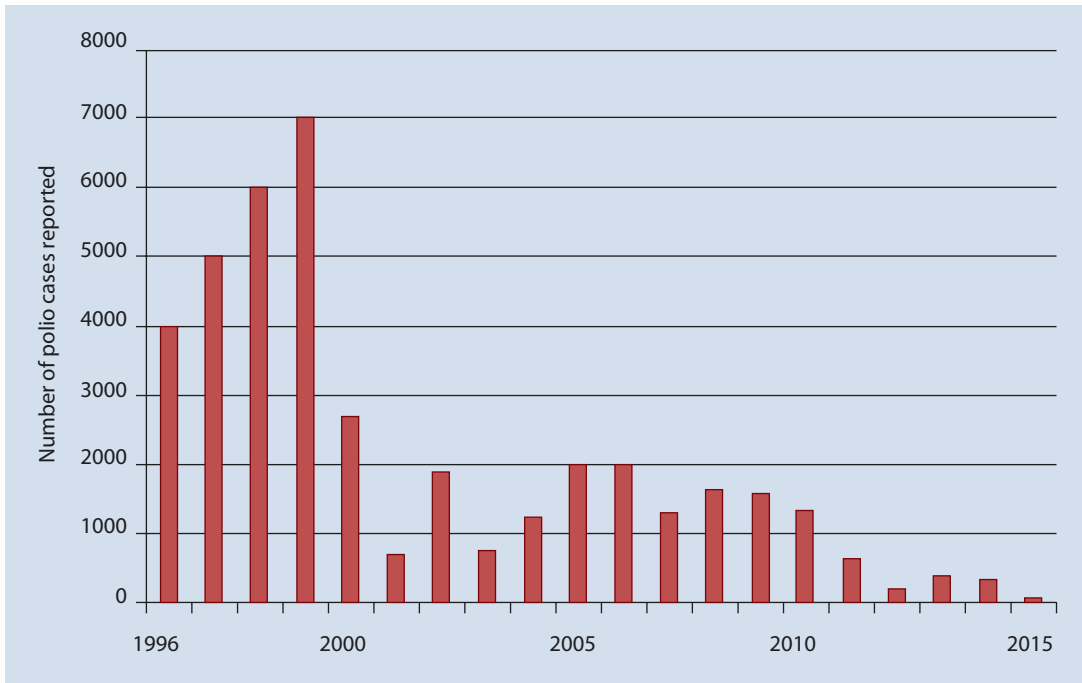
8.4 Global Poliovirus Eradication Initiative

In the 1970s, the WHO had incorporated OPV into the six-disease-target Expanded Programme of Immunization recommended to all infants

worldwide. The coverage of the age-based immunization remained low in developing countries. In the Americas, the Pan-American Health Organization started to supplement the routine vaccination with annual OPV campaigns, so-called National Immunization Days (NID) during which all children younger than 5 years received a dose of OPV irrespective of previous immunization history. This principle had been successfully used in Cuba since the early 1960s. By the mid-1980s, the success of these campaigns in Latin American countries was so good that a desire for the global eradication of poliomyelitis emerged.

In 1988, the World Health Assembly accepted the resolution *WHA41.28 Global eradication of poliomyelitis by the year 2000*. The subsequently created program, the Global Polio Eradication Initiative (GPEI) is spearheaded by the WHO, the Centers for Disease Control and Prevention (CDC) of USA, UNICEF, and Rotary International, and includes guidelines for intensified immunization, standards for surveillance, a supporting worldwide laboratory network, and centralized reporting. In addition to routine infant immunizations, NIDs and other modes of supplementary immunization were recommended to guarantee maximal vaccination coverage. In surveillance, the starting point was a suspected case, a patient with acute flaccid paralysis (AFP). Both local health care personnel and ad hoc trained lay “reporters” were supposed to notify these cases to designated epidemiologists who examined the cases, collected stools for virus isolation, and reported the results to a national epidemiological center (NEC). Each country had a nominated national polio laboratory (NPL), which carried out the stool examination and sent possible poliovirus isolates forward to designated polio reference laboratories for further characterization. Both NECs and NPLs reported their results to WHO regional centers and the latter reported to the WHO Headquarters (HQ) in Geneva, Switzerland.

The original target of the GPEI was not reached, but the initial progress was dramatic: starting from an estimated number of 350,000 new cases in 1988, already 10 years of the program reduced the number of annual cases by more than 99% and drastically limited the number of countries with persisting wild poliovirus circulation. Since then, however, various factors have delayed the completion of the desired eradication (■ Fig. 8.3). These include – apart from



■ **Fig. 8.3** Decrease in reported polio cases over the last 20 years. Column heights indicate the annual number of polio cases globally reported to the World Health Organization (Source: GPEI/WHO)

the limited availability of resources – deteriorated national infrastructure and health care systems in some countries, civil wars, and political or religious intrigues that interfered with the systematic immunization of infants and thus reducing herd immunity to poliovirus transmission. These factors, combined with the inherent property of polioviruses for the long asymptomatic carrier state in humans, and increasing international and intercontinental travel, have resulted in surprising outbreaks in countries that have already had years of polio-free history. Yet, in the longer run, the number of countries and districts have progressively decreased over the years. This has also resulted in narrowing of the genetic diversity of the wild polioviruses that are still circulating, which is important, because it has finally allowed the use of RT-PCR tests in primary diagnostic procedures, because the decreased diversity has allowed the development of poliovirus-specific primers. (Previously, genetically closely related nonpolio enteroviruses common throughout the world had confounded direct detection, and the time- and resource-consuming cell culture isolation had to be used.)

8.5 Future Prospects

The global polio eradication initiative has been the broadest international health care action ever performed. In 2016, it was closer to the target than ever. Only 74 cases caused by wild poliovirus (WPV) were reported in 2015 worldwide, all of them by type 1 and all of them from the two remaining wild poliovirus-endemic countries (i.e., countries in which continuous wild poliovirus transmission never stopped), Afghanistan and Pakistan. Type 2 WPV had been eradicated before 2000, and a period of 3.5 years without any evidence for type 3 WPV circulation in humans suggests that only type 1 WPV might remain. Unfortunately, reaching and maintaining high coverage polio immunization in the two remaining endemic countries has turned out to be problematic because of difficult to reach remote villages that are harboring the virus, and hostile attacks toward the vaccinators.

From a European point of view, maintaining high-coverage polio immunizations with IPV remains very important even after the desired eradication of WPV. As mentioned above, the immunity obtained with IPV protects individuals

against the paralytic disease, but does not provide the population with a strong herd immunity. Various factors pose a risk of the putative return of epidemic poliomyelitis to the future IPV-immunized world.

8.5.1 Missed Circulation of WPV in Remote Populations

This risk is not, in principle, very high, as small remote populations cannot support the transmission of WPV for very long. The quality of AFP surveillance has remained high, especially in Africa and in the Eastern Mediterranean Region of the WHO, where the last WPV-endemic countries are located. Unfortunately, even a small risk may sometimes be realized, as shown, in summer 2016, by the discovery of wild-type 1 poliovirus in two paralytic children in Nigeria after 2 years of apparent absence of the virus from the country, as judged by negative results of large number of tested clinical and environmental specimens.

8.5.2 Circulating VDPVs

Outbreaks of paralytic disease caused by cVDPV have been stopped by active immunization campaigns with the corresponding monovalent OPV. Stocks of monovalent vaccines for all three serotypes are maintained by the WHO. So far, cVDPV outbreaks have not spread from the original OPV-immunized population to IPV-using neighboring countries, but this possibility cannot be excluded. This risk decreases over time when one serotype after the other is removed from the OPV vaccine used in routine immunizations.

8.5.3 Long-Term Shedding of Vaccine-Derived Poliovirus by Immune-Deficient Individuals

These individuals are rare, and stool surveys carried out on immune-deficient (ID) patients known to the health care system in several countries suggest that only a small fraction of ID patients presents with persistent shedding of the ID-type vaccine-derived polioviruses (iVDPV). So far, no outbreak caused by iVDPV has been described, even though the viruses are neuroviru-

lent. On the other hand, several environmental poliovirus isolates share the distinct genetic features of iVDPV. The isolates have been found in different countries in the absence of known poliovirus-shedding ID patients in the region. Thus, not all individuals with a possibly mild ID but enabling persistent poliovirus infection are obviously known to the health care systems. Hence, the risk of polio return from long-term iVDPV-shedding is difficult to estimate.

8.5.4 Escape of WPV or cVDPV from a Laboratory or Vaccine Plant

Handling of polioviruses and poliovirus-containing specimens in the WHO poliovirus laboratory network and among the vaccine manufacturers follows good laboratory practice, taking into account strict biosecurity principles. However, humans can make mistakes, and for instance, the transport of wild poliovirus from an IPV manufacturing plant to the community in Europe has been reported. Therefore, attempts to replace the WPV strains in IPV production with the attenuated Sabin strains or with genetically modified avirulent derivatives of the current IPV strains are being pursued. In general, a so-called “containment project” of the WHO includes limiting the future poliovirus-handling laboratories to a small number of “essential poliovirus facilities,” following strict rules when handling the specimens, and advising member countries to destroy all unnecessary poliovirus stocks and poliovirus-containing specimens.

8.5.5 Poliovirus from the Melting Permafrost?

Together with ongoing global warming, the northern permafrost is known to be slowly melting. Fortunately, the corresponding latitudes have been sparsely populated for millennia and there is no direct evidence that polioviruses would be present in the earth of the now melting regions. Thus, this is a theoretical risk. However, it is known that the permafrost can store infectious viruses for tens of thousands of years, and this possibility should be kept in mind.

Further Reading

- Chumakov MP, Voroshilova MK, Drozdov SG, et al. Some results of the work on mass immunization in the Soviet Union with live poliovirus vaccine prepared from Sabin strains. *Bull World Health Organ.* 1961;25:79–91.
- Enders JF, Weller TH, Robbins FC. Cultivation of the Lansing strain of poliomyelitis virus in cultures of various human embryonic tissues. *Science.* 1949;109:85–7.
- Heymann D, Ahmed Q. The polio eradication end game: what it means for Europe. *Euro Surveill.* 2014;19:20702.
- Minor P. Vaccine-derived poliovirus (VDPV): impact on poliomyelitis eradication. *Vaccine.* 2009;27:2649–52.
- Sabin AB. Oral poliovirus vaccine: history of its development and use and current challenge to eliminate poliomyelitis from the world. *J Infect Dis.* 1985;151:420–36.
- Salk JE. Poliomyelitis vaccine in the fall of 1955. *Am J Public Health Nations Health.* 1956;46:1–14.
- Salk D, van Wezel AL, Salk J. Induction of long-term immunity to paralytic poliomyelitis by use of non-infectious vaccine. *Lancet.* 1984;2:1317–21.
- Thomassen YE, et al. Next generation inactivated polio vaccine manufacturing to support post polio-eradication biosafety goals. *PLoS One.* 2013;8(12):e83374.

Further Reading

- <http://www.polioeradication.org>.

Measles–Mumps–Rubella Vaccine

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The measles–mumps–rubella (MMR) vaccine is currently the exclusive tool for the prevention of measles, mumps, and rubella in Europe. The description below of single measles, mumps, and rubella vaccines is given for historical reasons and for the characterization of their properties as components of the MMR vaccine. Previous immunization programs with single vaccines, usually given as a single dose, were generally less effective than the current MMR immunization programs in early childhood, with the vaccine given in two doses.

A full description of measles, mumps, and rubella diseases is beyond the scope of this chapter. In the following, the description of the manifestations and the burden of these diseases is limited to make the argument in favor of immunization. With the success of the MMR vaccination programs in some European countries, the respective diseases are seldom seen and the rationale for vaccination may be forgotten or not appreciated by the public and health care personnel. In many European countries outbreaks of measles and rubella continue to occur.

9.1 Measles and Measles Vaccine

Measles is a systemic viral infection transmitted via airborne droplets and characterized by respiratory symptoms and rash. Common complications are pneumonia, otitis, diarrhea. Measles is sometimes regarded as an ordinary childhood disease that children should preferably experience to “strengthen” their immune system. However, it is important to remember that measles is a serious and potentially fatal disease, with 2–8% mortality in developing countries. Historically, measles carried a significant risk for mortality in Europe as well; even in the recent outbreaks of measles in Europe, there have been fatalities.

The development of a vaccine against measles became possible after isolation of the measles virus by Enders and Peebles in 1954. A nonlive measles vaccine was developed and used for a few years in the USA in the 1960s. The vaccine was withdrawn because some vaccinated children upon exposure to measles developed atypical and severe forms of measles.

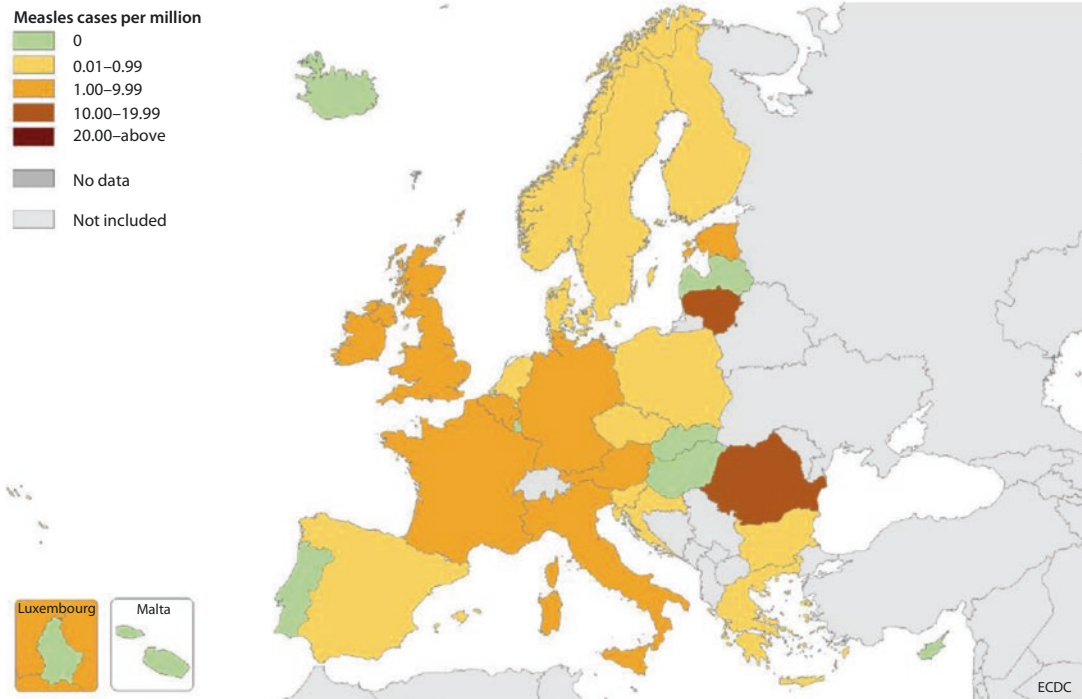
Isolation of the measles virus paved the way to attenuation and live vaccine development. The first vaccine strain was called Edmonston after the

boy from whom the virus was isolated. Most measles vaccines in the world and all those in Europe are derived from the Edmonston isolate. The two currently available vaccine strains, Schwarz (Edmonston A) and Moraten (Edmonston B), represent two different cell culture passage branches of the original, but are at practically the same attenuation level. Several studies have addressed possible differences in the immunogenicity and safety profile of these vaccines. The level of attenuation of the measles vaccine is a carefully chosen balance between sufficient immunogenicity and minimal (although still substantial) reactogenicity.

A less attenuated version of Edmonston B strain, Edmonston–Zagreb (E–Z), was developed and used in the former Yugoslavia. The E–Z strain was cultured in WI-38 human fibroblast cells. The vaccine was regarded as a more potent (than the current one) measles vaccine that could be given at the of age 4–6 months in the presence of maternal antibody, thereby contributing to measles elimination efforts in developing countries. However, it was found that this measles vaccine, initially endorsed by the WHO, was associated with increased all-cause mortality in girls, and the approach was withdrawn. Subsequently, widespread use of the present measles vaccines has shown that a more potent vaccine is not needed, but measles can be eliminated with the currently available vaccines if used extensively.

A Russian measles vaccine, Leningrad 16 strain, which was not derived from Edmonston, was given on a large scale in Eastern European countries over many decades. Therefore, measles immunity in adults in those countries is reliant on vaccination with Leningrad 16, which is no longer used in Europe.

In the 1970s, when measles vaccination was being introduced into Europe, mortality from measles was already low. Major arguments for the introduction of vaccine included the prevention of complications, notably meningoencephalitis, which occurs at a rate of 1:1,000–1:2,000 and may leave permanent sequelae. Another measles-related problem is subacute sclerosing panencephalitis (SSPE), which occurs several years after measles at an early age at a rate of 1:100,000 and is invariably fatal; preventing SSPE is an important goal of measles vaccination. Less serious complications such as pneumonia and otitis media are very common after measles. All the complications



■ Fig. 9.1 Countries reporting cases of measles in 2015–2016

combined make an argument in favor of measles vaccination in Europe.

Still, these arguments regarding measles vaccination were not compelling enough at that time to convince all physicians and other health care workers, and the coverage of single measles vaccination until the introduction of MMR vaccination programs remained at 60–70% in many Western European countries. This level of immunization reduced the epidemics, but postponed the acquisition of measles to adolescent age, resulting in many cases of serious disease and even deaths in young people. In contrast, in many Eastern European countries with mandatory measles vaccination programs, measles was virtually eliminated. In Western European countries, the elimination of measles only started with the introduction of two-dose programs of MMR vaccine.

As measles is targeted for eradication by the WHO, Europe will have to do its share in the process, which adds another compelling reason for measles immunization. Globally, measles-associated deaths have decreased from about 1.5 million a year to 134,200 in 2015, owing to vaccinations. As measles is highly infectious, over 95% vaccine coverage is needed for control, and only a very high global coverage can

result in eradication. Elimination of indigenous measles in Latin America is a strong indication that it can be done, and Europe should be able to accomplish the same. The WHO strategic plan is to eliminate measles in at least five WHO regions.

Cases of measles and rubella continue to occur sporadically and in outbreaks in European countries (■ Fig. 9.1). In 2011, there were 30,000 reports of measles to the European Centre for Disease Prevention and Control; the true number is probably higher. In the last 1-year period, there have been 1,818 reported cases of measles (■ Fig. 9.1).

9.2 Mumps and Mumps Vaccine

Mumps is a generalized viral infection transmitted via airborne droplets or direct contact with infected saliva. Transmission depends on the close contact and increases in overcrowded conditions. The classical manifestation is the swelling of one or both parotid glands. Other manifestations of mumps may be viral meningitis, encephalitis, pancreatitis, mastitis, orchitis, and arthropathy. Orchitis in post-pubertal males may result in sterility.

The case for mumps vaccination is less compelling than for measles and rubella, but still strong enough to justify the inclusion of the mumps component in the MMR vaccine. Since the introduction of vaccination, the incidence of mumps has decreased dramatically. In pre-vaccine era mumps was characterized by 4- to 5-year epidemic cycles. Natural mumps infection is thought to confer lifelong protection. Mumps is typically a mild childhood disease that begins with nonspecific symptoms followed by a unilateral or bilateral swelling of the parotid glands. Meninges, pancreas, and testes are other targets. An illustrative example of the clinical course and significance of mumps comes from a naïve population on St. Lawrence Island where an epidemic of mumps resulted in clinical disease in 65% and subclinical infection in 35%; of those with clinical mumps, 11% had meningitis and 25% of post-pubertal men had orchitis. Prevention of such complications is the reason for mumps vaccination.

In the past, inactivated mumps vaccines were developed and used in targeted populations such as the military in Finland. The protection conferred by the inactivated mumps vaccine against orchitis was good, but not as durable as that induced by a live mumps vaccine. In contrast to measles, no atypical forms of mumps have been reported in the recipients of a killed vaccine.

A live attenuated mumps vaccine was developed by the serial passaging in chicken embryo of fibroblast cells. The vaccine strain is called Jeryl Lynn, according to the patient from whom the virus was isolated; the developer was Maurice Hilleman at Merck Research Laboratories. Only this one mumps vaccine strain survives in the current major MMR vaccines. The Jeryl Lynn strain barely causes any adverse reactions. However, the single mumps vaccine is no longer available. Present strategies to control mumps are closely integrated with existing goals of measles and rubella control or elimination, and the MMR vaccine is used as a common tool.

A Japanese mumps vaccine strain Urabe AM9 was incorporated in an early version of GSK's MMR vaccine, but was withdrawn as it was found to cause meningitis at a rate of 1 in 50,000 recipients. Afterward, GSK re-isolated a mumps vaccine virus from the Jeryl Lynn vaccine preparation, by choosing only one plaque variant of the two present in the original Jeryl Lynn. The isolate was

called RIT4385 and is now incorporated in GSK's MMR vaccine. Comparative studies of the RIT 4385 and Jeryl Lynn vaccines showed a high safety level and similar seroconversion rates.

The Leningrad-3 mumps vaccine strain was developed in the former Soviet Union. This strain was further attenuated in Croatia, named Leningrad-Zagreb, and used for vaccine production in Croatia and India. The Rubini strain was first licensed in Switzerland in 1985. However, substantially lower rates of seroconversion and effectiveness among recipients of Rubini strain vaccine compared with those vaccinated with Jeryl Lynn or Urabe Am9 strains were observed. Therefore, the WHO recommends that the Rubini strain vaccine should not be used in national immunization programs.

9.3 Rubella and Rubella Vaccine

Rubella is a systemic viral infection that is highly contagious. In children, rubella is characterized by a mild fever and a short-living rash. Rubella is a mild disease and may be unrecognized or misdiagnosed in young children. Furthermore, up to 50% of rubella infections may be subclinical. These cases are still contagious in contact with unvaccinated and non-immune pregnant women. If a non-immune pregnant woman gets infected, the rubella virus may be transmitted to the fetus and induce serious birth defects described as congenital rubella syndrome (CRS).

The association of rubella in early pregnancy with congenital cataract in the infant was described by Norman Gregg in Australia in 1941, but it was not until 1964 and a major epidemic of rubella in the USA that resulted in an estimated 20,000 babies with damage, that the disease was fully appreciated and vaccine development started. The rubella virus had been isolated just 2 years earlier independently by Weller and Neva, and by Parkman and co-workers in the USA. The case for rubella vaccination lies primarily in the prevention of CRS. Systematic vaccination against rubella, usually in combination with measles, has eliminated both the congenital and acquired infections from some industrialized countries and Latin America.

Although CRS is very rare today, it should be kept in mind as a motivation for vaccination. CRS is limited to cases of maternal rubella in the first

trimester of pregnancy, although cases of hearing loss may occur up to 16 weeks of pregnancy. In the first ≤ 11 weeks of pregnancy, the rubella virus crosses the placenta in 90% of cases, and results in clinical sequelae in almost all, even though the severity of CRS varies. The classical triad is heart–eye–ear. Cardiovascular anomalies typically include pulmonary stenosis and patent ductus arteriosus. Ocular manifestations include retinopathy, cataract, and glaucoma, and may result in blindness. Hearing loss is the most common single manifestation of CRS and may be bilateral or unilateral. In addition to isolated organ damage, the full-blown CRS includes generalized infection of the newborn, with enlarged liver and spleen, purpura, jaundice, and CNS involvement. After mid-pregnancy, the rubella virus may still be transmitted to the fetus in about 50% of the cases, but does not cause any clinical damage.

Several live attenuated rubella vaccines were developed and licensed after isolation of the rubella virus in the 1960s. Early licensed vaccines included the Cendehill strain grown in rabbit kidney cells and the HPV77 strain isolated in monkey kidney cells and grown in duck embryo fibroblasts. RA27/3 was discovered in 1969 and is the only strain that survives today, as all previously registered vaccines were less immunogenic and more reactogenic than RA27/3.

The RA27/3 strain was isolated from a rubella-related abortion and the virus was attenuated in WI38 human fibroblast cells. Therefore, the passage history is entirely “human”. RA27/3 is highly immunogenic and nearly a 100% seroconversion rate is reached with a single rubella vaccination or in the MMR combination. Adverse effects attributable to the rubella component in MMR vaccination in children are rare. A notable adverse event is thrombocytopenia, which may manifest in about 1:50,000 vaccine recipients. In adult vaccinees, the rubella vaccine may be associated with joint pain or even arthritis; however, these were much more common in association with the early rubella vaccines than with RA27/3.

Currently, the only existing strategy to prevent CRS is the elimination of rubella by vaccination of all infants and children with the MMR vaccine. Previously, the single rubella vaccine was used for targeted vaccination of women and girls. The target groups included women post-partum (after the birth of the first child) or pre-pubertal girls. Neither strategy ever reached a high coverage and

both were ineffective in the prevention of CRS. Moreover, an inadequate level of rubella immunization of adolescent girls increases the number of sero-susceptible women at childbearing age and enhances the risk of rubella in pregnancy. For example, CRS increased in Greece and Romania after outbreaks of rubella. Rubella is targeted for elimination in Europe, but continues to occur in many European countries.

9.4 Measles–Rubella Vaccine

Measles and rubella are targeted for global elimination/eradication, whereas mumps is not. Not all countries consider mumps a priority for vaccination and prefer to use the measles–rubella (MR) vaccine instead. Globally, an Indian-made MR vaccine is being used extensively (150 million doses distributed), but is not available in Europe.

9.5 Measles–Mumps–Rubella Vaccine

Live attenuated measles, mumps, and rubella vaccines were combined into the MMR vaccine. Merck’s MMR vaccine was first introduced in the USA in 1971, and the composition was changed to the current one in 1978 (MMRII[®]). The MMRII[®] vaccine contains the Moraten strain of the measles vaccine, the Jeryl Lynn strain of the mumps vaccine, and the RA27/3 strain of the rubella vaccine. In Europe, the same vaccine has been marketed as MMR VaxPro[®] (SP-MSD). The first licensed MMR vaccine contained the HPV77/DE5 strain of the rubella vaccine, but this was replaced in 1978 by RA27/3 to make the MMRII[®] vaccine.

Although MMR vaccinations were started in Europe later than in the USA, the practice of giving two doses of MMR was initiated in Sweden and Finland in 1982. The introduction of the two-dose MMR vaccination programs offered a new tool for the elimination of measles in Europe. The two-dose program was purely empirical, but had the following rationale: (1) filling the immunogenicity gap in those who may remain susceptible after the first dose; (2) A booster effect in a proportion of the children who have taken the first dose; (3) single-dose MMR vaccination policy inevitably misses a certain proportion of infants and a second dose



■ Fig. 9.2 Measles-mumps-rubella vaccination schedules in Europe. European Centre for Disease Prevention and Control (ECDC) ▶ <http://vaccine-schedule.ecdc.europa.eu/Pages/Scheduler.aspx>

catches those individuals who have not received their primary MMR dose. Subsequently, it was realized that an important mechanism by which a second dose of MMR vaccine enhances protection (at least against measles) is the increased avidity of IgG antibodies. In Sweden, the second dose of MMR was given at the age of 12 years with the idea of maximizing protection against mumps and rubella just before puberty; this practice remains in some countries (■ Fig. 9.2). In Finland, the second dose is given at 6 years of age. At present, the timing of the second dose varies greatly from country to country (■ Fig. 9.2). With the two-dose program, Finland became the first country to eliminate indigenous measles, mumps, and rubella by 1994. To reach this target, the coverage had to be over 95% for the two doses.

In some countries with a lower coverage, but with the intention of eliminating measles, a much shorter interval between the two doses of MMR vaccine is being recommended. The practice of giving the doses of MMR at a short, even only 3-month interval was started in Germany and has spread to other European countries. Currently, there are a multitude of MMR vaccination schedules in Europe (■ Fig. 9.2). The “short” interval schedules are specifically targeted at the elimination of measles, whereas the “long” interval schedules target boosting the immunity for durable protection into adolescence and beyond. There is no longer any justification for a single-dose policy.

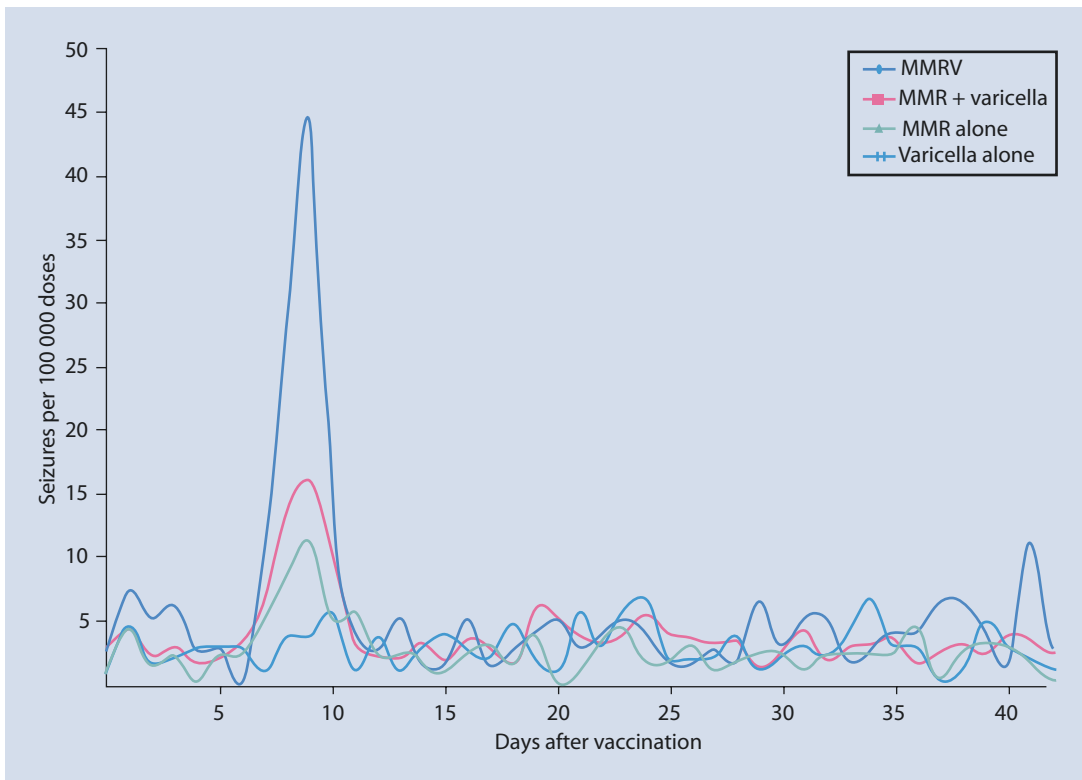
The MMR vaccine manufactured by GSK originally contained the Urabe AM9 strain of the mumps vaccine and was withdrawn in 1986. GSK

replaced the mumps component with a one-plaque variant of the Jeryl Lynn strain called RIT 4385. Thus, the present vaccine, Priorix®, contains the Schwarz strain of measles, the RIT 4385 strain of mumps, and the RA27/3 strain of rubella. Although the measles component is comparable with and the rubella component the same as in MMR-II, the mumps component was initially less immunogenic. However, with dose adjustments, the immunogenicity of RIT 4385 has been improved, and in general, the two vaccines are quite similar with regard to immunogenicity. In European countries, the two MMR vaccines may be used interchangeably. Both vaccines are safe and effective for the prevention of measles, mumps, and rubella/CRS.

Numerous clinical studies performed before the registration demonstrated the safety of currently used MMR vaccines. This safety is also confirmed in post-marketing surveillance. The most significant adverse reaction following MMR administration is fever. The timing of the fever is

characteristic and occurs 7–10 days after vaccination (■ Fig. 9.3). A rash may appear during the second week after vaccination, but other adverse events are very rare. No cases of viral meningitis have been reported after MMR vaccines currently used in Europe.

In 1998, Wakefield et al. published a paper in *The Lancet* reporting that MMR vaccination, but not the single measles vaccine, was associated with autism. The paper was later retracted by *The Lancet*. Still, the so-called “Wakefield hypothesis” endured. In 2001, a review by the Institute of Medicine (USA) concluded that the evidence rejected a causal relationship at the population level between the MMR vaccine and autistic spectrum disorder (ASD). The hypothesis has been tested in a number of observational and epidemiological studies and the main conclusion of the studies is that the MMR vaccination is not associated with an increased risk of ASDs.



■ Fig. 9.3 Measles–mumps–rubella–varicella combination vaccine and the risk of febrile seizures (Reproduced with permission. Klein et al. 2010)

9.6 MMR Vaccination of Special Groups

In exceptional cases, the MMR vaccine can be given to 6- to 9-month-old infants if they are at a high risk of becoming infected, e.g., during a measles outbreak or travel. These children may not respond to this early dose because of residual maternal antibodies; therefore, they need the two standard MMR doses, first at 12–13 months of age and a second dose later.

The MMR vaccine is recommended to non-pregnant women of child-bearing age if they have no proof of immunity against rubella or if this information is not available. It cannot be given to pregnant women, but it can be given after delivery. However, in cases of inadvertent vaccination during pregnancy there is no need to take any measures, as transmission of the rubella vaccine virus to the fetus is rare and is not known to cause clinical harm to the fetus.

If vaccination against measles, mumps or rubella is needed outside the current childhood immunization programs, the MMR vaccine should be given, rather than any of the single components. There is no harm in administering “extra” doses of MMR to previously immunized persons. In any case, single measles, mumps or rubella vaccines are no longer available in Europe.

The MMR vaccine cannot be given to persons who have had an anaphylactic reaction to a previous dose of the MMR vaccine or a component of it. MMR is a live virus vaccine and it should not be given to persons with an impaired immune response, e.g., those treated with high-dose steroids, those treated for cancer with chemotherapy or radiotherapy, or those who are on immunosuppressive drugs after an organ transplant.

9.7 Measles–Mumps–Rubella–Varicella Vaccine

The MMRV vaccine is also described in the ► Chap. 10. The main rationale of a combined MMRV vaccine is obviously the easier administration, with one injection only, of both MMR and varicella vaccines. Although the development of a combined MMRV vaccine began in the 1980s, it took almost two decades to get the vaccine licensed. The reason was twofold: (1) if a standard dose of varicella vaccine was combined with

MMR, it was not sufficiently immunogenic, and (2) if the titer of the varicella component was increased for sufficient immunogenicity, it also increased the reactogenicity of the combination in comparison with MMR. The current licensed MMRV vaccines represent a compromise and balance between these two issues.

The immunogenicity of MMRV for varicella zoster virus is clearly higher than that of a single varicella vaccine (■ Table 10.2). The immunogenicity for measles, mumps, and rubella may also be slightly higher, but the difference is not critical and not an argument in favor of the use of MMRV instead of MMR. Altogether, for the sake of the elimination of measles and rubella and for the control of mumps, there is no reason to use MMRV instead of MMR vaccine. Rather, MMRV should be seen as a tool for varicella vaccination in countries that have achieved good control of the MMR diseases.

The reactogenicity of the MMRV vaccine after primary vaccination has become a significant issue (■ Fig. 9.3). The fever rate is more than double that of MMR (and varicella given separately in a different arm at the same time) and the risk of febrile convulsions increases in the same proportion.

After the licensure of the MMRV vaccine in 2006, the ACIP in the USA recommended that it be the choice for varicella vaccination. However, because of the issue of febrile convulsions, the recommendation was changed to no-recommendation, i.e., the physician could choose between MMRV and separate MMR and varicella vaccination. In practice, the separate administration of MMR and varicella vaccines has become more common for primary vaccination, whereas for the second dose, MMRV is often chosen for convenience. Fever and febrile convulsions are not an issue for the booster vaccination, no matter how long the interval is between the first and second doses.

In Europe, febrile convulsions are not regarded as the same kind of problem as in the USA and MMRV is given for the primary vaccination in parts of Italy (Lombardy, Sicily) that have implemented varicella vaccination, and in Germany and Greece. In any case, febrile reactions can be anticipated because of the well-known timing, and anti-febrile medication can be initiated to prevent seizures.

Two licensed MMRV vaccines are available in Europe: Priorix-Tetra® (GSK) and Pro-Quad (Merck). The descriptions of the vaccines are shown in ■ Table 9.1.

Table 9.1 Descriptions of vaccines

Component	Priorix-Tetra	ProQuad
Measles	Schwarz strain	Enders' Edmonston
Mumps	RIT 4385 strain, derived from Jeryl Lynn strain	Jeryl Lynn™ (Level B)
Rubella	Wistar RA 27/3 strain	Wistar RA 27/3 strain
Varicella	Oka strain	Oka/Merck strain

Further Reading

- Bolotovskii V, et al. Immunization of 6 and 9 month old infants with AIK-C, Edmonston-Zagreb, Leningrad-16 and Schwarz strains of measles vaccine. *Int J Epidemiol*. 1994;23(5):1069–77.
- Bottiger M, et al. Swedish experience of two dose vaccination programme aiming at eliminating measles, mumps and rubella. *BMJ*. 1987;295:1264–7.
- Davis NF, et al. The increasing incidence of mumps orchitis: a comprehensive review. *BJU Int*. 2010;105(8):1060–5.
- Edmunds WJ, et al. The pre-vaccination epidemiology of measles, mumps and rubella in Europe: implications for modelling studies. *Epidemiol Infect*. 2000;125(3):635–50.
- Enders J. Measles virus. Historical review, isolation, and behavior in various systems. *Am J Dis Child*. 1962;103:282–7.
- Enders JF, Peebles TC. Propagation in tissue cultures of cytopathic agents from patients with measles. *Proc Soc Exp Biol Med*. 1954;86:277–86.
- Garenne M, et al. Child mortality after high-titre measles vaccines: prospective study in Senegal. *Lancet*. 1991;338(8772):903–7.
- Ikić DM. Edmonston-Zagreb strain of measles vaccine: epidemiologic evaluation in Yugoslavia. *Rev Infect Dis*. 1983;5(3):558–63.
- Immunization Safety Review Committee, Immunization safety review. Measles and autism. 2004, Washington, DC: National Academies Press. p. 214.
- Klein N, Fireman B, Yih K, et al. Measles-mumps-rubella-varicella combination vaccine and the risk of febrile seizures. *Pediatrics* 2010;126(1):e1–8.
- Lievano F, et al. Measles, mumps, and rubella virus vaccine (M–M–R™II): a review of 32 years of clinical and post-marketing experience. *Vaccine*. 2012;30(48):6918–26.
- Mercader S, Garcia P, Bellini WJ. Measles virus IgG avidity assay for use in classification of measles vaccine failure in measles elimination settings. *Clin Vaccine Immunol* 2012;19:1810–1817.
- Peltola H, et al. The elimination of indigenous measles, mumps, and rubella from Finland by a 12-year, two-dose vaccination program. *N Engl J Med*. 1994;331(21):1397–402.
- Peltola H, et al. Measles, mumps, and rubella in Finland: 25 years of a nationwide elimination programme. *Lancet Infect Dis*. 2008;8:796–803.
- Plotkin SA. The history of rubella and rubella vaccination leading to elimination. *Clin Infect Dis*. 2006;43(Suppl 3):S164–8.
- Plotkin S, Mortimer E. *Vaccines*. 2nd ed. Philadelphia, London, Toronto, Montreal, Sydney, Tokyo: W.B. Saunders Company; 1994.
- Plotkin SA, et al. Attenuation of RA27/3 rubella virus in WI-38 human diploid cells. *Am J Dis Child*. 1969;118:178–85.
- Stratton K, et al. Measles-mumps-rubella vaccine and autism. *Immunization Safety Review*. National Academy Press, Washington, DC; 2001.
- Usonis V, et al. Comparative study of reactogenicity and immunogenicity of new and established measles, mumps and rubella vaccines in healthy children. *Infection*. 1998;26(4):222–6.
- Usonis V, et al. Reactogenicity and immunogenicity of a new live attenuated combined measles, mumps and rubella vaccine in healthy children. *Pediatr Infect Dis J*. 1999;18(1):42–8.
- Vesikari T, Sadzot-Delvaux C, Rentier B, Gershon A. Increasing coverage and efficiency of measles, mumps, and rubella vaccine and introducing universal varicella vaccination in Europe: a role for the combined vaccine. *Pediatr Infect Dis J*. 2007;26(7):632–8.
- Wakefield AJ, Murch SH, Anthony A, et al. Ileal-lymphoid-nodular hyperplasia, non-specific colitis, and pervasive developmental disorder in children. *Lancet* 1998;351:637–41.
- WHO Euro. Measles and rubella elimination. package for accelerated action 2013–2015.
- WHO Position Paper. Mumps virus vaccines. *Wkly Epidemiol Rec*. 2007;82(7):51–60.

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10.1 Burden of Varicella Disease

Primary varicella zoster virus (VZV) infection, or chickenpox, is characterized by generalized pruritic rash, which rapidly progresses from macular to papular and finally vesicular before crusting. In an unvaccinated population it affects almost all persons and usually manifests between 1 and 9 years of age.

Varicella usually occurs in healthy children as an uncomplicated disease. However, severe disease may occur, especially among adolescents, adults, pregnant women, and the immunocompromised patients, but also in children without underlying disease. About one half of the severe cases are in subjects without any known risk factor.

Severe bacterial skin infection is a common complication of varicella. CNS manifestations include febrile or cerebral convulsions, cerebellar ataxia, encephalitis, Guillain-Barré syndrome, facial palsy, and cerebral vasculitis. Other frequent viral complications are pneumonia, hepatitis, thrombocytopenia, nephrotic syndrome, and pancreatitis. Severe complications include bacteremia and toxic shock syndrome, Reye syndrome (encephalopathy and hepatic failure following aspirin treatment in children with varicella), and necrotizing fasciitis, purpura fulminans, and disseminated coagulopathy are rare, but associated with significant mortality. Neonatal varicella, occurring in newborns between the 5th and the 12th day of life, is associated with mortality in up to 20% of cases. Congenital VZV infection resulting from varicella in the first 26 weeks of pregnancy causes severe abnormalities of the skin, CNS, eyes, and other organs in the fetus.

Prevention of severe varicella and its complications is a major goal of varicella vaccination. The two main reasons why varicella vaccination should be targeted at all healthy children are: (1) severe and complicated cases of varicella may occur in persons without risk factors; (2) children with risk factors such as primary or secondary

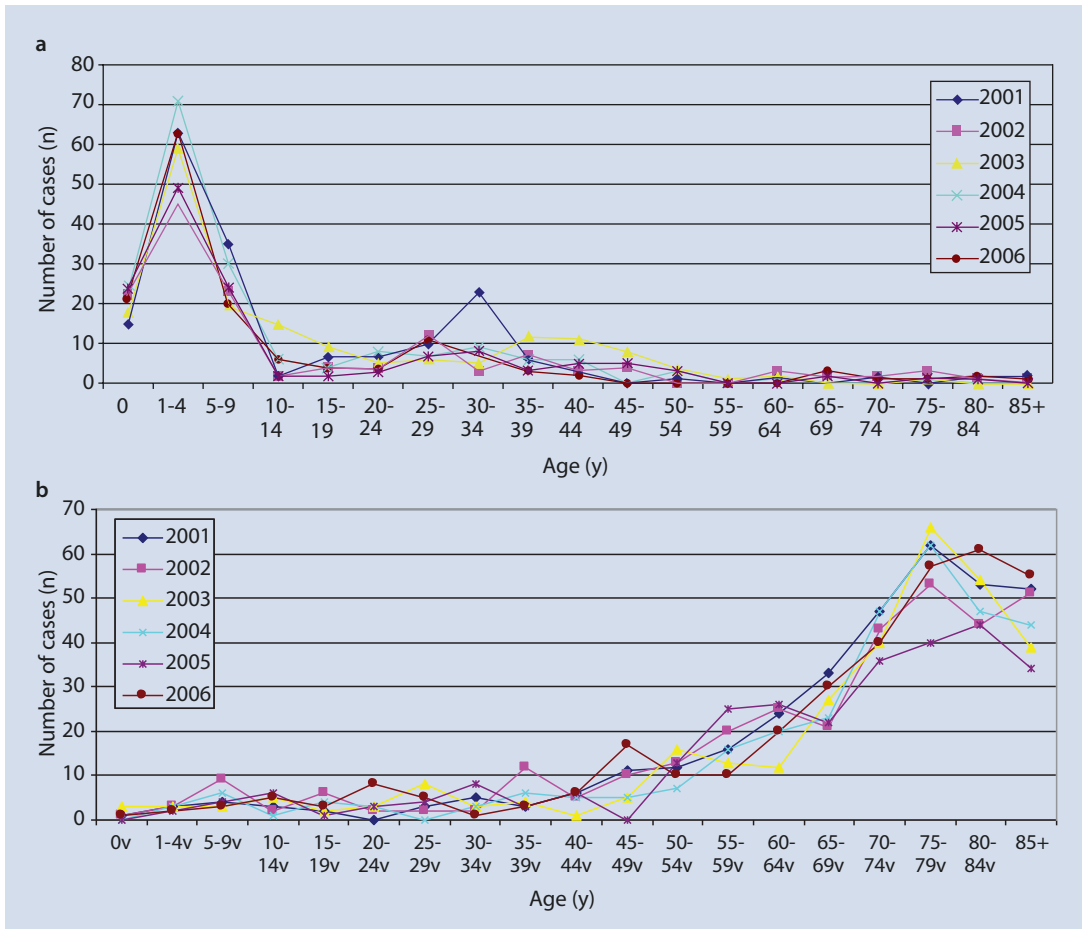
immunodeficiency (due to for example cancer or corticosteroid therapy) should not be vaccinated with a live attenuated varicella vaccine, and therefore can only be protected by herd protection resulting from high vaccine coverage in the healthy population.

10.2 VZV Epidemiology

The VZV is transmitted by respiratory secretions or vesicle fluid. The incubation period is usually 14–16 days, with a range of 10–21 days. Transmission from person to person may occur from 1–2 days before the onset of the rash until the fifth day after the onset of the rash, or until all lesions have crusted.

In the absence of vaccination, the annual number of varicella cases in Europe is close to the country's birth cohort, and nearly 95% of the population will have been infected by VZV by the age of 10 years. In the USA before the introduction of general varicella vaccination most cases occurred in children aged 5–9 years, whereas in central and northern European countries the age group of 1–4 years was (and is) mainly affected (■ Fig. 10.1a). The highest hospitalization rates are reported among those under 1 year of age followed by immunocompromised subjects and pregnant women. Before varicella vaccinations, there were about 100 deaths from varicella in the USA and 4–10 deaths in Germany each year.

Following primary infection, the virus establishes latency in the sensory nerve ganglia and can reactivate when natural immunity wanes, leading to herpes zoster (HZ). The risk of HZ increases with age starting from about 50 years of age (■ Fig. 10.1b). One third of HZ patients over 70 develop post-herpetic neuralgia (PHN), which may be refractory to various treatments, such as antivirals, nonsteroidal anti-inflammatory agents, corticosteroids, and tricyclic antidepressants. This is a major argument in favor of the development and use of vaccines against HZ.



■ Fig. 10.1 a Outpatient clinic visits for varicella, Finland 2001–2006. b Hospital admissions for zoster, Finland 2001–2006 (From: ▶ <https://www.julkari.fi/bitstream/handle/10024/103011/2008b40.pdf?sequence=1>)

10.3 Varicella Vaccines

Live attenuated varicella vaccine was developed by Takahashi in 1974. The viral strain that has been used as a vaccine was isolated from a Japanese child with varicella, named Oka, and was then attenuated through sequential passages in tissue cultures and at a low temperature. The “Oka” strain is currently used for the production of all licensed varicella vaccines worldwide.

In Europe, the first varicella vaccine (Varilrix, GSK) was licensed for high-risk children in 1984 and, with a higher virus titer, for healthy children in 1995. For varicella vaccination of healthy children, Varilrix and Varivax (Merck) vaccines are available in Europe. Varivax is reported to have a

virus concentration of 1,350 Oka/Merck plaque forming units (pfu) and Varilrix $10^{3.3}$ Oka/RIT pfu (representing about 1,995 pfu), i.e., the concentrations do not differ very much.

Live attenuated varicella vaccine induces a natural-like immunity, which is mediated through VZV-specific cellular and humoral immune responses. Evidence for the protective role of serum antibodies is indicated by a correlation between circulating VZV-specific antibody concentration and the probability of breakthrough varicella. Passive immunoprophylaxis by varicella zoster immunoglobulin after exposure to varicella also indicates the protective role of antibodies. VZV-specific cellular immunity is also associated with protection from VZV reactivation.

Table 10.1 Efficacy of a single dose of high-titer and low-titer varicella vaccine (Varilrix®) with a follow-up of 2.5 years

Vaccine	Efficacy against varicella	
	Any severity	Moderately severe
High titer (7,940 pfu)	88% (72–96)	100%
Low titer (2,540 pfu)	55% (31–72)	

From: Varis and Vesikari (1996)

Pre-licensure studies of GSK's varicella vaccine showed that protection was dose-dependent, with a higher dose of varicella vaccine conferring higher protection and the protection being better against severe disease (Table 10.1). A more recent post-licensure efficacy trial found the efficacy of one dose of Varilrix to be 65%. In real-life outbreaks, the protection has been even lower. The post-introduction experience has resulted in the introduction of a second vaccine dose in most countries with a general varicella vaccination program.

In 2006 and 2009, two doses of varicella vaccine to healthy children were recommended in the US and in Germany respectively. Two-dose vaccine recipients achieve up to 20-fold higher antibody levels and higher seroconversion rates than subjects receiving a single dose, and the booster effect is achieved irrespective of the time interval between administration of the first and second doses. The efficacy of two doses of either the Oka/Merck or the Oka/RIT strain is over 95% for any severity of varicella disease, at least for the first years after vaccination.

10.4 Vaccine Safety

The safety profile of the varicella vaccine in healthy subjects comes from preclinical studies and extensive post-marketing worldwide experience. The varicella vaccine may cause injection site reactions, including zoster-like localized rash in about 3–5% of immunized children. Additionally, a mild and transient generalized varicella-like rash may be seen. Rashes occur typically 5–26 days after immunization, and usually consist of two

to five lesions, mostly maculopapular rather than vesicular. However, most rashes that occur within the first 2 weeks after varicella immunization are due to wild-type VZV. Fever is common.

■ Description of Varilrix

► <https://www.medicines.org.uk/emc/medicine/9787>

Varilrix^{®c} contains 10^{3.3} pfu (representing about 1,995 pfu) of live attenuated varicella-zoster (Oka strain) virus propagated in MRC5 human diploid cells. The vaccine contains amino acids, human albumin, lactose, mannitol, and sorbitol. The solvent for reconstitution is Water for Injections. Two doses (each consisting of 0.5 ml of reconstituted vaccine) should be given, with an interval between doses of at least 6 weeks, but in no circumstances less than 4 weeks. One dose of Varilrix may be administered after a first dose of another varicella-containing vaccine.

■ Description of Varivax

The lyophilized vaccine contains sucrose, hydrolyzed gelatin, urea, sodium chloride, monosodium L-glutamate, anhydrous disodium phosphate, potassium dihydrogen phosphate, and potassium chloride.

When vaccination is initiated between 9 and 12 months of age, a second dose is needed and should be given after a minimum interval of 3 months.

In individuals from aged between 12 months and 12 years, at least 1 month must elapse between the first and second doses.

Individuals 13 years of age and older should receive two doses with an interval of 4–8 weeks. If the interval between doses exceeds 8 weeks, the second dose should be given as soon as possible.

10.5 Post Licensure Effectiveness of Varicella Vaccine (Live) (Oka/Merck Strain) in the USA

Five cross-sectional long-term surveys on varicella incidence, each from a random sample of approximately 8,000 children and adolescents 5–19 years of age, were conducted over a period of 15 years in the USA, from 1995 (pre-vaccine) through 2009. Results showed a gradual decline in varicella incidence rates by 90–95% overall (approximately 10- to 20-fold) from 1995 to 2009 in all age groups, both in vacci-

nated and unvaccinated children and adolescents. In addition, a decrease by approximately 90% (approximately 10-fold) in varicella hospitalization rates was observed in all age groups. The estimated vaccine effectiveness (largely one dose only) over the study period was between 73% and 90%.

10.6 Post Licensure Varicella Vaccine Effectiveness in Europe

In Europe, the greatest experience with post-licensure effectiveness data comes from Germany, which was the first European country to introduce universal varicella immunization (UVV) and at the same time have an active surveillance system to monitor the disease and its complications. Surveillance data indicate that in the first years after nationwide varicella vaccine implementation in 2004, the overall incidence of varicella decreased in two independent studies by 76–84% in children less than 19 years of age. Varicella hospitalization rates in the general population decreased between 2005 and 2012 by 60% in children and 40% in the adult population. Overall varicella vaccine effectiveness in preventing varicella disease (mild or severe) was 86% after dose 1 and 94% after dose 2. Moreover, sentinel data from April 2005 to May 2009 showed a reduction of 55% of varicella cases in all age groups, 63% in the age group 0–4 years and 38% in 5- to 9-year-olds.

The very significant reductions in the incidence of varicella and varicella-associated complications observed in Germany have also been

confirmed by regional data from other European countries that have implemented UVV programs, especially in those that have implemented a two-dose schedule coupled with a catch-up program and achieved a very high vaccination coverage. In all countries with a high vaccine coverage leading to a fast reduction of VZV circulation in the community, a significant reduction was observed in unvaccinated children younger than 1 year of age and older populations, indicating herd protection.

As of December 2016, Austria, Germany, Greece, and Luxembourg have UVV recommendations and programs at the national level and Italy at the regional level. Spain had implemented UVV in a few regions, but recently changed its policy and currently recommends the vaccine only for high-risk groups. Sixteen countries recommend targeted vaccination of susceptible adolescents and/or risk groups, 13 countries recommend vaccination for susceptible health care workers, and 4 for susceptible day-care personnel. Finland is starting UVV with a two-dose program combined with catch-up in 2017.

10.7 MMRV Vaccine

The combination of MMR plus varicella vaccine has been available since 2006. To make a proper combination, the titer of the varicella component in Merck's MMRV vaccine was increased from 1,350 pfu to 9,972 pfu for greater immunogenicity. Thus, the immunogenicity for VZV is higher after combined MMRV than after a single varicella vaccine (■ Table 10.2). In GSK's MMRV vaccine the

■ **Table 10.2** Immune responses to two doses of a quadrivalent measles, mumps, rubella, and varicella vaccine in healthy children

	Pro-Quad <i>n</i> = 381		MMR and varicella separately <i>n</i> = 390	
	Seroconversion (%)	GMT	Seroconversion (%)	GMT
Measles	100	747	99.7	253
Mumps	100	286	99.7	97
Rubella	100	254	98.6	128
Varicella	99	469	93.1	16.5

From: Shinefield et al. (2005)
MMR measles–mumps–rubella, GMT geometric mean titer

varicella component has the same titer as in single varicella vaccine (1,995 pfu). Two preparations of MMRV available are Priorix-Tetra® (GSK) and Pro-Quad® (Merck).

The reactogenicity of the MMRV combination is higher than after MMR vaccine given alone or separately, but concomitantly together with varicella vaccine (■ Fig. 9.3). This is true for skin reactions, but particularly for high fever that occurs around days 5–12 after vaccination. In line with more frequent and higher fever, febrile convulsions also occur more frequently after MMRV than after MMR and varicella vaccine given separately. It is not clear if the two preparations of MMRV differ in this respect.

Universal varicella immunization programs may use monovalent varicella vaccine for the first dose to avoid the increase in febrile seizures associated with MMRV administration. MMRV is preferred for the second dose. The timing of the second dose of MMRV is more frequently determined by the MMR vaccination schedule. Germany and certain parts of Italy administer the second dose of MMRV at a 3-month interval. Such immunization schedules could enhance vaccine effectiveness, especially in the first years of UVV implementation because they can reduce more effectively the circulation of the VZV virus in the community and prevent breakthrough. It has been speculated that solid immunity against VZV might also be more likely to block subclinical infection by wild-type VZV, with ensuing latency.

10.8 Shift of Varicella Disease to Older Ages

Reduced circulation of wild-type VZV in the community owing to the use of varicella vaccine could be associated with an undesirable age shift of the incidence of varicella, associated with an increased severity of the disease, expected in older children, adolescents, pregnant women, and adults infected by VZV. So far, surveillance of varicella disease following the implementation of the two-dose schedules in European countries is reassuring; data show that the overall rates of varicella among adolescents and adults are declining and no age shift of varicella has been

observed. However, data from seroprevalence studies indicate significant VZV immunity gaps among adolescents. Therefore, efforts at identifying susceptible adolescents for subsequent catch-up vaccination is critical to avoid the undesirable age shift of varicella to older ages, when varicella disease is more severe.

10.9 Impact of UVV on HZ

A significant concern with regard to the universal use of varicella vaccine is a possible effect of vaccination on the incidence of HZ, among both vaccinated and unvaccinated subjects.

In the vaccinated subjects, the vaccine virus may cause latent infection and remain in the dorsal root ganglia. The pathogenesis of HZ from the vaccine strain could be associated with a high concentration of the vaccine virus infecting the nerves at the vaccination site. It has been observed that the HZ rash in vaccinated children occurs more commonly in the dermatomes corresponding to the sites where the varicella vaccine was given.

However, real-life data from European countries have shown that the risk of developing zoster among the vaccinated population is significantly lower compared with that reported in children infected by wild-type varicella. This finding could be attributed to the lower viral loads in the vaccine and to the reduced pathogenic capacity of the OKA strain compared with the wild-type virus. Nevertheless, long-term surveillance for HZ is required to confirm that the two-dose schedule establishes effective and long-lasting cellular immunity that will reduce the incidence of HZ among vaccinated subjects.

More significant concerns have been associated with a possible increase in HZ among subjects that have already been infected with the wild-type virus. Re-exposure to VZV through contact with an infected person may boost VZV cellular immunity and increase protection against HZ, and in areas with UVV, the incidence of HZ could increase owing to reduced exposure to varicella.

Nevertheless, real-life experience has indicated only a slight increase or no increase in HZ incidence in areas where universal varicella vaccination has been implemented, comparable with countries where no UVV has been implemented.

Such a discrepancy between the predicted increase in HZ and the real-life situation suggests that silent reactivation of the wild-type virus might be associated with endogenous boosting of cellular immunity and might be more important in maintaining latency rather than immunity from exogenous boosting.

10.10 Varicella Vaccine Recommendations for Special Groups

After licensure, the varicella vaccine was primarily intended for the vaccination of high-risk groups, such as children with leukemia or cancer. However, live varicella vaccine today is contraindicated in individuals with immunosuppression because of the high rate of adverse effects and because it is necessary to temporarily stop chemotherapy for varicella vaccination. Instead, varicella vaccine is now recommended for:

- Childhood candidates for solid organ transplant with no history of chickenpox (or unclear) 6 months before surgery, with undetectable antibodies
- Seronegative subjects in remission from malignancies
- Adolescents 12–18 years or older and women of childbearing age with no history of varicella
- People in contact with immunosuppressed patients
- Health care workers
- Child care workers
- Laboratory staff
- As post-exposure prophylaxis (given within 72 h of exposure)

10.11 Contraindications to Varicella Vaccine

Varicella vaccine is contraindicated in:

- Subjects with primary or acquired immunodeficiency states with a total lymphocyte count less than 1,200 per mm³
- Severe humoral or cellular primary immunodeficiencies, e.g., severe combined immunodeficiency

- Subjects with a lack of cellular immune competence, such as leukemia, lymphoma, blood dyscrasia
- Individuals receiving immunosuppressive therapy including high-dose corticosteroids
- Patients who clinically manifest AIDS or symptomatic HIV infection or have low age-specific CD4+ T-lymphocyte counts
- Active untreated tuberculosis
- Pregnancy and breast-feeding (pregnancy should be avoided for 1 month following vaccination)

Transmission of the vaccine virus from vaccinees to susceptible contacts has rarely been shown to occur and has been associated with vaccine-associated cutaneous lesions. Therefore, contact with high-risk individuals must be avoided if the vaccinee develops a cutaneous rash likely to be vaccine-related within 4–6 weeks of the first or second dose and until this rash has completely disappeared. In the absence of a rash in the vaccinee, the risk of transmission of the vaccine viral strain is deemed non-existent.

10.12 Herpes Zoster Vaccine

The first vaccine against HZ (Zostavax®, Merck) was licensed in 2006. The vaccine is essentially a concentrated form of Varivax® containing 14 times more live VZV. In addition, it contains an unknown quantity of nonlive varicella antigenic material. However, because of the live virus, Zostavax® cannot be given to immunocompromised persons.

The efficacy of Zostavax against HZ in the age group 50–59 years is about 70%, and decreases with increasing age. Several European countries, including the UK, France, and two federal states of Germany recommend the use of Zostavax® in various older age groups.

A new nonlive vaccine against HZ (Shingrix®), constituting of VZV glycoprotein E combined with an adjuvant, was recently developed by GSK. This vaccine, given in two doses, has shown efficacy of 90–97%. As a nonlive vaccine, Shingrix® could be given to immunocompromised subjects, and may also become available for pediatric use in selected patients who experience HZ at an early age. The licensure of Shingrix® is expected in 2017.

Further Reading

- Baxter R, Ray P, Tran TN, Black S, Shinefield HR, Coplan PM, et al. Long-term effectiveness of varicella vaccine: a 14-year. Prospective Cohort Study *Pediatrics*. 2013;131(5):e1389–96.
- Bonanni P, Breuer J, Gershon A, Gershon M, Hryniewicz W, Papaevangelou V, et al. Varicella vaccination in Europe – taking the practical approach. *BMC Med*. 2009;7:26.
- Bonanni P, Gershon A, Gershon M, Kulcsar A, Papaevangelou V, Rentier B, et al. Primary versus secondary failure after varicella vaccination: implications for interval between 2 doses. *Pediatr Infect Dis J*. 2013;32(7):e305–13.
- Bozzola E, Tozzi AE, Bozzola M, Krzysztofak A, Valentini D, Grandin A, et al. Neurological complications of varicella in childhood: case series and a systematic review of the literature. *Vaccine*. 2012;30(39):5785–90.
- European Medicines Agency. Monovalent and multivalent measles, mumps, rubella and/or varicella vaccines 2012 [cited 2013]. Available from: http://www.ema.europa.eu/ema/index.jsp?curl=pages/medicines/human/referrals/MMRV/human_referral_000334.jsp&mid=WC0b01ac05805c516f#documents.
- Galea SA, Sweet A, Beninger P, Steinberg SP, Larussa PS, Gershon AA, et al. The safety profile of varicella vaccine: a 10-year review. *J Infect Dis*. 2008;197(Suppl 2):S165–9.
- Klein NP, Fireman B, Yih WK, Lewis E, Kulldorff M, Ray P, et al. Measles-mumps-rubella-varicella combination vaccine and the risk of febrile seizures. *Pediatrics*. 2010;126(1):e1–8.
- Knuf M, Zepp F, Meyer CU, Habermehl P, Maurer L, Burow HM, et al. Safety, immunogenicity and immediate pain of intramuscular versus subcutaneous administration of a measles-mumps-rubella-varicella vaccine to children aged 11–21 months. *Eur J Pediatr*. 2010;169(8):925–33.
- Lal H, Cunningham AL, Godeaux O, et al. Efficacy of an adjuvanted herpes zoster subunit vaccine in older adults. *N Engl J Med*. 2015;372:2087–96.
- Liese JG, Grote V, Rosenfeld E, Fischer R, Belohradsky BH, v Kries R. The burden of varicella complications before the introduction of routine varicella vaccination in Germany. *Pediatr Infect Dis J*. 2008;27(2):119–24.
- Ogunjimi B, Van Damme P, Beutels P. Herpes zoster risk reduction through exposure to chickenpox patients: a systematic multidisciplinary review. *PLoS One*. 2013;8(6):e66485.
- Oxman MN, Levin MJ, Johnson GR, et al. A vaccine to prevent herpes zoster and postherpetic neuralgia in older adults. *N Engl J Med*. 2005;352:2271–84.
- Seward JF, Marin M, Vazquez M. Varicella vaccine effectiveness in the US vaccination program: a review. *J Infect Dis*. 2008;197(Suppl 2):S82–9.
- Shinefield H, Black S, Williams WR, et al. Dose-response study of a quadrivalent measles, mumps, rubella and varicella vaccine in healthy children. *Pediatr Infect Dis J*. 2005;24(8):670–5.
- Siedler A, Arndt U. Impact of the routine varicella vaccination programme on varicella epidemiology in Germany. *Euro Surveill*. 2010;15(13)
- Spackova M, Muehlen M, Siedler A. Complications of varicella after implementation of routine childhood varicella vaccination in Germany. *Pediatr Infect Dis J*. 2010;29(9):884–6.
- Takahashi M, Otsuka T, Okuno Y, Asano Y, Yazaki T. Live vaccine used to prevent the spread of varicella in children in hospital. *Lancet*. 1974;2(7892):1288–90.
- Varis T, Vesikari T. Efficacy of high-titer live attenuated varicella vaccine in healthy young children. *J Infect Dis*. 1996;174(Suppl 3):S330–4.

Rotavirus Vaccine

Timo Vesikari

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11.1 Burden of Rotavirus Disease

The clinical characteristics of severe rotavirus (RV) gastroenteritis (RVGE) include watery diarrhea, frequent vomiting, and high fever. About 20–30% of all children experience a clinically manifest episode of RVGE, and 10–20% of these (2–3% of all) are severe. Prevention of severe RVGE is the primary target of RV vaccination. In Europe, RV causes about one half of severe acute gastroenteritis (GE) in childhood requiring hospitalization. On average, RVGE is more severe than gastroenteritis caused by other viruses.

Moreover, it is now recognized that RV often causes a systemic infection and RV antigen and RNA can be detected in the circulation. RV vaccination also prevents some 20% of all febrile seizures. Rather than gastroenteritis, it is more appropriate to talk about RV disease. Prevention of RV disease by vaccination is a neutral term that puts RV vaccine in the same category as other viral vaccines, in contrast to being a “diarrheal disease vaccine.”

Still, the first target of RV vaccination in Europe is the prevention of severe RVGE and, specifically, hospitalizations for RVGE. Hospital admission is also the major factor (about 90%) in calculations of financial burden associated with RVGE. The number of annual hospitalizations in Europe was at least 87,000 before RV vaccination was introduced. The rate of hospitalizations may vary according to local clinical practices, but there are probably also true differences between countries. For Europe, it has been estimated that the risk of hospitalization for RVGE before the age of 5 is 1 in 54, with a high of 1 in 33 in Finland and low of 1 in 67 in Denmark. It is plausible that in countries with long, cold winters, the RV season is longer and severe RVGE more common.

Some countries with a relatively low incidence of RVGE, such as Denmark and the Netherlands, have considered that there is no need to introduce RV vaccination into the immunization program. However, even if a country has decided not to introduce universal RV vaccination, at an individual level, the risk of severe RVGE in any European country is high enough to warrant prevention by vaccination.

Deaths from RVGE are rare in Europe (a 2006 estimate was 231 for European Union countries), but deaths may occur in cases of delayed admission to care. RVGE is still a potentially fatal dis-

ease in Europe, and the low mortality is only attributable to the availability of good case management at outpatient and hospital facilities.

Globally, RV is a major cause of childhood mortality. A recent estimate put the number of RV-associated deaths at 197,000 a year, down from 453,000 a few years earlier when the estimate was based on another method. Of individual countries, India has the highest number of deaths, followed by Nigeria, Pakistan, Bangladesh, and Indonesia. Introduction of RV vaccination in the high-mortality countries is a global public health priority.

11.2 RV Epidemiology

Almost all RVs causing disease in humans belong to group A, determined by the common inner core group antigen VP6. VP6 is the most abundant protein in the RV particle and a powerful immunogen, and immune reaction against this antigen is likely the major mechanism of protection against severe RV disease. Protection may be induced by natural RV infection or vaccination alike. It takes two or three infections, or “hits” to induce solid protection against severe disease; the “hits” may also be administered in two or three doses of oral vaccine, and the protection is limited to RV disease and not infection. Protection against RV infection depends on immunity against the VP7 and VP4 surface antigens, and such protection is more variable and not durable.

The two surface antigens VP7 and VP4 determine the G- and P-types of RVs respectively, and induce neutralizing antibodies. Although a large number of G- and P-combinations are possible, in reality a few fixed combinations prevail. The most common RV types are G1P[8], followed by G2P[4], G3P[8], G4P[8], G9P[8], and more recently, G12P[8]. Altogether, RV diversity has increased after RV vaccinations, but this has not reduced the effectiveness of the vaccine against severe RVGE, which is largely not dependent on immunity to G- or P-types. The surface antigen-induced antibodies protect against RV infection and have an effect on the RV strains that are prevalent in circulation, but the serotype-specific antibodies are not critical for the protection against severe RV disease.

Although the predominant RV types vary by the year, no single type is predominant in the

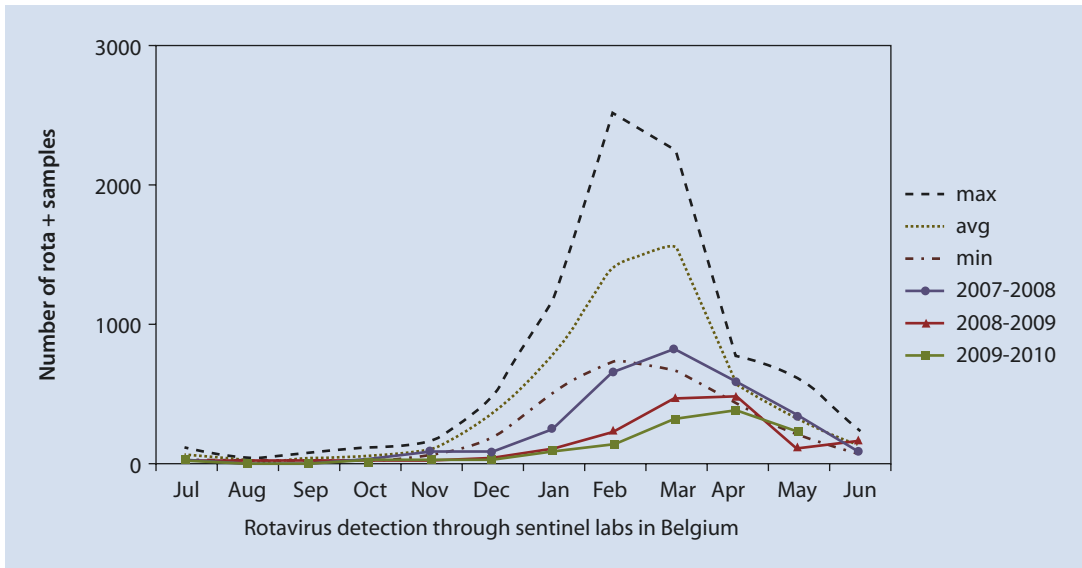


Fig. 11.1 Rotavirus epidemic season with a winter peak before vaccination and the blunted peak after vaccinations in Belgium

whole of Europe at the same time. Rather, there are multiple types of RV circulating at the same time in different regions. Thus, the rotavirus epidemic (season) does not have a single origin either, but RVs become prevalent in the winter season at various locations independently. Still, the seasonal pattern was very predictable until the introduction of universal RV vaccinations. In the countries with a high coverage of vaccinations, the RV season has shifted from peak winter toward spring and summer (Fig. 11.1).



Fig. 11.2 Oral administration of the rotavirus (RV) vaccine (RotaTeq®)

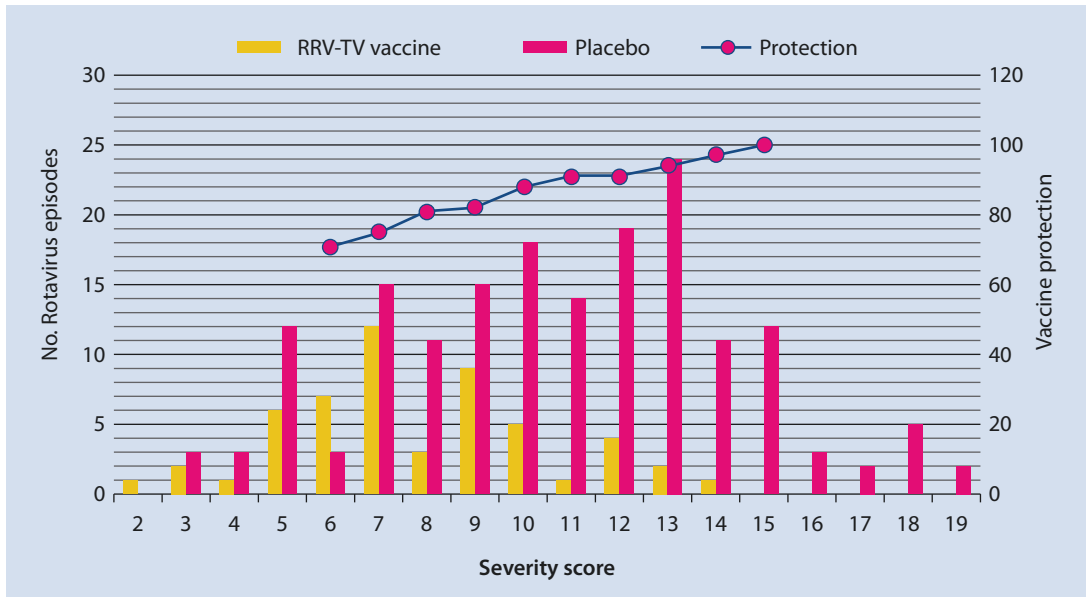
11.3 RV Vaccines

All RV vaccines are live attenuated tissue culture-grown RVs of human or animal origin or reassortants of human and animal RVs. RV vaccines are given orally (Fig. 11.2) to multiply in the intestine and to mimic asymptomatic wild-type RV infection. Vaccine virus infection is likely to induce natural-like immunity against RV disease, even if the mechanism of protection is not fully known.

The first experimental RV vaccine was a bovine rotavirus that was found to infect humans and to induce a high level of cross-protection against severe human RVGE in spite of having “non-human” G- and P-types. The early studies of bovine RV vaccine in the 1980s established some

general principles of RV vaccination, which have been confirmed subsequently in numerous studies with other live RV vaccines: (1) vaccine-induced protection is higher against severe RVGE than any (including mild) RV disease; (2) oral RV vaccine needs to be given with a buffer because gastric acid may inactivate RV and reduce the uptake of RV vaccine; (3) breast milk or breast feeding (despite RV IgA in the breast milk) does not negatively affect the uptake of RV vaccine; (4) simultaneous administration of oral poliovirus vaccine (OPV) may interfere with live RV vaccine.

The first licensed RV vaccine (RotaShield®, Wyeth) in 1998 in the USA was a rhesus-human



■ Fig. 11.3 Rotavirus disease severity and rhesus rotavirus tetravalent vaccine protection. (Joensuu et al. 1997; Ruuska et al. 1990)

reassortant “tetravalent” vaccine, which contained three reassortants of rhesus rotavirus with human G-types G1, G2, and G4 plus the rhesus RV (G3) itself. This vaccine was given in three doses and after a full series induced a high level of protection, as shown in ■ Fig. 11.3. With the use of a 20-point severity scale (“Vesikari scale”), the protection level against different severities of RVGE was determined with a greater accuracy. The protection reached 100% against disease with a severity score of 15/20; using the most commonly applied cut-off score of 11/20 for severe RVGE, the protection was about 90%. The same scale has been used to measure protection of other RV vaccines as well.

RotaShield® induced febrile reactions in about one third of the recipients and about 3% had high fever. After a million doses given in the USA by 1999, the vaccine was found to be associated with intussusception (IS), and was withdrawn. Other rotavirus vaccines are not reactogenic like RotaShield®. Still, the current RV vaccines may also cause IS, even though the risk is lower than that associated with RotaShield®.

The current major licensed RV vaccines are human RV vaccine (Rotarix™, GSK) and bovine–human reassortant RV vaccine (RotaTeq®, Merck),

both of which are available and widely used in Europe and globally. The recommendations of the European Society for Pediatric Infectious Diseases (ESPID) takes the position that both vaccines can be recommended to protect European children from RVGE and that the performance of the vaccines in Europe is equal. No formal head-to-head comparison of the vaccines has been done.

11.4 Human RV Vaccine Rotarix™

Human RV vaccine (Rotarix™, GSK), also termed RV1, is the most extensively used RV vaccine today. It was derived from a G1P[8] RV isolate in Cincinnati, passaged 33 times in cell culture and designated 89–12. The strain was acquired by GSK, cloned (by plaque purification) and passaged another 12 times in MRC-5 cells. In this process, the virus lost its residual reactogenicity and is generally regarded as nonreactogenic for humans. Rotarix™ multiplies effectively in humans, as characterized by a high rate of shedding (60% or even more) after the first dose, but does not cause diarrhea or systemic reactions; in other words, it is highly attenuated for its original host.

Description of Rotarix™ According to the Summary of Product Characteristic (SPC)

► https://www.gsksource.com/pharma/content/dam/GlaxoSmithKline/US/en/Prescribing_Information/Rotarix/pdf/ROTARIX-PI-PIL.PDF

ROTARIX, for oral administration, is a live, attenuated rotavirus vaccine derived from the human 89–12 strain, which belongs to the G1P[8] type. The rotavirus strain is propagated on Vero cells. After reconstitution, the final formulation (1 mL) contains at least 10⁶ median Cell Culture Infective Dose (CCID50) of live, attenuated rotavirus.

The lyophilized vaccine contains amino acids, dextran, Dulbecco's Modified Eagle Medium

(DMEM), sorbitol, and sucrose.

DMEM contains the following ingredients: sodium chloride, potassium chloride, magnesium sulfate, ferric (III) nitrate, sodium phosphate, sodium pyruvate, D-glucose, concentrated vitamin solution, L-cystine, L-tyrosine, amino acids solution, L-glutamine, calcium chloride, sodium hydrogencarbonate, and phenol red.

In the manufacturing process, porcine-derived materials are used. Porcine circovirus type 1 (PCV-1) is present in ROTARIX. PCV-1 is not known to cause disease in humans.

The liquid diluent contains calcium carbonate, sterile water,

and xanthan. The diluent includes an antacid component (calcium carbonate) to protect the vaccine during passage through the stomach and prevent its inactivation owing to the acidic environment of the stomach.

ROTARIX is available in single-dose vials of lyophilized vaccine, accompanied by a prefilled oral applicator of liquid diluent. The tip caps of the prefilled oral applicators may contain natural rubber latex; the vial stoppers are not made with natural rubber latex.

ROTARIX contains no preservatives.

Rotarix™ is given in two doses. The uptake and immunogenicity are excellent (90%) even after the first dose when given in the presence of a low level of maternal antibody, such as in European populations. The uptake of the second dose may be prevented by the antibodies induced after the first dose, as indicated by the lack of shedding and lack of a booster response after the second dose. Therefore, the second dose mainly fills the immunity gap remaining after the first dose, but does not induce an increase in the level of antibodies if the first dose has been successful. The pivotal safety and efficacy trial for licensure was carried out in 60,000 children in Latin America. Before licensure in Europe, the vaccine was tested in five European countries. Rotarix™ was the first new RV vaccine to be licensed after the withdrawal of RotaShield®, with European licensure in 2006.

The results of the major European efficacy trial of Rotarix™ are illustrative for the performance of this vaccine. The primary end-point was severe RVGE, as defined by score 11/20. Against such severe RVGE, the efficacy for 2 years was 91%, with 96% efficacy in the first season and 86% in the second season, showing a decline over time. Against any RVGE the efficacy was 78% and 68% in the first and second year respectively, for a total efficacy of 72% over 2 years. The efficacy against severe

RVGE by G-type ranged from 96% for G1P[8] to 86% for G2P[4], these differences were not statistically significant (■ Fig. 11.4a). For any RVGE, the efficacy point estimates were higher for G1, G3, G4, and G9 with P[8] than for with G2P[4] with 58%. The interpretation would be that a G1P[8] vaccine cannot well control the circulation of G2P[4] RV, but remains efficacious against severe RVGE caused by this “fully heterotypic” RV. G2P[4] has become more prevalent after universal RV vaccination with Rotarix, particularly in Latin America.

11.5 Bovine–Human Reassortant RV Vaccine, RotaTeq®

The “pentavalent” bovine–human reassortant RV vaccine (RotaTeq®, Merck, also termed RV5) is a combination of four G-type reassortants (for G1–G4) and one P-type (P[8]) reassortant on the WC-3 bovine RV genetic backbone. As WC-3 is a G6P[5] virus, these bovine G and P types are also present in the vaccine. The terms “pentavalent” and RV5 refer to the five mono-reassortant strains in the vaccine. However, it is now well established that the protection against severe RVGE induced by the vaccine is not limited to the G or P types contained in the product (see below).

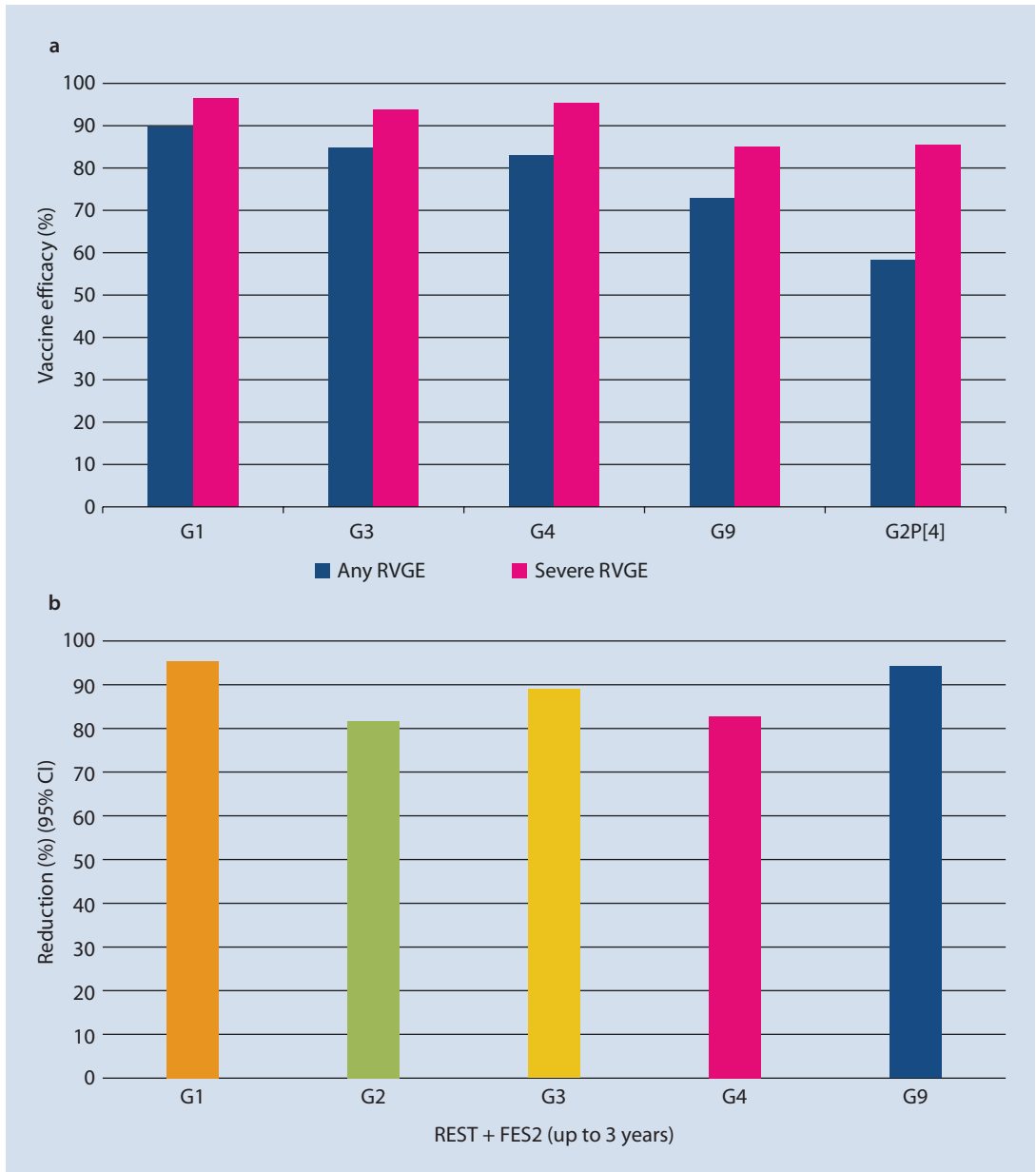


Fig. 11.4 **a** European efficacy trial of Rotarix. Vaccine efficacy against rotavirus gastroenteritis (RVGE) caused by specific RV types. **b** Finnish Extension Study: serotype-

specific efficacy of RV5 against hospitalizations and emergency department visits

Description of RotaTeq® According to the SPC

► http://www.merck.com/product/usa/pi_circulars/r/rotateq/rotateq_pi.pdf

RotaTeq is a live, oral pentavalent vaccine that contains five live reassortant rotaviruses. The rotavirus parent strains of the reassortants were isolated from human and bovine hosts. Four reassortant rotaviruses express one of the outer capsid proteins (G1, G2, G3, or G4) from the human rotavirus parent strain and the attachment protein (serotype P7) from the bovine rotavirus parent strain. The fifth reassortant

virus expresses the attachment protein, P1A (genotype P[8]), herein referred to as serotype P1A[8], from the human rotavirus parent strain and the outer capsid protein of serotype G6 from the bovine rotavirus parent strain.

The reassortants are propagated in Vero cells using standard cell culture techniques in the absence of antifungal agents.

The reassortants are suspended in a buffered stabilizer solution. Each vaccine dose contains sucrose, sodium citrate, sodium phosphate monobasic monohydrate, sodium

hydroxide, polysorbate 80, cell culture media, and trace amounts of fetal bovine serum. RotaTeq contains no preservatives.

In the manufacturing process for RotaTeq, a porcine-derived material is used. DNA from porcine circoviruses (PCV) 1 and 2 has been detected in RotaTeq. PCV-1 and PCV-2 are not known to cause disease in humans.

RotaTeq is a pale yellow clear liquid that may have a pink tint.

The plastic dosing tube and cap do not contain latex.

The RotaTeq® vaccine is given in three doses. This was determined early on to accommodate the US childhood immunization program (2, 4, and 6 months of age), but has an additional basis in the demonstration of incremental immunogenicity and protection by each dose. RotaTeq® vaccine virus is also shed after vaccination, and the shedding may rarely be associated with diarrhea. The G1 and P[8] reassortants included in the RotaTeq® vaccine may re-reassort with each other and form vaccine-derived (vd) double reassortants on the bovine RV VP6 core, which may be more virulent than the original single reassortant vaccine viruses, and vdG1P[8] may be responsible for most of the diarrhea seen after vaccination in about 1% of the vaccine recipients.

The efficacy and safety of the RotaTeq® vaccine were established in the large (70,000 infants) Rotavirus Efficacy and Safety Trial (REST). The overall efficacy against severe RVGE as determined by health care utilization (combined endpoint of hospital admission and outpatient clinic treatment) was 95% (■ Fig. 11.4b). An extension study of the REST in Finland involving 21,000 children confirmed that RotaTeq was efficacious against severe RVGE associated not only with G1, G3, and G4, all P[8], but also against G9P[8], which is not among the G-types in the vaccine, and G2 P[4], with a dif-

ferent P-type. RotaTeq® was licensed in 2006 and is now one of the two major RV vaccines used globally.

11.6 Comparative Efficacy

Both the Rotarix™ and the RotaTeq® vaccines have been tested for efficacy in different environments, from developed to “intermediate” to developing countries. In general, the overall and serotype-specific efficacy against severe RVGE of the two vaccines are remarkably similar in all settings, being highest in Europe (around 95%) followed by Latin America (80–85%) and Africa (50–70%). No formal head-to-head comparative efficacy trial has been conducted. In a recent comparative immunogenicity study in the USA, three doses of RotaTeq® was more immunogenic by RV IgA response than two doses of Rotarix™. The same study showed that a mixed schedule of two doses of RotaTeq® and one dose of Rotarix™ was even more immunogenic.

11.7 Real-Life Effectiveness

Studies on the real-life effectiveness of RV vaccines after the introduction of immunization programs have been conducted in several countries

and areas. On the whole, there seems to be a similar gradient in vaccine effectiveness to that in prelicensure efficacy trials among developed, “intermediate,” and developing countries.

In Europe, the examples of Finland and Belgium are representative. In these countries, which have reached a high coverage with RV5 (Finland) and RV1 (Belgium) respectively, the real-life vaccine effectiveness in the target population has been well above 90% against hospitalization for RVGE. In Austria, with coverage of 72–74%, the reduction of RVGE hospitalizations in the target age group was 81–84% and this was sustained for up to 3 years. The direct impact of RV vaccination in the target age group has shifted the occurrence of RVGE to older unvaccinated children.

The indirect effect of RV vaccinations on unvaccinated children remains unsettled. In Austria, there was initially an indirect effect on unvaccinated children, but after 3 years this was followed by an increase in RVGE hospitalizations in 5- to 14-year-old children. In Finland, with an RV vaccination coverage of 95%, the reduction in cases of RVGE seen in hospitals was 94% in a period of 4 years after vaccination, but specifically in the age group 5–14 years, no significant reduction was seen over this period. It seems that large-scale RV vaccinations interrupt the circulation of wild-type RVs after initial introduction, but do not eliminate RV circulation. Over time, the circulating wild-type RVs find susceptible individuals and some of these will come down with severe RVGE. Still, the cases in older children do not have the same clinical significance as those in infants.

The impact of vaccines on all hospitalizations due to acute gastroenteritis depends on the share of RV in all severe gastroenteritis and the vaccine coverage. At best, the total reduction of hospitalizations from any gastroenteritis may be as high as 70%, as observed in Finland over a period of 4 years.

11.8 Introduction of RV Vaccination

After Austria (both vaccines), Belgium (Rotarix™), and Finland (RotaTeq® exclusively), there was a gap of a couple of years until Germany started universal vaccinations state by state. The most

significant recent step forward is perhaps the introduction into the UK in 2014. The map in **Fig. 11.5** shows the status of universal RV vaccinations in Europe in 2016.

No country that has initiated a universal program has stopped it. However, in 2015, France recalled the recommendation for RV vaccination over concerns of safety (IS) and is unlikely to relaunch a universal RV vaccination program. Spain has withdrawn the Rotarix vaccine for concerns over porcine circovirus (PCV-1) contamination (see below).

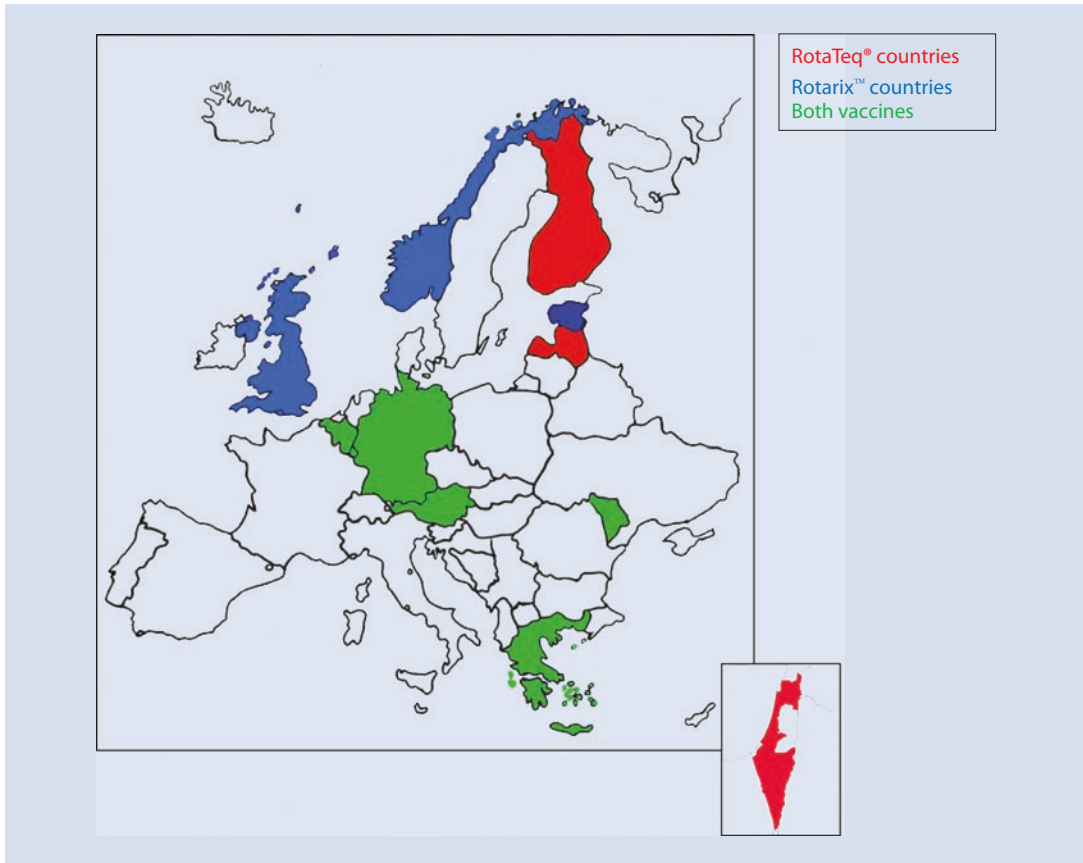
11.9 Intussusception

Intussusception is the most important adverse effect of RV vaccination. Association with IS led to the withdrawal of the first licensed RV vaccine, RotaShield®, in 1999. IS occurred mostly 3–7 days after the first dose of RotaShield®, and the attributable risk was estimated at 1:10,000. However, the risk of IS was shown to be age-dependent, and most of the cases occurred in the catch-up vaccination program in infants who were over 90 days of age at the time of the first dose.

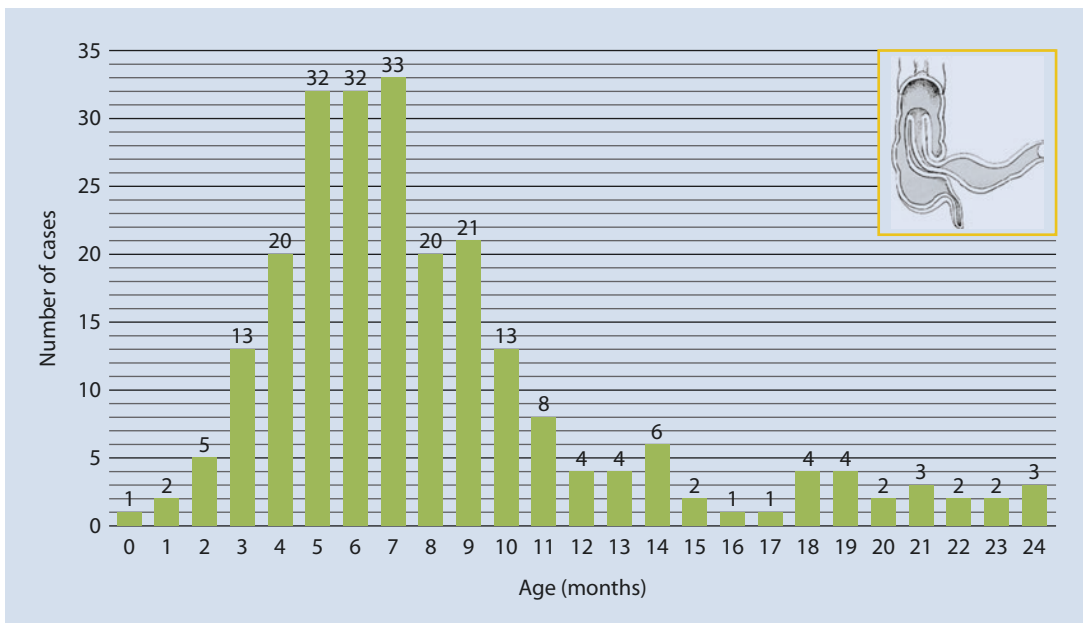
Both of the leading licensed RV vaccines, Rotarix™ and RotaTeq®, are also associated with IS, albeit with a lower risk than RotaShield®. The prelicensure trials did not detect the risk, as they were designed to rule out a risk of IS of similar magnitude to that with RotaShield®. Later, in a post-marketing surveillance study, the risk estimates of IS for both vaccines are between 1:50,000 and 1:80,000 after the first dose.

The age pattern of RV vaccine-associated IS, whether by RotaShield® or the current vaccines, may follow that of naturally occurring IS (**Fig. 11.6**) Therefore, it is important not to administer the first dose of any RV vaccine after 90 days of age, but it is prudent to follow the current ESPID recommendation and give the first dose of RV vaccine as early as possible, i.e., at 6–8 weeks of age.

The small risk of IS is often weighed against the benefits of RV vaccination, and this comparison comes out in favor of vaccination in developed countries as well. However, everything should be done to minimize the risk, and early administration of the first dose is of key importance.



■ Fig. 11.5 Implementation in Europe. Universal RV vaccination in Europe



■ Fig. 11.6 Age distribution of naturally occurring intussusception in Finnish children

11.10 Porcine *Circovirus*

In 2010, both licensed RV vaccines were found to have porcine *Circovirus* (PCV) as a contaminant. PCV is not known to infect humans, and the WHO and European Medicines Agency have held that RV vaccines may continue to be used. Some European countries withdrew Rotarix™ temporarily, but this position is maintained only in Spain. In Rotarix™, PCV contamination was traced to virus seed, but the manufacturer is committed to providing a PCV-free vaccine in the future. In RotaTeq®, the source of contamination was traced to batches of trypsin used in the manufacturing process and with changes in the process, PCV-free vaccine should be available. However, at the present time neither RV vaccine is explicitly PCV-free.

11.11 RV Vaccine Recommendations

In Europe, there is no formal recommendation-issuing body, but the pediatric societies, the ESPID and the European Society for Paediatric Gastroenterology Hepatology and Nutrition issued recommendations in 2008 that were updated as ESPID recommendations in 2015. The US Advisory Committee on Immunization Practices recommendations are also widely followed. Globally, the most important one is the WHO position for universal recommendation.

All major recommendations hold that RV vaccination should be given to all children, because no special “risk groups” for RVGE can be identified. Knowledge of the safety and efficacy of RV vaccine in many special groups has accumulated slowly, and some are summarized in the following.

11.11.1 Premature Infants

Both RotaTeq® and Rotarix™ vaccines can be given to prematurely born infants regardless of gestational age, following the recommendations according to calendar age. If the infant is still in hospital, a possible risk of transmission of the vaccine virus must be considered.

11.11.2 HIV Infected Children

Asymptomatic HIV-infected infants can be vaccinated normally according to calendar age without any safety issues using either Rotarix™ or RotaTeq®. Screening for maternally acquired HIV infection can often be done by the time of RV vaccination at 6–8 weeks of age, but the result is not needed for decision-making on RV vaccination.

11.11.3 Immunodeficiency

The RV vaccine causes symptomatic disease (prolonged diarrhea and viral shedding) in children with severe combined immunodeficiency, and therefore vaccination is contraindicated and exposure to RV vaccine shedders should be avoided in such children. Other immunodeficiencies may be regarded similarly. Selective IgA deficiency may result in the prolonged shedding of the RV vaccine, but does not constitute a safety problem and, in any case, is usually not diagnosed by the time of RV vaccination.

11.11.4 Short Gut Syndrome and Intestinal Failure

The RV vaccine may cause substantial symptoms in children with short bowel, but given the severity of the wild-type RV infection, they should nevertheless be vaccinated under close observation.

11.12 Nonlive RV Vaccines

The need and rationale for the development of nonlive RV vaccines as alternatives to live oral RV vaccines are based on efficacy and safety concerns. IS remains a serious safety concern, although the magnitude of the problem is regarded as tolerable. Also, the possibility of contamination by adventitious agents such as PCV is associated with live vaccines. As for efficacy, all live RV vaccines have shown a relatively (in comparison with developed countries) low efficacy in developing countries for reasons that may not be easily remedied. Parenteral immunization may induce a higher level of protection against RV disease bypassing the intestinal obstacles.

The types of nonlive RV vaccines under consideration include three main categories: (1) inactivated whole RVS; (2) RV virus-like particles (VLPs) of double-layered (VP2/VP6) or triple-layered (2/6/7 or 2/4/6/7) composition; (3) recombinant RV proteins such as VP8 or VP6.

Rotavirus VP6 alone forms tubular structures or spheres under appropriate conditions, and particulate forms of VP6 are strong immunogens. VP6 is also the simplest possible RV candidate vaccine consisting of only a single protein, which is considered a group antigen common to all group A rotaviruses. A whole new scenario might be a combined immunization against RV and norovirus GE using a RV VP6–norovirus VLP vaccine (see ► Chap. 25).

Further Reading

- Bishop RF, Davidson GP, Holmes IH, Ruck BJ. Virus particles in epithelial cells of duodenal mucosa from children with acute non-bacterial gastroenteritis. *Lancet*. 1973;2(7841):1281–3.
- Blazevic V, Lappalainen S, Nurminen K, Huhti L, Vesikari T. Norovirus VLPs and rotavirus VP6 protein as combined vaccine for childhood gastroenteritis. *Vaccine*. 2011;29:8126–33.
- Braeckman T, Van Herck K, Meyer N, RotaBel Study Group, et al. Effectiveness of rotavirus vaccination in prevention of hospital admissions for rotavirus gastroenteritis among young children in Belgium: case-control study. *BMJ*. 2012;345:e4752.
- Hemming M, Räsänen S, Huhti L, Paloniemi M, Salminen M, Vesikari T. Major reduction of rotavirus, but not norovirus, gastroenteritis in children seen in hospital after the introduction of RotaTeq vaccine into the National Immunization Programme in Finland. *Eur J Pediatr*. 2013;172(6):739–46.
- https://www.gsksource.com/pharma/content/dam/GlaxoSmithKline/US/en/Prescribing_Information/Rotarix/pdf/ROTARIX-PI-PIL.PDF.
- http://www.merck.com/product/usa/pi_circulars/r/rotateq/rotateq_pi.pdf.
- Joensuu J, Koskeniemi E, Pang XL, Vesikari T. Randomised placebo-controlled trial of rhesus-human reassortant rotavirus vaccine for prevention of severe rotavirus gastroenteritis. *Lancet*. 1997;350(9086):1205–9.
- Libster R, McNeal M, Walter EB, et al. Safety and immunogenicity of sequential rotavirus vaccine schedules. *Pediatrics*. 2016;137(2):e20152603.
- Murphy TV, Gargiullo PM, Massoudi MS, Nelson DB, Jumaan AO, Okoro CA, Zanardi LR, Setia S, Fair E, LeBaron CW, Wharton M, Livengood JR, Rotavirus Intussusception Investigation Team. Intussusception among infants given an oral rotavirus vaccine. *N Engl J Med*. 2001;344(8):564–72.
- Payne DC, Boom JA, Staat MA, et al. Effectiveness of pentavalent and monovalent rotavirus vaccines in concurrent use among US children <5 years of age, 2009–2011. *Clin Infect Dis*. 2013;57(1):13–20.
- Ruiz-Palacios GM, Pérez-Schael I, Velázquez FR, et al. Safety and efficacy of an attenuated vaccine against severe rotavirus gastroenteritis. *N Engl J Med*. 2006;354(1):11–22.
- Ruuska T, Vesikari T. Rotavirus disease in Finnish children: use of numerical scores for clinical severity of diarrhoeal episodes. *Scand J Infect Dis*. 1990;22(3):259–67.
- Simonsen L, Viboud C, Elixhauser A, Taylor RJ, Kapikian AZ. More on RotaShield and intussusception: the role of age at the time of vaccination. *J Infect Dis*. 2005;192(Suppl 1):S36–43.
- Soriano-Gabarro M, Mrukowicz J, Vesikari T, Verstraeten T. Burden of rotavirus disease in European Union countries. *Pediatr Infect Dis J*. 2006;25(Suppl):7–11.
- Svensson L, Sheshberadaran H, Vesikari T, Norrby E, Wadell G. Immune response to rotavirus polypeptides after vaccination with heterologous rotavirus vaccines (RIT 4237, RRV-1). *J Gen Virol*. 1987;68(Pt 7):1993–9.
- Velazquez FR, Matson DO, Calva JJ, Guerrero L, Morrow AL, Carter-Campbell S, et al. Rotavirus infections in infants as protection against subsequent infections. *N Engl J Med*. 1996;335(14):1022–8.
- Vesikari T, Isolauri E, D'Hondt E, Delem A, André FE, Zissis G. Protection of infants against rotavirus diarrhoea by RIT 4237 attenuated bovine rotavirus strain vaccine. *Lancet*. 1984;1:977–81.
- Vesikari T, Matson DO, Dennehy P, Van Damme P, Santosham M, Rodriguez Z, et al. Safety and efficacy of a pentavalent human-bovine (WC3) reassortant rotavirus vaccine. *N Engl J Med*. 2006;354:23–33.
- Vesikari T, Itzler R, Karvonen A, Korhonen T, Van Damme P, Behre U, Bona G, Gothefors L, Heaton PM, Dallas M, Goveia MG. RotaTeq, a pentavalent rotavirus vaccine: efficacy and safety among infants in Europe. *Vaccine*. 2009;28(2):345–51.
- Vesikari T, Van Damme P, Giaquinto C, Dagan R, Guarino A, Szajewska H, Usonis V. European Society for Paediatric Infectious Diseases consensus recommendations for rotavirus vaccination in Europe: update 2014. *Pediatr Infect Dis J*. 2015;34:635–43.
- WHO position paper. Rotavirus vaccines. *Wkly Epidemiol Rec*. 2013;88:49–64.

Hepatitis A Vaccines

Pierre Van Damme

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12.1 The Disease

Hepatitis A is a liver disease caused by the hepatitis A virus (HAV). The incubation period of hepatitis A is usually 14–28 days. Symptoms of hepatitis A range from mild to severe, and can include fever, malaise, loss of appetite, diarrhea, nausea, abdominal discomfort, dark-colored urine, and jaundice (a yellowing of the skin and whites of the eyes). Infected children under 6 years of age do not usually experience noticeable symptoms, and only 10% develop jaundice. Among older children and adults, infection usually causes more severe symptoms, with jaundice occurring in more than 70% of cases. Because of the often asymptomatic or subclinical course of hepatitis A infection, incidence rates are often underestimated. Review data from 1990 to 2005 suggest a global increase from 117 million HAV infections in 1990 to 121 million infections in 2005.

Hepatitis A sometimes relapses. The person who just recovered falls sick again with another acute episode. This is, however, followed by recovery. Unlike hepatitis B and C, hepatitis A infection does not cause chronic liver disease and is rarely fatal.

The estimated case–fatality ratio of hepatitis A varies with age and ranges from 0.1% among children <15 years of age to 0.3% among persons 15–39 years of age, and is 2.1% among adults aged ≥40 years. In Argentina, 0.4% of pediatric patients developed fulminant hepatitis, 60% of which were fatal. Recent reports from South America and the Republic of Korea have raised concerns that the incidence of fulminant hepatitis A might be rising, particularly in children.

There is no specific treatment for hepatitis A. Recovery from symptoms following infection may be slow and may take several weeks or months.

12.2 Epidemiology

Hepatitis A occurs sporadically and in epidemics worldwide, with a tendency toward cyclic recurrences. The hepatitis A virus is one of the most frequent causes of foodborne infection. The HAV persists in the environment and can withstand food production processes routinely used to inactivate and/or control bacterial pathogens.

The HAV is transmitted primarily via the fecal–oral route; that is, when an uninfected person ingests food or water that has been contami-

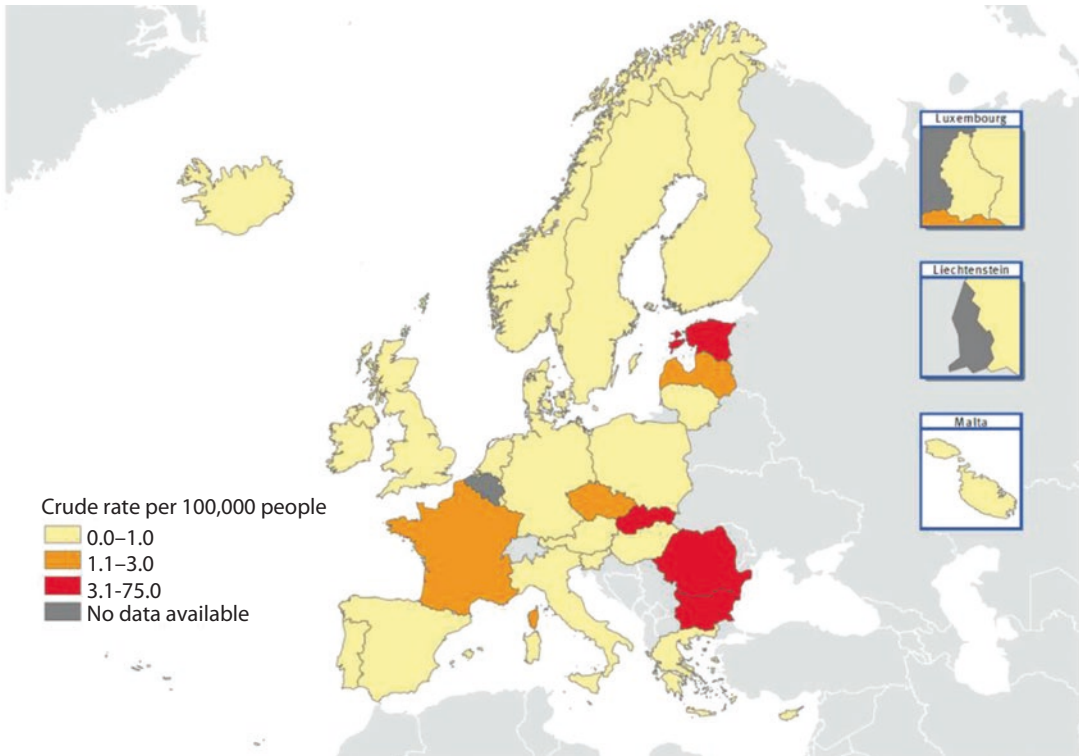
nated with the feces of an infected person. In families, this may happen when an infected person prepares food for family members with dirty hands. Waterborne outbreaks, though infrequent, are usually associated with sewage-contaminated or inadequately treated water. The virus can also be transmitted through close physical contact with an infectious person, although casual contact among people does not spread the virus.

In developing countries with poor sanitary conditions and hygienic practices, most children (90%) are infected with the HAV before the age of 10 years, mostly with no noticeable symptoms. Epidemics are uncommon because older children and adults are generally immune. Symptomatic disease rates in these areas are low and outbreaks are rare.

In middle income countries, often developing countries with transitional economies, and regions where sanitary conditions are variable, children often escape infection in early childhood and reach adolescence or adulthood without immunity. Ironically, these improved economic and sanitary conditions may lead to accumulation of adolescence and adults who have never been infected and who have no immunity. This higher susceptibility in older age groups may lead to higher disease rates and large outbreaks can occur in these communities.

In industrialized countries with good sanitary and hygienic conditions, infection rates are low. Disease may occur among adolescents and adults in high-risk groups, such as injecting drug users, men who have sex with men (MSM), people travelling to areas of high endemicity, and in isolated populations, such as closed communities. Seroprevalence and surveillance in Europe illustrate the large variability in hepatitis A endemicity across the WHO-EURO region, ranging from very low in Scandinavian countries and low in Western Europe (reaching 50% somewhere between 35 and 70 years) to intermediate and high in Central Europe and the Newly Independent States¹ (with 50% seropositivity

1 Newly Independent States (NIS): The NIS is a collective reference to 12 republics of the former Soviet Union: Russia, Ukraine, Belarus (formerly Byelorussia), Moldova (formerly Moldavia), Armenia, Azerbaijan, Uzbekistan, Turkmenistan, Tajikistan, Kazakhstan, Kirgizstan (formerly Kirghizia), and Georgia. Following dissolution of the Soviet Union, the distinction between the NIS and the Commonwealth of Independent States (CIS) was that Georgia was not a member of the CIS. That distinction dissolved when Georgia joined the CIS in November 1993.



EEA: European Economic Area; EU: European Union.

Fig. 12.1 Distribution of hepatitis A crude notification rates in EU/EEA countries, 2011 (Changing hepatitis A epidemiology in the European union ref.: Gossner et al. 2015)

reached during childhood or by the age of 20). More recent data (2005) show a further overall trend of decreasing incidence, with seroprevalence rates in Europe still increasing from west to east. Recent ECDC data based on notification from 1997 to 2011 mention a decrease from 10.0 to 2.5/100,000 population, with 21 of the 28 countries reporting rates less than or equal to 1/100,000 (Fig. 12.1).

12.3 Prevention

Improved sanitation, food safety, and immunization are the most effective ways of combating hepatitis A. The spread of hepatitis A can be reduced by adequate supplies of safe drinking water; proper disposal of sewage within communities; and personal hygiene practices such as regular hand-washing with safe water.

12.4 HAV Vaccines

Several inactivated and live attenuated vaccines against hepatitis A were developed in the 1980s and

licensed for use in the early 1990s. These vaccines are safe and well-tolerated, they are highly immunogenic, and they provide long-lasting protection against hepatitis A disease in children and adults. Four formalin-inactivated, cell-culture-produced, whole-virus vaccines are available internationally: Havrix (HM 175 strain, GlaxoSmithKline Biologicals, Rixensart, Belgium), Vaqta (CR326F strain, Merck, West Point, PA, USA), Epaxal (RG SB strain, Crucell [Janssen vaccines], Leiden, Netherlands), and Avaxim (GBM strain, Sanofi Pasteur, Lyon, France) are licensed in most parts of the world.

Other hepatitis A vaccines are produced with limited or local distribution. These include for instance a Chinese live attenuated vaccine (H2 strain, Zhejiang Academy of Medical Sciences, Hangzhou, People's Republic of China), and a vaccine manufactured by Vaccine and Bio-product Company in Vietnam since 2004.

Several types of combination vaccines containing an inactivated hepatitis A vaccine have been developed to protect individuals against more than one infectious disease when traveling

Table 12.1 Dosage and schedule for inactivated monovalent hepatitis A vaccines (in chronological order)

Vaccine	Antigen content (HAV strain)	Volume (ml)	Two-dose schedule (months)
Havrix™720 Junior	720 EI.U (HM 175)	0.5	0, 6–12
Havrix™1440 Adult	1440 EI.U (HM 175)	1	0, 6–12
Vaqa®	25 U (CR326 F)	0.5	0, 6–18
Vaqa®	50 U (CR326 F)	1	0, 6–18
Epaxal® Junior	12 IU (RG SB)	0.25	0, 6–12
Epaxal®	24 IU (RG SB)	0.5	0, 6–12
Avaxim® 80 U Pediatric	80 antigen units (GBM)	0.5	0, 6–12
Avaxim® 160 U	160 antigen units (GBM)	0.5	0, 6–12

HAV hepatitis A virus, EI.U ELISA units

to endemic countries. Such vaccines include Twinrix (GlaxoSmithKline Biologicals, Rixensart, Belgium), the only combined vaccine against both hepatitis A and hepatitis B infections, licensed since 1996; other combined vaccines include Hepatyrix (GlaxoSmithKline Biologicals, Rixensart, Belgium) and ViATIM (Sanofi Pasteur, Lyon, France), both protecting against hepatitis A and typhoid fever.

Inactivated hepatitis A vaccines all contain HAV antigen, but the content per vaccine dose is expressed in different units by various manufacturers (Table 12.1). Recommended vaccination schedules, ages for which the vaccine is licensed, and whether there is a pediatric and adult formulation also vary. All vaccines are licensed from 1 year of age in most countries. The inactivated vaccines are produced according to similar manufacturing processes involving whole-virus preparations of HAV strains growing in human MRC-5 diploid cell cultures, with subsequent virus purification and inactivation with formaldehyde. Havrix (HM175 strain), Vaqa (CR326F strain), and Avaxim (GBM strain) are adjuvanted with alum, whereas Epaxal (RG SB strain) contains a liposome adjuvant in the form of immunopotentiating reconstituted influenza virosomes (IRIV). Havrix and Avaxim contain 2-phenoxyethanol as a preservative, whereas the other vaccines are preservative-free formulations. All vaccines are administered via intramuscular injection, according to varying dosages and schedules, as described in Table 12.1.

If medically indicated, such as in hemophiliacs or in patients under anticoagulation, all four vaccines can be given subcutaneously.

12.5 Vaccine Tolerability

To date, several million doses of hepatitis A vaccines have been administered to children and adults worldwide, with no serious adverse event ever statistically linked to their use. The safety profile of inactivated hepatitis A vaccines has been extensively reviewed and results from clinical trials, and those from post-marketing surveillance studies, have demonstrated that the vaccines are all safe and well-tolerated. The most commonly reported adverse events included mild and transient local site reactions, such as pain, swelling, and redness (21% in children and 52% in adults); Epaxal has a two to three times lower rate of local reactions in comparison to alum-adsorbed hepatitis A vaccines. General reactions such as fever, fatigue, diarrhea, vomiting, and headache were reported in less than 5% of subjects.

12.6 Vaccine Immunogenicity and Protective Efficacy

The absolute minimum level of anti-HAV antibodies required to prevent HAV infection has not been defined. Experimental studies in chimpanzees have shown that low levels of passively transferred

antibody (<10 mIU/mL) obtained from vaccinated persons do not protect against infection, but do prevent clinical hepatitis and virus shedding. In the absence of an absolute lowest protective level of antibody required to prevent HAV infection, the lower limit of detection of the specific assay used in a study is generally considered as an accepted correlate of protection, i.e., 20 mIU/ml or 33 mIU/ml by ELISA in clinical studies with Havrix; 20 mIU/ml by ELISA with Avaxim and Epaxal, and 10 mIU/ml by ELISA for Vaqta.

Currently licensed inactivated hepatitis A vaccines have proven highly immunogenic in extensive clinical studies, conferring protective immunity against the disease 2–4 weeks after administration of the first dose. Recent data have shown that most individuals seroconvert within 2–4 weeks of vaccination, with rates ranging from 95–100% in children and adults. Administration of the second dose of the primary schedule (6–18 months after the first dose) ensures long-term protection. Review of the immunogenicity data for each vaccine and results from several comparative clinical trials demonstrate the equally high immunogenicity and interchangeability of hepatitis A vaccines.

The protective efficacy of inactivated hepatitis A vaccines against clinical disease has been documented in several controlled clinical efficacy trials. The cumulative protective efficacy of the vaccination course with Havrix in more than 40,000 Thai children aged 1–16 years was 95%. The observed protective efficacy of Vaqta was 100% after one vaccine dose in a trial involving more than 1,000 children aged 2–16 years from a highly endemic community in the USA. In a trial involving 274 Nicaraguan children aged 1.5–6 years, the protective efficacy of a single dose of Epaxal was also 100%.

The presence of passively transferred antibodies from previous maternal HAV infection has been shown to result in reduced antibody response to hepatitis A vaccination in infants. However, in spite of lower antibody concentrations observed after primary vaccination of infants born to anti-HAV seropositive mothers, several studies have indicated that priming and immune memory were induced, as demonstrated by the anamnestic response at the time of the booster. This was the case after a second vaccine dose administered at 12 months to 300 infants born to either anti-HAV seronegative or seropositive mothers in a study

conducted in Israel. Similarly, in a study conducted in Turkey with children who had received primary vaccination at 2, 4, and 6 months of age, all subjects showed anamnestic response after booster vaccination at 4 years of age. At 15 months of age, protective levels of antibody were also present in 93% of American Indian infants born to anti-HAV positive mothers, who had received primary immunization at 2, 4, and 6 months or at 8 and 10 months of age.

12.7 Co-administration

Such findings relating to hepatitis A vaccine immunogenicity in children younger than 2 years of age, in addition to studies showing that hepatitis A vaccine may be effectively and safely co-administered with other pediatric vaccines, such as diphtheria–tetanus–acellular pertussis, inactivated and oral polio, *Haemophilus influenzae* type b vaccine, and hepatitis B vaccines are of particular importance in the implementation of prevention strategies involving routine childhood vaccination programs. Other studies in adults have demonstrated effective and safe co-administration of hepatitis A vaccine with traveler vaccines, including hepatitis B, polio, diphtheria, tetanus, typhoid fever, yellow fever, rabies, cholera, and Japanese encephalitis.

12.8 Flexibility of Schedule

Hepatitis A vaccine has a recommended a two-dose schedule, with the second dose being administered at 6–12 months in the case of Havrix, Avaxim, and Epaxal, and at 6–18 months in the case of Vaqta. However, timing of the second dose is flexible since an anamnestic response has been shown to be triggered by a second dose when administered several years after the first vaccine dose in children and adults. Flexible two-dose vaccination schedules with a “delayed” second dose are of critical importance because travelers often miss the second dose and present some years later with a new/repeated indication for hepatitis A vaccination. In addition, a flexible schedule may help to introduce hepatitis A vaccines into established childhood routine vaccination programs. For example, a vaccination schedule for infants/children with the first dose administered during the 2nd year of life and a second dose given at

school entry at the age of 5–6 seems worth investigating. Also, additional long-term follow-up studies of individuals who have received a single vaccine dose should help to formulate future recommendations in terms of dosing schedule.

12.9 Early Protection and Duration of Protection

Hepatitis A vaccines confer early protection, as confirmed by recent data showing that most individuals seroconvert within 2 weeks of vaccination, well within the 28-day incubation period of the virus. Travelers receiving the vaccine any time before departure may thus be expected to be protected against the disease.

With regard to the duration of immunity, long-term follow-up studies have shown persistence of protective anti-HAV antibodies for at least 17 years in children and up to more than 20 years in adults, post-vaccination. Mathematical models using data from vaccinated adults have estimated protective antibodies to persist for at least 40 years in more than 90% of vaccinees.

12.10 Field Effectiveness of Routine Vaccination Programs

Hepatitis A routine immunization of young children has proven effective in rapidly reducing disease incidence, and maintaining very low incidence levels among vaccine recipients and across all other age groups, thus demonstrating the development of herd immunity, in a number of settings. A national toddler immunization program in place in Israel since 1999 has also demonstrated vaccine effectiveness, with a decrease in the annual incidence rate of hepatitis A disease from 50.4 per 100,000 (1993–1998) to 2.2–2.5 per 100,000 (2002–2004), representing more than a 95% reduction. This marked decline was seen in targeted vaccine recipients (85–90% coverage), and in all other age groups, thus demonstrating the effectiveness of hepatitis A vaccination, and the development of herd immunity. Mass vaccination programs also proved effective in localized regions of intermediate to high HAV endemicity of industrialized nations with otherwise low endemicity levels, such as the Puglia region of Italy, the Catalonia region of Spain, and in North Queensland, Australia.

In 2005, public health authorities in Argentina began a universal immunization program in 12 month-old children based on a single-dose schedule of inactivated HAV vaccine. In 2007, with vaccination coverage of 95%, the incidence of symptomatic viral hepatitis A had dropped by >80% in all age groups. Six years after implementation of this country-wide single-dose program, no hepatitis A cases have been detected among vaccinated individuals, whereas among the unvaccinated a number of cases have occurred, confirming continued circulation of HAV in the Argentinian population. An increasing number of countries in Latin America are currently implementing such a one-dose schedule.

12.11 Field Effectiveness of Post-exposure Administration and in an Outbreak Control Situation

Studies in chimpanzees, further supported by randomized trials in humans, have shown that hepatitis A vaccine is effective in preventing HAV infection when administered post-exposure. Although the post-exposure window for successful vaccination is yet to be defined, there is increasing evidence for the efficacy of hepatitis A as a valid alternative to passive post-exposure prophylaxis with immune globulin (no longer available in most countries), allowing, in particular, for a better control of outbreak situations. Results from studies conducted in chimpanzees have also shown that vaccinated animals did not shed HAV once exposed to the wild-type virus, thus demonstrating that the use of vaccines is effective at controlling the spread in the case of outbreak.

The effectiveness of hepatitis A vaccination to control outbreak situations has been reported in various settings in the USA, including rural communities from Alaska, and Europe, including Slovakia, Croatia, the UK, and Italy.

12.12 Immunization Programs

12.12.1 Risk Group Approach

Based on the transmission of HAV, several risk groups have been identified, for whom prevention by vaccination is recommended by official

Box 12.1 Summary of Current ACIP, WHO, and VHPB Recommendations for Hepatitis A Vaccination

Persons at increased risk for HAV who should be routinely vaccinated:

- Persons travelling to or working in countries that have high or intermediate endemicity of infection
- MSM
- Intravenous drug users
- Persons who have an occupational risk for infection
- Persons who have clotting factor disorders
- Day-care centre children and staff
- Persons in residential institutions
- Food handlers
- Health care workers

Vaccination of persons who have chronic liver disease:

- Susceptible persons who have chronic liver disease or who are either awaiting or have received liver transplants should be vaccinated

Hepatitis A vaccination during outbreaks:

- Vaccination for outbreak control should take into consideration the characteristics of hepatitis A epidemiology in the community and existing hepatitis A vaccination programs

Sources: ACIP, World Health Organization, VHPB.

institutions such as the World Health Organization (WHO), the Advisory Committee on Immunization Practices (ACIP) of the US Centers for Disease Control and Prevention, and the Viral Hepatitis Prevention Board. These risk groups can either be at increased risk for HAV infection (e.g., travelers to endemic regions) or have a higher probability of developing severe complications if a HAV infection were to occur (e.g., chronic liver disease patients; see ► Box 12.1).

12.13 Universal Immunization Programs

Vaccination against hepatitis A should be part of a comprehensive plan for the prevention and control of viral hepatitis. Planning for large-scale

immunization programs should involve careful economic evaluations and consider alternative or additional prevention methods, such as improved sanitation, and health education for improved hygiene practices.

Whether to include the vaccine in routine childhood immunizations depends on the local context. The proportion of susceptible people in the population and the level of exposure to the virus should be considered. Generally speaking, countries with intermediate endemicity benefit the most from the universal immunization of children. Countries with low endemicity may consider vaccinating high-risk adults. In countries with high endemicity, the use of vaccine is limited as most adults are naturally immune.

As of June 2016, a total of 16 countries used hepatitis A vaccine in the routine immunization of children nationally (including 6 countries in the American region, 3 in the Eastern Mediterranean region, 4 in the European region, and 3 in the Western Pacific region).

In the WHO-EURO region, Israel started a nation-wide universal vaccination program in 1999, thereby offering two doses of HAV vaccine to toddlers at 18 and 24–30 months of age, with coverage rates reaching 85–90%. Italy and Spain (Catalonia) have regional universal HAV vaccination programs. In Puglia, Italy, the HAV vaccine has been offered to children aged 15–18 months since 1997, and the existing hepatitis B vaccination program for 12-year-old adolescents simultaneously started using the combined vaccine against hepatitis A and B; in Catalonia, Spain, 12-year-old adolescents have also been offered the combined hepatitis A and B vaccine since 1998–1999. In addition, Greece and Turkey recently introduced a universal immunization program in toddlers.

Regarding immunization for outbreak response, recommendations for hepatitis A vaccination should also be site-specific. The feasibility of rapidly implementing a widespread immunization campaign needs to be included. Vaccination to control community-wide outbreaks is most successful in small communities, when the campaign is started early and when high coverage of multiple age groups is achieved. Vaccination efforts should be supplemented by health education to improve sanitation, hygiene practices, and food safety.

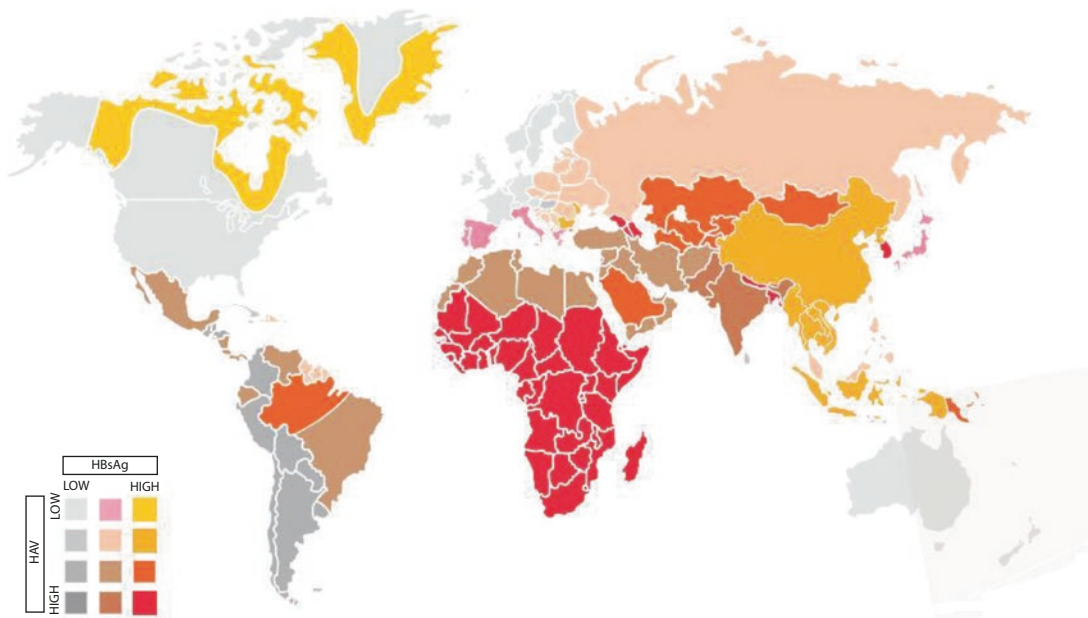


Fig. 12.2 Combined map of hepatitis B surface antigen (HBsAg; date not specified) and estimated prevalence of hepatitis A virus (HAV; 2005) (Adapted from Jacobsen and Wiersma, 2010; Plotkin and Orenstein, 2013)

12.14 Combined Hepatitis A and B Vaccine

Infections caused by the HAV and hepatitis B virus (HBV), which occur across the globe, are associated with significant morbidity and mortality, and inflict a considerable health care burden (Fig. 12.2). Vaccination is the most effective method of conferring long-term protection against both viruses and, together with improved sanitation and hygiene, has resulted in a steady reduction in global infection.

Monovalent vaccines against hepatitis A and B are immunogenic and well-tolerated with long-term immunogenic benefits observed in clinical studies with up to 20 years' follow-up. Because of the considerable overlap of risk factors and areas of high endemicity for both diseases, a combined vaccine against both viruses represents a pragmatic approach that reduces the number of vaccine administrations, in particular for travelers, patients with chronic liver disease, patients infected with HCV, or persons at increased risk of sexually transmitted infections (e.g., MSM).

Three presentations of the combined vaccine against hepatitis A and B are available (Twinrix, Twinrix Pediatric, and Ambirix; GSK Vaccines, Belgium; Table 12.2). These bivalent vaccines

Table 12.2 Three presentations of combined vaccine against hepatitis A and B

Vaccine	Target population	Formulation	Schedule
Twinrix	Adults	1.0 ml–720 EI.U HAV–20 µg HBsAg	3 doses
Twinrix pediatric	Children (1–11 years)	0.5 ml–360 EI.U HAV–10 µg HBsAg	3 doses
Ambirix	Children and adolescents (1–15 years)	1.0 ml–720 EI.U HAV–20 µg HBsAg	2 doses

HBsAg surface antigen of the hepatitis B virus

are widely available, with a safety and immunogenicity profile demonstrated as being comparable with that of the respective monovalent vaccines alone. These vaccines confer concurrent protection against the two infections while reducing the number of injections, associated costs, and other logistic issues, offering greater convenience to the vaccinee and health care provider.

After complete vaccination with these combined hepatitis A and B vaccines, the rate of

seropositivity for anti-HAV ranged from 96% to 100% in adults, children, and adolescents. The rate for seroprotection for hepatitis B surface antibody (anti-HBs) ranged from 92% to 100%, with decreasing immunogenicity response with increasing age. Immunogenicity results were equal to or higher for both anti-HAV and anti-HBs following Twinrix vaccination compared with monovalent hepatitis A and B vaccination. Long-term kinetics of the combined vaccine-induced hepatitis A and B antibodies perfectly mimics what was respectively demonstrated with the monovalent hepatitis A and B vaccines, both in terms of long-term persistence of vaccine-induced antibodies (at least 20 years shown in the adult population) and immune memory: the latter was demonstrated by mounting a strong anamnestic response after a challenge dose of HAV or HBV vaccine, indicative of the induction and persistence of immune memory.

Co-administration of Twinrix pediatric or Ambirix with other routine childhood vaccines was immunologically non-inferior to administration of the combined hepatitis A and B vaccine alone and did not significantly alter the safety profile. Safety profiles of the combined versus monovalent hepatitis A and B vaccines were similar.

Further Reading

- André F, Van Damme P, Safary A, Banatvala J. Inactivated Hepatitis A vaccine: immunogenicity, efficacy, safety and review of official recommendations for use. *Expert Rev Vaccines*. 2002;1(1):9–23.
- Bakker M, Bunge E, Marano C, de Ridder M, De Moerlooze L. Immunogenicity, effectiveness and safety of combined hepatitis A and B vaccine: a systematic literature review. *Expert Rev Vaccines*. 2016;15:829–51.
- Bell BP, Feinstone SM. Hepatitis A vaccine. In: Plotkin SA, Orenstein WA, editors. *Vaccines*. 4th ed. Philadelphia: Saunders; 2004. p. 269–97.
- Beran J, Beutels M, Levie K, Van Damme P, Dieussaert I, Gillet M, et al. A single dose, combined vaccine against typhoid fever and hepatitis A: consistency, immunogenicity and reactogenicity. *J Travel Med*. 2001;7(5):246–52.
- Dagan R, Amir J, Mijalovsky A, Kalmanovitch I, Bar-Yochai A, Thoelen S, et al. Immunization against hepatitis A in the first year of life: priming despite the presence of maternal antibody. *Pediatr Infect Dis J*. 2000;19:1045–52.
- Dagan R, Leventhal A, Anis E, Slater P, Ashur Y, Shouval D. Incidence of hepatitis A in Israel following Universal Immunization of Toddlers. *JAMA*. 2005;294:202–10.
- Gossner CM, et al. Changing hepatitis A epidemiology in the European Union: new challenges and opportunities. *Eurosurveillance*. 2015;20(16):1–6.
- Hadler SC. In: Hollinger FB, Lemon SM, Margolis HS, editors. *Global impact of hepatitis A virus infection: changing patterns*. Baltimore: Williams & Wilkins; 1991. p. 14–20.
- Hens N, Habteab Ghebretinsae A, Hardt K, Van Damme P, Van Herck K. Model based estimates of long-term persistence of inactivated hepatitis A vaccine-induced antibodies in adults. *Vaccine*. 2014;32(13):1507–13. doi:10.1016/j.vaccine.2013.10.088.
- Innis BL, Snitbhan R, Kunasol P, Laorakpongse T, Poopatanakool W, Kozik CA, et al. Protection against hepatitis A by an inactivated vaccine. *JAMA*. 1994;271:1328–34.
- Jacobsen KH, Koopman JS. Declining hepatitis A seroprevalence: a global review and analysis. *Epidemiol Infect*. 2004;132:1005–22.
- Jacobsen KH, Wiersma ST. Hepatitis A virus seroprevalence by age and world region, 1990 and 2005. *Vaccine*. 2010;28:6653–7.
- Letson GW, Shapiro CN, Kuehn D, Gardea C, Welty TK, Krause DS, et al. Effect of maternal antibody on immunogenicity of hepatitis A vaccine in infants. *J Pediatr*. 2004;144:327–32.
- Loutan L, Bovier P, Althaus B, Glück R. Inactivated virosome hepatitis A vaccine. *Lancet*. 1994;343:322–34.
- Mayorga O, Bühler S, Jaeger VK, Bally S, Hatz C, Frösner G, Protzer U, Van Damme P, Egger M, Herzog C. Single-dose Hepatitis A immunization: 7.5-year observational pilot study in Nicaraguan children to assess protective effectiveness and humoral immune memory response. *J Infect Dis*. 2016;214(10):1498–506.
- Ott JJ, Wiersma ST. Single-dose administration of inactivated hepatitis A vaccination in the context of hepatitis A vaccine recommendations. *Int J Infect Dis*. 2013;17(11):e939–44. doi:10.1016/j.ijid.2013.04.012. Review.
- Ott JJ, Irving G, Wiersma ST. Long-term protective effects of hepatitis A vaccines. A systematic review. *Vaccine*. 2012;31(1):3–11. doi:10.1016/j.vaccine.2012.04.104. Review.
- Plotkin S, Orenstein W, Offit P. *Vaccines*. 6th (ed). 2013 Saunders. ISBN: 9781455737987. <https://www.elsevier.com/books/vaccines/plotkin/978-1-4557-0090-5>
- Theeten H, Van Herck K, Van Der Meer O, Crasta P, Van Damme P, Hens N. Long-term antibody persistence after vaccination with a 2-dose Havrix (inactivated hepatitis A vaccine): 20 years of observed data, and long-term model-based predictions. *Vaccine*. 2015;33(42):5723–7. doi:10.1016/j.vaccine.2015.07.008.
- Van Damme P, Van Herck K. A review of the efficacy, immunogenicity and tolerability of a combined hepatitis A and B vaccine. *Expert Rev Vaccines*. 2004a;3(3):249–67.
- Van Damme P, Van Herck K. A review of the efficacy, immunogenicity and tolerability of a combined hepatitis A and B vaccine. *Expert Rev Vaccines*. 2004b;3(3):249–67.
- Van Damme P, Van Herck K. Effect of hepatitis A vaccination programs. *JAMA*. 2005;294:246–8.
- Van Damme P, Thoelen S, Cramm M, De Groote K, Safary A, Meheus A. Inactivated hepatitis A vaccine: reactogenicity, immunogenicity, and long-term antibody persistence. *J Virol Med*. 1994;44(4):446–51.

- Van Damme P, Banatvala J, Fay O, Iwarson S, McMahon B, Van Herck K, et al. Consensus statement: hepatitis A booster vaccination: is there a need? *Lancet*. 2003;362:1065–71.
- Van Damme P, Leroux-Roels G, Crasta P, Messier M, Jacquet JM, Van Herck K. Antibody persistence and immune memory in adults, 15 years after a three-dose schedule of a combined hepatitis A and B vaccine. *J Med Virol*. 2012;84(1):11–7. doi:10.1002/jmv.22264.
- Van Damme P, Leroux-Roels G, Suryakiran P, Folschweiller N, Van Der Meeren O. Persistence of antibodies 20 y after vaccination with a combined hepatitis A and B vaccine. *Hum Vaccin Immunother*. 2017;13:972–80.
- Vidor E, Dumas R, Porteret V, Bailleux F, Veitch K. Aventis Pasteur vaccines containing inactivated hepatitis A virus: a compilation of immunogenicity data. *Eur J Clin Microbiol Infect Dis*. 2004;23(4):300–9.
- Viral Hepatitis Prevention Board. Hepatitis A: disease and epidemiology. *Hepatitis A update*. *Viral Hepatitis*. 1997;6: 5–14.
- Wasley A, Samandari T, Bell B. Incidence of hepatitis A in the United States in the era of vaccination. *JAMA*. 2005;294:194–201.
- Werzberger A, Mensch B, Kuter B, Brown L, Lewis J, Sitrin R, et al. A controlled trial of a formalin-inactivated hepatitis A vaccine in healthy children. *N Engl J Med*. 1992;327(7):453–7.
- World Health Organization. Hepatitis A vaccines WHO position paper. *Weekly Epidemiology Record*. 2012;875:261–76. <http://www.who.int/wer>.

Hepatitis B Vaccines

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13.1 The Disease

Hepatitis B virus, previously called the Dane particle, is a 42-nm DNA virus that belongs to the *Hepadnaviridae* family. HBV is primarily hepatotropic and the liver damage is produced by the cellular immune response to viral proteins in infected hepatocytes. Infection with HBV causes a broad spectrum of liver disease, including sub-clinical infection, acute, clinically overt self-limited hepatitis, and fulminant hepatitis. The clinical manifestations of acute hepatitis B are indistinguishable from other causes of viral hepatitis; a definitive diagnosis requires serological testing. The average incubation period is 90 days (range, 60–150 days) from exposure to onset of jaundice, and 60 days (range, 40–90 days) from exposure to onset of abnormal alanine aminotransferase (ALT) levels. Persons infected with HBV can also develop persistent infection, which can lead to chronic liver disease and death from cirrhosis or hepatocellular carcinoma (HCC). The age at acquisition of HBV infection is the main determining factor in the clinical expression of acute disease and the development of chronic infection (■ Fig. 13.1). Fewer than 10% of children younger than 5 years who become infected have initial clinical signs or symptoms of disease (i.e., acute hepatitis B), compared with 30–50% of older children and adults. The risk for developing

chronic HBV infection varies inversely with age: approximately 90% of infants infected during the 1st year of life develop chronic infection, compared with 30% of children infected between the ages 1 and 4 years and less than 5% of persons infected as adults.

Persons who have persistence of HBsAg in serum for at least 6 months are classified as having chronic infection. HBV replication persists throughout the course of chronic HBV infection, and the natural history of chronic HBV infection is determined by the interaction between virus replication and host immune response. Persons with chronic HBV infection are at a high risk for developing HCC.

13.2 Burden of Hepatitis B

Hepatitis B virus infection is a highly prevalent infection around the globe, the frequency and burden of which vary by region and subpopulation. Approximately 30% of the world's population (i.e., about 2 billion persons) have serological evidence for HBV infection, and of these, more than 240 million persons are living with chronic infection.

Hepatitis B virus causes significant morbidity and mortality worldwide. In 2013, approximately 686,000 HBV-infected persons died from causes

■ Fig. 13.1 Studies evaluating the risk for chronic hepatitis B virus infection by age at infection. *Filled squares* represent data from developing countries; *open squares* represent data from developed countries (From Edmunds et al. 1993, with permission)

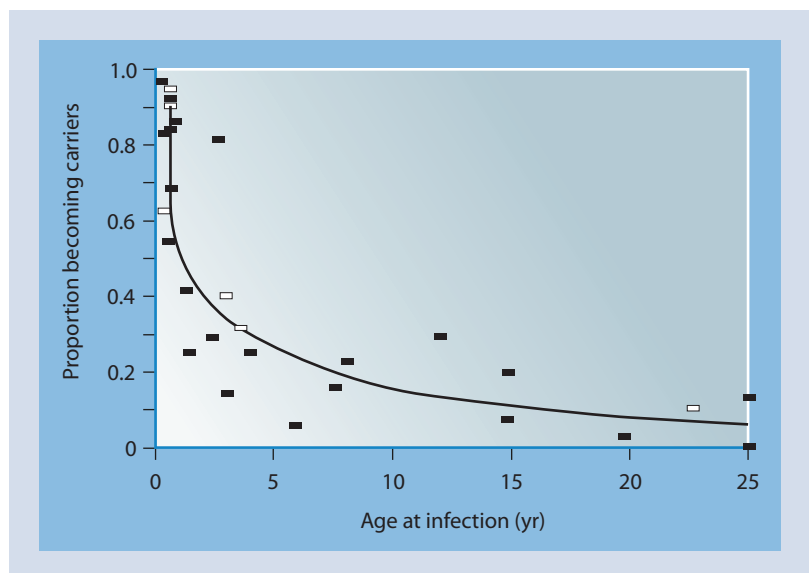
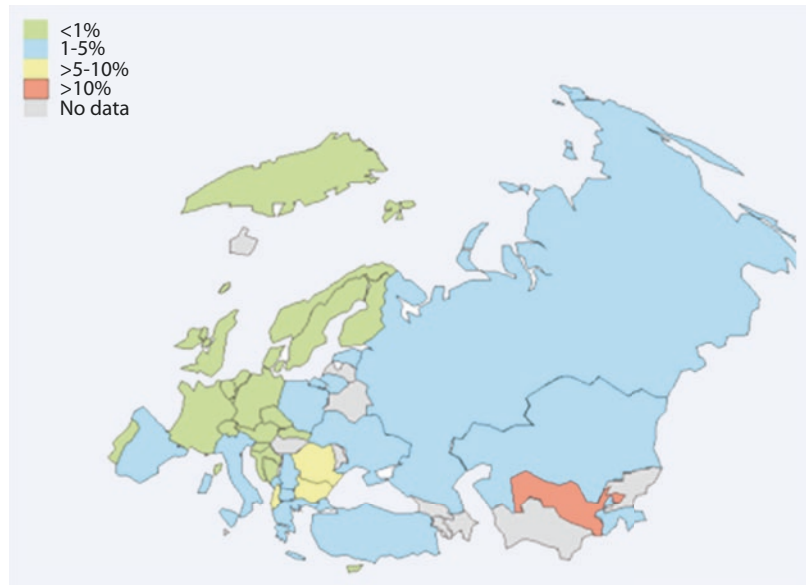


Fig. 13.2 Endemicity of hepatitis B in Europe (WHO EURO Region) (Estimated prevalence of carriage of hepatitis B surface antigen, WHO European Region, 2013 ▶ <http://www.euro.who.int/en/health-topics/communicable-diseases/hepatitis/data-and-statistics/fact-sheet-hepatitis-b>)



related to acute infection (69,000 deaths), cirrhosis (317,000 deaths), and HBV-associated liver cancer (300,000 deaths), the sixth most common cancer globally and the second leading cause of cancer-related death worldwide. Of all cases of primary liver cancer, 70–90% are caused by HCC, of which HBV is a major cause. The lifetime risk for HCC in a chronically infected person is approximately 10–25%, which is 15–20 times greater than that for persons without HBV infection.

Approximately 13 million people in the WHO European Region are chronically infected with hepatitis B, leading to approximately 60,000 deaths per year from hepatitis B-related liver cancer and cirrhosis. The epidemiology of hepatitis B in the Region is diverse, with a prevalence of hepatitis B surface antigen ranging from extremely low (<0.1% in Hungary) to high (13% in Uzbekistan;

▶ Fig. 13.2).

13.3 Epidemiology

Hepatitis B virus is transmitted by percutaneous (i.e., puncture through the skin) or mucosal (i.e., direct contact with mucous membranes) exposure to infectious blood or body fluids. All hepatitis B 's' antigen (HBsAg)-positive persons are potentially infectious, but those who are also hepatitis B 'e' viral protein (HBeAg)-positive are *more* infectious

because their blood contains high concentrations of HBV (typically 10^7 – 10^9 virions/ml). Although HBsAg has been detected in multiple body fluids, only serum, saliva, semen, and vaginal fluid have been demonstrated to be infectious. Primary sources of HBV infection are perinatal exposure from infected mothers, nonsexual person-to-person contact, sexual contact, and percutaneous exposure to blood or infectious body fluids. HBV is not transmitted by air, food, or water. The frequency and patterns of HBV transmission vary markedly in different parts of the world. In highly endemic countries, most infections are acquired during the perinatal period and early childhood, when the risk for developing chronic infection is greatest. In areas of intermediate endemicity, the lifetime risk for HBV infection is 20–60%, and infections occur in all age groups. Most HBV infections in areas of low endemicity, such as Europe, occur in adults in relatively well-defined risk groups, but a high proportion of chronic infections may occur as a consequence of perinatal and early childhood exposures. Persons considered at risk for hepatitis B are: persons at risk through sexual exposure, contacts of persons with chronic HBV infection, hemodialysis patients, incarcerated persons, injection-drug users, persons at risk for occupational exposure, developmentally disabled persons in long-term care facilities, and travelers to regions with moderate or high HBV endemicity.

13.4 Prevention of Hepatitis B

13.4.1 Passive Immunization

A major use of hepatitis B immune globulin (HBIG; a specific immune globulin containing high concentrations of anti-HBs) is as an adjunct to hepatitis B vaccine in preventing perinatal HBV transmission. Untreated, 70–90% of infants born to HBeAg-positive mothers become infected at birth and develop chronic HBV infection. Immunoprophylaxis with both HBIG and hepatitis B vaccine confers an efficacy of preventing perinatal HBV transmission from 85% to 95% and provides long-term protection. The standard dose of HBIG is 0.5 ml for post-exposure prophylaxis of infants born to HBsAg-positive mothers and 0.06 ml/kg for all other indications. HBIG should be administered intramuscularly and may be administered simultaneously with hepatitis B vaccine, but at a different injection site. HBIG is also recommended for post-exposure prophylaxis (often in combination with hepatitis B vaccine) in specific settings.

13.4.2 Hepatitis B Vaccines

The first available vaccines were produced by harvesting HBsAg (the 22-nm particle) from the plasma of persons with chronic HBV infection, the so-called plasma-derived vaccines. Nowadays, these vaccines are no longer on the market. The development of recombinant DNA technology to express HBsAg in other organisms offered the potential to produce unlimited supplies of vaccine, and recombinant DNA vaccines have now completely replaced the plasma-derived vaccines. Hepatitis B vaccines are formulated to contain 2.5–40 µg of HBsAg protein and an aluminum phosphate or aluminum hydroxide adjuvant: 0.25 mg in pediatric dose vaccines, and 0.5 mg in adult dose vaccines.

13.4.3 Combination Vaccines

Several vaccine manufacturers have produced combination vaccines containing a hepatitis B vaccine component. These combination vaccines include diphtheria and tetanus toxoids and whole-cell

pertussis (DTwP)–hepatitis B vaccine; DTwP–*Haemophilus influenzae* type b conjugate (Hib)–hepatitis B vaccine; diphtheria and tetanus toxoids and acellular pertussis (DTaP)–hepatitis B vaccine; DTaP–Hib–inactivated poliovirus vaccine (IPV)–hepatitis B vaccine; DTaP–IPV–hepatitis B vaccine; Hib–hepatitis B vaccine; and hepatitis A–hepatitis B vaccine. For each of these combination vaccines, the manufacturer has shown that the components remain sufficiently immunogenic to elicit protective levels of anti-HBs (see ► Chap. 20).

13.4.4 Dosage and Route of Administration

The quantity of HBsAg protein per dose that induces a protective immune response in infants and children varies by manufacturer (range 2.5–10 µg) and by composition of the envelope protein(s), and is partially related to the vaccine production processes. In general, the vaccine dosage for infants and adolescents is 50% lower than that required for adults. There is no international standard of vaccine potency expressed in micrograms of HBsAg protein.

13.4.5 Vaccine Immunogenicity and Schedules

Historically, the standard three-dose hepatitis B vaccine series has consisted of two priming doses administered 1 month apart and a third dose administered 6 months after the first dose. Multiple schedules have been used successfully: at birth and at 1 and 6 months of age; at 2, 4, and 6 months; at 3, 5 and 11 months; at 8, 12, 16 weeks, and 12 or 15 months; and at 6, 10, and 14 weeks (in the World Health Organization's [WHO's] Expanded Program on Immunization [EPI] schedule).

13.4.6 Infants and Children

A variety of hepatitis B vaccine schedules have been shown to induce levels of seroprotection of greater than 95% in infants (see ► Sect. 4.5). Programmatically, there is an advantage to administering the three doses of hepatitis B vaccine

at the same time as the three doses of other childhood vaccines (e.g., DTP, Hib, IPV), and these schedules accommodate the use of DTP-, IPV- and Hib-containing combination vaccines. To prevent perinatal HBV transmission in settings where combination vaccines are used, a four-dose hepatitis B vaccination schedule is needed, with the first dose administered at birth. Use of four-dose hepatitis B vaccine schedules, including schedules with a birth dose, has not increased vaccine reactogenicity. Certain premature infants with low birthweights (i.e., <2,000 g) may have decreased seroconversion rates after administration of hepatitis B vaccine at birth. However, by the age of 1 month, all premature infants, regardless of initial birthweight or gestational age, have a response to vaccination that is comparable to that of full-term infants.

13.4.7 Adolescents

Hepatitis B vaccine schedules that have been demonstrated to induce seroprotection rates of greater than 95% in adolescents include doses administered at 0, 1, and 6 months; 0, 2, and 4 months; and 0, 12, and 24 months. In addition, for adolescents aged 11–15 years, the adult dose of hepatitis B vaccine can be used for administration at 0 and at 4–6 months. This two-dose schedule produces anti-HBs concentrations equivalent to those obtained with the pediatric dose administered on a three-dose schedule.

13.4.8 Adults

Hepatitis B vaccine induces a protective antibody response in approximately 30–55% of healthy adults aged less than 40 years after the first dose, in 75% after the second dose, and in more than 90% after the third dose. In adults older than 40 years, response rates decline with age, and by age 60 years, protective levels of antibody develop in only 75% of vaccinated persons.

13.4.9 Correlates of Protection

An anti-HBs concentration of 10 mIU/ml or more measured 1–3 months after administration of the last dose of the primary vaccination

series is considered a reliable marker of protection against infection. In vaccine efficacy studies, immunocompetent persons who developed anti-HBs concentrations of 10 mIU/ml or higher after vaccination had virtually complete protection against both acute disease and chronic infection for decades, even if subsequently, over time, anti-HBs concentrations declined to less than 10 mIU/ml. Indeed, the protective efficacy of hepatitis B vaccination is related to the induction of anti-HB antibodies, but it also involves the induction of memory B and T cells. Routine postvaccination testing for immunity is not necessary, but it is recommended for high-risk persons whose subsequent clinical management depends on knowledge of their immune status. Persons at increased risk for hepatitis B found to have anti-HBs concentrations of less than 10 mIU/ml after the primary vaccine series should be revaccinated. Administration of three doses on an appropriate schedule, followed by anti-HBs testing 1–2 months after the third dose, is usually more practical than serological testing after one or more doses of vaccine.

13.4.10 Duration of Protection and Need for Booster Doses

After primary immunization with hepatitis B vaccine, anti-HBs concentrations decline rapidly within the first year and more slowly thereafter. Among children who respond to a primary three-dose vaccination series with anti-HB concentrations of 10 mIU/ml or greater, 15–50% have low or undetectable concentrations of anti-HBs 5–15 years after vaccination. Protection has been shown to outlast the presence of vaccine-induced antibodies, conferring effective long-term protection against acute disease and development of HBsAg carriage for a minimum of 25–30 years. Based on currently available scientific evidence, the WHO, in addition to advisory groups in the USA and Europe do not recommend routine booster doses of hepatitis B vaccine or periodic serological testing to monitor anti-HBs concentrations for immunocompetent persons who have responded to vaccination or in universal immunization programs.

13.4.11 Vaccine-Associated Adverse Events

Adverse events after immunization against hepatitis B are infrequent and generally mild. With the exception of localized pain, placebo-controlled studies have revealed that reported events (e.g., myalgia and transient fever) occur no more frequently among vaccinees than among persons receiving placebo (<10% among children, 30% among adults). Data from numerous long-term studies fail to causally link serious adverse events to hepatitis B vaccination. Reports of severe anaphylactic reactions are very rare, and data do not indicate a causal association between hepatitis B vaccine and Guillain-Barré syndrome or demyelinating disorders, including multiple sclerosis.

Hepatitis B vaccine is contraindicated only for persons with a history of allergic reactions to yeast or any of the vaccine's components. Neither pregnancy nor lactation is a contraindication for use of this vaccine. Both premature infants and HIV-positive persons can receive this vaccine.

13.5 Recommendations for Hepatitis B Vaccination

13.5.1 Vaccination of Infants at Birth

Because perinatal and early postnatal transmission are primary causes of chronic infections globally, the first dose of hepatitis B vaccine should be given as soon as possible (<24 h) after birth, regardless of whether a country has low, intermediate, or high HBV endemicity. Some countries augment universal vaccination of newborns with maternal screening for HBsAg and the administration of HBIG and a dose of hepatitis B vaccine to infants born to HBsAg-positive mothers. Among infants born to HBsAg-positive mothers, a birth dose of hepatitis B vaccine reduces the risk for perinatal HBV transmission by 72% and by >90% when combined with HBIG. The timely delivery of a birth dose of hepatitis B vaccine should be a performance measure for all immunization programs.

13.5.2 Full Immunization of Infants by Routine Immunization Programs

To complete the primary hepatitis B vaccine series, the birth dose should be followed by two or three additional doses of vaccine administered at least 4 weeks apart. To help ensure completion of the vaccine series, doses should be given concurrently with DTP or other routine infant vaccinations. For older children and adults, the primary series of three doses with appropriate intervals applies.

13.5.3 Public Health Considerations and the Impact of Worldwide Hepatitis B Vaccination Programs

Routine infant immunization has become a long-standing practice in more than 95% of countries, providing evidence for the effectiveness of hepatitis B immunization in significantly reducing or eliminating HBV transmission. In general, studies conducted in areas of high HBV endemicity have demonstrated declines in the prevalence of chronic HBV among children to less than 2% after routine infant immunization. Countries that adopted and implemented universal hepatitis B immunization early include Taiwan (1984), Bulgaria (1989), Malaysia (1990), the Gambia (1990), Italy, Spain, the USA (1991), and Israel (1992).

Taiwan is perhaps the best example of an area with previously high endemicity showing a substantial decrease in the burden of hepatitis B and HBV-related diseases after the 1984 mass vaccination of newborns. The HBsAg prevalence in individuals less than 20 years of age decreased from 9.8% in 1984 to 1.3% in 1994, and to 0.6% in 2004. The annual average incidence of HCC among children aged 6–14 years decreased from 0.7 per 100,000 in 1981 through 1986, to 0.36 per 100,000 in 1990 through 1994. In 2004, the HCC incidence for age groups of 6–9, 10–14, and 15–19 years decreased to 0.15, 0.19, and 0.16 per 100,000 person-years respectively, clearly indicating the hepatitis B vaccine to be the first vaccine against a major human cancer.

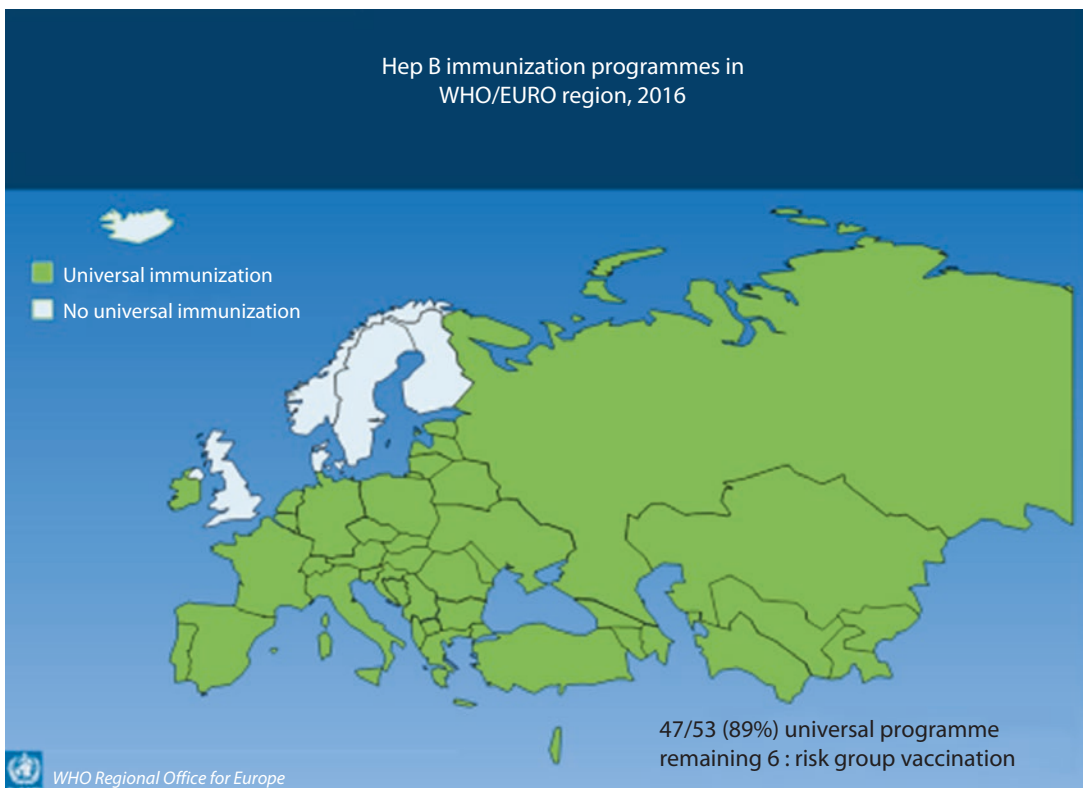
Surveillance data from Italy, where universal vaccination started in 1991 in infants and in adolescents, have shown a clear overall decline in the incidence of acute hepatitis B, from 5/100,000 in 1990 to 0.9/100,000 in 2010. This decline was even more striking in individuals aged 15–24 years, in whom the morbidity rate per 100,000 inhabitants fell from 17 in 1990 to less than 0.5 in 2010. Moreover, a generation of children and young adults (at present, 32 age cohorts in 2011) is emerging with virtually no markers of HBV infection.

13.5.4 Introduction of Hepatitis B Vaccination Programs

Despite the availability of hepatitis B vaccine since the early 1980s, barriers at that time impeded efforts to immunize infants and children against hepatitis B, for example, high vaccine costs, ill-founded concerns about using a plasma-derived vaccine during the first years of the AIDS epidemic, and the lack of global vaccine policies. By 1991, only 20 countries had implemented routine

infant immunization against hepatitis B. In the following decades, hepatitis B vaccination coverage grew rapidly and by 2010, hepatitis B vaccination coverage among infants had reached an estimated 75% worldwide. By the end of 2014, a total of 184 countries have integrated hepatitis B vaccine into their national childhood immunization systems. Global coverage with three doses of hepatitis B vaccines is estimated at 82% and is as high as 92% in the Western Pacific.

In the WHO European Region as of 2012, a total of 47 of the 53 European countries (89%) had implemented a universal hepatitis B vaccination program (■ Fig. 13.3). The most recent countries to follow the recommendation were Ireland (in 2008), the Netherlands (in 2011), and the UK (expected for 2017). Still, five countries (Denmark, Finland, Iceland, Norway, and Sweden) adopt vaccination targeting risk groups only, instead of adding a universal vaccination program. However, changing demography, increasing immigration, and the current vaccine costs make the cost–benefit ratios in these remaining low endemicity countries strongly in favor of universal HBV vaccination.



■ Fig. 13.3 Universal hepatitis B immunization programs in the WHO European Region, 2014

On 17 September 2014, the Regional Committee for Europe approved a resolution, EUR/RC64/R5, to adopt the “European Vaccine Action Plan 2015–2020” (EVAP), which defines a regional vision and goals for immunization and control of vaccine-preventable diseases and outlines priority actions to achieve them. One of the major EVAP goals is to strengthen hepatitis B control through immunization. The EVAP states “The Regional Office commits itself to prepare a program and action plan for the control of hepatitis B infection and identify targets for 2020.” The overriding vision of hepatitis B control is that the children in all countries in the Region would be free of chronic hepatitis B infection, defined as an HBsAg prevalence of 0.5% or lower in children 5–10 years of age by 2020.

Further Reading

- Banatvala JE, Van Damme P. Hepatitis B vaccine: do we need boosters? *J Viral Hepat.* 2003;10:1–6.
- Beasley RP. Hepatitis B virus: the major etiology of hepatocellular carcinoma. *Cancer.* 1988;61:1942–56.
- Beasley RP, Hwang LY, Lin CC, et al. Hepatocellular carcinoma and hepatitis B virus: a prospective study of 22 707 men in Taiwan. *Lancet.* 1981;2:1129–33.
- Blumberg BS, Alter HJ, Visnich S. A “new” antigen in leukemia sera. *JAMA.* 1965;191:541–6.
- Dane DS, Cameron CH, Briggs M. Virus-like particles in serum of patients with Australia-antigen-associated hepatitis. *Lancet.* 1970;1:695–8.
- Edmunds WJ, Medley GF, Nokes DJ, et al. The influence of age on the development of the hepatitis B carrier state. *Proc Biol Sci.* 1993;253:197–201.
- European Consensus Group on Hepatitis B Immunity. Are booster immunisations needed for lifelong hepatitis B immunity? *Lancet.* 2000;355:561–5.
- European Vaccination Action Plan., available at: ► <http://www.euro.who.int/en/health-topics/disease-prevention/vaccines-and-immunization/publications/2014/european-vaccine-action-plan-20152020>. Accessed 4 Sept 2015.
- Goldstein ST, Zhou F, Hadler SC, et al. A mathematical model to estimate global hepatitis B disease burden and vaccination impact. *Int J Epidemiol.* 2005;34:1329–39.
- Hilleman MR, Buynak EB, Roehm RR, et al. Purified and inactivated human hepatitis B vaccine: progress report. *Am J Med Sci.* 1975;270:401–4.
- Hurie MB, Mast EE, Davis JP. Horizontal transmission of hepatitis B virus infection to United States-born children of Hmong refugees. *Pediatrics.* 1992;89:269–73.
- Institute of Medicine. Immunization safety review: hepatitis B vaccine and demyelinating neurological disorders. www.iom.edu/Reports/2002/Immunization-Safety-Review-Hepatitis-B-Vaccine-and-Demyelinating-Neurological-Disorders.aspx. May 2002.
- Jilg W, Schmidt M, Deinhardt F. Vaccination against hepatitis B: comparison of three different vaccination schedules. *J Infect Dis.* 1989;160:766–9.
- Langer-Gould A, Qian L, Tartof S, et al. Vaccines and the risk of multiple sclerosis and other central nervous system demyelinating diseases. *JAMA Neurol.* 2014. doi:10.001/jamaneurol.2014.2633.
- Lernout T, Hendrickx G, Vorsters A, et al. A cohesive European policy for hepatitis B vaccination, are we there yet? *Clin Microbiol Infect.* 2014;20(Suppl 5):19–24.
- Leuridan E, Van Damme P. Hepatitis B and the need for a booster dose. *Clin Infect Dis.* 2011;53:68–75.
- Romano L, Paladini S, Van Damme P, et al. The worldwide impact of vaccination on the control and protection of viral hepatitis B. *Dig Liver Dis.* 2011;43(Suppl 1):S2–7.
- Stevens CE, Taylor PE, Tong MJ, et al. Yeast-recombinant hepatitis B vaccine: efficacy with hepatitis B immune globulin in prevention of perinatal hepatitis B virus transmission. *JAMA.* 1987;257:2612–6.
- Trépo C, Ghan HL, Lok A. Hepatitis B virus infection. *Lancet.* 2014;384:2953–63.
- Van Damme P, Leuridan E, Hendrickx G, Vorsters A, Theeten H, Leino T, Salminen M, Kuusi M. Should Europe have a universal hepatitis B vaccination programme? *BMJ.* 2013;347:f4057. doi:10.1136/bmj.f4057.
- World Health Organization. Hepatitis B vaccines. *Wkly Epidemiol Rec.* 2009;84:405–19.

Influenza Vaccines

Timo Vesikari and Susanna Esposito

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Influenza is caused by influenza viruses that belong to the *Orthomyxoviridae* family and have a segmented RNA genome. Influenza virus types A and B cause more than 99.9% of all the influenza cases that occur every winter season in countries with a temperate climate; influenza type C is not a significant pathogen. The incidence varies from year to year, as group A viruses may change the prevalent subtype (e.g., from H1N1 to H3N2 or vice versa), or because of antigenic “drift” within the subtype. Point mutations on genes encoding the two surface proteins of influenza viruses, hemagglutinin (HA) and neuraminidase (NA), are called antigenic drift and allow the viruses to evade immune defenses developed by individuals as a result of previous infections or vaccination. Variability due to antigenic drift is significantly more common among A viruses, in particular, the A/H3N2 subtype. Influenza B viruses are more stable with regard to antigenic drift, but they frequently switch the prevalent lineage for the epidemic season (see below). Major mutations (antigenic shift) that occur only in influenza A viruses by reassortment of the RNA genome can cause pandemics because previous immunity is not effective against such a completely different virus. Examples of antigenic shift are the emergence of “Asian influenza” in 1957 (H2N2), “Hong Kong flu” in 1968 (H3N2), and “swine flu” in 2009 (H1N1sw or H1N1pdm09).

14.1 Influenza in the Pediatric Age

Influenza causes medical, social and economic problems in children younger than 5 years of age, the elderly, pregnant women, and individuals with severe chronic medical conditions independently of age. Approximately 5% to 15% of the world population suffer from seasonal influenza every year, with three to five million severe cases and more than 500,000 deaths. Medical visits, hospitalization rates, admissions to the intensive care unit, and the prescription of drugs, antipyretics, and antibiotics are increased during influenza season, with a related impact on health care expenditure. School absenteeism not only has an impact on children, but contributes to an average loss of 3 working days for the parent, who must remain at home with the child.

Children have the highest incidence of influenza each year. Children also shed the virus in greater amounts and for a longer time than older subjects, and are considered the main contributors to the transmission of infection in the community. Although influenza in children is frequently a mild respiratory infection, it has been clearly demonstrated that influenza in healthy children may be very severe and lead to death. In a study carried out in the USA on the influenza seasons from October 2004 to September 2012, during which 830 pediatric influenza-associated deaths were reported, it was found that 43% of children who died had no high-risk medical conditions. Moreover, contrary to what was generally thought, influenza was found potentially severe not only in children younger than 5 years, but also in older children and adolescents. Although the highest risk of death was associated with the first years of life (including the first 6 months), a large number of deaths occurred in children aged over 5 years.

Pregnant women are at-risk for severe influenza, its complications, and death. Vaccination during pregnancy is safe and well-tolerated, does not induce fetal complications and is highly effective in reducing the risk of influenza in young infants up to the age of 6 months (■ Fig. 6.2).

In the USA, the recommendation is to vaccinate all children from the age of 6 months. In Europe, only Finland and the UK have influenza vaccination as part of the national immunization program. In Finland, the program is for 6- to 36-month-old children and in the UK (using intranasal vaccine) from age 2 years up. Reduction of the burden of influenza in children can be obtained only by vaccination. Two different types of influenza vaccines are presently available: inactivated influenza vaccines (IIVs), which are given via the intramuscular and intradermal routes, and live attenuated influenza vaccines (LAIVs), which are given intranasally.

14.2 Nonlive Influenza Vaccines

The technology for the first influenza vaccines dates back to 1941 and used whole influenza viruses grown in embryonated eggs and inactivated by formalin. Whole viruses have been largely replaced by split-virion vaccines or sub-

unit (HA and NA) vaccines. Most of the influenza vaccines used in the world are still based on egg-grown virus, but are split virion or subunit types. Cell culture-grown influenza vaccines have also been licensed, but are in the minority.

Until 2013, IIVs contained three inactivated (or split or subunit) viruses: representatives of type A/H1N1 and type A/H3N2, and one of the two genetic lineages of type B virus (Yamagata or Victoria), which had been recognized since the 1990s. Such a combination is called trivalent influenza vaccine (TIV). Specific strains to be included in the vaccine formulation are chosen every year by the WHO considering the epidemiology of virus circulation in the previous year. Inaccurate prediction of the predominant influenza B lineage left many vaccinated individuals with suboptimal protection against influenza B disease caused by the influenza B lineage not being included in the TIV. In Europe, a B-mismatch between vaccine and circulating strains occurred in 5 out of 10 seasons between 2001 and 2011. This led to a modification of the conventional composition of the influenza vaccine with the inclusion of both B lineages for a quadrivalent (or tetravalent) influenza vaccine (QIV). The quantity of HA in the vaccine is usually 15 µg per antigen. Thus, TIV contains a total of 45 µg and QIV 60 µg of HA.

Cell culture produced IIVs are not more efficacious than egg-based vaccines, but cell culture is seen as a competitive production platform for the future. Moreover, cell-culture IIVs allow the problem of egg allergy to be overcome, although the risk of severe reactions following administration of traditionally prepared IIVs to patients with an egg allergy is very low. A recent study found that the rate of anaphylaxis after all influenza vaccines, including both IIVs and LAIVs, was only 1.31 per 1 million vaccine doses given. Consequently, it was stated that influenza vaccine may be administered to patients with previous egg-associated hives without any precaution. Persons who report symptoms other than hives, such as angioedema, respiratory distress, lightheadedness, or recurrent emesis; or who required epinephrine or another emergency medical intervention, may still receive influenza vaccine under close control.

Conventional TIV (or QIV) is effective, but has limitations. It has been calculated that on

average prevention of influenza occurs in about 60% of vaccinated healthy adults when the circulating viruses match those in the vaccine and in approximately 40% in case of virus mismatch. Protection in older children is similar and lower in young children. Naïve children 6–36 months of age have a moderately good response to the influenza A components delivered in two half doses of TIV, but the response to B viruses is lower. All responses are poor in immunocompromised subjects or subjects with a severe chronic underlying disease with some degree of immune system deficiency. Further limitation of IIVs is the inability to evoke high antibody titers against heterovariant viruses with resulting low protection in the case of mismatch between circulating strains and strains included in the vaccines.

Usually, a half dose (7.5 µg HA) is given to infants and children aged 6–36 months and the full dose (15 µg of each HA) is used for older children. The recommendation is to give two injections to vaccine-naïve children and a single dose annually thereafter.

A higher dose (full adult dose) of HA of each virus included in the vaccine yields a better response and greater protection in the age group 6–36 months (■ Table 14.1), but is not approved by the regulatory authorities. Regardless, Finland is recommending the full adult dose for its program in 6- to 36-month-old children. However, even with a higher dose, the protection against B-strains remains low.

■ **Table 14.1** Adult dose trivalent influenza vaccine (TIV) for young children in the Finnish National Immunization Program

Full dose TIV effectiveness by the strain

	Influenza A	Influenza B
All children	84% (40–96)	45% (–34–78)
≤2 years of age	79% (21–95)	28% (–212–84)

From Heinonen et al. (2011)
Full-dose TIV for children aged 6–36 months (off-label) is efficacious against well-matched A-strains, but not B-strains

All influenza vaccine recommendations for children start at 6 months of age. For the protection of infants aged <6 months the only available option is maternal immunization in pregnancy (see Fig. 6.2).

14.3 Adjuvanted IIVs

Oil-in-water emulsion adjuvants increase the immunogenicity of IIVs. The best-known adjuvant is MF59, which contains squalene. MF59 has been extensively studied and is currently licensed for use in the elderly in many EU countries. The MF59-adjuvanted trivalent seasonal influenza vaccine (aTIV) has been evaluated in young children for immunogenicity, safety, and efficacy. MF59-adjuvanted vaccine was safe and well-tolerated with only a small, clinically marginal increase in local adverse events. aTIV was highly efficacious in all children under 6 years of age and significantly more efficacious than a TIV comparator (Fig. 14.1). In the specific age group 6–24 months, aTIV was efficacious whereas TIV was not. Immune responses against B strains were high after two doses of aTIV. MF59 adjuvant also increases the heterovariant immune responses to A strains not included in the vaccine.

Despite the promising results, aTIV was not licensed in the EU for children. However, studies have been continued with a quadrivalent formula-

tion of MF59-adjuvanted vaccine (aQIV), and there is a reasonable expectation that this vaccine may be licensed for EU children in the near future.

Virosomes, which are reconstituted viral envelopes including membrane lipids and viral spike glycoproteins, but devoid of viral genetic material, were used for preparation of adjuvanted influenza vaccines until few years ago. Several studies showed a significant improvement in the immune response in comparison with conventional IIVs in subjects of any age. However, the virosome vaccine was withdrawn from the market, mainly because its administration in younger children was followed by high fever in a non-negligible number of subjects.

14.4 Pandemic H1N1sw Vaccine and Narcolepsy

In 2009, with the emergence of “swine flu” of the H1N1sw pandemic, vaccines against this strain were hastily produced and implemented with a minimal delay. Conventional split virion or subunit vaccines were not sufficiently immunogenic, whereas whole-virion vaccine was reasonably immunogenic, but of limited supply. MF59-adjuvanted H1N1sw vaccine was used to some extent in Europe, but more extensively outside. In contrast, a vaccine with a “stronger” adjuvant, AS03, which contains both squalene

Fig. 14.1 Efficacy of MF59-adjuvanted (aTIV) vaccine in children according to age in a multicenter trial in Finland and Germany. * Statistically significant result, † *Post hoc* analysis (From Vesikari et al. 2011)

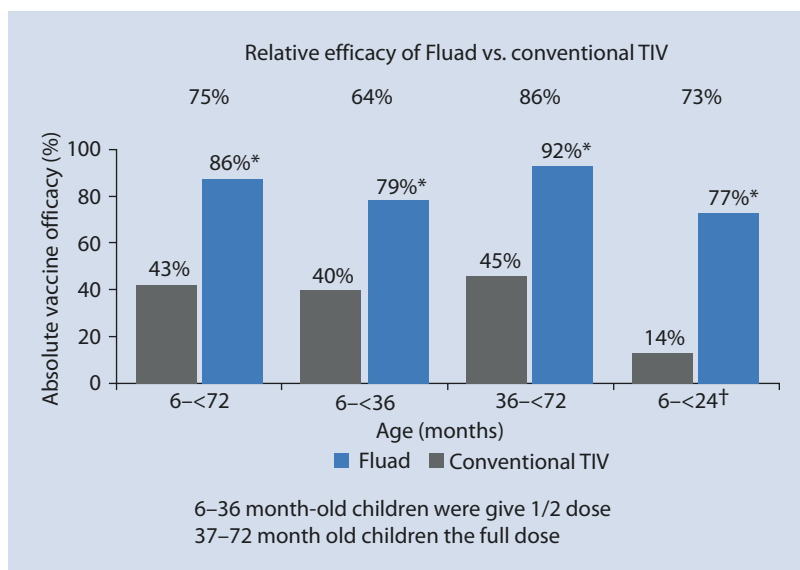
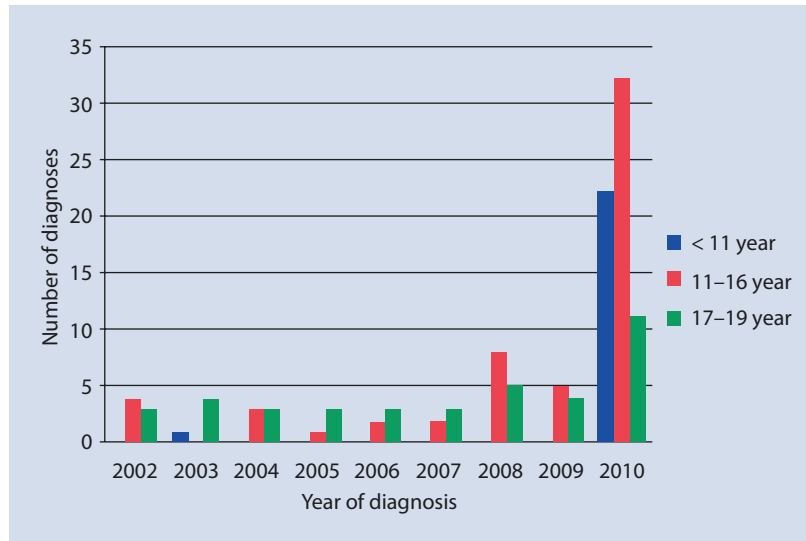


Fig. 14.2 Increase in newly diagnosed cases of narcolepsy in 4- to 19-year-old children and adolescents in Finland in 2010 after extensive vaccinations against H1N1pdm09 virus in the fall of 2009 using an AS03-adjuvanted vaccine (Partinen et al. 2012)



and α -tocopherol, was introduced into several European countries. Such a vaccine was highly immunogenic, but also reactogenic in children. It was used extensively and showed high effectiveness in all age groups.

In 2010, the AS03-adjuvanted H1N1sw vaccine (Pandemrix) was found to be associated with narcolepsy, which is one of the greatest vaccine disasters of modern times. Narcolepsy is a permanent and debilitating condition. First reported in Sweden and Finland, narcolepsy was seen in many other countries using the Pandemrix vaccine (but not in connection with other vaccines). The vaccine increased the risk of narcolepsy in genetically susceptible subjects, mainly in the age range of 5–19 years, to at least 13-fold the background risk (Fig. 14.2). A similar increase in narcolepsy was not seen in Canada, where another AS03-adjuvanted H1N1sw vaccine (Aripanax) was used.

The underlying mechanism may be related to the production process of the split virion vaccine in Europe, resulting in a high content of the influenza nucleoprotein (NP) antigen in the vaccine. NP may be polymerized and in the presence of a strong adjuvant such as AS03 a very strong immune response in young people is induced not only against the HA and NA antigens, but also NP, which in turn may result in the induction of auto-immune reaction in susceptible individuals, with cross-reactivity against hypocretin, leading to its deficiency and clinical narcolepsy.

14.5 Live Attenuated Influenza Vaccine

In the past few years, LAIVs became an option for annual immunization against seasonal influenza in children. The current vaccine (the only one available in Europe) is based on cold-adapted (*ca*) temperature sensitive (*ts*) mutants that were developed by HF Maassab in 1966. The *ca*, *ts* parent strains for influenza A and B are reassorted with the HA and NA genes of current seasonal influenza viruses to make 6:2 reassortants of influenza A and B respectively, that contain six genes from the *ca* and *ts* parents and retain the characteristics of the parent strain. The parent strains grow well in embryonated eggs, which are used for vaccine production.

The parent strains for *ca* and *ts* influenza virus strains were developed separately for influenza A and B viruses in primary chicken kidney cells by serial passages at successive (down to 25 °C) temperatures. The parent strains are stable and retain the mutations responsible for *ca* and *ts* phenotypes upon serial passages in animals and after replication in humans. The *ca* phenotype refers to the ability to grow at 25 °C and *ts* to no growth at 39 °C for influenza A and 37 °C for influenza B. In practice, this means that LAIV vaccine viruses are able to multiply on mucous membranes of the upper airways, but not in the lungs.

Traditionally the *ca*, *ts* parent strains were reassorted with the HA and NA genes of epidemic influenza viruses by co-infection in eggs, to create 6:2 reassortants for influenza A and B vaccines respectively. Since 2006, a new technology, reverse genetics, has been used instead. This technology enables modification of the HA gene before production and incorporation in the vaccine, and has been used to improve the yield and thermostability of the vaccine strains.

14.5.1 Efficacy

A trivalent composition of LAIV (CAIV-T, LAIV3) was tested in a number of efficacy trials before licensure in 2007. The current quadrivalent formulation LAIV4 was not tested for efficacy before licensure in 2013, but real-life effectiveness data are available from several sources, and, in fact, suggest that LAIV4 has not performed so well in 2013–2016 as LAIV3 did before.

A placebo-controlled efficacy trial in 8- to 36-month-old day care children was conducted in six European countries in 2000–2001, and forms the basis for expectations of LAIV performance. The study lasted for two influenza epidemic seasons. Before the first season, all children received two doses of LAIV3, and before the second season one dose. This is how the LAIV vaccine should be administered, but often is not. Laboratory-confirmed influenza occurred in over 10% of the subjects in the first year and about 20% in the second year, indicating the high incidence of influenza in young children and hence the need to vaccinate.

The composite vaccine efficacy in the first and second years respectively was 85.4% and 88.7%. The strain-specific efficacy is shown in [Table 14.2](#).

Table 14.2 Efficacy of live attenuated influenza vaccines (LAIV)3 in a European multicenter trial in 8–36 month-old day care children

Strain	Vaccine efficacy
A/H1N1, season 2001	91.8% (80.8, 97.1)
A/H3N2, season 2002	90.3% (82.9, 94.9)
B, season 2002 Mixed Yamagata and Victoria	81.7% (53.7, 93.9)

Modified from Vesikari et al. (2006a)

Efficacy against A-strains was at least 90% and against B-strains 70–80%, even though some of the circulating B-strains were of a different lineage.

Several other LAIV3 efficacy trials were performed against the TIV comparator and each showed a greater efficacy in children. The pivotal trial carried out in 8,475 children aged 6–59 months is shown in [Fig. 14.3](#). LAIV3 given in two doses to vaccine-naïve children (about two thirds of the study population) showed 54% greater efficacy than TIV; if it was assumed that the efficacy of TIV might have been about 50%, then the efficacy of LAIV3 amounts to around 80% against any strain.

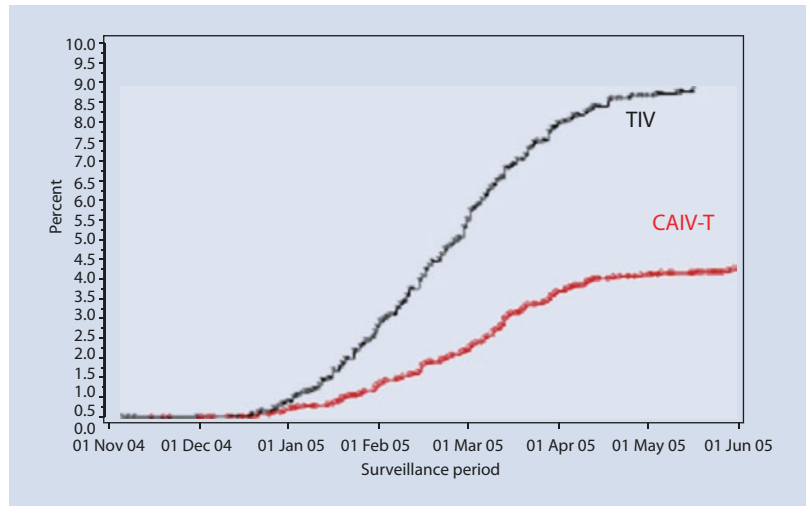
14.5.2 Real-Life Effectiveness

Because of safety issues (see below) LAIV is licensed only for children above 2 years of age. Accordingly, LAIV has been used increasingly in the USA and Europe in children above this age. In the UK, a recommendation was issued in 2013 to give a single dose of LAIV to children aged 2–17 years; in practice, the introduction of this program reached the age groups 2–7 years by the season 2015–2016. In Finland, LAIV has been given since 2014 to children aged 24–36 months as part of the national immunization program. In most cases in the USA and exclusively in the UK and Finland, a single dose of LAIV has been given, which is not what had been studied in prelicensure trials of LAIV.

Matters were complicated by two issues:

1. In 2009, the H1N1pdm09 pandemic strain was introduced into the LAIV vaccine to replace an earlier H1N1 component. The HA of the H1N1pdm09 turned out to be thermolabile, which may have been the reason for reduced vaccine effectiveness discovered several years later;
2. A quadrivalent composition of LAIV (LAIV4) was introduced in 2013. This vaccine contained two B-strains representing Victoria and Yamagata lineages. The logic behind this was the same as for nonlive influenza vaccines: to increase coverage against influenza B. However, the real-life value of the quadrivalent composition was not tested in an efficacy trial, but the US Food and Drugs Administration approved the LAIV4 vaccine based on immunogenicity only. In Europe, the European Medicines Agency, unlike for QIV, did not require an efficacy trial for LAIV4 either.

Fig. 14.3 Cumulative occurrence of culture-confirmed influenza during one influenza season in 6- to 59-month-old children vaccinated with two doses of TIV or LAIV (CAIV-T) (From Belshe et al. 2007)



The real-life effectiveness follow-up in the USA (US Flu Vaccine Effectiveness Network) was lower than expected, compared with prelicensure efficacy. Since 2013, the LAIV4 composition did not show any efficacy against influenza A (largely H1N1) at all. This led to withdrawal of the recommendation of LAIVs by the Advisory Committee on Immunization Practices in June 2016.

In Europe, a case-control study of LAIV4 in the UK in the age group 2–7 years showed an adjusted vaccine efficacy of 57.6% (25.1–76%) against laboratory-confirmed influenza. Strain-specific efficacy is not available. In Finland, an overall vaccine effectiveness of 46.5% was found in the 2- to 3-year-olds; the numbers were too small to draw any conclusions on strain-specific effectiveness.

14.5.3 Safety

Live-attenuated influenza vaccines continued to be used in 2016–2017 season in the UK and Finland immunization programs despite their relatively low efficacy. In general, it would seem prudent to follow the US recommendation not to continue the use of LAIVs until the reasons for low performance have been fully elucidated and the problems corrected.

Three safety issues are related to LAIVs:

1. Flu-like symptoms associated with the multiplication of live attenuated viruses in the upper respiratory tract. This issue is also related to the shedding and potential trans-

mission of vaccine viruses to susceptible subjects in the environment.

2. Provocation of asthma or asthma-like wheezing in asthma-prone children.
3. Increased hospitalizations, owing to respiratory problems and other reasons in subjects under 12 months of age.

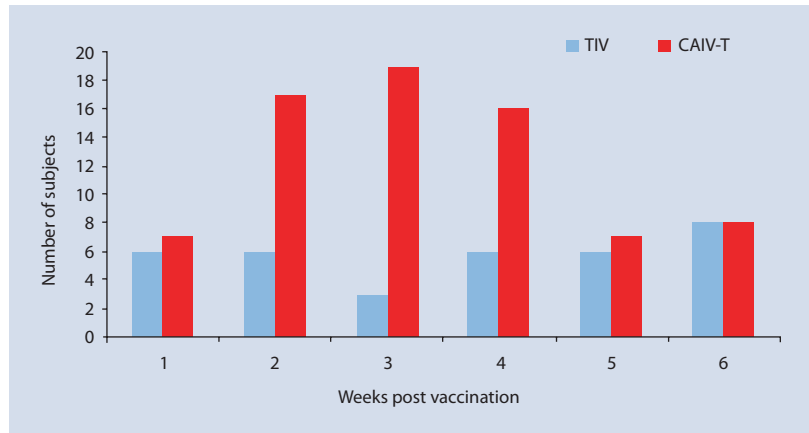
Respiratory symptoms occur in about 10% of naïve children 3–7 days after administration of LAIVs and usually last 2–3 days. The symptoms include runny and stuffy nose and mild fever. The vaccine seems to protect against itself: flu-like symptoms are rare after the second dose.

Shedding of the vaccine virus may be detected in up to 70% of susceptible children. The B-strain is dominant over A-strains. Shedding peaks between days 3 and 10, but may last up to 3 weeks. Transmission is, however, rare. Of note, if the vaccine virus is transmitted, it retains the *ca*, *ts* phenotype and does not cause significant symptoms in the recipients.

Asthma-like wheezing is the best-known side effect of LAIVs, which has limited the use of this vaccine in children aged 24 months and older. Characteristically, the wheezing period occurs 7–14 days after administration of LAIVs (Fig. 14.4).

Wheezing is mainly limited to asthma-prone children under 24 months of age; although it may occur in older children, the rate is not significantly higher than in controls. In younger children, about one half of the wheezing episodes are mild, but the other half are not and some may

Fig. 14.4 Episodes of wheezing in children aged 6–59 months vaccinated with TIV or LAIV (CAIV-T)



require hospitalization. Therefore, the current LAIVs should not be given to children younger than 24 months of age.

The increased rate of hospitalization in recipient children aged 6–11 months is largely unexplained. It is likely related to the insufficient attenuation of LAIV for the youngest infants.

14.5.4 Other LAIVs

Another LAIV, based on the cold-adapted H2N2 virus backbone, was developed in Russia (“Leningrad strain”), and recently licensed to manufacturers in China and India. The intranasal vaccine has been tested and extensively used in Russia, but not outside that country. The H2N2 backbone has been used to generate 6:2 reassortants of a variety of seasonal influenza A-strains and pandemic influenza strains. The vaccine can be produced in cell culture.

Further Reading

- Ambrose CS, Yi T, Falloon J. An integrated, multistudy analysis of the safety of Ann Arbor strain live attenuated influenza vaccine in children aged 2–17 years. *Influenza Other Respir Viruses*. 2011;5:389–97.
- Belshe RB, Edwards KM, Vesikari T, et al. Live attenuated versus inactivated influenza vaccine in infants and young children. *N Engl J Med*. 2007;356:685–96.
- Centers for Disease Control and Prevention. Influenza (Flu). Flu vaccine and people with egg allergies. Available at: <https://www.cdc.gov/flu/protect/vaccine/egg-allergies.htm>. Accessed on: 12 March 2017.

Chen J, Deng YM. Influenza virus antigenic variation, host antibody production and new approach to control epidemics. *Virology*. 2009;6:30.

Durando P, Icardi G, Ansaldo F. MF59-adjuvanted vaccine: a safe and useful tool to enhance and broaden protection against seasonal influenza viruses in subjects at risk. *Expert Opin Biol Ther*. 2010;10:639–51.

Esposito S, Marchisio P, Ansaldo F, Bianchini S, Pacei M, Baggi E, et al. A randomized clinical trial assessing immunogenicity and safety of a double dose of virosomal-adjuvanted influenza vaccine administered to unprimed children aged 6–35 months. *Vaccine*. 2010;28:6137–44.

Heinonen S, Silvennoinen H, Lehtinen P, Vainionpää R, Ziegler T, Heikkinen T. Effectiveness of inactivated influenza vaccine in children aged 9 months to 3 years: an observational cohort study. *Lancet Infect Dis*. 2011;11:23–9.

Jon H, Chen Z. Production of live attenuated influenza vaccines against seasonal and potential pandemic influenza viruses. *Curr Opin Virol*. 2014;6:34–9.

Maassab HF, DeBorde DC. Development and characterization of cold-adapted viruses for use as live virus vaccines. *Vaccine*. 1985;3:355–69.

Murphy BR, Coelingh K. Principles underlying the development and use of live attenuated cold-adapted influenza A and B virus vaccines. *Viral Immunol*. 2002;15:295–323.

Osterholm MT, Kelley NS, Sommer A, Belongia EA. Efficacy and effectiveness of influenza vaccines: a systematic review and meta-analysis. *Lancet Infect Dis*. 2012;12:36–44.

Partinen M, Saarenpää-Heikkilä O, Ilveskoski I, et al. Increased incidence and clinical picture of childhood narcolepsy following the 2009 H1N1 pandemic vaccination campaign in Finland. *PLoS One*. 2012;7:e33723.

Peasah SK, Azziz-Baumgartner E, Breese J, Meltzer MI, Widdowson MA. Influenza cost and cost-effectiveness studies globally – a review. *Vaccine*. 2013;31:5339–48.

Principi N, Esposito S. Adjuvanted influenza vaccines. *Hum Vaccin Immunother*. 2012;8:59–66.

- Principi N, Esposito S. Protection of children against influenza: emerging problems. *Hum Vaccin Immunother.* 2017;27:1–8: doi: [10.1080/21645515.2017.1279772](https://doi.org/10.1080/21645515.2017.1279772).
- Rudenko LG, Slepishkin AN, Monto AS, et al. Efficacy of live and inactivated influenza vaccine in school children and their unvaccinated contacts in Novgorod, Russia. *J Infect Dis.* 1993;168:881–997.
- Sakala IG, Honda-Okubo Y, Fung J, Petrovsky N. Influenza immunization during pregnancy: benefits for mother and infant. *Hum Vaccin Immunother.* 2016;12:3065–71.
- Vesikari T, Fleming DM, Aristegui JF, et al. Safety, efficacy, and effectiveness of cold-adapted influenza vaccine-trivalent against community-acquired, culture-confirmed influenza in young children attending day care. *Pediatrics.* 2006a;118:2298–312.
- Vesikari T, Karvonen A, Korhonen T, et al. A randomized, double-blind study of the safety, transmissibility and phenotypic and genotypic stability of cold-adapted influenza virus vaccine. *Pediatr Infect Dis J.* 2006b;25:590–5.
- Vesikari T, Pellegrini M, Karvonen A, Groth N, Borkowski A, O'Hagan DT, et al. Enhanced immunogenicity of seasonal influenza vaccines in young children using MF59 adjuvant. *Pediatr Infect Dis J.* 2009;28:563–71.
- Vesikari T, Knuf M, Wutzler P, et al. Oil-in-water emulsion adjuvant with influenza vaccine in young children. *N Engl J Med.* 2011;365:1406–16.
- WHO. Vaccines against influenza WHO position paper – November 2012. *Weekly Epidemiol Rec.* 2012;87: 461–76.

Human Papillomavirus Vaccines

Paolo Bonanni

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15.1 Burden of HPV-related Diseases

Human papillomaviruses (HPVs) include more than 100 viral types, with tropism for mucosa or skin. Infection with HPVs may become persistent, progress to precancerous lesions and eventually to invasion, causing cancers in a variety of sites, including the uterine cervix, vulva, vagina, penis, anus, oral cavity, oropharynx, and possibly the skin in patients with epidermodysplasia verruciformis. HPV infections are estimated to account for 5.2% of all cancers in the world, being responsible for 3% of mouth, 12% of oropharynx, 40% of penis, 40% of vulva/vagina, and virtually 100% of uterine cervix cancers. In particular, HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, and 59 are classified as group 1 carcinogens by the International Agency for Research on Cancer (IARC). Cervical cancer is the fourth most common cancer in women, and the seventh overall, with an estimated 528,000 new cases in 2012. Most (around 85%) of the global burden occurs in Low and Middle Income Countries (LMIC), where it accounts for almost 12% of all female cancers. There were an estimated 266,000 deaths from cervical cancer worldwide in 2012, comprising 7.5% of all female cancer deaths. Almost 9 out of 10 (87%) cervical cancer deaths occur in the LMIC regions. Mortality varies 18-fold among the different regions of the world, with rates ranging from less than 2 per 100,000 in Western Asia, Western Europe, and Australia/New Zealand to more than 20 per 100,000 in Melanesia (20.6), Middle (22.2) and Eastern (27.6) Africa. In addition to causing malignant cancers, HPV are also the cause of genital warts (GWs), histologically benign lesions that represent the most common sexually transmitted disease in many countries. Several million cases of GWs occur every year in the world in both females and males, with a peak incidence between 20 and 24 years of age for women, and between 25 and 29 years among men. HPVs are also responsible for a very rare but extremely debilitating disease, juvenile onset recurrent respiratory papillomatosis (JORRP), characterized by the growth of recurrent tumors in the respiratory tract, which results from a vertical transmission of HPV from mother to child. Virology studies have substantiated

the link between genital condylomas and JORRP. HPV types 6 and 11, which are responsible for 80–90% of the condylomas, are responsible for nearly 100% of JORRP.

European data (2012) (► <http://www.hpvcentre.net/statistics/reports/XEX.pdf>) confirm that the disease burden due to HPV infection is impressive: more than 58,000 new cervical cancer cases are estimated to be diagnosed annually, i.e. the 6th cause of female cancer in Europe overall, and the 2nd most common female cancer in women aged 15–44 years. Looking at mortality, more than 24,000 new cervical cancer deaths occur annually in Europe, i.e. the 7th cause of female cancer death overall, and the 2nd most common cause of female cancer death in women aged 15–44 years.

Data on other HPV-related cancers are more difficult to obtain, owing to their relatively lower incidence and to a lack of standardization of registries. However, estimates performed using reliable information available for 26 countries in Europe (EU countries not including Greece, Hungary, Luxemburg, and Romania plus data from Iceland, Norway, and Switzerland) show an incidence of about 2700 vulvar cancers, 1100 vaginal cancers, 4600 anal cancers (2900 in females, 1700 in males), 15,200 head and neck cancers (2500 in females, 12,700 in males), and almost 1100 penile cancers. In the same countries, 23,200 cervical cancer cases are estimated to occur every year. Overall, this means that, of the 48,000 HPV16- and -18-related cancers occurring each year in the selected European countries, 30% are in men. Excluding cervical cancer, of the approximately 23,000 cancer cases due to HPV16/18, most are seen in men owing to the incidence of head and neck cancers, which are fivefold more frequent in males than females. New cases of GWs attributable to HPV types 6/11 in the same countries are estimated at between 615,000 and 675,000 each year, with an equal sex distribution.

15.2 Epidemiology and Ways of Transmission

The association between persistent HPV infection and cervical cancer is one of the strongest known in epidemiology, meaning that cervical cancer is

necessarily linked to such an infection. HPV types 16 and 18 are responsible for >70% of cervical cancers in the world, the remaining less than 30% being due to the other carcinogenic types. The fraction of noncervical cancers attributable to HPV is variable, being about 83% for anal cancer, 60% for vaginal cancer, 42% for penile cancer, 31% for vulvar cancer, and 22% for oropharyngeal cancer. HPV16 is also the single most important type to which almost all noncervical cancers due to HPV are attributable.

HPV6 and -11 are responsible for >90% of genital warts and JORRP cases in the world.

Several co-factors linked to the possible evolution from persistent infection toward precancerous and cancerous lesions have been recognized: smoking, parity, use of oral contraceptives, HIV infection, other sexually transmitted infections. Male circumcision has been shown to decrease the risk of cervical cancer in female partners.

Transmission of HPV occurs primarily through sexual intercourse, not necessarily implying penetration. As a matter of fact, infection has also been described following manual–genital or oral–genital contact. Condom use may reduce the risk of infection, but is not completely protective.

In addition, nonsexual routes are possible, the most important being mother-to-child vertical transmission, which is a rare but possible event. Transmission through contaminated objects (i.e., surgical gloves or biopsy forceps) has been hypothesized, but has never been definitely proven.

15.3 Human Papillomavirus Vaccines

The development of HPV prophylactic vaccines started after the demonstration of the possibility of producing virus-like particles (VLPs) through self-assembly of antigens codified by the genomic regions L1 and L2 (virus capsid proteins). This property is one of the reasons for the high immunogenicity of HPV vaccines, as recombinant L1 proteins produced in yeast or insect cells reconstitute the external shell structure of the virus.

Infection with HPV is an exclusively mucosal event (no viremia) that does not cause inflamma-

tion or cell death. Consequently, natural immunity following infection is usually weak, and re-infection with the same HPV type may occur. Vaccination is given intramuscularly, and a strong primary and secondary response (including immunological memory induction) is obtained after a complete course of immunization.

The mechanism of protection is based on neutralizing antibodies able to prevent virus entry into the target mucosal cell. It is postulated that anti-HPV antibodies produced following active immunization transudate into the cervical mucosal basal layer and into the cervical mucus, where virions are neutralized. However, no minimum protective level of antibodies (correlate of protection) has been defined, also as a consequence of the excellent protection afforded by vaccines. The lack of such a correlate implies that the protective effect of vaccines needs to be defined clinically. As it is not possible to measure the efficacy of HPV vaccines against cervical and other cancers in clinical trials for evident ethical and temporal reasons, it was necessary to find a surrogate marker of protection afforded by vaccination. Persistent infection with HPV is a possible outcome, but viral clearance can occur spontaneously. Prevention of cervical intraepithelial neoplasia (CIN) grade 2 or higher (CIN2+) is considered the best surrogate, as spontaneous reversion to normal histology, although possible, is very rare.

The demonstration of immunogenicity, efficacy, and safety of the first prototype monovalent vaccine against HPV16, paved the way for the development and availability of first-generation vaccines, i.e., the quadrivalent vaccine (containing HPV types 6, 11, 16, and 18) and the bivalent vaccine (containing HPV types 16 and 18). More recently, a nine-valent HPV vaccine (containing HPV types 6, 11, 16, 18, 31, 33, 45, 52, and 58) has been approved for use in Europe.

15.4 Bivalent HPV Vaccine

The bivalent vaccine, Cervarix (GSK), is produced in insect cells (derived from the butterfly *Trichoplusia ni*) by recombinant DNA techniques and adjuvanted with the AS04 system, which is composed of aluminum hydroxide and

Table 15.1 Efficacy of the nine-valent human papilloma virus (HPV) vaccine against HPV31/33/45/52/58 (cervical/vulvar/vaginal disease, persistent infection) – women aged 16–26 years; received all three vaccinations within year of enrolment

Per protocol efficacy population (median follow up 40 months post dose 33)			
Endpoint	9vHPV Vaccine No. of cases/ <i>n</i>	qHPV Vaccine No. of cases/ <i>n</i>	Efficacy (95% CI)
≥CIN2/3, VIN2/3, VaIN2/3	1/6016	30/6017	96.7% (80.9, 99.8)
All CIN, VIN, VaIN2	3/6016	103/6017	97.1% (91.8, 99.2)
6-month persistent infection	35/5939	810/5953	96.0% (94.4, 97.2)

1. Joura et al. (2015)

2. Supplement for Joura et al. (2015)

3. Bautista O. V503–001 MEMO – Median Follow-up Time for Efficacy. Data on file.

Dr. A Luxembourg ACIP February 2014 Meeting ► <http://www.cdc.gov/vaccines/acip/meetings/slides-2016-02.html>

monophosphoryl-lipid A, a lipopolysaccharide from the cell wall of *Salmonella* spp. bacteria (Table 15.1). Efficacy data were evaluated in a young adult female population after a three-dose schedule at 0, 1, and 6 months of the vaccine or placebo. Efficacy was primarily in the total vaccinated cohort, which included women who were HPV-DNA-negative and seronegative for HPV16 and HPV18 at study entry, and who had received at least one dose of vaccine or placebo. Women who had a baseline high-grade lesion or lacking cytology data were excluded from the analysis. The efficacy of the bivalent vaccine in the prevention of CIN2+ associated with HPV16 and/or HPV18 was 90.4% (97.9% confidence interval [CI] 53.4–99.3%). Efficacy against the single types was 93.3% (97.9%; CI 47.0–99.9%) for HPV16, and 83.3% (97.9% CI –78.8–99.9%) for HPV18. When the analysis was also based on HPV16 or HPV18 in the lesion and in preceding cytology samples (post-hoc analysis with attribution of the lesion to specific HPV types), efficacy values all became 100% (97.9% CI 74.2–100% for type 16/18; 64.5–100% for type 16; –49.5 to 100% for type 18). The bivalent HPV vaccine showed a

cross-protective efficacy, especially against types 31 and 45. An overall efficacy against CIN3+ lesions (irrespective of HPV type in the lesion) of 93.2% (95% CI 78.9–98.7%) was reported at year 4 of follow-up for women involved in the PATRICIA clinical trial. However, it is not possible to define the duration of such a cross-protective effect, also because the clinical trials of HPV vaccines were not powered with the aim of measuring cross-protection. A comparative study on the immunogenicity of the bivalent and the quadrivalent HPV vaccines showed significantly higher levels of antibodies against both HPV16 and HPV18 following administration of the bivalent versus the quadrivalent vaccine. The meaning of such data for long-term protection is as yet unknown.

Following a specifically designed clinical trial to compare the immunogenicity of two doses of bivalent vaccine in girls 9–14 years of age vs three doses given to young women aged 15–25 years, which demonstrated that GMTs after two doses in girls were not inferior to three doses in women, a change occurred in the recommended schedule for young girls, which now foresees the adminis-

tration of two doses at 6 months apart for subjects aged <15 years.

15.5 Quadrivalent HPV Vaccine

The quadrivalent vaccine (Gardasil, Merck) is produced in yeast cells by recombinant DNA techniques and adjuvanted with amorphous aluminum salts (■ Table 15.1). The phase 3 clinical trial was performed in 13 different countries, involved about 12,000 women, randomly assigned to receive HPV vaccine or placebo according to a three-dose schedule (0, 2, and 6 months). The composite efficacy result (CIN2, CIN3, adenocarcinoma in situ) after an average 3-year follow-up was 98% (95% CI 86–100%) in the per-protocol susceptible population, and 44% (95% CI 26–58%) in the intention-to-treat population, where women already infected were also represented.

Immunological memory against the quadrivalent L1-encoded HPV antigens was demonstrated by a challenge dose administered 5 years after the first dose in fully vaccinated women. A booster response was elicited even if the woman had lost detectable antibodies to some antigen. Interestingly, in the same study, it was possible to highlight that no case of breakthrough infection occurred in women of the vaccine group who became seronegative in the 5-year time interval, whereas 10 cases of infection occurred in women belonging to the placebo group. It is not clear whether this means that vaccinated women retained a protective, but undetectable level of antibodies to the L1 antigen, or if they were protected through an anamnestic response at the mucosal level. Women belonging to the placebo group were immunized at year 5, and this intervention prevented the possibility of having long-term efficacy data through the comparison of vaccinated vs unvaccinated women. However, data from originally vaccinated women followed through cancer registries in Nordic countries showed no breakthrough infection after 9 years of follow-up. The quadrivalent vaccine showed a good degree of cross-protection in clinical trials, especially against HPV31 and -33, the duration of

which needs to be further investigated. An independent study comparing the antibody response obtained after two doses administered 6 months apart in girls aged 9–13 years versus young women aged 16–26 years receiving three doses at 0, 2, and 6 months showed non-inferiority of the two-dose schedule and the two-dose schedule was approved for the quadrivalent vaccine given at age 9–13 years.

15.6 Nonavalent HPV Vaccine

The nine-valent HPV vaccine was developed based on the heritage of the quadrivalent vaccine, with which it shares the same production process and the same adjuvant. It includes the additional HPV types 31, 33, 45, 52, and 58. The foreseen direct impact of the new vaccine is an increase in prevented overall HPV-related cancers from 75% to 89%. About 90% of cervical cancer cases would be directly preventable using the nine-valent vaccine (vs 72% with the quadrivalent vaccine), whereas the increase in prevented cases would be more limited for anal cancer (from 87% to 90%).

In a double blind, randomized, multicenter study, over 14,000 women were randomly assigned to receive three doses of either the nonavalent or the quadrivalent HPV vaccine (comparator) at months 0, 2, and 6. The nonavalent vaccine turned out to have overlapping (and non-inferior) seroconversion rates and geometric mean titers (GMTs) 1 month after the third dose (month 7). The efficacy of the nonavalent vaccine against precancerous lesions and persistent infection due to HPV types 31, 33, 45, 52, and 58 was directly measured in the trial, as the quadrivalent vaccine lacks VLPs of such HPV types. The overall efficacy data against different endpoints for the five types are reported in ■ Table 15.1, and was invariably >90%, mostly >95%. ■ Table 15.2 reports the 6-month efficacy against persistent infection for the single additional types of the nine-valent vaccine, which ranged from 94.8% to 99.1%.

The nine-valent vaccine was recently also approved for use with a two-dose schedule at 0–6/12 months in girls and boys aged 9–14 years,

Table 15.2 Efficacy of the nine-valent HPV vaccine against 6-month persistent infection (PI) due to types 31, 33, 45, 52, and 58. Per protocol population

Endpoint 6-month PI	9vHPV No. cases/total	qHPV No. cases/total	Efficacy (95% CI)
HPV 31	7/5251	150/5198	95.5% (90.7, 97.9)
HPV 33	1/5553	106/5560	99.1% (95.2, 100)
HPV 45	4/5649	124/5658	96.8% (92.1, 98.9)
HPV 52	11/5263	387/5160	97.3% (95.5, 98.7)
HPV 58	12/5297	225/5284	94.8% (91.0, 97.1)

15.7 Effectiveness of HPV Vaccines: From Trials to the Real World

Ten years after HPV vaccination was implemented in several countries, a considerable amount of disease impact data is available. HPV vaccination programs have been proven to reduce incident and prevalent HPV-related conditions and diseases even a couple of years after vaccination implementation. As those data come from ecological studies, results must be interpreted with care. Below, some of the available data on HPV vaccination effectiveness are reported.

The first diseases on which immunization have an impact are GWs.

In Australia, 5 years after implementation of HPV vaccination, a 93% reduction of GWs irrespective of vaccination status was registered in women aged <21 years, and a 100% reduction in women who declared that they had been vaccinated. GW incidence in heterosexual men also decreased by indirect effect. Such an effect was not visible in the homosexual male population. In Denmark, after introduction of the vaccination

program, the incidence ratio (IR) of GWs in 16- and 17-year-old women between 2008 and 2013 decreased from 1071 to 58 per 100,000 person-years, and was reduced from 365 to 77 per 100,000 person-years in men.

Also, the prevalence of vaccine-type HPV DNA decreased significantly in Australian females aged 18–24 years: 4vHPV prevalence decreased from 29% to 7% in partially and to 2% in fully vaccinated women; a lower prevalence of vaccine-targeted types in unvaccinated women (19%) suggested herd immunity. Furthermore, in a country using the bivalent vaccine, such as Scotland, from a total of 4679 samples tested, a significant reduction in prevalence of HPV16 and -18 from 29.8% (95% CI 28.3, 31.3%) to 13.6% (95% CI 11.7, 15.8%) was registered in the 5 years after vaccination implementation.

Pre-cancerous lesions have also decreased significantly following implementation of immunization strategies. Australian data updated to March 2014, with a vaccine coverage around 70% for three doses, showed a reduction of high-grade precancerous lesions (CIN2/3) of 50% in women aged <21 years. In Sweden, a study on 1,333,691 women aged 13–29 years compared women who declared that they had been vaccinated or not vaccinated. Effectiveness against CIN2+ was 75% (incidence rate ratio [IRR] = 0.25, 95% CI = 0.18–0.35) for those initiating vaccination before age 17, and 46% (IRR = 0.54, 95% CI = 0.46–0.64) and 22% (IRR = 0.78, 95% CI = 0.65–0.93) for those initiating vaccination at ages 17–19 and at ages 20–29 respectively. In Scotland, a significant reduction of CIN diagnoses in women who received three doses of vaccine vs those not vaccinated was registered: for CIN 1, adjusted RR was 0.71 (95% CI 0.58–0.87; $P = 0.0008$). For CIN 2, adjusted RR was 0.5 (95% CI 0.4–0.63; $P = 0.0001$) and for CIN 3, adjusted RR was 0.45 (95% CI 0.35–0.58; $P = 0.0001$). In younger women (birth cohort 1992), with a vaccination coverage of 81.5% (74.1% with three doses), the adjusted RR vs age cohort 1988 (prevaccination) was 0.49 (95% CI 0.34–0.71 $P = 0.0002$) independently of vaccination status.

15.8 Safety of HPV Vaccines

All HPV vaccines showed a good safety profile in clinical trials. Local reactions (pain, swelling, induration, redness, etc.) are frequently reported side effects. Systemic reactions included fever, headache, vertigo, and nausea.

Post-marketing surveillance data include, as expected, reports of a wide range of adverse events following immunization (AEFIs). Causality assessment is a complex process that implies the verification of the simultaneous presence of different criteria, and not simply temporal association.

Several diseases of uncertain etiology have been reported after HPV vaccination; however, none of them was demonstrated to be causally associated with immunization.

For the quadrivalent vaccine, a review of 15 published post-marketing studies based on both passive and active surveillance showed an excellent record of safety on >1 million vaccinated subjects around the world. The US Institute of Medicine published a review on HPV vaccination, autoimmune disease and acute disseminated encephalomyelitis (ADEM), which stated that the vaccine is not associated with an increased risk of multiple sclerosis or other demyelinating diseases.

The most recent threat to HPV vaccination programs was the report of some cases of complex regional pain syndrome (CRPS) and of postural orthostatic tachycardia syndrome (POTS) in vaccinated girls, following which vaccination coverage dramatically fell in Japan. Following a Danish request of review for evidence of possible causality, the Pharmacovigilance Risk Assessment Committee (PRAC) of the European Medicine Agency (EMA) stated that available evidence does not show any causal relationship between HPV vaccination and the two syndromes. Such a conclusion was endorsed officially by the EMA on 20 November 2015. In December 2015, the WHO Global Advisory Committee on Vaccine Safety confirmed that no such causal association exists, calling for efforts by Japan health authorities to restore vaccination coverage.

15.9 Vaccination Programs in the World

Vaccination for HPV has been recommended and implemented in the adolescent female population of several countries for about 10 years, starting from industrialized areas, and achieving different coverage results (■ Fig. 15.1).

Today, recommendations and implementation of HPV vaccination for adolescent or pre-adolescent girls are a public health priority in all countries of the world. Countries with limited resources have been involved in vaccination demonstration projects and, in some cases, have launched a national program with the help of international agencies and alliances (■ Fig. 15.2). Extension of the immunization offer to adolescent male subjects has become an important additional opportunity for several countries, also because the progressive decrease of vaccine costs and the possibility of administering two doses only in adolescents has made universal HPV immunization a cost-effective option in many instances. Special attention is needed for homosexual men with HIV infection, who are at a particularly increased risk for HPV-related diseases and deaths. However, it seems unlikely that a high vaccination coverage is reached in such a risk group, universal (female and male) adolescent programs being the real solution. An extension of female age groups involved in the active offer of immunization to include young adults would allow a faster impact of vaccination programs on HPV-related cancers and pre-cancers.

Furthermore, it must not be forgotten that reaching high coverage with HPV vaccines can have a deep impact on the organization of screening programs. In the presence of a high coverage against HPV vaccine types in the population, it would be possible to extend HPV DNA testing as a primary screening test, and fewer screening rounds during a woman's lifetime would be sufficient to provide almost complete protection against HPV.

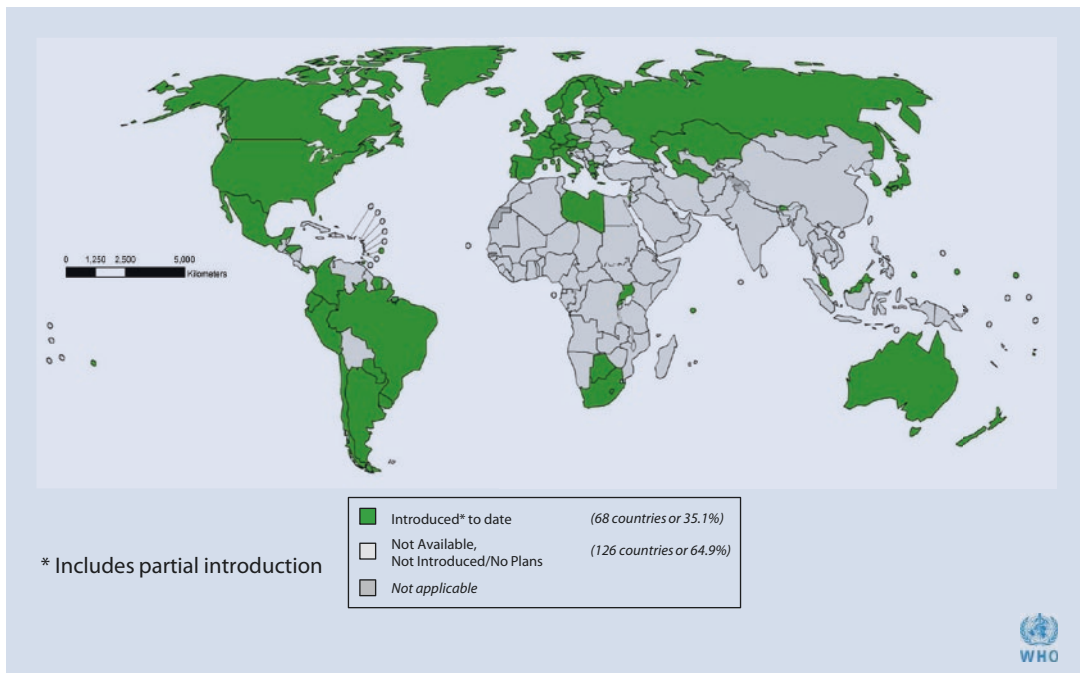


Fig. 15.1 Countries that include human papilloma virus (HPV) vaccine in their national immunization program (Data source: WHO/IVB Database, as of 10 January 2017. Map production Immunization Vaccines and Biologicals (IVB), World Health Organization; The boundaries and names shown and the designations used on this map do not imply the expression of any opinion

whatsoever on the part of the World Health Organization concerning the legal status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries. Dotted lines on maps represent approximate border lines for which there may not yet be full agreement. ©WHO 2017. All rights reserved)

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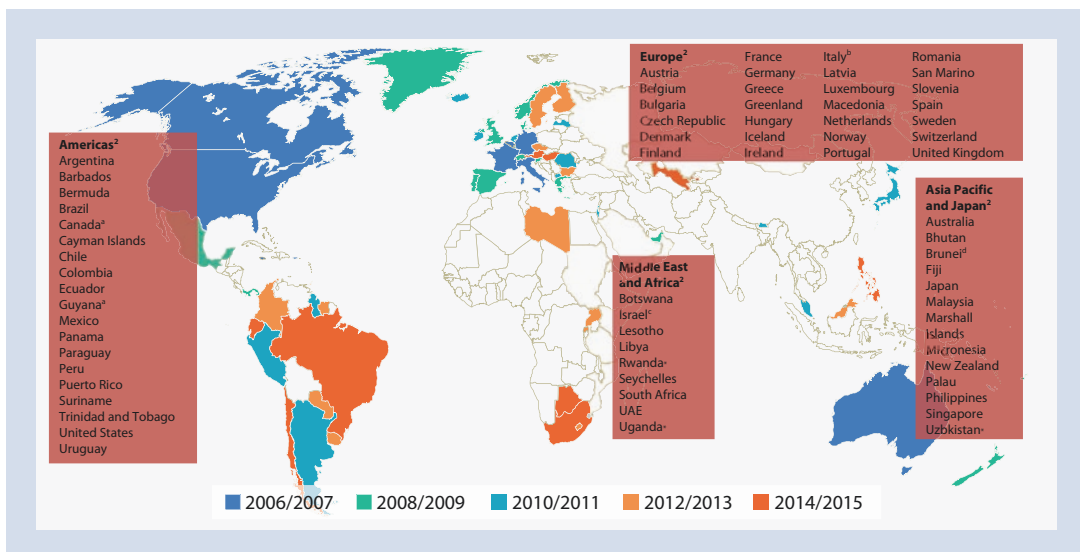


Fig. 15.2 The HPV vaccine in a national immunization program, by year of introduction *Gavi eligible. ^aAs an extended dosing schedule of 0, 6, and 60 months in Mexico in 2008; and from 2007 to 2009 in Quebec and

British Columbia. ^bFrom 2007 to 2008. ^cIntroduced for females in 2011 and males in 2015. ^dFrom 2012 to 2015. ¹Garland et al. (2016), ²Bruni et al. (2016)

	Bivalent vaccine	Quadrivalent vaccine	Nonavalent vaccine
Antigens(Virus-like particles – VLPs)	20 µg HPV-16 20 µg HPV-18	40 µg HPV-16 20 µg HPV-18 20 µg HPV-6 40 µg HPV-11	60 µg HPV-16 40 µg HPV-18 30 µg HPV-6 40 µg HPV-11 20 µg HPV-31 20 µg HPV-33 20 µg HPV-45 20 µg HPV-52 20 µg HPV-58
Expression system	Baculovirus expression vector system in <i>Trichoplusia ni</i> Rix4446 cell substrate	<i>Saccharomyces cerevisiae</i> yeast	<i>Saccharomyces cerevisiae</i> yeast
Adjuvant	AS04 Adjuvant system [50 µg MPL and 500 µg Al(OH) ³]	225 µg amorphous aluminum hydroxyphosphate sulfate	500 µg amorphous aluminum hydroxyphosphate sulfate
Administration schedule	2 doses 5–13 months apart from 9 to 14 years 3 doses at month 0, 1, 6 in subjects ≥15 years	2 doses at month 0 and 6 from 9 to 13 years 3 doses at month 0, 2, 6 in subjects ≥14 years	2 doses 5–13 months apart from 9 to 14 years 3 doses at month 0, 2, 6 in subjects ≥15 years

Further Reading

- Ali H, Donovan B, Wand H, Read TR, Regan DG, Grulich AE, Fairley CK, Guy RJ. Genital warts in young Australians five years into national human papillomavirus vaccination programme: national surveillance data. *BMJ*. 2013;346:f2032.
- Bollerup S, Baldur-Felskov B, Blomberg M, Baandrup L, Dehlendorff C, Kjaer SK. Significant reduction in the incidence of genital warts in young men 5 years into the Danish Human Papillomavirus vaccination program for girls and women. *Sex Transm Dis*. 2016; 43:238–42.
- Bonanni P, Bechini A, Donato R, Capei R, Sacco C, Levi M, Boccalini S. Human papilloma virus vaccination: impact and recommendations across the world. *Ther Adv Vaccines*. 2015;3:3–12.
- Bosch FX, Robles C, Díaz M, Arbyn M, Baussano I, Clavel C, et al. HPV-FASTER: broadening the scope for prevention of HPV-related cancer. *Nat Rev Clin Oncol*. 2016; 13:119–32.
- Brotherton JM, Saville AM, May CL, Chappell G, Gertig DM. Human papillomavirus vaccination is changing the epidemiology of high-grade cervical lesions in Australia. *Cancer Causes Control*. 2015;26:953–4.
- Bruni L, et al. Global estimates of human papillomavirus vaccination coverage by region and income level: a pooled analysis. *Lancet Glob Health*. 2016;4:e453–63.
- Castellsagué X, Bosch FX, Muñoz N. Environmental co-factors in HPV carcinogenesis. *Virus Res*. 2002a;89:191–9.
- Castellsagué X, Bosch FX, Muñoz N, Meijer CJ, Shah KV, de Sanjose S, et al. Male circumcision, penile human papillomavirus infection, and cervical cancer in female partners. *N Engl J Med*. 2002b;346:1105–12.
- De Flora S, Bonanni P. The prevention of infection-associated cancers. *Carcinogenesis*. 2011;32:787–95.
- Dobson SR, McNeil S, Dionne M, Dawar M, Ogilvie G, Kraiden M, et al. Immunogenicity of 2 doses of HPV vaccine in younger adolescents vs 3 doses in young women: a randomized clinical trial. *JAMA*. 2013;309:1793–802.
- Einstein MH, Baron M, Levin MJ, Chatterjee A, Edwards RP, Zepp F, et al. Comparison of the immunogenicity and safety of Cervarix and Gardasil Human Papillomavirus (HPV) cervical cancer vaccines in healthy women aged 18–45 years. *Hum Vaccin*. 2009;5:705–19.
- European Medicines Agency. Assessment report. Human Papilloma Virus (HPV) vaccines: 11 November 2015. http://www.who.int/vaccine_safety/committee/GACVS_HPV_statement_17Dec2015.pdf?ua=1.
- FUTURE II Study Group. Quadrivalent vaccine against human papillomavirus to prevent high-grade cervical lesions. *N Engl J Med*. 2007;356:1915–27.
- Garland SM, Kjaer SK, Muñoz N, et al. Impact and effectiveness of the quadrivalent human papillomavirus vaccine: a systematic review of 10 years of real-world experience. *Clin Infect Dis*. 2016;63(4):519–27.

- Hartwig S, Syrjänen S, Dominiak-Felden G, Brotons M, Castellsagué X. Estimation of the epidemiological burden of human papillomavirus-related cancers and non-malignant diseases in men in Europe: a review. *BMC Cancer*. 2012;12(3):30. doi:10.1186/1471-2407-12-30.
- Herweijer E, Sundström K, Ploner A, Uhnöo I, Sparén P, Arnheim-Dahlström L. Quadrivalent HPV vaccine effectiveness against high-grade cervical lesions by age at vaccination: a population-based study. *Int J Cancer*. 2016;138:2867–74.
- IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Biological agents. IARC Monogr Eval Carcinog Risks Hum. 2010;100B. ► <http://globocan.iarc.fr/old/FactSheets/cancers/cervix-new.asp>.
- ICO Information Centre on HPV and Cancer. Human papillomavirus and related diseases report – Europe. Available at: ► <http://www.hpvcentre.net/statistics/reports/XEX.pdf>. Accessed 30 January 2017.
- Iversen OE, Miranda MJ, Ulied A, Soerdal T, Lazarus E, Chokeyhaibulkit K, et al. Immunogenicity of the 9-valent HPV vaccine using 2-dose regimens in girls and boys vs a 3-dose regimen in women. *JAMA*. 2016;316:2411–21.
- Joura EA, Giuliano AR, Iversen OE, Bouchard C, Mao C, Mehlsen J, et al. A 9-valent HPV vaccine against infection and intraepithelial neoplasia in women. *N Engl J Med*. 2015;372:711–23.
- Kavanagh K, Pollock KG, Potts A, Love J, Cuschieri K, Cubie H, Robertson C, Donaghy M. Introduction and sustained high coverage of the HPV bivalent vaccine leads to a reduction in prevalence of HPV 16/18 and closely related HPV types. *Br J Cancer*. 2014;110(11):2804.
- Kirnbauer R, Booy F, Cheng N, Lowy DR, Schiller JT. Papillomavirus L1 major capsid protein self-assembles into virus-like particles that are highly immunogenic. *Proc Natl Acad Sci USA*. 1992;89:12180–4.
- Kjaer SK, Chackerian B, van den Brule AJ, Svare EI, Paull G, Walbomers JM, et al. High-risk human papillomavirus is sexually transmitted: evidence from a follow-up study of virgins starting sexual activity (intercourse). *Cancer Epidemiol Biomark Prev*. 2001;10:101–6.
- Lehtinen M, Paavonen J, Wheeler CM, et al. Overall efficacy of HPV-16/18 AS04-adjuvanted vaccine against grade 3 or greater cervical intraepithelial neoplasia: 4-year end-of-study analysis of the randomised, double-blind PATRICIA trial. *Lancet Oncol*. 2012;13:89–99.
- Olsson SE, Villa LL, Costa RL, et al. Induction of immune memory following administration of a prophylactic quadrivalent human papillomavirus (HPV) types 6/11/16/18 L1 virus-like particle (VLP) vaccine. *Vaccine*. 2007;25:4931–9.
- Paavonen J, Jenkins D, Bosch FX, Naud P, Salmerón J, Wheeler CM, et al. Efficacy of a prophylactic adjuvanted bivalent L1 virus-like-particle vaccine against infection with human papillomavirus types 16 and 18 in young women: an interim analysis of a phase III double-blind, randomised controlled trial. *Lancet*. 2007;369:2161–70.
- Parkin DM. The global health burden of infection-associated cancers in the year 2002. *Int J Cancer*. 2006;118:3030–44.
- Patel H, Wagner M, Singhal P, Kothari S. Systematic review of the incidence and prevalence of genital warts. *BMC Infect Dis*. 2013;13:39.
- Pollock KG, Kavanagh K, Potts A, Love J, Cuschieri K, Cubie H, et al. Reduction of low- and high-grade cervical abnormalities associated with high uptake of the HPV bivalent vaccine in Scotland. *Br J Cancer*. 2014;111:1824–30.
- Romanowski B, Schwarz TF, Ferguson LM, Peters K, Dionne M, Schulze K, et al. Immunogenicity and safety of the HPV-16/18 AS04-adjuvanted vaccine administered as a 2-dose schedule compared with the licensed 3-dose schedule: results from a randomized study. *Hum Vaccin*. 2011;7:1374–86.
- Sinisgalli E, Bellini I, Indiani L, Sala A, Bechini A, Bonanni P, Boccalini S. HPV vaccination for boys? A systematic review of economic studies. *Epidemiol Prev*. 2015;39(Suppl 1):51–8.
- Vichnin M, Bonanni P, Klein NP, Garland SM, Block SL, Kjaer SK, et al. An overview of quadrivalent human papillomavirus vaccine safety: 2006 to 2015. *Pediatr Infect Dis J*. 2015;34:983–91.
- Winer RL, Lee SK, Hughes JP, Adam DE, Kiviat NB, Koutsky LA. Genital human papillomavirus infection: incidence and risk factors in a cohort of female university students. *Am J Epidemiol*. 2003;157:218–26.

Tick-Borne Encephalitis Vaccines

Herwig Kollaritsch and Ulrich Heininger

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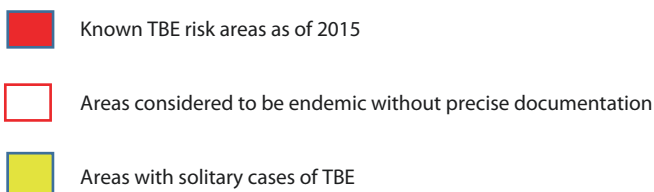
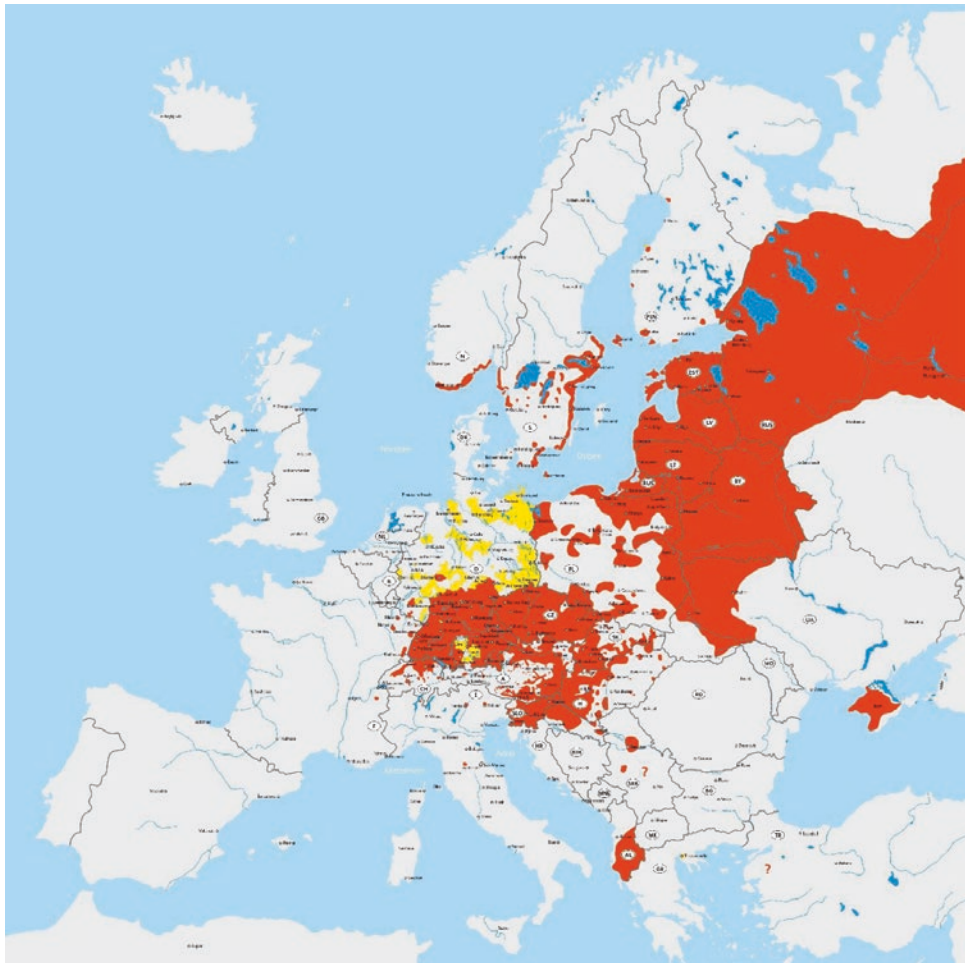
16.1 Tick-Borne Encephalitis Disease

Tick-borne encephalitis (TBE) is caused by the TBE virus (TBEV), a member of the *Flavivirus* family. There are three antigenetically very closely related subtypes of the virus: the European subtype (TBEV-Eu), the Siberian subtype (TBEV-Sib), and the Far East subtype (TBEV-Fe). The virus is inoculated to the host by a sting (frequently and erroneously referred to as a “bite”) from an infected tick and the virus then replicates locally, followed

by viremia of 2–7 days and facultative invasion of the central nervous system.

Viremia occurs in all patients with TBEV infections, but approximately two thirds of them remain asymptomatic and only one third get clinical symptoms.

Tick-borne encephalitis is only rarely exported to other countries and a recent review on travel-associated TBE presented evidence that in 2012 only 39 cases of TBE were documented in Central and Western Europe among international travelers (■ Figs. 16.1 and 16.2).



■ Fig. 16.1 Endemic areas of tick-borne encephalitis (TBE) in Europe (©Pfizer with permission)

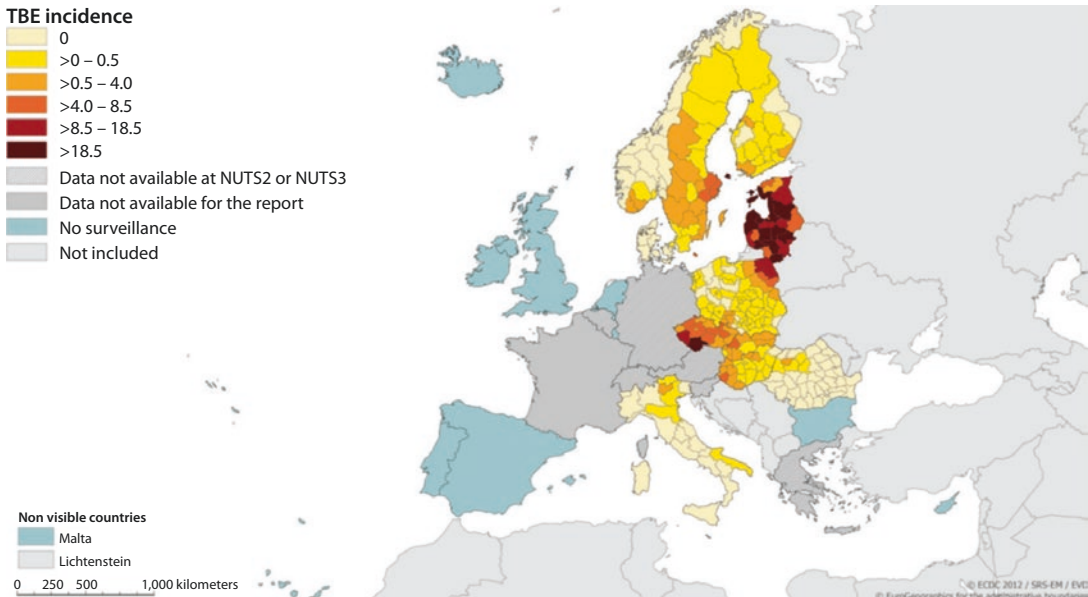


Fig. 16.2 The average annual incidence rate of TBE per 100,000 inhabitants in the EU/European Free Trade Association area at a lower administrative level Nomen-

clature of Territorial Units for Statistics (NUTS) 2 (Italy) or NUTS 3 (European Centre for Disease Prevention and Control, 2012)

Box 16.1: Criteria for Tick-Borne Encephalitis (TBE) Case Confirmation and Consecutive Case Classification

Clinical criteria

Any person with symptoms of inflammation of the central nervous system (e.g., meningitis, meningoencephalitis, encephalomyelitis, encephaloradiculitis).

Laboratory criteria¹

1. Confirmed case

At least one of the following five:

- TBE-specific immunoglobulin M (IgM) and immunoglobulin G antibodies in blood
- TBE-specific IgM antibodies in cerebrospinal fluid
- Sero-conversion or four-fold increase in TBE-specific antibodies in paired serum samples
- Detection of TBE viral nucleic acid in a clinical specimen
- Isolation of the TBE virus from a clinical specimen

2. Probable case:

- Detection of TBE-specific IgM antibodies in a unique serum sample

Epidemiological criteria

- Exposure to a common source (unpasteurized dairy products)

Case classification

- i. *Possible case*: not applicable
- ii. *Probable case*
 - Any person meeting the clinical criteria and the laboratory criteria for a probable case, or
 - Any person meeting the clinical criteria and with an epidemiological link
- iii. *Confirmed case*
 - Any person meeting the clinical and laboratory criteria for case confirmation

¹ Serological results should be interpreted according to the vaccination status and previous exposure to other flaviviral infections. Confirmed cases in such situations should be validated by serum neutralization assay or other equivalent assays.

Nevertheless, TBE has to be taken into account in the differential diagnosis of aseptic meningitis in patients who had stayed in a TBE-endemic area in the previous 4 weeks, especially in the warm season (► Box 16.1).

16.2 TBE Vaccines

Vaccines Currently, four different inactivated whole-virus alum-adsjuvanted vaccines against TBE are available, two of them regionally in Russia (based on TBEV-Sib) and two in Europe (based on TBEV-Eu). The two European vaccines, Encepur[®] (available since 1994) and FSME-Immun[®] (available since 1976) are formulated in two separate preparations: one for children and one for adults. Both vaccine brands underwent a series of modifications up to the beginning of the millennium, when the actual preparations (■ Table 16.1) were introduced to the market. FSME-Immun 0.25 ml junior[®] is licensed for use in children ≥ 1–15 years of age. It contains

■ **Table 16.1** Specific characteristics and composition of European tick-borne encephalitis (TBE) vaccines

Name and producer	FSME-IMMUN [®] ; Pfizer	Encepur [®] ; GSK
<i>Antigen details</i>		
Strain	TBEV-Eu Neudörfl	TBEV-Eu K23
Passages	PCEC	PCEC
Production	PCEC	PCEC
Amount of antigen (adults/children)	2.4 µg/ 1.2 µg	1.5 µg/ 0.75 µg
<i>Excipients</i>		
Adjuvant	Al(OH) ₃	Al(OH) ₃
Preservative	No	No
Stabilizer	HSA	Sucrose
<i>Presentation</i>		
Formulation (adults/children)	0.5/0.25 ml, liquid	0.5/0.25 ml, liquid
Packaging	Prefilled syringe	Prefilled syringe
Shelf-life	30 months (2–8 °C)	30 months (2–8 °C)

Adapted from WHO (2011)
TBEV-Eu tick-borne encephalitis virus European strain, HSA human serum albumin, PCEC primary chicken embryonic cells, Al(OH)₃ aluminum hydroxide

1.2 mcg of inactivated TBEV-Eu strain Neudörfl. Encepur 0.25 ml children[®] is licensed according to the Summary of Product Characteristics for children ≥ 1–11 years of age and contains 0.75 mcg inactivated TBE virus strain Karlsruhe 23. The respective virus strains are cultivated on primary chicken embryonic cells. The vaccines contain either sucrose (Encepur[®]) or human serum albumin (HSA; FSME-Immun[®]) as stabilizers, both without any preservative (■ Table 16.1).

Adult formulations of the TBE vaccines simply contain twice the amount of antigen of the respective vaccines for children in 0.5 ml volume and should be used for intramuscular administration in individuals 12 (Encepur[®]) and 16 years of age (FSME-Immun[®]) onward respectively.

Immunization Schedules The conventional basic immunization schedule of TBE vaccines consists of two doses given at an interval of 1–3 months, followed by a third dose 5 or 9–12 months later, ideally applied before the tick season starts. The first booster dose should be applied 3 years later and then boosting is recommended in 5-year intervals to maintain circulating neutralizing antibodies. Some national recommendations differ from this licensed schedule, e.g., the Austrian authorities recommend a 3-year booster interval for persons ≥ 60 years of age and in Switzerland booster doses are recommended at intervals of 10 years.

For both vaccines accelerated immunization schedules are licensed, consisting of a series of three doses on days 0–7–21, followed by a fourth dose after 12–18 months (Encepur[®]) and two doses 14 days apart with a third dose after 5–12 months (FSME-Immun[®]; ■ Table 16.2).

Irregular Vaccination Schedules If a person has received only one dose of TBE vaccine, a second dose should be applied within 1 year after the first one; otherwise immune response is not guaranteed (seroconversion 94% rather than >95%). After at least two doses, a single further dose administered up to 20 years (and beyond) later leads to a sufficient anamnestic response, indicating a robust immune memory.

Interchangeability FSME-Immun[®] and Encepur[®] can be administered alternately for boosting, whereas the primary series (at least doses 1 and 2) should be performed with either product, as data on interchangeability during basic immunization are scarce.

Table 16.2 Immunization schedules for TBE vaccines^a according to the Summary of Product Characteristics, intervals given in months unless indicated otherwise

	Basic immunization conventional schedule; (dose 1 on day 0)		Basic immunization rapid schedule; (dose 1 on day 0)			First booster (years)	Subsequent boosters (years)
	2nd dose (months)	3rd dose (months)	2nd dose	3rd dose (months)	4th dose (months)		
FSME-Immun [®]	1–3	5–12	14 days	5–12	–	3	5 ^b
Encepur [®]	1–3	9–12	7 days	21 days	12–18 ^{c,d}	3	5 ^b

Adapted from WHO (2011)

^aSchedules apply for both preparations (children's and adults' preparation)

^bIn persons 50 years of age and older, an interval of 3 years (Austria: persons 60 years of age and older); Switzerland: 10-year intervals, independent of age

^cConsidered as first booster

^dAlternatively, as with FSME-Immun[®], the interval between the first doses may be reduced to 14 days, followed by a third dose 9–12 months later

Table 16.3 TBE vaccination after a tick sting

Vaccination history (written documentation)	Interval between last immunization and tick sting	Interval between tick sting and physician visit ^a	Recommendation
Unvaccinated or unknown	Not applicable	<4 weeks	Wait until ≥ 4 weeks after sting, then initiate immunization series
1 dose	≤ 14 days	Not relevant	Wait until ≥ 4 weeks after sting, then administer 2nd dose
		15 days – 1 year	Administer 2nd dose immediately
	≥ 1 year	≥ 48 h	Wait until ≥ 4 weeks after sting, then administer 2nd dose ^b
		<48 h	Administer 2nd dose immediately ^b
≥ 2		≥ 48 h	Wait until ≥ 4 weeks after sting, then administer 2nd dose ^b
			Additional vaccination according to regular schedule

Adapted from BMG (2016)

^aIf time elapsed is not to be determined, use schedule ">48 h after tick sting"

^bControl of antibody response recommended. If not possible, count this vaccination as the first one in the basic immunization schedule

TBE Vaccination After a Tick Sting There are no generally accepted post-exposure procedures in persons without or with incomplete immunizations against TBE in the case of a tick sting in a TBE-endemic area. However, the Austrian Health

Authorities published a useful schedule in their national vaccination recommendations (Table 16.3) that may be followed. Basically, a first dose of immunization should be avoided in a previously unimmunized patient during the TBE incu-

bation period after a tick sting, as it is not expected to be efficient, but may cause concern if it interferes with natural TBE infection.

16.3 TBE Vaccines: Immunogenicity and Effectiveness

Immunogenicity Encepur[®] and FSME-Immun[®] have been registered based on immunogenicity and safety study data, but no controlled trials with clinical efficacy endpoints have been conducted. For both vaccines, ample data on their immunogenicity are available, demonstrating high seroconversion rates of close to 100% and robust neutralizing antibody titers in healthy subjects. Persistence of neutralizing antibodies after primary and/or booster vaccinations indicate long-term protection in healthy persons. Of note, age at initiation of the immunization series plays an important role, with higher seroconversion rates and mean antibody values in addition to more prolonged antibody persistence in children and adolescents compared with adults.

Focusing on the pediatric use of the two TBE vaccines, a number of studies have evaluated the immunogenicity of the preparations and consistently found high seroconversion rates of 98–100% after a primary course (conventional or accelerated dosing schedule) of vaccinations and appropriate persistence of neutralizing antibodies to support the recommended boosting intervals. A few studies show evidence that antibody persistence in children may be even longer than expected. There is no convincing evidence that one vaccine would induce a superior immune response or lead to better protection against disease than the other.

Cross Protection TBEV-Eu-containing vaccines induce some cross-protection against the other TBEV subtypes, indicating that FSME-Immun[®] and Encepur[®] are also protective against TBEV-Sib and TBEV-Fe.

Immunocompromised Patients and Low Responders The TBE vaccines induce a strong and robust immune memory in healthy persons. However, there is some recent evidence that in immunocompromised patients or those with certain underlying chronic diseases the immune response may be impaired. Primary low responsiveness after vaccination seems to occur rarely: recent investigations of “low responders” after TBE vaccination show

that low cellular, humoral, and cytokine response levels, particularly IL-2 and IFN- γ , correlate with each other. Although immune response may be impaired, there is consensus that TBE vaccination with the available vaccines will do no harm in immunocompromised patients.

Field Effectiveness Field effectiveness of TBE vaccines has been investigated systematically in Austria, where vaccination coverage reached a sufficient level to obtain robust data. A first calculation, covering the period 2000–2006, yielded a field effectiveness of 99% for regularly vaccinated (mainly with FSME-Immun[®]) persons and reached 95.5% for those with irregular immunization schedules. A further analysis covering the years 2010–2011 (including approximately one third of subjects vaccinated with Encepur[®]) showed similar results: effectiveness was 98.7% for regularly vaccinated subjects and 92.5% in those with irregular schedules respectively. These data are in accordance with only few reports of vaccine failure in fully vaccinated individuals during the last few decades.

It is estimated that in Austria around 4000 cases of TBE were prevented by vaccination between 2000 and 2011 and the yearly reported number of TBE cases fell to 10–15% compared with the prevaccination era levels.

16.4 TBE Vaccines: Adverse Events and Contraindications

Reactogenicity The formulations of the TBE vaccines have been refined several times over the past decades and this has significantly reduced their reactogenicity. WHO and Cochrane reviews on safety attested the two TBEV-Eu-based vaccines to be safe. Pharmacovigilance data from both manufacturers, including about 72 million doses of vaccines of both brands distributed from 2001 to 2009, indicate a combined rate of severe adverse events of 1.6–1.9 per 100,000 doses. These include a range of entities, usually coinciding with immunization, but not necessarily causally related.

Typical systemic adverse events in children include mild and short-lasting fever mainly associated with the first dose and with a very low frequency of less than 0.5% (medically accompanied cases) of vaccinated individuals in a cohort of more than 25,000 vaccinees. Other systemic adverse events include headache, fatigue, malaise,

Table 16.4 Safety and reactogenicity of Encepur® and FSME-Immun® (Source: SPC)

Vaccines and probability of occurrence (adverse events/doses)	≥1/10	≥1/100 <1/10	≥1/1000 <1/100	≥1/10,000 <1/1000	Not known (based on single cases or small case series)
<i>FSME-Immun</i> ® First dose: <i>n</i> = 3512 Second dose: <i>n</i> = 3477 Third dose: <i>n</i> = 3277	Local reaction at injection site: Redness, swelling, induration	Headache, nausea Myalgia, arthralgia Malaise, fatigue	Lymphadenopathy Vertigo Vomiting Fever (only exceptionally >39 °C)	Acute allergic reactions Somnolence Diarrhea, abdominal pain	Aggravation of autoimmune disease Visual impairment, photophobia, meningismus, epilepsy, encephalitis, neuritis, tachycardia Urticaria, pruritus, exanthema Flu-like symptoms, weakness, edema
<i>Encepur</i> ® (pooled data from clinical studies and postmarketing surveillance)	Transient pain at injection site; general malaise, myalgia Headache	Redness, swelling at injection site Flu-like symptoms Nausea, arthralgia	Arthralgia and myalgia (neck)	Granuloma at injection site Lymphadenopathy Neuritis-like symptoms Diarrhea Systemic allergic reactions such as urticaria, dyspnea, bronchospasm, hypotension	Extremely rare: Guillain-Barré syndrome

muscle pain, and joint pain. Local redness, injection site pain, and itching may also occur. For details see [Table 16.4](#). Allergic reactions to vaccine components occur only occasionally.

Contraindications The TBE vaccines are contraindicated in persons with acute diseases. Allergies to vaccine components also constitute contraindications; in the case of egg protein allergy, contraindication is restricted to severe forms, i.e., with anaphylactic reactions. In all other patients, appropriate precautionary measures and supervision after immunization should be applied. Chronic diseases, including those affecting the central nervous system, are not a contraindication for TBE vaccination. However, the phenomenon of coinciding changes in the natural course of such underlying diseases should be discussed with the patients or their parents before immunization.

16.5 TBE Vaccination Recommendations

With more than 30 years of experience with the use of TBE vaccines in Europe, there is ample evidence for their positive public health impact. Most European countries, especially those with endemic TBE areas, do recommend TBE vaccination for their populations at risk, including travelers to endemic areas outside the country. In accordance with the labeled licensure of the vaccines, this includes children 1 year of age or older. In contrast, Austria is the only European country to date with a universal vaccination recommendation, reflecting the high burden of TBE in the pre-immunization era and the wide distribution of endemic areas in that country. The current recommendations for selected European countries are listed in [Table 16.5](#).

Table 16.5 Vaccination recommendation for TBE

Country	Recommendation status
Albania	No policy
Austria	Universal recommendation \geq 1 year of age
Bulgaria	No policy
Croatia	For highly endemic areas and occupational exposure
Czech Republic	For highly endemic areas and occupational exposure
Estonia	Recommended for populations at risk ^a
Finland	Recommended for endemic areas
Germany	Recommended for endemic areas
Hungary	Recommended for populations at risk ^a
Italy	Recommended on a district level
Latvia	Recommended for populations at risk ^a
Lithuania	Recommended for populations at risk ^a
Norway	Recommended for high-risk populations
Poland	Recommended for high-risk populations
Romania	No policy
Serbia	No policy
Slovakia	For highly endemic areas and occupational exposure
Slovenia	For highly endemic areas and occupational exposure
Sweden	Recommended for highly and moderately endemic districts, above 1 or 3 years of age respectively
Switzerland	Recommended for individuals living in or travelling to endemic areas, in general \geq 6 years of age

Adapted from Hombach et al. (2016)

^aNo detailed specifications available

In addition, there is a tendency in Europe to limit vaccination recommendations to older age groups, as pediatric TBE cases tend to be less severe, although there is growing evidence that TBE cases in children may also take a severe clinical

course and long-term outcome after TBE in children is underestimated. Cost–benefit calculations of TBE vaccination for endemic areas is mostly not available; only one study predicted savings of \$80million for Austria from a general vaccination recommendation. More recent data from Slovenia clearly indicated the cost-effectiveness of TBE vaccination for a highly endemic country.

Further Reading

- BMG. Nationaler Impfplan der Republik Österreich. Bundesministerium für Gesundheit, 2016. ► <http://bmg.gv.at/cms/home/attachments/2/8/1/CH1100/CMS1452867487477/impfplan.pdf>.
- Cizman M, et al. Severe forms of tick-borne encephalitis in children. *Wien Klin Wochenschr.* 1999;111(12):484–7.
- Demicheli V, Debalini MG, Rivetti A. Vaccines for preventing tick-borne encephalitis. *Cochrane Database Syst Rev.* 2009(1):CD000977.
- ECDC. Epidemiological situation of tick-borne encephalitis in the European Union and European Free Trade Association countries. ECDC: Stockholm; 2012.
- Fowler A, et al. Biomarkers in cerebrospinal fluid of children with tick-borne encephalitis. *Pediatr Infect Dis J.* 2016;35(9):961–6. doi:10.1097/INF.0000000000001210.
- Heinz FX, et al. Vaccination and tick-borne encephalitis, central Europe. *Emerg Infect Dis.* 2013;19(1):69–76.
- Kaiser R. Tick-borne encephalitis. *Infect Dis Clin N Am.* 2008;22(3):561–75.
- Kollaritsch H, et al. Vaccines and vaccination against tick-borne encephalitis. *Expert Rev Vaccines.* 2012;11(9):1103–19.
- Lindquist L. Tick-borne encephalitis. *Handb Clin Neurol.* 2014;123:531–59.
- Loew-Baselli A, et al. Prevention of tick-borne encephalitis by FSME-IMMUN® vaccines: review of a clinical development programme. *Vaccine.* 2011;29:7307–19.
- Orlinger KK, et al. A tick-borne encephalitis virus vaccine based on the European prototype strain induces broadly reactive cross-neutralizing antibodies in humans. *J Infect Dis.* 2011;203(11):1556–64.
- Paulke-Korinek M, et al. Factors associated with seroimmunity against tick borne encephalitis virus 10 years after booster vaccination. *Vaccine.* 2013;31(9):1293–7.
- Plotkin, SA, Orenstein, W, Offit, PA, Edwards, KM. *Plotkin's Vaccines.* 7th Edition. Philadelphia, PA: Elsevier; 2017
- Pollabauer EM, et al. Clinical evaluation to determine the appropriate paediatric formulation of a tick-borne encephalitis vaccine. *Vaccine.* 2010a;28(29):4558–65. doi:10.1016/j.vaccine.2010.04.075.
- Pollabauer EM, et al. Comparison of immunogenicity and safety between two paediatric TBE vaccines. *Vaccine.* 2010b;28(29):4680–5. doi:10.1016/j.vaccine.2010.04.047.
- Prymula R, et al. Antibody persistence after two vaccinations with either FSME-IMMUN® Junior or ENCE®(R) Children followed by third vaccination with FSME®MUN(R) Junior. *Hum Vaccin Immunother.* 2012;8(6):736–42.

- Schösser R, et al. Irregular tick-borne encephalitis vaccination schedules: the effect of a single catch-up vaccination with FSME-IMMUN. A prospective non-interventional study. *Vaccine*. 2014;32(20):2375–81.
- Schuler M, et al. Epidemiology of tick-borne encephalitis in Switzerland, 2005 to 2011. *Euro Surveill*. 2014;19(13): pii: 20756.
- WHO. Vaccines against tick-borne encephalitis. WHO position paper. *WER*. 2011;24(86):241–56.
- Wittermann C, Petri E, Zent O. Long-term persistence of tick-borne encephalitis antibodies in children 5 years after first booster vaccination with Encepur Children. *Vaccine*. 2009a;27(10):1585–8.
- Wittermann C, Schondorf I, Gniel D. Antibody response following administration of two paediatric tick-borne encephalitis vaccines using two different vaccination schedules. *Vaccine*. 2009b;27(10):1661–6.
- Zent O, et al. Safety, immunogenicity and tolerability of a new pediatric Tick-Borne Encephalitis (TBE) vaccine, free of protein-derived stabilizer. *Vaccine*. 2003;21(25–26): 3584–92.



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Tuberculosis Vaccines

Federico Martinon-Torres and Carlos Martin

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17.1 Introduction

Tuberculosis (TB) is one of the oldest human diseases known. Today, it is the leading cause of death worldwide exceeding those attributed to HIV. Despite all the efforts to fight it since the discovery of *Mycobacterium tuberculosis* by Robert Koch in 1882, it is still responsible for nearly two million deaths per year worldwide.

Tuberculosis is primarily a pulmonary disease caused by *M. tuberculosis* and transmitted via the respiratory route, which could present different manifestations and affect bones, the central nervous system, and lymph nodes, and the progression of the disease can have several outcomes, largely determined by the response of the host immune system. TB is a major contributor to mortality in under 5-year-olds in TB-endemic settings.

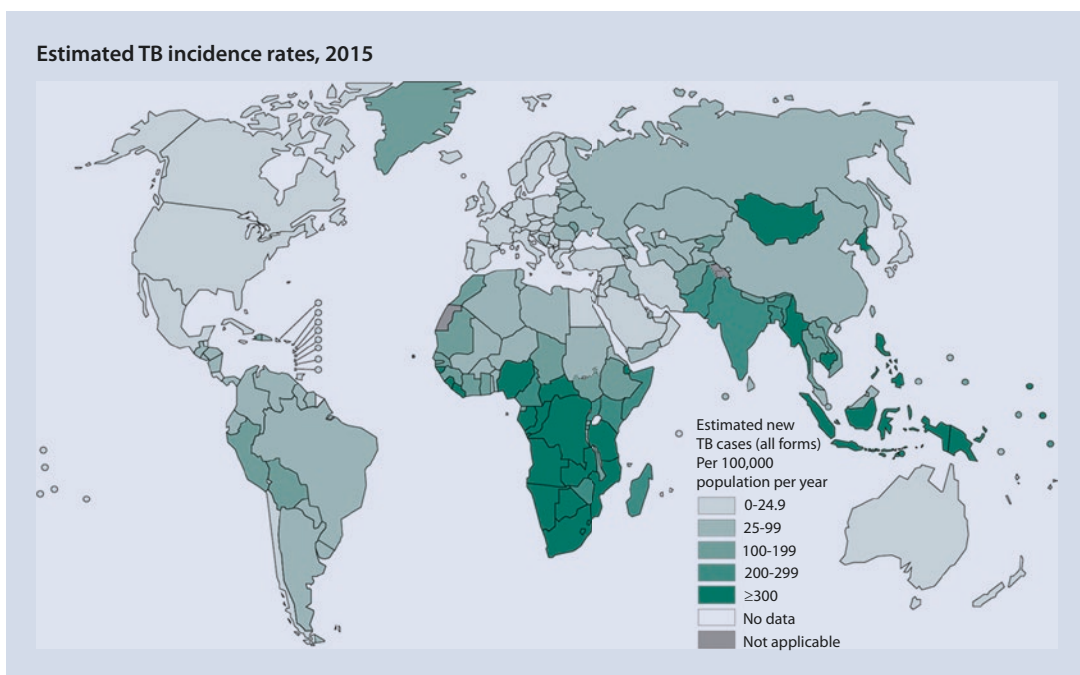
Bacillus Calmette–Guérin (BCG) is the current vaccine against TB. BCG provides a strong protection against disseminated forms of the disease in infants and young children, but confers very limited protection against pulmonary forms of TB, which are responsible for the transmission of the disease. Today, BCG is included in the immunization schedule for TB-endemic countries, with a global coverage at birth close to 90% worldwide. In Europe, some countries, such as Portugal and Ireland, cur-

rently recommend BCG vaccination for everyone, whereas others such as France used to recommend BCG vaccination for everyone, but not any longer (for information concerning country's BCG policies and practices visit ► <http://www.bcgatlas.org/> and for Europe ► <http://vaccine-schedule.ecdc.europa.eu/Pages/Scheduler.aspx>).

The goals of the World Health Organization's (WHO) 2015 End TB Strategy include a 95% reduction in TB deaths and a 90% global disease reduction by 2035; for this, it is necessary to develop a comprehensive and appropriate approach that includes new and more effective vaccines, in addition to improved diagnostics and treatment.

17.2 Tuberculosis Epidemiology

In 2016, the WHO estimated that out of the 10.4 million incident cases of TB in 2015, a total of 1.8 million people died from TB. Approximately one million deaths occurred among children under 15. Approximately 60% of these cases occurred in six countries (China, India, Indonesia, Nigeria, Pakistan, and South Africa) with the highest TB burden (■ Fig. 17.1). Furthermore, 480,000 people had developed multidrug-resistant TB by 2015. Also in 2017, the European Union/European



■ Fig. 17.1 Worldwide tuberculosis data (WHO report 2016). Estimated new TB cases (any form) per 100,000 population corresponding to the year 2015



■ Fig. 17.2 Tuberculosis notification rates of new tuberculosis cases and relapses per 100,000 population, European Region, 2015 (from 2017 WHO/ECDC tuberculosis report)

Economic Area (EU/EEA), the European Centre for Disease Control (ECDC), and the WHO reported on the TB situation according to 2015 data, stating 32,000 estimated deaths from TB and around 323,000 new TB cases, with an average notification rate of 34.5 cases per 100,000 population (■ Fig. 17.2). The WHO 2017 Report estimated an incidence for Europe of 35.5 cases per 100,000 population (range 32.9–38.3) and the multi-drug-resistant (MDR) TB of 14 per 100,000 population (range 12–15) and with a mortality of 3.5 per 100,000 population (range 3.4–3.6).

17.3 BCG Vaccine

The BCG vaccine is currently the only licensed vaccine against TB in use today and is one of the most widely administered vaccines in the world. Around four billion BCG doses have been administered worldwide in history, principally in the setting of routine newborn immunization (as recommended by the WHO). Today, global immunization BCG coverage at birth is estimated to be close to 90% (■ Figs. 17.3 and 17.4).

The original strain of *Mycobacterium bovis* BCG strain was developed in 1921 at the Pasteur Institute with attenuation through serial passage of an isolate from a cow with tubercular mastitis. This isolate was subsequently distributed to several laboratories in the world and a number of strains developed. Before the adoption of freeze-drying in the 1960s, the different laboratories preserved their strain by repeated sub-culture passages, and this resulted in the appearance of different BCG substrains that became designated by the laboratory. Genomic analysis of BCG strains has documented multiple molecular changes. The main reason for BCG attenuation is the loss of the region of difference 1 (RD1) associated with subsequent loss of the immunodominant virulence factor, the early secretory antigen of 6 kDa (ESAT-6) and CFP10, both used in that interferon- γ release assay (IGRA), to differentiate BCG vaccination from *M. tuberculosis* infection. Multiple other deletions probably contribute to phenotypic differences between BCG strains and although there are clear reactogenicity differences, it is not clear whether strain differences are a significant factor contributing to the variable efficacy of BCG observed in clinic.

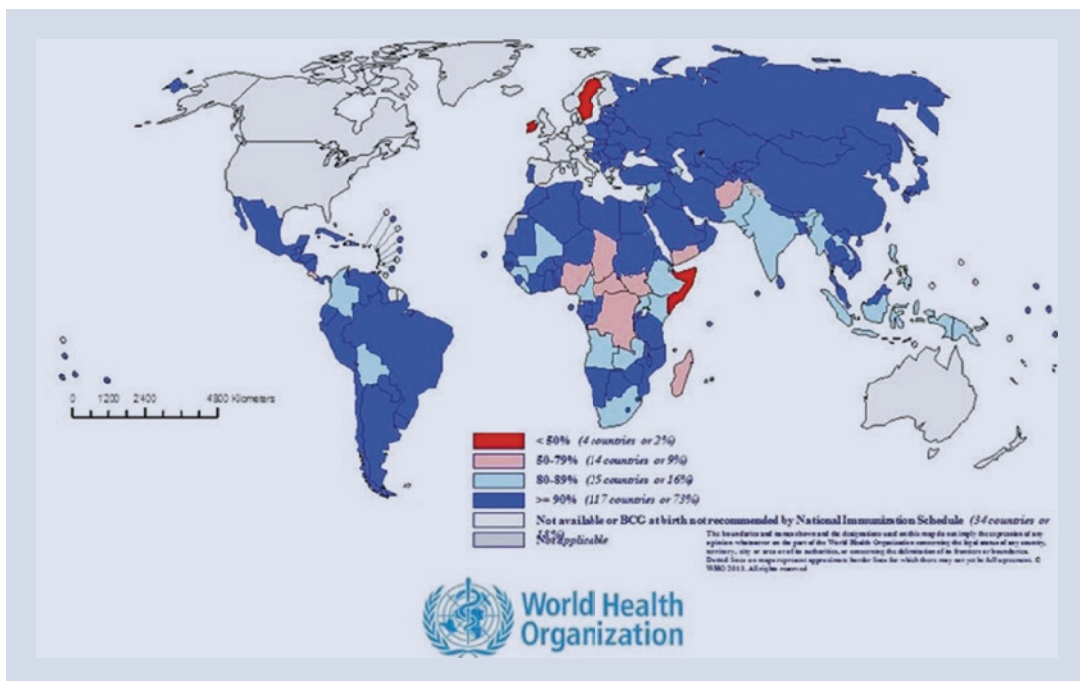
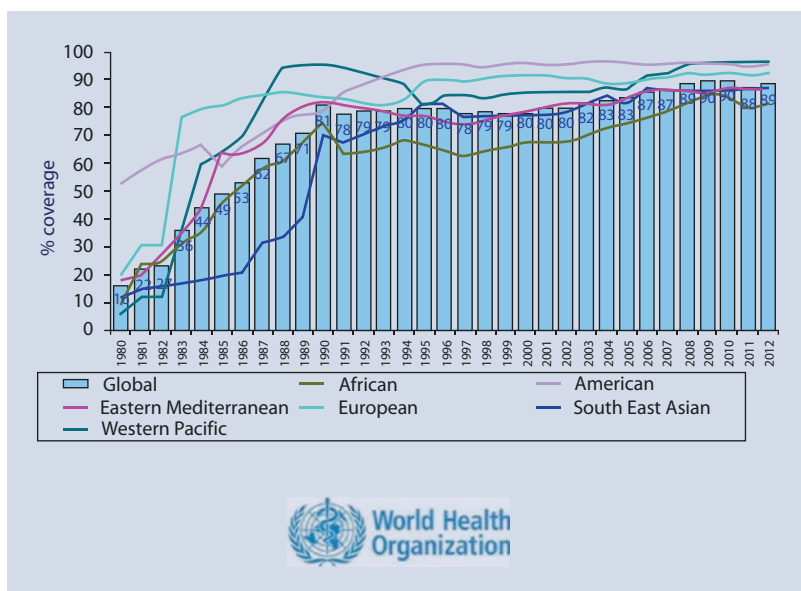


Fig. 17.3 Map of the immunization coverage with BCG at birth in 2012 (Source WHO; WHO/UNICEF coverage estimates 2012 revision, July 2013. 194 WHO Member

States. Map production: Immunization Vaccines and Biologicals (IVB). World Health Organization)

Fig. 17.4 Global immunization 1980–2012, BCG coverage at birth, global coverage at 89% in 2012 (Source WHO; Source: WHO/UNICEF coverage estimates 2012 revision. July 2013 Immunization Vaccines and Biologicals (IVB), World Health Organization)



Currently, five main strains account for more than 90% of the vaccines in use worldwide in international immunization programs, each strain possessing different characteristics. The agreed terminology for the strains include the Pasteur 1173 P2, the Danish 1331, the Glaxo 1077 (derived from

the Danish strain), the Tokyo 172-1, the Russian BCG-I; the Moreau RDJ strain is used mainly in Brazil. BCG vaccine shortages have been reported in many countries. These shortages started in 2013 and continued into 2015. The United Nations Children’s Fund (UNICEF) is the main supplier

of BCG vaccine to TB-endemic countries. Two of its four suppliers, the Statens Serum Institut in Denmark and the Serum Institute of India experienced technical difficulties that resulted in reduced production capacity. Global demand for BCG is estimated at 260 million doses per year. UNICEF reported shortages of 8 million doses in 2013, of 23 million doses in 2014, and of 17 million doses in 2015.

The BCG vaccine is administered intradermally, after reconstitution of a lyophilized composition. After reconstitution, every 1 ml of vaccine contains $2\text{--}8 \times 10^5$ cfu of *M. bovis* live attenuated BCG. The vaccine should be stored between 2° and 8 ° C. When reconstituted it should be protected from light.

Vaccine dosage:

- For adults and children >12 months, 0.1 ml of the reconstituted vaccine is recommended.
- For infants <12 months, 0.05 ml of the reconstituted vaccine is recommended.

Currently, there are new vaccines against tuberculosis under development, some of which are designed to boost the effects of BCG and others as BCG replacement vaccines (see further).

17.4 Methods of Administration

The injection site should be dry and clean. If the skin is swabbed with an antiseptic (such as alcohol), this should be allowed to evaporate completely before the injection is given.

The BCG vaccine should be administered by personnel trained in the intradermal technique, using a syringe of 1 ml subgraduated into hundredths of a milliliter (1/100 ml) fitted with a short bevel needle (25G/0.50 mm or 26G/0.45 mm). Jet injections or multiple puncture devices should not be used.

The vaccine should be injected strictly intradermally in the arm, over the distal insertion of the deltoid muscle onto the humerus (approximately one third down the upper arm) as follows (■ Fig. 17.5):

- The skin is stretched between thumb and forefinger.
- The needle should be almost parallel to the skin surface and slowly inserted (bevel upward), approximately 2 mm into the superficial layers of the dermis.



■ Fig. 17.5 Administration of the BCG vaccine. The skin is stretched between thumb and forefinger. The needle should be almost parallel with the skin surface and slowly inserted (bevel upward), approximately 2 mm into the superficial layers of the dermis. The needle should be visible through the epidermis during insertion. The injection is given slowly. A raised, blanched bleb is a sign of correct injection (Picture courtesy of Dr Jesper Kjærgaard, Copenhagen University Hospital)

- The needle should be visible through the epidermis during insertion.
- The injection is given slowly.
- A raised, blanched bleb is a sign of correct injection.

The injection site is best left uncovered to facilitate healing.

17.5 Efficacy of BCG

The efficacy of the current TB vaccine BCG is consistent against the severe forms of the disease (meningeal and miliary TB), but is limited against pulmonary forms of the disease; this disease manifestation is responsible for transmission – fueling the growing epidemic worldwide. The most controversial aspect of BCG is its variable efficacy when used in different trials, with variable, geographically dependent, efficacy against pulmonary disease. BCG does not seem to protect against disease when given to people already infected or sensitized to environmental mycobacteria, which could explain the geographic variation. Furthermore, until recently, it was not possible to establish whether the protective effect of BCG vaccination against disease stemmed from its action in preventing acquisition of infection, or limited to the prevention of progression from infection to clinical disease. A systematic review and meta-analysis conducted in 2014 demonstrated that the

BCG vaccine reduced infections by 19–27% and progression to active TB by 71%.

Primary vaccination of newborns and infants appears to confer better protection than older children and adults, the absence of prior *M. tuberculosis* infection or sensitization with environmental mycobacteria is associated with higher efficacy of BCG against pulmonary tuberculosis and possibly against miliary and meningeal tuberculosis. In contrast, the immune response to mycobacterial antigens in older children and adults living in areas that are endemic for tuberculosis appear to have higher background immunity than those living in non-endemic areas. This could have an influence on the relative efficacy of BCG when administered to older children and adults living in TB-endemic regions.

The possible reasons for variable efficacy include:

- Genetic variability of the population.
- Environmental factors as suggested by the relatively good efficacy seen in temperate regions compared with tropical regions of the globe.
- Background exposure to the disease TB: previous exposure may both limit the replication of BCG (“blocking”) and/or confer protection equivalent to BCG (“masking”).
- Nonspecific immune responses against TB mycobacteria by non-TB mycobacteria.
- Exposure to parasites that skew the immune response toward a Th2 type of response rather than a Th1 type of response, the latter believed to be most important for immunoprotection.
- The use of different strains of BCG with potentially different efficacy. However, a recent meta-analysis of trials, including 18 studies reporting on protection against pulmonary and six reporting on protection against miliary or meningeal tuberculosis, showed no evidence that the efficacy of BCG was influenced by vaccine strains.

17.6 Immune Response to BCG

The immune response to primary BCG immunization has been evaluated in different studies in children demonstrating that there is a BCG-

associated induction of CD4⁺ and CD8⁺ T cells, interferon (IFN)- γ a⁺, interleukin (IL)-2⁺, tumor necrosis factor (TNF)- α ⁺, and polyfunctional CD4⁺ T cells.

As there is a lack of correlate of protection for TB, immunological studies of infant BCG immunization cannot currently be applied to determine immunization policy because a surrogate marker of BCG-induced protection is yet to be identified.

The BCG vaccine is administered intradermally. As natural infection and sensitization to *M. tuberculosis* in humans usually occurs via the respiratory route, research is being conducted on the respiratory administration of BCG.

The BCG vaccination reduces rates of *M. tuberculosis* infection and provides strong protection against disseminated forms of the disease in infants and young children; TB is a major contributor to under-5 mortality in TB-endemic settings. In the last few years, epidemiological and trial evidence in humans have supported the conclusion that BCG vaccination leads to several beneficial heterologous effects on all-cause mortality. BCG vaccination reduces all-cause mortality through beneficial nonspecific effects on the immune system; the importance of these effects has been formally recognized by the WHO. These “non-specific” beneficial effects suggest improved survival as the result of enhanced immune protection against nonrelated infections. Mechanisms for this heterologous effect are identified as immune alternative cross-reactivity and the recently described “immune training” effect of vaccination. This “training” targeted on the cells of the innate immune system could be related to epigenetic reprogramming of innate cells through a NOD2-related mechanism. Trained cells become more efficient at immune response against non-related pathogens after vaccination (see also ► Chap. 1).

17.7 Vaccination Schedules and Indications

The International Union Against Tuberculosis and Lung Disease (IUATLD) has suggested a number of criteria according to which it may be reasonable for a country to move from a policy of systematic vaccination with BCG to the selective vaccination of high-risk groups.

The IUATLD and WHO recommend suspension of systematic BCG only if:

- There is an effective reporting system and the average annual notification rate of smear-positive pulmonary tuberculosis is <5 per 100,000.
- The average annual notification rate of tuberculous meningitis is <1 per ten million inhabitants in the last 5 years.
- The average annual risk of tuberculosis infection is <0.1%.

The BCG vaccination is considered strictly necessary in the following cases:

- Newborn vaccination is recommended in countries with high incidence of TB.
- Children without PPD exposed to smear-positive patients with poor compliance or refusal of treatment, or when the treatment does not get the negative sputum (persistently smear-positive patients).
- Children without PPD who move to live in highly TB-endemic countries, especially where control programs and access to appropriate treatment is not possible and where the prevalence of multidrug-resistant TB is high.

17.7.1 Administration of BCG in HIV Patients

In countries with a high prevalence of TB and HIV, it is important to exercise caution when BCG is administered routinely owing to the risk of disseminated BCG in HIV-infected infants (ranges of 400–1300 per 100,000 doses administered). Therefore, BCG vaccination is not appropriate for infants or adults with known HIV infection (or other immunodeficiency) or for those patients with a high degree of suspicion for HIV infection, even if unconfirmed by laboratory results.

The BCG vaccination should be administered to asymptomatic infants born to mothers with unknown HIV status in countries with a high TB prevalence. However, for asymptomatic infants with unknown HIV status born to mothers known to be infected with HIV, the optimal approach to BCG vaccination is uncertain. At present, the WHO recommends that routine childhood BCG

immunization be continued until all elements of an HIV-testing program can be implemented.

In countries with a low incidence of TB, BCG immunization may be considered in children ≤5 years in the following circumstances:

- The child is continuously exposed to an untreated or ineffectively treated patient who has infectious pulmonary TB and neither separation from the infectious patient nor long-term primary preventive therapy is feasible.
- The child is continuously exposed to a patient who has infectious pulmonary TB caused by *M. tuberculosis* strains resistant to isoniazid and rifampin, and separation from the infectious patient is not feasible.
- Children moving to Europe from endemic countries.

17.7.2 Exposure to MDR-TB

The efficacy of BCG vaccination for persons who are travelling to endemic areas with expected exposure to drug-resistant tuberculosis is uncertain. However, given the potentially significant risk of multidrug-resistant TB treatment failure, together with the relatively low rate of complications related to BCG vaccination in immunocompetent individuals, some favor administering BCG vaccination to unvaccinated, tuberculin-negative individuals exposed to multidrug-resistant TB. Further studies are needed to reconcile the protective efficacy of BCG vaccination in the setting of multidrug-resistant TB exposure among older children and adults.

17.8 Administration with Other Vaccines or Products

The BCG vaccine can be administered concomitantly with other vaccines without increasing side effects. The immunogenicity obtained is similar to that obtained with separate administration. The main limitation is the need for administration in different anatomical sites.

Co-administration with any other vaccine is possible (including other live vaccines). BCG enhances T and B cell responses to unrelated vaccine antigens. Unexpectedly, BCG vaccination

has affected responses to various vaccines differently whether administered at the time of priming, boosting, or even before priming. BCG enhances both Th1 and Th2 cytokine responses to unrelated antigens and extended its influence on antibody responses to oral polio vaccine (see ▶ Chap. 1).

Regional lymphadenitis cases have been reported after administering other vaccines in the same place in which BCG vaccination was applied. Therefore, it is not recommended to administer any other vaccine in the same limb within 3 months of BCG administration.

It is also recommended not to administer BCG vaccine if the patient has been treated with antibiotics during the previous 30 days.

17.9 Safety

Overall, the BCG vaccine is well tolerated. After 2–6 weeks of receiving the vaccine a small papule appears that increases in size and changes into an ulcer. The lymphatic nodules in cervical and axillary areas may be temporarily enlarged.

After a period of about 3 months, a scar appears, which is permanent (■ Fig. 17.6).

The safety of BCG vaccination has been widely proven because more than four billion units have been administered all over the world since 1921. The most common complication found with its use is the occurrence of regional lymphadenitis with or without suppuration (■ Table 17.1). Other types of local reactions, such as abscess or ulcers, are rarely seen and are more related to the administration technique, which must be carried out under strictly aseptic conditions and always intradermally.

17.10 Contraindications of the BCG Vaccine

- Immunocompromised patients, given that BCG vaccination is a live vaccine: congenital or acquired immunodeficiency due to immunosuppressive drugs such as corticosteroids, alkylating antineoplastic agents, radiation... Patients with HIV (with the exceptions as mentioned above).



■ Fig. 17.6 Scarring after BCG vaccination. After 1–6 weeks, a small, red blister may appear where the injection was given. This should heal in a few weeks. After 6–12 weeks, the blister may turn into a small, weeping sore. The sore may take up to 3 months to heal, and leave a small scar (picture courtesy of Dr Jesper Kjærgaard, Copenhagen University Hospital)

■ Table 17.1 Common local and systemic reactions to BCG (expressed as a percentage)

Systemic reactions	
Anorexia	<5%
Fever	<1%
Systemic reaction to vaccine	<0.003%
Asthenia	<5%
Osteitis	<0.0001%
Local	
Abscess	<0.01%
Lymphadenopathy	1–2%
Keloid	2–4%
Pain	95%
Erythema	95%
Ulceration	95% after 14 days
Pustule	95%
Swelling	95%
Pain	95%
Scar	95%

- Patients with a positive PPD skin test or with TB.
- BCG should be avoided in pregnancy, especially in the first trimester, as it is a live vaccine.
- Hypersensitivity to the BCG vaccine or any of its components.
- Burns patients.
- Malnourished children.
- Active infection.
- Preterm infants with a birth weight less than 2.5 kg.
- Patients with blood diseases.
- Oncology patients.
- Patients who are already undergoing TB treatment.
- Patients with skin diseases. The area of insertion of the vaccine should be free of lesions.

17.11 General Warnings

The BCG vaccine, when administered at birth, is highly reactogenic but safe. Swelling and scarring are common.

Anaphylactic reactions are seen only rarely, but their management should be prepared in advance and the patient should be closely observed for 15–30 min after administration.

The vaccine should be administered intradermally. Deeper administration could produce lymphadenitis or abscesses.

If an overdose of the vaccine is given, it may lead to suppurative lymphadenitis or, rarely, systemic infection.

17.12 New Tuberculosis Vaccines

The most successful, licensed vaccines available today induce neutralizing antibodies that provide protective immunity. Animal and human studies of TB, however, suggest that a robust cellular immune response is required for protection against *M. tuberculosis* infection and disease. For this reason, most current clinical TB vaccine candidates are based on a variety of vectors, adjuvants, and antigens that induce classical TH1 cytokines such as

IFN- γ or TNF- α from either CD4+ or CD8+ T cells.

The current global TB vaccine portfolio consists of three main types of vaccine strategies, which are either preventive or therapeutic. The preventive strategies embrace the priming BCG replacement vaccines and subunit BCG boosts (or enhancers). Therapeutic candidates that have reached clinical development to date comprise inactivated forms of mycobacteria being developed for patients with active TB. They receive TB drug therapy also in addition to this vaccine to shorten the duration of the therapy and to reduce the likelihood of recurrence after completion of treatment.

There are two main strategies for which research on prophylactic vaccines for TB prevention is focused:

- A better vaccine than the current BCG: more efficacious and longer lasting, or preventive of TB infection and disease in infants who have not been infected with *M. tuberculosis* (BCG replacement strategy).
- A BCG booster vaccine: for use as a heterologous boost in BCG-primed individuals, where BCG is given at birth and then boosting is applied with specific *M. tuberculosis* antigens. This strategy may be indicated in those patients who are latently infected, preventing infection and/or progression to active disease. Subunit vaccines are based on one or a few *M. tuberculosis*-specific protein antigens using viral vectors or adjuvants as the delivery system.

Replacement strategies for BCG are divided into two classes of live vaccines, namely recombinant BCG (rBCG) and live-attenuated *M. tuberculosis*. The rBCG candidates are designed to improve the efficacy of BCG by the insertion of other genes. Rationally attenuated *M. tuberculosis* of human origin is considered a classical Pasteurian approach to human vaccinology, expected to mimic natural infection without causing disease (Table 17.2). Two BCG replacement vaccines are in the advanced stages of development: rBCG VPM1002 and MTBVAC. rBCG VPM1002 (rBCG Δ UreC::hly) expresses listeriolysin (hly) from *Listeria monocytogenes* with deletion of

Table 17.2 Description of different TB vaccine candidates studied in clinical trials at the time of writing

Candidate name/identifier	Developer	Type	Phase I	Phase IIa	Phase IIb	Phase III
Ad5 Ag85A	McMaster University, CanSino	Human adenovirus antigen	X			
TB/Flu-04 L	RIBSP	Attenuated influenza	X			
DAR-901	Dartmouth, Aeras	Heat-killed NTM	X			
ChAdOx1.85A/MVA85A	Oxford	Chimp adenovirus/modified vaccine	X			
MTBVAC	TBVI, University of Zaragoza, Biofabri	Live attenuated TB	X			
VPM 1002	Max Planck, VPM, TBVI, Serum Institute of India	Modified recombinant BCG		X		
ID93 + GLA-SE	IDRI, Aeras	4 Ag adjuvanted fusion protein		X		
H1 + IC31	SSI, TBVI, EDCTP	2 Ag adjuvanted fusion protein		X		
RUTI	Archivel Farma	Lysate of MTB		X		
H4/Aeras-404 + IC31	SSI, Sanofi-Pasteur, Aeras, Intercell	2 Ag adjuvanted fusion protein		X		
H56/Aeras-456 + IC31	SSI, Aeras, Intercell	3 Ag adjuvanted fusion protein		X		
Crucell Ad35/Aeras-402	Crucell, Aeras	h Adenovirus 35 3 antigen		X		
M72 + AS01E	GlaxoSmithKline, Aeras	2 Ag adjuvanted fusion protein			X	
<i>M. vaccae</i>	Anhui Zhifei Longcon, China	Lysate of NTM				X

NTM nontuberculous mycobacteria, *MTB Mycobacterium tuberculosis*, *RIBSP* Research Institute for Biological Safety Problems, *TBVI* Tuberculosis Vaccination Initiative, *VPM* Vakzine Projekt Management, *IDRI* Infectious Disease Research Institute, *SSI* Statens Serum Institut, *EDCTP* European & Developing Countries Clinical Trials Partnership

the urease C (*ureC*) gene, MTBVAC is a live, rationally attenuated derivative of a human *M. tuberculosis* isolate, which belongs to lineage 4 (European–African–American), one of the most widespread lineages of *M. tuberculosis*. MTBVAC contains all the genes present in *M. tuberculosis* strains, including the genes that are deleted in *M. bovis* and BCG. MTBVAC contains two independent stable deletion mutations in the virulence genes *phoP* and *fadD26*. These deletions were generated in the absence of antibiotic

resistance markers, fulfilling the Geneva consensus requirements for progressing live mycobacterial vaccines to clinical trials. MTBVAC is the first live-attenuated *M. tuberculosis* vaccine to enter clinical trials and has completed a phase I trial; the safety and immunogenicity results of the phase I trial conducted at the University of Lausanne in PPD and HIV-negative adults were satisfactory and immunological results encouraging. When given at the same dose as BCG (5×10^5 cfu), MTBVAC has shown a comparable safety

profile to BCG and there were more responders in the MTBVAC group than in the BCG group, with a greater frequency of polyfunctional CD4⁺ central memory T cells. A notable finding in the first trial was the absence of ESAT-6- and CFP-10-specific T cell responses at the end of the study, suggesting that interferon- γ release assays (IGRAs) could be utilized as study endpoints in future efficacy trials to test efficacy against *M. tuberculosis* infection. The immunogenicity data show that MTBVAC is at least as immunogenic as BCG. Taken together, these data supported the advanced clinical development in high-burden countries where TB is endemic. MTBVAC is currently in clinical development, with the primary target population being newborns (BCG replacement vaccine) and the secondary target being adolescents and adults (BCG revaccination).

Today, there are 13 different TB vaccines being studied in clinical trials (WHO Tb Report 2016), and many more in preclinical development. Five of these trials are based on whole cell mycobacteria. The rest of them are various subunit-based approaches in which *M. tuberculosis* antigens are expressed as recombinant proteins that are either formulated with adjuvants or presented in recombinant viral vectors.

At present, there are no accepted correlates of protection that, unto themselves, could support a decision to license a TB vaccine. Robust safety and immunogenicity data are needed for future efficacy trials of new TB vaccines. Therefore, the development of new vaccines against the pulmonary forms of TB are urgently needed for the control of TB.

■ ■ Potential Conflicts of Interest

CM is co-inventor on a composition of matter patent “tuberculosis vaccine” at the University of Zaragoza. There are no other conflicts of interest

Further Reading

Black GF, Weir RE, Floyd S, et al. BCG-induced increase in interferon-gamma response to mycobacterial antigens and efficacy of BCG vaccination in Malawi and the UK: two randomised controlled studies. *Lancet*. 2002;359:1393.

Blok BA, Arts RJ, van Crevel R, Benn CS, Netea MG. Trained innate immunity as underlying mechanism for the long-term, nonspecific effects of vaccines. *J Leukoc Biol*. 2015;98(3):347–56. doi:10.1189/jlb.5RI0315-096R.

Colditz GA, Brewer TF, Berkey CS, et al. Efficacy of BCG vaccine in the prevention of tuberculosis. Meta-analysis of the published literature. *JAMA*. 1994;271:698.

European Centre for Disease Prevention and Control (ECDC)/World Health Organization Regional Office for Europe. Tuberculosis surveillance and monitoring in Europe 2015. Stockholm. 2015.

Evans TG, Schragger L, Thole J. Status of vaccine research and development of vaccines for tuberculosis. *Vaccine*. 2016;34:2911–4.

Fine PE, Carneiro IA, Milstien JB, Clements CJ. Issues relating to the use of BCG in immunization programs: a discussion document. Geneva, Switzerland: Department of Vaccines and Biologicals, World Health Organization 1999. p.1. World Health Organization. 2011. The Immunological Basis for Immunization Series. Module 5: Tuberculosis.

Hoft DF. Tuberculosis vaccine development: goals, immunological design, and evaluation. *Lancet*. 2008;372:164.

Horvath CN, Shaler CR, Jeyanathan M, et al. Mechanisms of delayed anti-tuberculosis protection in the lung of parenteral BCG-vaccinated hosts: a critical role of airway luminal T cells. *Mucosal Immunol*. 2012;5:420.

Jensen KJ, Larsen N, Biering-Sørensen S, Andersen A, Eriksen HB, Monteiro I, Hougaard D, Aaby P, Netea MG, Flanagan KL, Benn CS. Heterologous immunological effects of early BCG vaccination in low-birth-weight infants in Guinea-Bissau: a randomized-controlled trial. *J Infect Dis*. 2015;211(6):956–67. doi:10.1093/infdis/jiu508.

Kay AW, Blish CA. Delayed BCG. Vaccination – time to take a shot. *J Infect Dis*. 2015:211–5.

Kleinnijenhuis J, van Crevel R, Netea MG. Trained immunity: consequences for the heterologous effects of BCG vaccination. *Trans R Soc Trop Med Hyg*. 2015;109(1):29–35. doi:10.1093/trstmh/tru168.

Mangtani P, Abubakar I, Ariti C, Beynon R, Pimpin L, Fine PEM, et al. Protection by BCG vaccine against tuberculosis: a systematic review of randomized controlled trials. *Clin Infect Dis*. 2014;58:470–80.

Marais BJ, Seddon JA, Detjen AK, van der Werf MJ, Grzemska M, Hesselting AC, et al. Interrupted BCG vaccination is a major threat to global child health. *Lancet Respir Med*. 2016;4:251–3.

Marinova D, Gonzalo-Asensio J, Aguilo N, Martin C. Recent developments in tuberculosis vaccines. *Expert Rev Vaccines*. 2013;12:1431–48.

Ota MOC, Vekemans J, Schlegel-Haueter SE, Fielding K, Sanneh M, Kidd M, et al. Influence of *Mycobacterium bovis* bacillus Calmette-Guérin on antibody and cytokine responses to human neonatal vaccination. *J Immunol*. 2002;168:919–25.

Roy A, Eisenhut M, Harris RJ, Rodrigues LC, Sridhar S, Habermann S, Snell L. Effect of BCG vaccination against *Mycobacterium tuberculosis* infection in children: systematic review and meta-analysis. *BMJ*. 2014;349:g4643. doi: <http://dx.doi.org/10.1136/bmj.g4643>

Soares AP, Kwong Chung CK, Choice T, et al. Longitudinal changes in CD4(+) T-cell memory responses induced by BCG vaccination of newborns. *J Infect Dis*. 2013;207:1084.

Spertini F, Audran R, Chakour R, et al. Safety of human immunisation with a live-attenuated Mycobacterium tuberculosis vaccine: a randomised, double-blind, controlled phase I trial. *Lancet Respir Med*. 2015;3:953–62.

Wilson ME, Fineberg HV, Colditz GA. Geographic latitude and the efficacy of bacillus Calmette-Guérin vaccine. *Clin Infect Dis*. 1995;20:982.

European Centre for Disease Prevention and Control/WHO Regional Office for Europe. Tuberculosis surveillance and monitoring in Europe 2017. Available at: http://www.euro.who.int/_data/assets/pdf_file/0020/334703/tuberculosis-surveillance-and-monitoring-in-europe-2017.pdf.

Pertussis Vaccines

Ulrich Heininger

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18.1 Burden of Pertussis Disease

The clinical characteristics of pertussis disease are highly dependent on the host's basic immunity. Most if not all neonates and young infants (i.e., <3 months of age) of mothers who were not immunized against pertussis during pregnancy develop a cough when exposed to *Bordetella pertussis*, the causative agent of pertussis (or whooping cough). At this young age, infants are highly vulnerable for complicated disease, which includes apnea (in 49–58% of affected individuals), the need for supplemental oxygen (59–100%) and/or mechanical ventilation (27–100%), and pulmonary hypertension (11–39%). In accordance, most deaths due to *B. pertussis* infection occur in neonates and young infants with a case fatality rate of 1–3%.

Typical pertussis is a three-stage disease and usually occurs in unimmunized older infants and children, less frequently in adolescents or adults: after an incubation period of 7–10 days the catarrhal phase begins with nonspecific nasal congestion, rhinorrhea, conjunctivitis, mild sore throat, and cough. Fever is uncommon. One to 2 weeks later, the paroxysmal stage follows. It is characterized by worsening coughs, cumulating in frequent paroxysmal spells, occurring day and night, with viscous secretions, vomiting, and the characteristic whoops terminating the coughing spell, but sometimes directly leading to the next one. Between these paroxysms, the patient appears well. After several weeks, the final convalescent stage of highly variable duration brings relief, with decreasing frequency and severity of coughing spells and accompanying symptoms.

Leukocytosis due to lymphocytosis is a hallmark of typical pertussis and the basis for most pulmonary complications, which may lead to respiratory failure with the need for exchange transfusion of extracorporeal membrane oxygenation.

In contrast, the clinical presentation of pertussis in immunized individuals and adolescents and adults, even if unimmunized, is frequently less typical, i.e., a nonspecific cough of variable duration. It frequently lasts for several weeks and because of the lack of characteristic signs such as

paroxysms, vomiting, and whooping, it often remains undiagnosed unless it is linked to a case of typical pertussis or a knowledgeable physician considers pertussis in the differential diagnosis and applies appropriate diagnostic tests.

Complications are rare with atypical pertussis. In contrast, in patients with typical pertussis, severe coughing episodes, pneumothorax, rib fracture, herniated intervertebral disc, epistaxis, subconjunctival hemorrhage, subdural hematoma, hernia, rectal prolapse, urinary incontinence, and carotid artery dissection have been reported to be consequences of increased intrathoracic pressure.

Importantly, all patients with pertussis – typical or less typical – are contagious and therefore play an important role in transmission chains.

18.2 Pertussis Epidemiology

As has been shown in longitudinal seroprevalence studies (with anti-pertussis toxin IgG antibody values as a sensitive and the only specific marker of infection), most *B. pertussis* infections (affecting up to 20% of any population per year) remain asymptomatic. Fewer individuals, 0.5–1% (or 500–1000 per 100,000 of the population per year), develop a cough of ≥ 2 weeks' duration due to *B. pertussis* infection and this is only detected in prospective studies. Among those, a variable fraction – depending on the basic immunity (see ► Sect. 8.1 above) – develop classic pertussis. In passive surveillance systems, the basis for nationwide mandatory reporting in many European countries, the yearly incidence of pertussis varies greatly, with values ranging from 0.01 to 96 per 100,000, where most countries report an incidence of approximately 10. These differences by all likelihood are not real, but can be explained by the heterogeneity of surveillance systems in place and their associated case definitions. In the future, it is hoped that all European countries, or at least all European Union Member States will use the pertussis case definition proposed by the European Centre for Disease Prevention and Control (ECDC; ► Box 18.1).

Box 18.1 Pertussis Case Definition and Case Classification Proposed by the European Centre for Disease Prevention and Control (ECDC)

Pertussis (*Bordetella pertussis*)

Clinical criteria:

Any person with a cough lasting at least 2 weeks and at least one of the following three features:

- Paroxysms of coughing
- Inspiratory “whooping”
- Post-tussive vomiting

Or

Any person diagnosed as having pertussis by a physician

Or

Apneic episodes in infants

Laboratory criteria:

At least one of the following three:

- Isolation of *Bordetella pertussis* from a clinical specimen
- Detection of *Bordetella pertussis* nucleic acid in a clinical specimen
- *Bordetella pertussis*-specific antibody response

Serology results need to be interpreted according to the vaccination status

Epidemiological criteria:

- An epidemiological link due to human-to-human transmission

Additional information:

Incubation period 6–20 days, most often 10 days

Case classification:

- A. Possible case
 - Any person meeting the clinical criteria
- B. Probable case
 - Any person meeting the clinical criteria and with an epidemiological link
- C. Confirmed case
 - Any person meeting the clinical and laboratory criteria

Note: The case definition and classification is that stipulated by the EU Commission Decision of 8 August 2012

shortly after *B. pertussis* was isolated, and the first results regarding protection were reported from the USA in 1925. After an animal model had been established (“Kendrick’s mouse protection test”) in 1947, standardization of vaccine production was possible and consecutive field studies in Great Britain were performed in the 1940s and 1950s. They demonstrated that the potency of wP vaccines as determined in the mouse protection test correlated with their clinical effectiveness in children. After that, wP, in combination with diphtheria and tetanus toxoids (DTP), was introduced into national immunization programs in many countries worldwide.

In the 1970s, concerns were raised based on serious adverse events (i.e., sudden infant death syndrome and various neurological illnesses including “encephalopathy” and epilepsy), which were reported following the use of DTP and were erroneously attributed to the wP component of the combination vaccine. It took decades to demonstrate that these events were coincidental rather than causally connected to DTP. Yet, these concerns, along with notable local side effects and fever induced by wP, led to the development of new pertussis vaccines. Such new acellular vaccines (see below) were first developed and then generally introduced for use in infants in Japan in 1981. Large field efficacy trials in Europe and Senegal, performed during the early 1990s, demonstrated better tolerability and acceptable efficacy of aP vaccines (■ Tables 18.1 and 18.2) and have paved the way for the licensure of various aP combination vaccine products ever since.

In Europe, all countries except Poland and Serbia have switched from wP to aP vaccines for the primary immunization series in infants at some point in time between 1995 and 2010. There are two main disadvantages of wP vaccines compared with aP vaccines: higher reactogenicity, especially fever, and less standardized production, leading to highly variable lot-to-lot performance with regard to effectiveness (■ Tables 18.1 and 18.2). Yet, many countries outside Europe – especially low- and middle-income countries – still use wP vaccines in various combinations of diphtheria and tetanus toxoid (DTwP), with or without further components such as Hib, hepatitis B, and IPV.

18.3 Pertussis Vaccines

18.3.1 Whole-Cell Pertussis Vaccines

All whole-cell pertussis (wP) vaccines contain killed *B. pertussis* organisms of various genetic backgrounds. The first wP vaccines were developed

Table 18.1 Comparative reactogenicity of whole-cell and acellular pertussis vaccines, by doses 1–3, as established in the United States Nationwide Multicenter Acellular Pertussis Trial

Adverse events	DTaP ^a (frequency in %)			DTP (frequency in %)		
	Dose 1 n = 1814	Dose 2 n = 1774	Dose 3 n = 1717	Dose 1 n = 370	Dose 2 n = 358	Dose 3 n = 342
<i>Local</i>						
Redness, any	13.5	17.1	21.5	49.4	47.7	47.6
Redness, >2 cm	1.3	0.9	1.7	8.6	6.1	3.2
Swelling, any	8.7	12.1	13.3	39.7	34.1	35.7
Swelling, >2 cm	1.7	1.4	2.2	16.5	9.5	5.6
Pain, moderate or severe	3.8	2.0	2.1	27.3	18.7	15.8
Pain, severe	0.2	0.1	0.1	9.7	6.1	3.8
<i>Systemic</i>						
Fever (temperature ≥ 37.8 °C [100.1 °F])	4.2	11.3	15.8	27.3	34.1	37.7
Fever (temperature ≥ 38.4 °C [101.1 °F])	0.4	1.2	2.2	3.0	5.3	9.9
Fussiness, moderate or severe	6.6	7.7	6.7	20.6	23.5	17.3
Fussiness, severe	2.0	1.6	1.3	3.8	7.0	4.7
Drowsiness	29.9	17.6	12.9	43.5	31.0	24.6
Anorexia	9.3	8.9	8.9	19.5	16.5	14.3
Vomiting	6.3	4.5	4.2	7.0	4.5	5.3
Use of antipyretic	39.3	36.7	36.3	60.5	59.8	61.4

Modified after Decker et al. (1995)

DTaP diphtheria-tetanus-acellular pertussis, DTP diphtheria-tetanus-pertussis

^aPooled data from 13 different DTaP products

18.3.2 Acellular Pertussis Vaccines

In the late 1970s, and throughout the 1980s and early 1990s, several vaccine manufacturers developed aP vaccines with the goal of better tolerability and similar efficacy compared with conventional wP vaccines. I former goal has clearly been reached (Table 18.1), but the latter unfortunately has not. Although aP vaccines (formulated as DTaP) performed better than one lot of DTwP vaccine produced in the USA when tested in efficacy trials in Italy and Sweden (Stockholm), overall efficacy estimates of DTwP vaccine were approximately 10% higher than those of DTaP after three or four doses in 3 + 0 (all doses in infants, no booster in the 2nd year of life) and 3 + 1 (with a booster dose in the 2nd year of life) immunization schedules respectively (Table 18.2).

18.4 Safety and Reactogenicity

Tolerability of DTaP vaccines is good and not different from that of DT vaccines without the aP component. A detailed comparison of DTaP and DTwP reactogenicity profiles, as established in the United States Nationwide Multicenter Acellular Pertussis Trial is shown in Table 18.1.

In the 1970s and 1980s, wP vaccines were held responsible for allegedly having caused “pertussis vaccine encephalopathy” in infants to the order of 1 per 330,000 doses within 7 days of immunization. However, careful investigations later demonstrated that what had been thought to be specific wP vaccine damage was in reality the result of various underlying morbidity with diverse etiopathogenesis, including genetic disorders such as the recently discovered SCN1A gene mutation,

Table 18.2 Comparative whole-cell and acellular pertussis vaccine efficacy as established in prospective randomized clinical trials

Country/ region	Study design	Schedule			
		Vaccine ^a efficacy	No. doses (age)	Typical pertussis (%)	Mild and typical pertussis (%)
Germany, Erlangen	Prospective cohort	aP-4	4 doses (3, 4, 6 + 15–18 months)	83	72
		wP	As above	93	83
Germany, Mainz	Household contact	aP-3	3 doses (3, 4, 5 months)	89	81
		wP	As above	98	Not reported
Germany, Munich	Case control	aP-2	4 doses (2, 4, 6, 15–25 months)	93	Not reported
		wP	As above	96	Not reported
Italy, Rome	Double-blind, prospective cohort	aP-3a	3 doses (2, 4, 6 months)	84	71
		aP-3b	As above	84	71
		wP	As above	36	23
Senegal	Household contact	aP-2	3 doses (2, 4, 6, 15–25 months)	74	Not reported
		wP	As above	92	Not reported
Sweden, Gothenburg	Double-blind, prospective cohort	aP-1	3 doses (3, 5, 12 months)	71	54
Sweden, Stockholm	Double-blind, prospective cohort	aP-2	3 doses (2, 4, 6 months)	59	42
		aP-3	As above	85	78
		wP	As above	48	41

wP whole-cell pertussis vaccine

^aaP-1 = single component acellular pertussis vaccine, aP-2 = 2-component acellular pertussis vaccine, etc.

leading to Dravet syndrome. In other words, what was observed and reported was coincidence rather than cause and effect. Even before rare, specific underlying morbidities were discovered, it was epidemiologically shown that the increased risk of onset of central nervous system disease potentially leading to brain damage within 7 days of immunization was offset by a decreased relative risk over the subsequent 3-week period such that the overall result was no increased risk for serious neurological disease with wP vaccines. However, despite such clear evidence against it, the myth of

“pertussis vaccine damage” continues to prevail, especially on obscure internet fora.

Hypotonic–hypo-responsive episodes (HHEs) have been reported after many vaccines used in infants, with or without aP or wP components. However, the risk of HHEs is approximately tenfold higher with DTwP vaccine than with DTaP vaccine (approximately 1 per 15,000 vs 1 per 1500 doses).

For children from the age of 4 years onward, adolescents, and adults without any upper age limit, acellular pertussis vaccines in combination with tetanus and diphtheria toxoids with reduced

diphtheria and pertussis antigen content – therefore referred to as “Tdap” (tetanus–diphtheria–acellular pertussis) – have been licensed in Europe and elsewhere.

18.5 Real-Life Effectiveness

Investigations into the real-life effectiveness of aP vaccines (and to a lesser extent, wP vaccines) are being conducted on an ongoing basis, with new evidence arising constantly. After the introduction of aP immunization programs in Europe in 1995 and onward, duration of protection after three or four doses was the focus of investigations. When it became apparent that efficacy waned with time after the last of three or four doses in infants and young children, pre-immunization booster doses were introduced in several countries in the early 2000s. Population-wide implementation of a fifth dose, usually at pre-school age, is more difficult to achieve than doses 4 and especially 1–3, and low uptake contributes to the limited effectiveness. Moreover, it has recently become clear that even after five doses, protection against pertussis does not last very long: in a matched case–control study from Washington State, USA, adolescents and young adults (11–19 years of age) with suspected, probable, and confirmed pertussis were identified, and vaccine effectiveness was calculated based on pertussis immunization history. Among those individuals who had received only acellular pertussis vaccines, Tdap vaccine effectiveness was 73% at 1 year and 34% at 2–4 years following their last pertussis vaccine dose. Similarly, waning immunity was shown in a study in Wisconsin, where a pertussis outbreak had occurred in 2012. Tdap effectiveness in preventing laboratory-confirmed pertussis decreased with increasing time since receipt of the last Tdap vaccine, with values of 75%, 68%, 34%, and 12% among those who received their last Tdap dose in 2012, 2011, 2010, and 2009/2008 respectively. Therefore, the introduction of further booster doses in adolescents, and even adults, was the next step that some but by far not all countries in Europe have taken in recent years. However, with aP vaccines of suboptimal effectiveness, control of pertussis is challenging if not impossible. Recently, this has raised discussions about the re-introduction of wP, for example, as part of sequential wP/aP immunization schedules. Although some wP vaccines do appear to be

more efficacious than any aP vaccine, there is lot-to-lot inconsistency with poor efficacy (<50%) for some wP products (■ Table 18.2). As, unfortunately, there is a lack of a reliable serological correlate of vaccine protection and no reliable animal models that would allow wP vaccine performance to be predicted, their use in the field is a constant lot-to-lot lottery.

18.6 Pertussis Vaccine Recommendations

Currently, so-called “2 + 1” and “3 + 1” DTaP immunization schedules are used in 8 and 23 European countries respectively, organized under the umbrella of the ECDC (■ Fig. 18.1). The first figure stands for the number of priming doses in infants (i.e., 2 or 3) and the “+1” stands for the reinforcing last dose of the primary series, usually given around the first birthday. The apparent heterogeneity in time points reflects variable interpretations of data by national immunization technical advisory groups to the governments, variable histories of the development of such recommendations, and associations with scheduled health care visits such as the “well baby visits,” which again may vary from country to country.

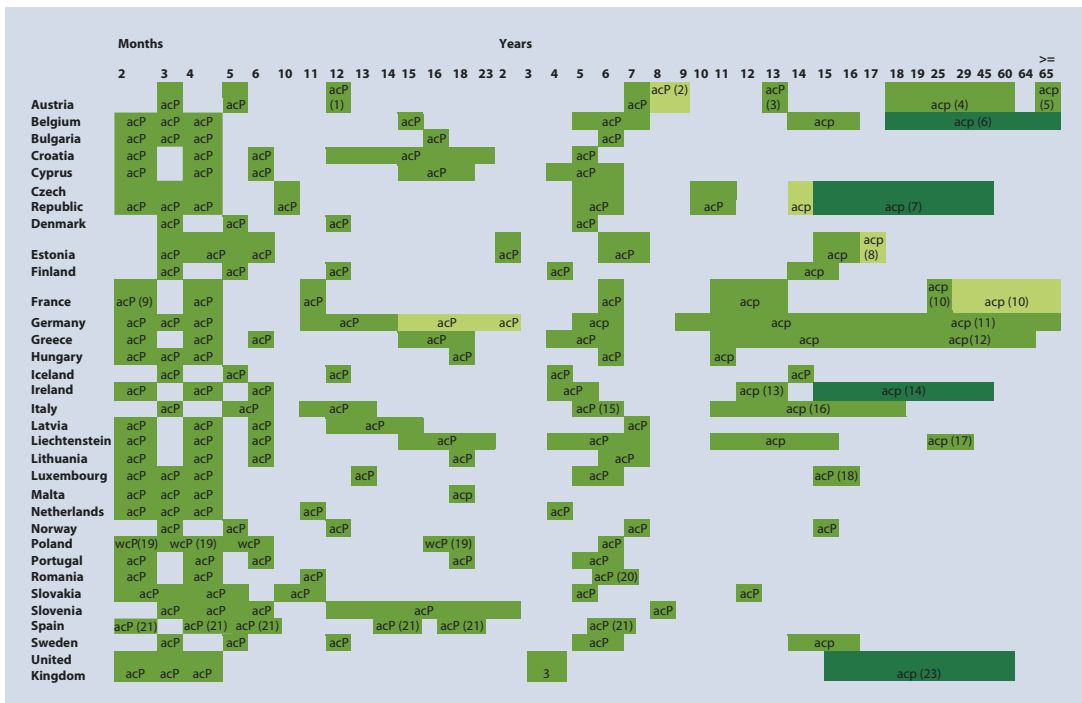
In addition to regularly scheduled doses throughout childhood, some countries do recommend pertussis immunization in specific situations, with the goal of decreasing the risk of transmission to young, vulnerable infants (“cocooning”). Unfortunately, several studies, including one from Switzerland, have shown that cocooning is extremely challenging from a logistic point of view and a complete “cocoon” around the newborn or young infant is hardly ever achieved, especially in large households.

Today, amongst various strategies of maternal and paternal immunization, the concept of immunization in pregnancy is probably most promising. Maternal and paternal immunization means that women’s and men’s pertussis immunization status is brought up to date as part of family planning before the woman’s pregnancy or catching up with pertussis immunizations after delivery, if they were missed before. Basically, this leads to cocooning of the young infant, as discussed above. In addition to this, immunizing a woman *during* pregnancy brings a new dimension of protection to the infant, i.e.,

direct protection via transplacental transfer of high quantities of maternal anti-pertussis toxin (PT) IgG antibodies. A case-control study, performed as part of a national vaccination program for pregnant women in the UK between October 2012 and July 2013, demonstrated that only 17% of mothers of infants (<8 weeks of age) with pertussis compared to 71% of mothers of healthy age-matched controls had been immunized against pertussis in pregnancy. This resulted in a protective effectiveness of immunization in pregnancy of 93%. It was further shown in Belgium that the average PT antibody levels in children whose mothers had been vaccinated against pertussis during pregnancy were much higher than those in children of unvaccinated mothers (101 vs 12 IU/ml and 16 vs 1 IU/ml at birth, and at the age of 2 months respectively). When measured again 4 weeks after completion of the primary immunization series at age 2–3–4 months, however, anti-PT values in infants of vaccinated mothers were lower than those in control children (29 vs 54 IU/ml) and this difference was still present after the fourth dose at 15 months of age (36 vs 57 IU/ml).

The clinical significance of this blunting of the child's immune response to aP vaccine is unclear because of the lack of a reliable serological correlate of immunity and must be further evaluated in prospective epidemiological studies. Given the benefit of significant protection during the first months of life in infants, these observations do not question pertussis immunization in pregnancy. So far, no safety concerns have arisen with regard to pertussis immunization in pregnant women. In an observational study based on the US Vaccine Safety Datalink, which accompanied the introduction of the immunization program in pregnant women in 2012 in the USA, no safety signals were detected. Today, in addition to England and Wales and the USA, an increasing number of countries recommend pertussis (Tdap) immunization for pregnant women.

However, given the suboptimal protective power of currently available aP vaccines, the search for “better” vaccines is ongoing. Intensive efforts are underway to identify biomarkers that would predict protection from *B. pertussis* infection and/or disease and would then promote the



■ Fig. 18.1 Recommended pertussis immunizations

Footnotes:

1: Earliest 6 month after the second dose

2: If not vaccinated at 7 years of age

3: For children who received only Td-IPV at 7-9 years

4: dTaP-IPV every 10 years between 18 and 60 years of age.

5: dTaP-IPV every 5 years from 65 years of age

6: One dose of dTTPa for all adults, with emphasis on cocoon vaccination

Vaccination of expectant mothers during every pregnancy with a pertussis-containing vaccine in week 24 to week 32 of pregnancy

7: A single dose of Tdap is recommended to be given in pregnancy, ideally in the third trimester, between pregnancy weeks 28 and 36.

In women who did not receive Tdap in pregnancy, a single dose of Tdap is recommended to be given immediately after delivery to minimise the risk of infection transmission to the neonate. see full recommendation at http://www.szu.cz/uploads/Epidemiologie/Pertuse/CR_Pertussis_Recommendation_for_pregnant_women.pdf

8: only to children born in the period 1990-1995 and previously vaccinated at the age 12 years by sixth dose of dT vaccine.

9: or 8 weeks of age

10: For those who did not receive a dose of pertussis containing vaccine during the past 5 years, a booster with a quadrivalent vaccine (dTacP-IPV) is recommended at the time of the Td-IPV booster at 25 years.

For those aged 25 years and above that did not receive a booster dose, catch-up with a dTAcP-IPV vaccine can be proposed until 39 years of age.

Recommendation to have an interval of 10 years in adults between a documented pertussis and pertussis re-vaccination.

11: Only one of the Td 10-yearly booster doses should be with a Tdap vaccine in adults. Subsequent booster doses are to be done with Td vaccines..

12: Td booster every 10 year. One of the booster dose should be with Tdap or Tdap-IPV. Td from 65 years of age

13: Booster dose

14: Tdap -Vaccination for pregnant women between 27-36 weeks gestation (introduced in September 2013). If the recipient does not have a medical card, they must pay administration cost of the vaccination out-of-pocket.

15: After seven years, a low-dose pertussis-containing dT vaccine should be used

16: To be given ten years after completing primary vaccination with DTaP-containing vaccines

17: Boosters at the age of 25-29, 45, 65, then every 10 years. First booster preferably before having first child, in order to protect the newborn against pertussis.

18: Subsequent Tdap-IPV booster every 10 years

19: An acellular pertussis component (aP) combination vaccine should be used for children with contraindications to vaccination with the whole cell pertussis vaccine and in children born before 37th week of pregnancy or born with birth weight less than 2500 g

20: DTacP-IPV at 6 years to begin in 2015

21: For more detail on review and recommendation for pertussis vaccination in Spain, please refer to <http://msc.es/profesionales/saludPublica/prevPromocion/vacunaciones/docs/TosFerina.pdf>

22: Either DTacP-IPV or dTAcP-IPV can be given depending on availability

23: Specific programme to vaccinate expectant mothers with a pertussis-containing vaccine from 28 weeks of pregnancy. for more information, refer to <http://immunisation.dh.gov.uk/pertussis-pregnant/>

The contents of this report are covered by the ECDC legal notice. See: <http://ecdc.europa.eu/en/pages/legalnotice.aspx>. The report reflects the state of submissions in the ECDC vaccination schedule platform as of 2016-09-17 at 16:42.

	General recommendation for Austria
	Recommendation for specific groups only for Austria
	Catch-up (e.g. if previous dosed missed) for Austria

development of new vaccines, which would elicit a protective immune response against *B. pertussis*. Although we may dream about a new generation of such better performing pertussis vaccines, the best use of aP (and wP, where still in use) vaccines should be made. This includes the optimization of vaccine coverage in the whole population and the timely administration of the recommended doses in infants and young children.

Further Reading

- Acosta AM, DeBolt C, Tasslimi A, Lewis M, Stewart LK, Misegades LK, Messonnier NE, Clark TA, Martin SW, Patel M. Tdap vaccine effectiveness in adolescents during the 2012 Washington State pertussis epidemic. *Pediatrics*. 2015;135:981–9.
- Berkovic SF, Harkin L, McMahon JM, Pelekanos JT, Zuberi SM, Wirrell EC, Gill DS, Iona X, Mulley JC, Scheffer IE. De-novo mutations of the sodium channel gene SCN1A in alleged vaccine encephalopathy: a retrospective study. *Lancet Neurol*. 2006;5:488–92.
- Büttcher M, Heininger U, Braun M, Bonhoeffer J, Halperin S, Heijbel H, de Menezes MR, Vermeer-de Bondt P, The Brighton Collaboration HHE Working Group. Hypotonic-Hyporesponsive Episode (HHE) as an adverse event following immunization in early childhood: case definition and guidelines for data collection, analysis, and presentation. *Vaccine*. 2007;25:5875–81.
- Cherry JD. Epidemic pertussis and acellular pertussis vaccine failure in the 21st century. *Pediatrics*. 2015;135:1130–2.
- Cherry JD, Heininger U. Pertussis and other *Bordetella* infections. In: Feigin RD, Cherry JD, Demmler-Harrison GJ, Kaplan SL, editors. *Textbook of pediatric infectious diseases*. 6th ed. Philadelphia: Saunders; 2009. p. 1683–706.
- Dabrera G, Amirthalingam G, Andrews N, Campbell H, Ribeiro S, Kara E, Fry NK, Ramsay M. A case-control study to estimate the effectiveness of maternal pertussis vaccination in protecting newborn infants in England and Wales, 2012–2013. *Clin Infect Dis*. 2015;60:333–7.
- Decker MD, Edwards KM, Steinhoff MC, Rennels MB, Pichichero ME, Englund JA, Anderson EL, Deloria MA, Reed GF. Comparison of 13 acellular pertussis vaccines: adverse reactions. *Pediatrics*. 1995;96:557–66.
- European Centre for Disease Prevention and Control (ECDC). Pertussis. Stockholm: ECDC. n.d.. Available from: http://ecdc.europa.eu/en/activities/surveillance/euavac/case_definition/Pages/pertussis.aspx.
- Forsyth KD, Campins-Marti M, Caro J, Cherry JD, Greenberg D, Guiso N, Heininger U, Schellekens J, Tan T, von König CH, Plotkin S, Global Pertussis Initiative. New pertussis vaccination strategies beyond infancy: recommendations by the Global Pertussis Initiative. *Clin Infect Dis*. 2004;39:1802–9.
- Heininger U, Cherry JD. Pertussis immunisation in adolescents and adults – *Bordetella pertussis* epidemiology should guide vaccination recommendations. *Expert Opin Biol Ther*. 2006;6:685–97.
- Heininger U, André P, Chlibek R, Kristufkova Z, Kutsar K, Mangarov A, Mészner Z, Nitsch-Osuch A, Petrović V, Prymula R, Usonis V, Zavadská D. Comparative epidemiologic characteristics of pertussis in 10 central and Eastern European Countries, 2000–2013. *PLoS One*. 2016;11(6):e0155949.
- Kharbanda EO, Vazquez-Benitez G, Lipkind HS, Klein NP, Cheetham TC, Naleway AL, Lee GM, Hambidge S, Jackson ML, Omer SB, McCarthy N, Nordin JD. Maternal Tdap vaccination: coverage and acute safety outcomes in the vaccine safety datalink, 2007–2013. *Vaccine*. 2016;34:968–73.
- Klein NP, Bartlett J, Rowhani-Rahbar A, Fireman B, Baxter R. Waning protection after fifth dose of acellular pertussis vaccine in children. *N Engl J Med*. 2012;367:1012–9.
- Koepke R, Eickhoff JC, Ayele RA, Petit AB, Schauer SL, Hopfensperger DJ, Conway JH, Davis JP. Estimating the effectiveness of tetanus-diphtheria-acellular pertussis vaccine (Tdap) for preventing pertussis: evidence of rapidly waning immunity and difference in effectiveness by Tdap brand. *J Infect Dis*. 2014;210:942–53.
- Maertens K, Caboré RN, Huygen K, Hens N, Van Damme P, Leuridan E. Pertussis vaccination during pregnancy in Belgium: results of a prospective controlled cohort study. *Vaccine*. 2016;34:142–50.
- McGirr A, Fisman DN. Duration of pertussis immunity after DTaP immunization: a meta-analysis. *Pediatrics*. 2015;135:331–43.
- Nieves D, Heininger U, Cherry J. *Bordetella pertussis* and other *Bordetella* spp. infections. In: Wilson CB, Nizet V, Maldonado YA, Remington JS, Klein JO, editors. *Remington and Klein's infectious diseases of the fetus and newborn infant*. 8th ed. Philadelphia, PA: Elsevier Saunders; 2016. p. 598–616.
- Urwyler P, Heininger U. Protecting newborns from pertussis – the challenge of complete cocooning. *BMC Infect Dis*. 2014;14(1):397. doi:10.1186/1471-2334-14-397.
- Winter K, Cherry JD, Harriman K. Effectiveness of prenatal Tdap vaccination on pertussis severity in infants. *Clin Infect Dis*. 2017a;64(1):9–14.
- Winter K, Nickell S, Powell M, Harriman K. Effectiveness of prenatal versus postpartum Tdap vaccination in preventing infant pertussis. *Clin Infect Dis*. 2017b;64(1):3–8.

***Haemophilus influenzae* type b (Hib) Vaccines**

Mary P.E. Slack

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19.1 Introduction

Haemophilus influenzae is a human-restricted pathogen that colonises the nose and throat and to a lesser extent the conjunctivae and genital tract.

H. influenzae was first identified as a pathogen by Koch in 1883, who described small gram-negative bacilli in conjunctivitis. In 1889–1892 a major outbreak of influenza swept across Europe. Pfeiffer examined sputum from patients suffering from influenza and reported “in every case ... a similar type of bacillus was found in absolutely pure culture... and in almost incredible numbers”. He had difficulty in growing the bacillus until he added blood to the culture medium.

Continued belief that *Bacillus influenzae* (or Pfeiffer’s bacillus) was the cause of influenza resulted in it being specifically named *Haemophilus influenzae*. In 1922 Kristensen proposed that this organism was a secondary invader and not the primary cause of influenza. In 1933 Smith, Andrewes and Laidlaw established that influenza was a viral infection, but the name of the bacterium has remained unchanged.

In 1933 Margaret Pittman differentiated *H. influenzae* into two major groups: encapsulated and non-encapsulated strains (more commonly described as non-typeable *Haemophilus influenzae*: NTHi). She also described the six antigenically and chemically distinct types of

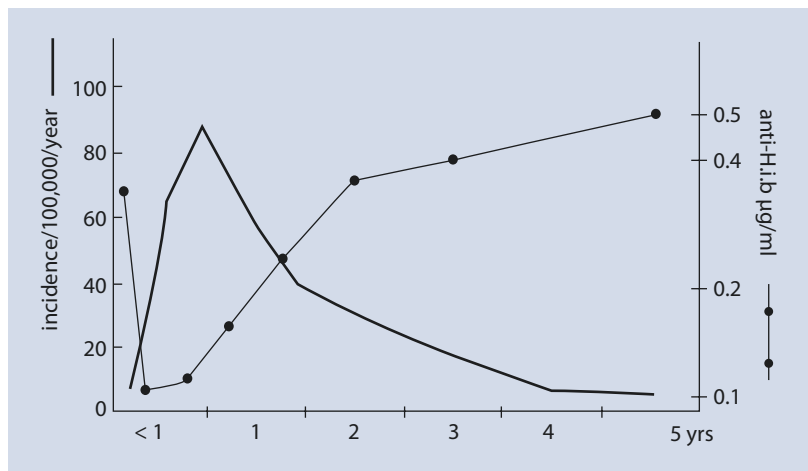
encapsulated strains, designated Pittman types a to f and identified type b (Hib) as of predominant importance in causing meningitis and other systemic haemophilus infections. The most virulent serotype is Hib.

In 1933 Fothergill and Wright showed that the blood of young children, aged less than 2 years, had absent or low levels of bactericidal activity against the *H. influenzae* type b polysaccharide capsule, whereas the blood from older children and adults did demonstrate bactericidal activity against Hib. They also noted that the majority of cases of Hib meningitis occurred in young children, leading them to speculate that naturally acquired antibodies to the polysaccharide capsule were protective against serious Hib infections. The rarity of infections in the first 2 months of life correlates with the presence of maternal antibodies to Hib and the occurrence of infection in early infancy with the absence of antibodies having such specificity. As the mean level of Hib antibodies in the population rises, so Hib infections decline (■ Fig. 19.1).

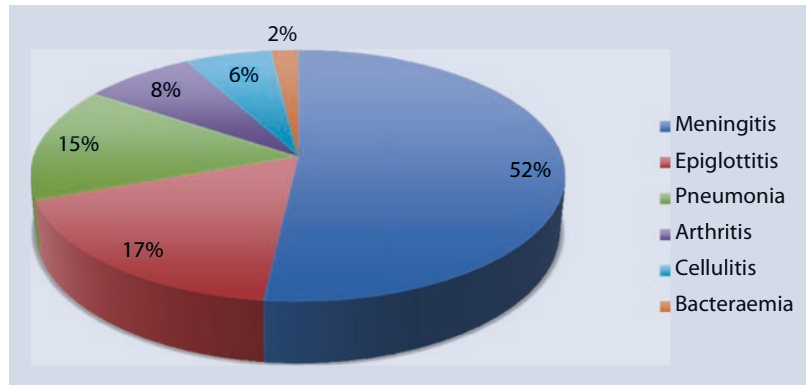
19.2 Burden of Disease

Haemophilus influenzae is responsible for significant morbidity and mortality in children aged less than 5 years and prior to the introduction of routine immunisation with Hib conjugate

■ Fig. 19.1 Incidence of *H. influenzae* meningitis (heavy solid line) during the first 5 years of life and the corresponding mean level of anti-*H. influenzae* type b (Hib) capsular polysaccharide antibodies (thin line) (Peltola et al. 1977)



■ **Fig. 19.2** Spectrum of invasive infections caused by Hib in the UK, prospective surveillance data in all ages for 2 years before the introduction of routine Hib immunisation (Anderson et al. 1995)



vaccines; 90% of serious *H. influenzae* infections were due to Hib. It has been estimated that Hib caused more than 8 million serious infections and 371,000 deaths worldwide in 2000.

Before the introduction of Hib conjugate vaccine, Hib was the most common cause of bacterial meningitis in children, 75% of whom were over the age of 2 months and under 3 years old. Hib meningitis had a case fatality ratio of 5–10%, with up to one third of survivors suffering significant sequelae, including deafness, intellectual impairment, cerebral palsy and epilepsy. Hib was also the most common cause of acute epiglottitis in children, which generally occurred in children aged between 2 and 4 years of age. Other manifestations of invasive Hib infection include bacteraemia, periorbital cellulitis, septic arthritis, osteomyelitis and pneumonia (■ Fig. 19.2).

The annual incidence/100,000 children aged <5 years of invasive Hib disease, prior to the introduction of Hib conjugate vaccine, was not uniform in all countries, ranging between 25 in Ireland and 31 in the UK (England and Wales); between 40 and 60 in Australia, New Zealand and Scandinavia; and between 60 and 130 in the USA. In the indigenous populations of the USA and Australia, the incidence was as high as 450/100,000 children under 5 years of age. In Europe higher incidences were seen in Northern European (■ Fig. 19.3) than in Southern European countries.

19.3 Pathogenesis

H. influenzae is transmitted by aerosols of respiratory secretions or by direct contact with contaminated material. The primary event is colonisation of the nasopharynx. Before the introduction of Hib vaccines, 3–5% of healthy pre-school children in industrialised countries were asymptomatic Hib carriers. The rate of non-typeable *H. influenzae* carriage is much higher. Asymptomatic Hib carriage can persist for up to 6 months.

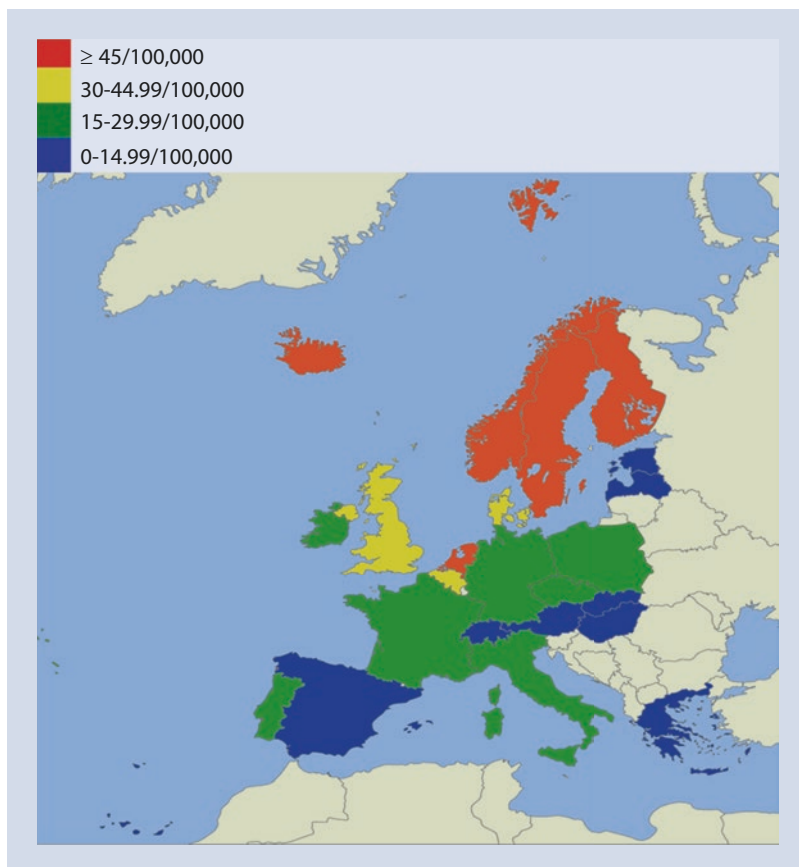
Prior infection with respiratory viruses, such as influenza, predisposes to nasopharyngeal carriage by several mechanisms, including obstruction to the outflow of respiratory secretions, depression of local immunity and suppression of mucociliary clearance. The rate of Hib carriage varies with age, crowding, geography and vaccine coverage in a population.

Invasive Hib disease follows invasion of the bloodstream. Recent viral infection is a risk factor for developing invasive Hib disease, by facilitating the attachment of Hib to the respiratory epithelium.

The risk of invasive Hib disease is increased in children with certain co-morbidities, including sickle cell disease, asplenia, malignancies and antibody deficiency syndromes.

The capsule of Hib is composed of polyribosylribitol phosphate (PRP). PRP is the single most important major virulence determinant for

Fig. 19.3 Annual incidence of invasive Hib disease/100,000 < 5 years in Europe before the introduction of routine Hib immunisation (data available at ► http://www.hpa-bioinformatics.org.uk/euibis/documents/hib_vaccine_evaluation.pdf) Hib catch-up campaign



invasion of the bloodstream because it resists phagocytosis and complement-mediated bacteriolysis and resists splenic clearance.

Studies on unimmunised individuals in Finland indicated that serum anti-PRP antibodies of $\geq 0.15 \mu\text{g/mL}$ were correlated with a decreased incidence of Hib meningitis. Further studies established that a concentration of $\geq 0.15 \mu\text{g/mL}$ provides short-term protection against invasive Hib disease, but long-term protection requires a concentration of $\geq 1.0 \mu\text{g/mL}$.

19.4 Hib Polysaccharide Vaccine

The first Hib vaccine was a PRP plain polysaccharide vaccine, which was used in a field trial in Finland. This trial involving 100,000 children aged 3 months to 5 years demonstrated an age-dependent response to PRP. PRP polysaccharide

vaccine had no demonstrable effect on nasopharyngeal carriage of Hib and thus did not interrupt transmission of Hib or produce herd protection. Polysaccharide vaccines activate B cells via a T-helper cell-independent pathway, which is poorly developed in children aged <18 months and characterised by lack of immune memory, short-lived antibody responses and poor immunogenicity.

19.5 Hib Conjugate Vaccines

In the late 1980s, conjugate Hib vaccines were developed and increased the immunogenicity of PRP polysaccharide. The polysaccharide-protein conjugate induced a T-cell-dependent response. T-cell-dependent responses develop much earlier in infants than T-cell-independent responses, and infants are able to respond to

polysaccharide-protein conjugate vaccines from the age of 6 weeks.

Protein antigens encourage class switching from IgM to IgG via T-helper cells. The IgG is predominantly IgG1 subclass which in vitro induces complement-mediated opsonic activity and bacteriolysis. In addition, immunising with a conjugate vaccine results in antibodies with higher avidity compared to those produced after immunisation with plain PRP polysaccharide, with the added benefit of avidity maturation.

Protein-polysaccharide conjugate vaccines also have a marked impact on nasopharyngeal carriage. By reducing nasopharyngeal carriage of Hib, transmission to other susceptible unimmunised children and adults is interrupted, reducing infection in other age groups. This effect is known as herd protection.

Four different Hib vaccines were initially developed (■ Table 19.1). They differ in the protein carrier used, the length of the PRP saccharide and the method of protein-polysaccharide conjugation. The four protein carriers were tetanus toxoid (PRP-TT), diphtheria toxoid (PRP-D), *Neisseria meningitidis* outer membrane protein complex (PRP-OMP) and a non-toxic mutant *Corynebacterium diphtheriae* protein CRM¹⁹⁷ (PRP-CRM).

Although the different Hib vaccines were equally immunogenic in adults, they elicited differing immune responses in children aged <2 years. The PRP-D conjugate was the least immunogenic in infants, eliciting an anti-PRP antibody titre ≥ 1.0 $\mu\text{g}/\text{mL}$ in $\approx 30\%$ of infants after 2 or 3 doses.

The other three vaccines are highly immunogenic in children aged >18 months. Their immunogenicity varies in children aged <18 months. PRP-OMP vaccine stimulates the highest antibody concentration with a single dose administered at 2 months of age eliciting antibody titres ≥ 1.0 $\mu\text{g}/\text{mL}$ in 70–80% of infants. For this reason PRP-OMP was the preferred vaccine for use in populations where there was a high burden of disease in very young infants, for example, indigenous Australian, Apache and Navajo and native Alaskan infants.

PRP-TT and PRP-CRM have similar immunogenicity, and there is no significant difference in the percentage of infants achieving anti-PRP antibody titres of ≥ 1.0 $\mu\text{g}/\text{mL}$ after 3 priming doses. A booster dose of any of the PRP vaccines administered in the second year of life results in seroprotective levels of anti-PRP antibodies, irrespective of the PRP-vaccine used for the primary immunisation series.

■ Table 19.1 *H. influenzae* conjugate vaccines

Vaccine	Manufacturer	Hib protein carrier	Amount of PRP (μg)
HibTITER	Wyeth (now Pfizer)	CRM ₁₉₇	10
Pedvax-Hib	Merck & Co Inc	OMP	7.5
Act-Hib	Sanofi Pasteur	PRP-TT	10
Hiberix	GSK	PRP-TT	10
ProHIBIT	Connaught laboratories	PRP-D	25
Menitorix	GSK	PRP-TT (Hib-MenC)	5

19.6 Combination Vaccines

Following the successful introduction of monovalent Hib conjugate vaccines, the Hib component was incorporated into a number of combination vaccines (see ► Chap. 20). However, the combination can result in a significantly reduced anti-PRP antibody response compared to that achieved by a separate Hib conjugate vaccine. The Hib component has been combined with diphtheria toxoid (D), tetanus toxoid (T), whole-cell pertussis (wP), acellular pertussis (aP), inactivated polio (IPV) and hepatitis B (HepB). It has also been combined with meningococcal group C – tetanus toxoid (MenC-TT) as a dual vaccine Hib-MenC-TT (■ Table 19.2).

Table 19.2 Some examples of Hib-containing vaccines available in Europe in 2017

Name	Characteristics	Manufacturer	Type
Act-Hib	Hib (PRP-TT)	Sanofi Pasteur	Mono
Hexacima	DTaP-HepB-IPV-Hib (PRP-TT)	Sanofi Pasteur	Combination
Hexyon	DTaP-HepB-IPV-Hib (PRP-TT)	Sanofi Pasteur	Combination
Hiberix	Hib (PRP-TT)	GSK	Mono
Infanrix + Hib	DTaP-Hib (PRP-TT)	GSK	Combination
Infanrix Hexa	DTaP-HepB-IPV-Hib (PRP-TT)	GSK	Combination
Infanrix IPV + Hib	DTaP-IPV-Hib (PRP-TT)	GSK	Combination
Pentavac	DTaP-IPV-Hib (PRP-TT)	Sanofi Pasteur	Combination
Vaxelis	DTaP-HepB-IPV-Hib (PRP-OMP)	Sanofi Pasteur	Combination

19.7 Introduction of Hib Conjugate Vaccines in Europe

Hib conjugate vaccine was included in the National Immunisation Programme of Finland in 1986 and over the next 20 years was added to the national infant immunisation schedule in all European countries.

19.8 Impact of Hib Conjugate Vaccines

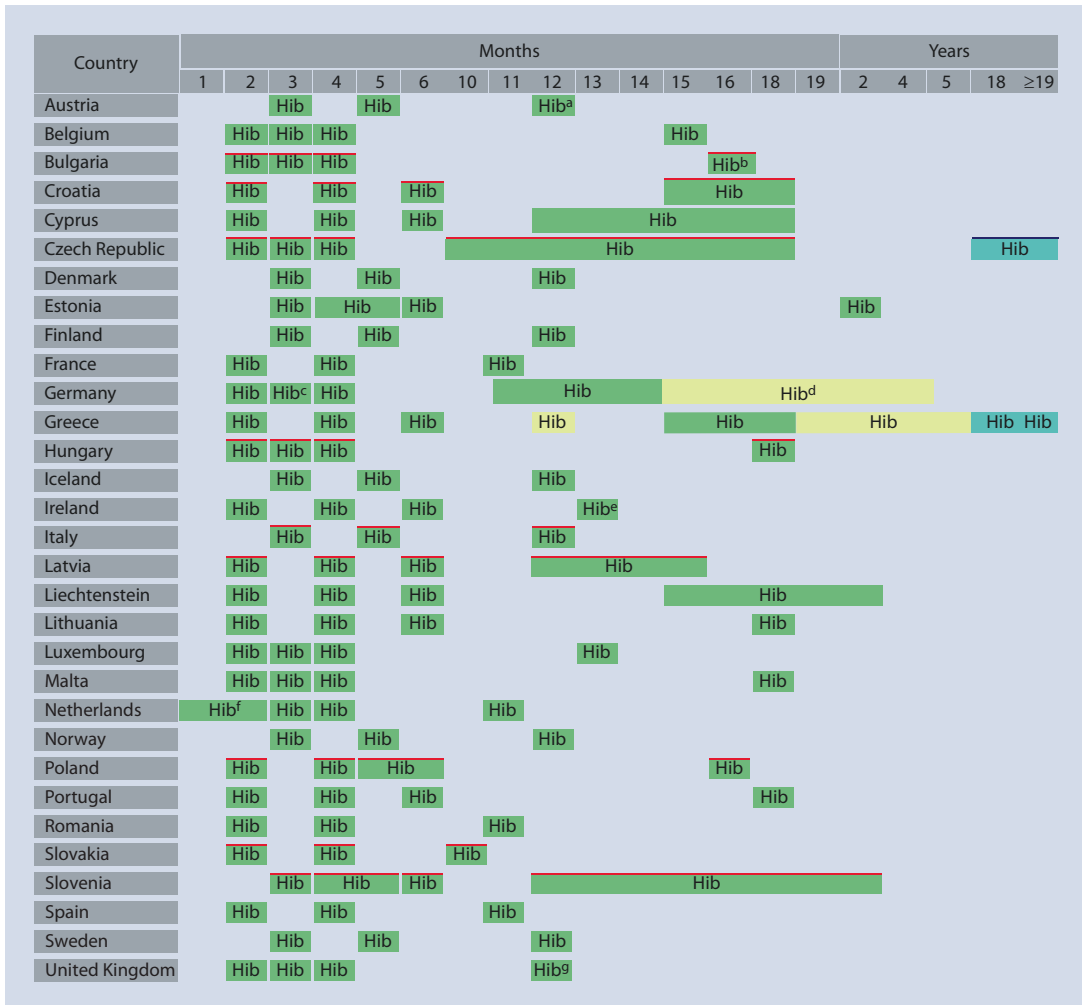
Following the introduction of routine Hib conjugate vaccination in many European countries, an international collaboration was established in 1996 to monitor the impact of Hib conjugate vaccine on the epidemiology of invasive *H. influenzae* disease. Data on invasive *H. influenzae* disease was collected from 25 European countries between 1999 and 2007 by the European Union Invasive Bacterial Infections Surveillance Network (EU-IBIS) (<http://www.hpa-bioinformatics.org.uk/euibis/>) funded by the European Commission DG Sanco. The countries initially participating were Austria, Belgium, Czech Republic, Denmark, Finland, France, Germany, Greece, Iceland, Ireland, Italy, Luxembourg, Malta, the Netherlands, Norway, Portugal, Spain, Sweden, Switzerland and the UK. From 2003 to 2007 data

was also provided by Estonia, Hungary, Latvia, Lithuania, Poland, Slovakia and Slovenia. In 2007 the surveillance activities were transferred to ECDC.

After 2000, all countries included in the EU-IBIS surveillance network had implemented Hib conjugate vaccination in their national immunisation programmes. By 2006 the overall annual incidence across Europe of invasive Hib disease had fallen to 0.58/100,000 < 5 years of age (http://www.hpa-bioinformatics.org.uk/euibis/documents/2006_hib.pdf), and the coverage of Hib conjugate vaccine varied from 80% to >95% across Europe.

Hib conjugate vaccine is now included in the national infant immunisation programme in all European countries. The schedules of vaccine administration vary, with some countries giving three doses in the first year, followed by a booster in the second year of life and others giving two doses in infancy plus a booster dose after the first birthday (■ Fig. 19.4).

Multiple Hib-containing vaccines have been used in Europe. Countries have changed the Hib vaccine used over time, but overall there has been a convergence towards the use of pentavalent or hexavalent combination vaccines. High levels of coverage for the Hib 3 dose (87–99%) were reported across Europe in 2015 (■ Fig. 19.4).



■ **Fig. 19.4** Recommended Hib immunisation schedules for European countries: 2017 Reproduced with permission from ► http://apps.who.int/immunization_monitoring/globalsummary/timeseries/tscoveragehib3.html; ► <http://vaccine-schedule.ecdc.europa.eu/Pages/Scheduler.aspx> ^aEarliest 6 months after second dose ^bNot earlier than 12 months after the 3rd dose; ^cOptional dosis if monovalent and other combination vaccines are used; ^dNum-

ber of doses necessary varies according to age; ^eHib/MenC combined vaccine; ^fShould be given at 6-9 weeks; ^g1 dose - Hib/MenC combined vaccine. *Green* General Recommendation, *Blue* Recommendation for specific groups only, *Yellow* Catch-up (e.g. if previous dosed missed), *Blue line above* Vaccination not found by the national Health System, *Red line above* mandatory Vaccination

19.9 Safety of Hib Conjugate Vaccines

All of the Hib conjugate vaccines have an excellent safety profile. Mild local reactions, including redness, induration and swelling, are reported to be more common with PRP-TT than with PRP-CRM or PRP-OMP.

19.10 Hib Vaccine Failures

Although Hib conjugate vaccines are highly effective, vaccine failures do occasionally occur. Clinical and immunological evaluation is therefore recommended for children who develop invasive Hib disease despite a full course of Hib vaccinations.

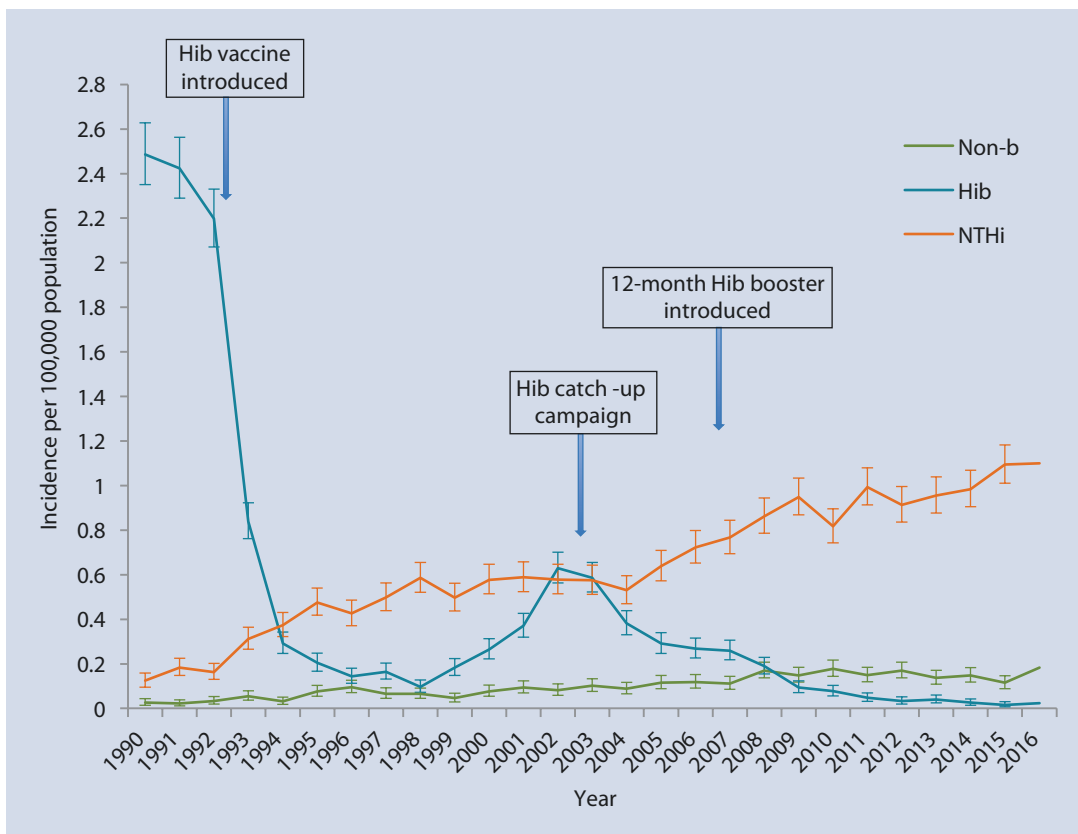
19.11 UK Hib Vaccine Experience: Lessons Learned

Routine Hib immunisation was introduced in the UK in October 1992. Three doses of Hib conjugate vaccine (PRP-TT; Pasteur Mérieux) given at 2, 3 and 4 months of age were offered to all infants <1 year old. There was no booster dose in the second year of life. It was believed that a booster dose would not be needed as immunological memory was expected to provide long-term protection. A catch-up programme of a single dose of PRP-CRM vaccine was offered to children aged 1–4 years over the first year of the national infant immunisation programme.

Following the introduction of Hib conjugate vaccination in the UK, there was a rapid and sustained decline in invasive Hib disease (■ Fig. 19.5) with the annual attack rate for invasive Hib disease in children <5 years falling from 23.8/100,000 in 1991–1992 to 1.8 /100,000 in

1993–1994. The decline in vaccinated age groups was soon followed by a decline in other age groups through indirect (herd) protection. By 1998 the incidence of invasive Hib disease in children aged <5 years had fallen to 0.63/100,000. There were estimated to be 2.2 vaccine failures/100,000 vaccinated children (95% CI, 1.8–2.7). Vaccine failures were uncommon. Although the vaccine effectiveness waned with time, it remained high (>95%) until the sixth year of life.

From 1999, there was a resurgence in cases of Hib infection in children (■ Fig. 19.5) with 134 cases in <5 year olds in 2002 vs 31 cases in 1996. There appear to be several reasons for this resurgence. The vaccine effectiveness among children immunised in infancy was lower than had been anticipated. Among children <5 years who developed invasive Hib infection between 1993 and 2000, the vaccine effectiveness (VE) was estimated to be 57% (95% CI, 43–67). The VE was lower in children immunised in infancy compared to those



■ Fig. 19.5 Incidence of invasive *H. influenzae* disease in England and Wales by serotype: 1990–2016 (Data from PHE)

who received a single dose of Hib vaccine as part of the catch-up programme, and the VE in those immunised in infancy declined significantly over time ($p = 0.004$), declining to zero after 1 year. This lower VE only became apparent when the direct and indirect protection provided by the catch-up campaign in children aged 1–4 years began to wane. By 1998 all children aged <5 years had only received routine infant immunisation in early infancy.

A further reason for the resurgence was a shortage of the DTwP-Hib vaccine that was being used in the UK, which led to approximately half of infants receiving an alternative combination vaccine containing acellular pertussis component. DTaP-Hib vaccines have been shown to have lower Hib immunogenicity, especially when used in an early accelerated infant schedule, as was the case in the UK. There is evidence that combination DTaP-Hib vaccines can elicit a significant reduction in the anti-PRP antibody titres, possibly through catalytic depolymerisation of PRP in the presence of aluminium hydroxide or because they lack the adjuvant effect of the whole-cell pertussis component on PRP.

Another potential cause of the resurgence was the concomitant introduction of MenC conjugate vaccine in 1999, which was given at the same time as the Hib conjugated vaccine. Most of the MenC conjugate used was CRM based, and there is evidence that the use of this vaccine together with DTaP-Hib also results in lower immunogenicity of the Hib component.

Control of the resurgence was achieved by the administration of a single dose of Hib vaccine to all children aged 6 months to 4 years in April 2003. In 2004 the DTwP-Hib conjugate was switched to routine use of DTaP-IPV-Hib conjugate, and a routine booster dose of Hib vaccine, administered as a Hib-MenC combination, at 12 months of age was added to the schedule in 2006. A second pre-school booster campaign was conducted in 2007 for children who were too old for the 12-month booster dose but too young for the 2003 booster campaign.

Following these measures, the number of cases of invasive Hib disease declined rapidly.

There was a similar resurgence in the Republic of Ireland, who had also introduced an infant Hib immunisation programme with a schedule of 3 doses at 2, 4 and 6 months without a booster dose.

The UK experience with Hib conjugate vaccines showed that immunological memory per se was not sufficient to confer clinical protection. The lower than expected vaccine effectiveness of an early accelerated infant immunisation schedule was masked for several years by the catch-up campaign which produced high levels of antibody and prolonged direct protection in older cohorts and contributed to high population immunity. Protection against Hib infection may depend on the level of serum anti-PRP antibodies at the time of acquisition of the organism in the nasopharynx. A booster dose in the second year of life produces high levels of serum anti-PRP antibodies, which are sustained above the protective threshold to provide protection against Hib infection in children <5 years of age.

19.12 Invasive *H. influenzae* Infections in Europe in the Era of Routine Hib Conjugate Vaccination

In 2014 there were 2799 cases of invasive *H. influenzae* disease reported by 29 European countries to the European Centre for Disease Prevention and Control. The overall incidence was 0.6/100,000 population (■ Fig. 19.6). 1706/2799 (61%) of the isolates were of known serotype; 1394/1706 (82%) were NTHi. NTHi was the most common *H. influenzae* reported in all age groups, and the majority of cases were in patients aged >65 years. There were 104 Hib reports (6%), and 57% of these infections occurred in individuals >25 years of age. Hif was the most common capsulated type with 150 reported cases. There were 47 Hie infections. Both Hif and Hie infections occurred mainly in patients >45 years of age. There were 8 Hia, 1 Hic and 2 Hid infections.

There had been a concern that other capsulated serotypes of *H. influenzae* might occupy the ecological niche formerly occupied by Hib and emerge as significant causes of invasive disease. This has not happened although there has been a small increase in the number of invasive Hif and Hie cases in Europe. However, there has been a significant year on year increase in the number of cases of invasive NTHi infection.

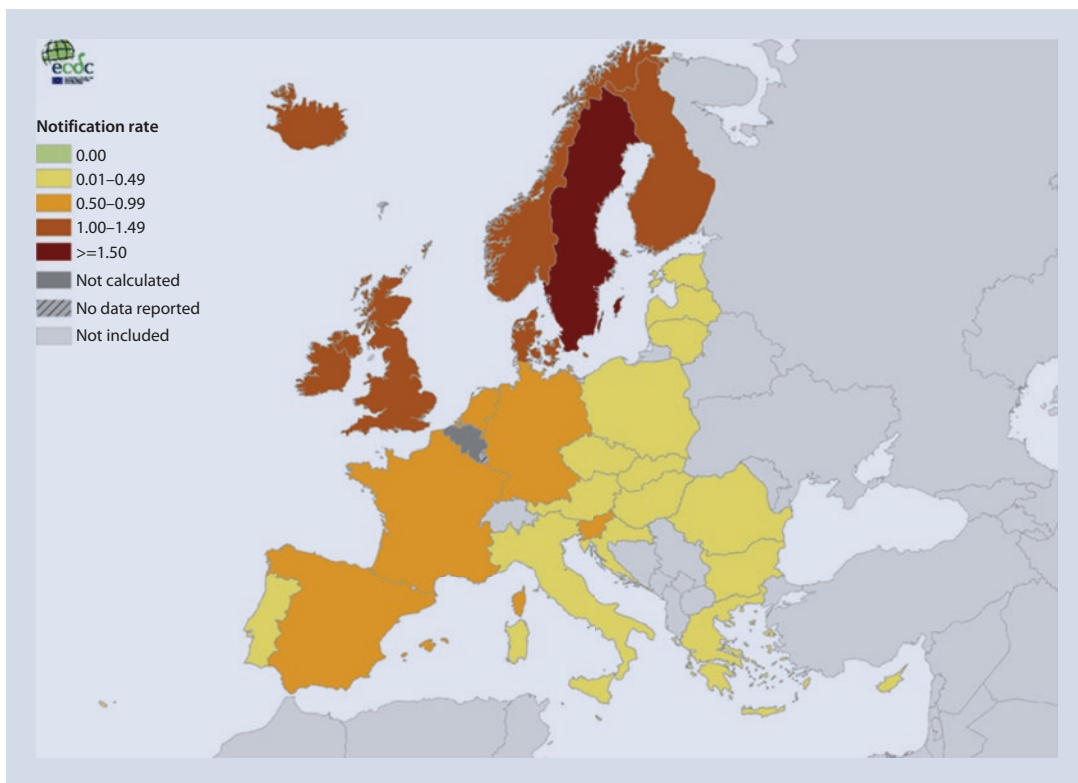


Fig. 19.6 Number of confirmed/reported cases of invasive *H. influenzae* disease/100,000 population in EU/EAA countries, 2014 (European Centre for Disease Prevention and Control 2016) (Source: Country reports from Austria, Belgium, Bulgaria, Croatia, Cyprus, the Czech

Republic, Denmark, Estonia, Finland, France, Germany, Greece, Hungary, Iceland, Ireland, Italy, Latvia, Lithuania, Malta, the Netherlands, Norway, Poland, Portugal, Romania, Slovakia, Slovenia, Spain, Sweden, the United Kingdom)

19.13 Conclusions

The introduction of Hib conjugate vaccine has been extremely successful, with the virtual elimination of invasive Hib disease in children and a significant reduction in cases in adults due to herd protection. NTHi has now emerged as the commonest cause of invasive *H. influenzae* infection, including pneumonia and bacteraemia. A vaccine effective against NTHi could be of value in preventing these infections. A 10-valent pneumococcal conjugate vaccine (PHid-CV; Synflorix; GSK) uses *H. influenzae* outer membrane lipoprotein D as its carrier protein, which is conserved among the majority of strains of *H. influenzae*. Immunisation results in high concentrations of anti-protein D antibodies, but has no effect on nasopharyngeal NTHi colonisation and to date has not had any demonstrable efficacy against invasive NTHi infections. The challenge is

to overcome the marked heterogeneity and phase variability of NTHi. Such a vaccine could be targeted at groups who have a high incidence of mucosal NTHi infections, including otitis media in indigenous children or adults with chronic obstructive pulmonary disease.

Further Reading

- Anderson EC, Begg NT, Crawshaw SC, Hargreaves RM, Howard AJ, Slack MP. Epidemiology of invasive Haemophilus influenzae infections in England and Wales in the pre-vaccination era (1990-2). *Epidemiol Infect.* 1995;115(1):89-100.
- Collins S, Ramsay M, Campbell H, Slack MP, Ladhani SN. Invasive Haemophilus influenzae type b disease in England and Wales: who is at risk after 2 decades of routine childhood vaccination? *Clin Infect Dis.* 2013;57(12):1715-21.
- Daum RS, Zenko CE, Given GZ, et al. Magnitude of interference after diphtheria-tetanus toxoid-acellular

- pertussis/Haemophilus influenzae type b capsular polysaccharide vaccination is related to the number of doses administered. *J Infect Dis*. 2001;184:1293–9.
- Dagan R, Poolman JT, Zepp F. Combination vaccines containing DTPaHib: impact of IPV and co-administration of CRM¹⁹⁷ conjugates. *Expert Rev Vaccines*. 2008;7:97–115.
- Decker MD, Edwards KM, Bradley R, Palmer P. Comparative trial in infants of four conjugate Haemophilus influenzae type b vaccines. *J Pediatr*. 1992;120(2 pt. 1):184–9.
- European Centre for Disease Prevention and Control. Annual epidemiological report: invasive Haemophilus influenzae. Stockholm ECDC. 2016. Available at: http://ecdc.europa.eu/en/healthtopics/Haemophilus_Influenzae_Infection/Pages/Annual-epidemiological-report-2016.aspx
- Käyhty H, Peltola H, Karanko V, Makela PH. The protective level of serum antibodies to the capsular polysaccharide of Haemophilus influenzae type b. *J Infect Dis*. 1983;147:1100.
- Käyhty H, Eskola J, Peltola H, et al. Antibody responses to four Haemophilus influenzae type b conjugate vaccines. *Arch Dis Child*. 1991;145(2):223–7.
- Kelly DF, Moxon ER, Pollard AJ. Haemophilus influenzae type b conjugate vaccines. *Immunology*. 2004;113:163–74.
- Ladhani S, Ramsay ME, Chandra M, Slack MP. EU-IBIS No evidence for Haemophilus influenzae serotype replacement in Europe after introduction of the Hib conjugate vaccine. *Lancet Infect Dis*. 2008;8(5):275–6.
- Ladhani S, Slack MP, Heath PT, von Gottberg A, Chandra M, Ramsay ME. Invasive Haemophilus influenzae disease, Europe, 1996–2006. European Union invasive bacterial infection surveillance participants. *Emerg Infect Dis*. 2010a;16(3):455–63.
- Ladhani S, Heath PT, Slack MP, McIntyre PB, Diez-Domingo J, Campos J, Dagan R, Ramsay ME, Participants of the European Union Invasive Bacterial Infections Surveillance Network. Haemophilus influenzae serotype b conjugate vaccine failure in twelve countries with established national childhood immunization programmes. *Clin Microbiol Infect*. 2010b;16(7):948–54.
- Ladhani SN, Ladhani SN, Collins S, Vickers A, Litt DJ, Crawford C, Ramsay ME, Slack MPE. Invasive Haemophilus influenzae serotype e and f disease in England and Wales. *Emerg Infect Dis*. 2012;18(5):725–32.
- McVernon J, Howard AJ, Slack MP, Ramsay ME. Long-term impact of vaccination on Haemophilus influenzae type b (Hib) carriage in the United Kingdom. *Epidemiol Infect*. 2004;132:765–7.
- Pace D. Glycoconjugate vaccines. *Expert Opin Biol Ther*. 2013;13(1):11–33.
- Paul Ehrlich Institute. Vaccines for Haemophilus influenzae type b. n.d.. Available at: <http://www.pei.de/EN/medicinal-products/vaccines-human/haemophilus-influenzae-type-b-hib/haemophilus-influenzae-type-b-hib-node.html>
- Peltola H, Käyhty H, Sivonen A, Mäkelä H. Haemophilus influenzae type b capsular polysaccharide vaccine in children : a double blind field study of 100,000 vaccinees 3 months to 5 years of age in Finland. *Pediatrics*. 1977;60:730–7.
- Peltola H. Worldwide Haemophilus influenzae type b disease at the beginning of the 21st century: global analysis of the disease burden 25 years after the use of the polysaccharide vaccine and a decade after the advent of conjugates. *Clin Microbiol Rev*. 2000;13:302–17.
- Poolman J, Kaufhold A, DeGrave D, Goldblatt D. Clinical relevance of lower Hib response in DTPa-based combination vaccines. *Vaccine*. 2001;19:2280–5.
- Slack MPE, Azzopardi HJ, Hargreaves RM, Ramsay ME. Enhanced surveillance of invasive Haemophilus influenzae disease in England, 1990 to 1996: impact of conjugate vaccine. *Pediatr Infect Dis J*. 1998;17:S204–7.
- Sturgess AW, Rush K, Charbonneau RJ, et al. Haemophilus influenzae type b conjugate vaccine stability: catalytic depolymerisation of PRP in the presence of aluminium hydroxide. *Vaccine*. 1999;17:1169–78.
- Takala A, Eskola J, Peltola H, Makela H. Epidemiology of invasive Haemophilus influenzae type b disease among children in Finland before vaccination with Haemophilus influenzae type b conjugate vaccine. *Ped Infect Dis J*. 1989;8:297–301.
- Takala AK, Meuman O, Kleemola M, et al. Preceding respiratory infection predisposing for primary and secondary invasive Haemophilus influenzae type b disease. *Ped Infect Dis J*. 1993;12:189–95.
- Van Eldere J, Slack M, Ladhani S, Cripps A. Non-typeable Haemophilus influenzae: an under-recognised pathogen. *Lancet Infect Dis*. 2014;14(12):1281–92.
- Vidor E, Offenbach A, Fletcher MA. Haemophilus influenzae type b vaccine: reconstitution of lyophilised PRP-T vaccine with a pertussis-containing paediatric combination vaccine, or a change in the primary series immunisation schedule, may modify the serum anti-PRP antibody responses. *Cur Med Res Open*. 2001;17:197–209.
- Watt JP, Wolfson LJ, O'Brien KL. Burden of disease caused by Haemophilus influenzae type b in children younger than 5 years: global estimates. *Lancet*. 2009;374:903–11.
- World Health Organisation. Third dose of Hib vaccine Reported estimates 2016. Available at: http://apps.who.int/immunization_monitoring/globalsummary/timeseries/tscveragehib3.html

Pediatric Combination Vaccines

Federico Martín-Torres

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20.1 Introduction: The Need, Challenges and Benefits of Combination Vaccines

Since the beginning of the vaccination era, the number of vaccine-preventable diseases has continued to increase at a fast rate. Traditionally, with each new vaccine included in the vaccination schedule, a new injection was required to administer the immunization, and this sparked multiple responses from different social sectors: on the one hand, general practitioners were confused by the ever-changing immunization schedules; on the other hand, parents were concerned about their children becoming “pincushions”. This problem, far from being solved, continued to worsen as the number of vaccines in development raised each year, making the situation more pressing.

Different approaches emerged to address the problem. One of these involved deferring additional injections until the next office visit. However, ultimately, this strategy backfired: the increasing costs and burden on staff associated with the scheduling of new visits, combined with the increased likelihood of vaccinations being missed, ended up jeopardizing vaccination coverages. In this context, the necessity for combination vaccines became acute.

Combination vaccines are individual preparations that include two or more antigens of different microorganisms. Combination vaccines have been used in adults and children alike for over half a century; in 1948, the combination of diphtheria, tetanus, and pertussis antigens into a single vaccine was first used to vaccinate infants and children. Since then, many new techniques have been developed and the number of components combined into a single product has risen greatly.

Combination vaccines have not only solved the burden of multiple injections. Other challenges such as the storage and shipment of vaccines, the increasing number of visits, the injection of more adjuvants, or the introduction of new vaccines into the calendar have been met, owing

to the availability of combination vaccines (■ Fig. 20.1).

20.2 The “Perfect” Combination Vaccine

An ideal combination vaccine needs to meet the following requirements:

- *Safety and efficacy*: a new combination vaccine should not be more reactive, less immunogenic or less efficacious than the individual components administered separately.
- *Fit the established immunization schedule*: a combination vaccine should include components that are normally administered at the same immunization visit and respect its established timing and interval, with only slight variations being acceptable.
- *Ease of use*: from the practical point of view, a combination vaccine should be easy to store and administer, and not increase the burden on staff.

20.3 Composition of Combination Vaccines

Commonly administered combination vaccines include as base the diphtheria and tetanus toxoid, used alone (DT or Td) or with whole cell (DTwP) or acellular (DTaP) pertussis component (■ Fig. 20.2). To this baseline product, a plethora of components can be added. Common combinations include inactivated poliovirus (IPV), *Haemophilus influenzae b* vaccine (Hib) and/or hepatitis B vaccine (HepB). An additional component may be hepatitis A vaccine (HA).

Another branch of combination vaccines is live attenuated measles–mumps–rubella vaccine (MMR), with a more recent addition of a varicella vaccine (V) component (see ► Chap. 9). Henceforth, this chapter focuses specifically on the pentavalent and hexavalent combination vaccines (■ Tables 20.1 and 20.2).

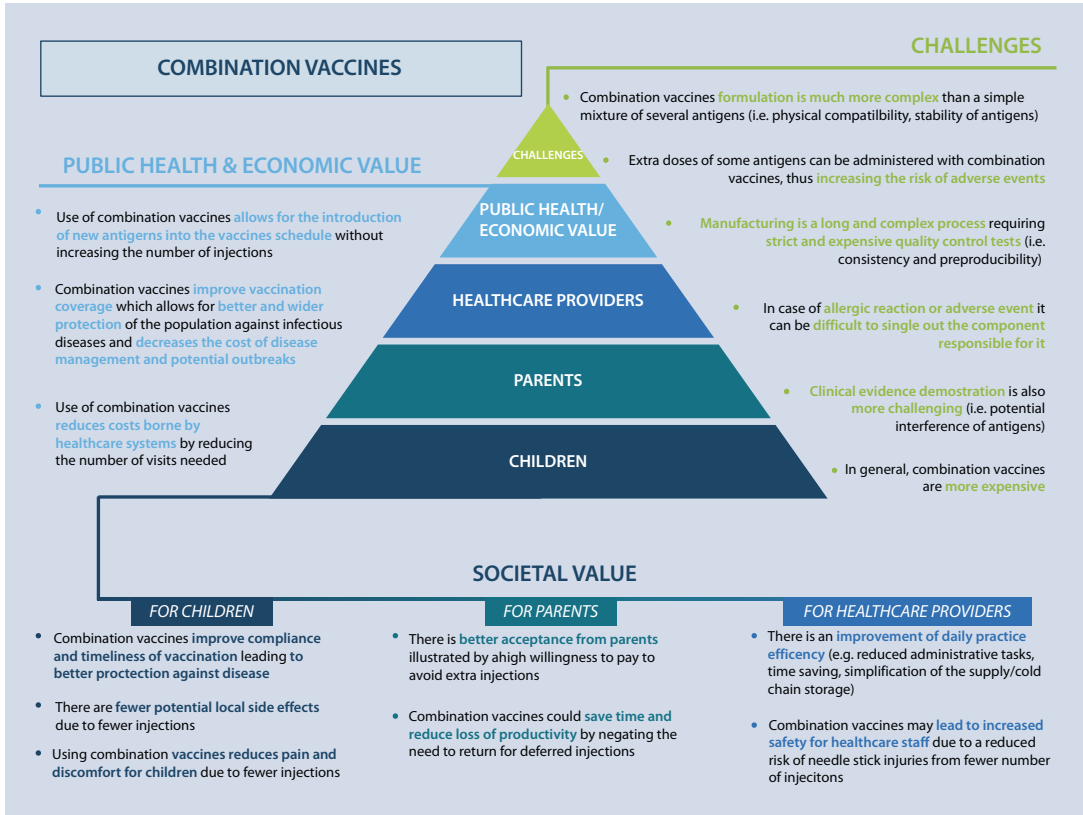


Fig. 20.1 Combination vaccines: from challenges to benefits (Adapted from Maman et al. 2015). Several key benefits from combination vaccines can be easily

identified, with societal and public health and economic categories being the most important. Also, important challenges should be considered

Fig. 20.2 Development of combination vaccines based on DTPa. (*) heptavalent vaccine with MenC under development

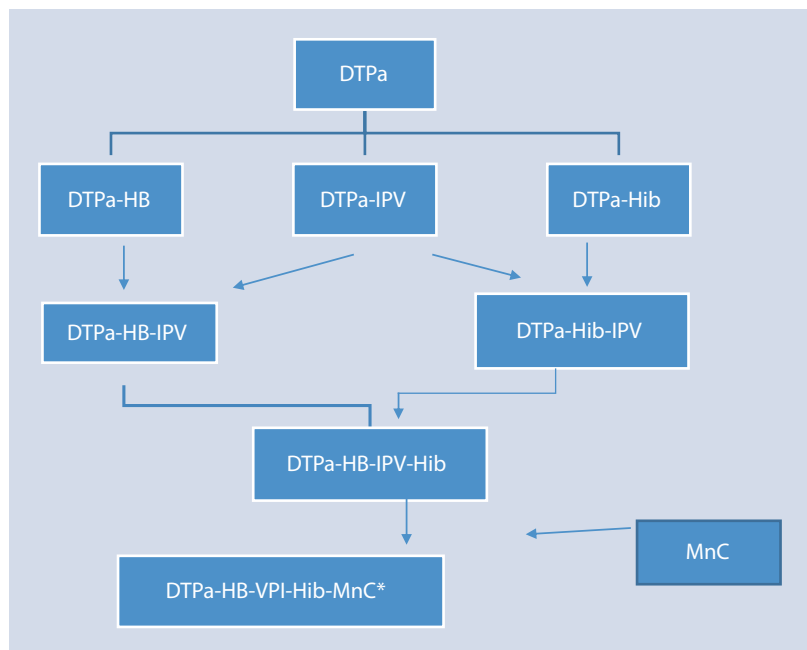


Table 20.1 Pentavalent combination vaccines. Summary of characteristics of main pentavalent vaccines available in Europe

Combination	Commercial name	Company	DT	TT	PT (mcg)	FHA (mcg)	PRN (mcg)	FIM (mcg)	HBs (mcg)	PRP-T (mcg)	Polio 1 (D)	Polio 2 (D)	Polio 3 (D)	Adjuvants
DTPa-IPV-Hib	Pediacel®	Sanofi Pasteur	30 IU	40 IU	20	20	3	5	–	10–20	40	8	32	Aluminum phosphate
	Infanrix IPV/Hib®	GSK	30 IU	40 IU	25	25	8	–	–	10–30	40	8	32	Aluminum hydroxide, hydrated
	Pentaxim® Pentavac®	Sanofi Pasteur	30 IU	40 IU	25	25	–	–	10	10	40	8	32	Aluminum hydroxide, hydrated

DT diphtheria toxoid, *TT* tetanus toxoid, *PT* pertussis toxoid, *FHA* filamentous hemagglutinin, *PRN* pertactin, *FIM* fimbriae type 2 and 3, *HBs* hepatitis B surface antigen, *PRP-T* polyribosylribitol phosphate conjugated to tetanus toxoid, *Polio 1* poliovirus (inactivated) type 1 (Mahoney), *Polio 2* poliovirus (inactivated), type 2 (MEE-1), *Polio 3* poliovirus (inactivated) type 3 (Saukett)

Table 20.2 Hexavalent combination vaccines. Summary of characteristics of main hexavalent vaccines available in Europe

Combination	Commercial name	Company	DT (IU)	TT (IU)	PT (mcg)	FHA (mcg)	PRN (mcg)	FIM (mcg)	HBs	PRP-T ^a PRP-OMC (mcg) ^b	Polio 1/2/3 (D)	Adjuvants	Presentation
DTaP5-HB-IPV-Hib	Vaxelis®	Sanofi Pasteur and MSD	20	40	20	20	3	5	10	3 / 50 mcg ^b	40 / 8 / 32	Aluminum phosphate Aluminum hydroxy-phosphate sulfate	Fully liquid 0.5 mL suspension (prefilled syringe) for intramuscular injection
DTaP2-HB-IPV-Hib	Hexyon® Hexacima® Hexaxim®	Sanofi Pasteur	20	40	25	25	-	-	10	12 / 22 to 36 mcg ^a	40 / 8 / 32	Aluminum hydroxide, hydrated	Fully liquid 0.5 mL suspension (prefilled syringe) for intramuscular injection
DTaP3-HB-IPV + Hib	Infanrix-Hexa®	GSK	30	40	25	25	8	-	10	10 / 25 mcg ^a	40 / 8 / 32	Aluminum hydroxide, hydrated Aluminum phosphate	Powder (Hib lyophilized) and suspension for 0.5 ml suspension for intramuscular injection

DT diphtheria toxoid, TT tetanus toxoid, PT pertussis toxoid, FHA filamentous hemagglutinin, PRN pertactin, FIM fimbriae type 2 and 3, HBs hepatitis B surface antigen, PRP-T polyribosylribitol phosphate conjugated to tetanus toxoid, PRP-OMC polyribosylribitol phosphate conjugated to meningococcal protein, Polio 1 poliovirus (inactivated) type 1 (Mahoney), Polio 2 poliovirus (inactivated), type 2 (MEF-1), Polio 3 poliovirus (inactivated) type 3 (Saukett)

^aPrepared with PRP-T

^bPrepared with PRP-OMC

20.4 Introduction to Pentavalent and Hexavalent Vaccination

With the new immunization recommendations made by the WHO, the number of routine vaccinations has grown from the initial 6 recommended EPI antigens – bacillus Calmette–Guérin, diphtheria, tetanus, pertussis, poliomyelitis, and measles – to the current 11 antigens, which additionally include HepB, Hib, pneumococcus, rotavirus, and rubella. This increase meant that the development of pentavalent and hexavalent combination vaccines fitting the routine vaccination schedules became a necessity. In this respect, Europe has taken the lead in comparison with other world regions, and routine vaccination with pentavalent and hexavalent combinations, including DTPa, Hib, HepB, and IPV, has been on European vaccination programs for more than 15 years. Since the marketing authorization of Hexavac® and Infanrix Hexa® in 2000, immunization schedules in most European countries have included hexavalent vaccines. With the introduction of combination vaccines, there has been an increase in acceptance and vaccination coverage, especially for HepB.

20.4.1 Pentavalent

1. DTaP-IPV-Hib (Pediace®l, Infanrix IPV-Hib®, Pentavac®/Pentaxim®)

Pediace®l (Sanofi Pasteur) is indicated for primary and booster vaccination against diphtheria, tetanus, pertussis, poliomyelitis, and invasive Haemophilus influenzae type b disease in infants and children from the age of 6 weeks up to the fourth birthday.

► <https://www.medicines.org.uk/emc/medicine/26217>

Infanrix IPV-Hib® (GSK) is indicated for active immunization against diphtheria, tetanus, pertussis, poliomyelitis, and Haemophilus influenzae type b disease from the age of 2 months.

► <https://www.medicines.org.uk/emc/medicine/28678>

Pentavac® / Pentaxim® (Sanofi Pasteur) is indicated for active immunization against diphtheria, tetanus, pertussis, poliomyelitis, and Haemophilus influenzae type b for primary vaccination in

infants, as a booster in children who have previously received a primary vaccination with this vaccine, or a diphtheria-tetanus-whole-cell or acellular pertussis-poliomyelitis vaccine, whether mixed or not with freeze-dried conjugate Haemophilus influenzae type b vaccine.

► <http://www.medicines.ie/printfriendlydocument.aspx?documentid=4541&companyid=202>

The pentavalent combination including DTaP, IPV, and Hib is the most widely distributed and used combination in Europe. This combination vaccine is available in 18 out of 33 European countries, either as the main pillar of the routine vaccination program, or to complement vaccination recommendations where the hexavalent would add an unnecessary additional HepB dose.

2. DTaP-Hib-HepB (This combination is not available on the European market)
3. DTaP-IPV-HepB (This combination is not available on the European market)

Some European countries, especially in eastern Europe, still use DTwP-containing combination vaccines in their routine vaccination programs. The human immune responses against aP vaccines are directed against purified protein virulence factors whereas in wP vaccines, it is directed against an array of antigens of the whole bacterial cells. However, changes in effectiveness of wP have occurred without being noticed in the production or lot release process, which has not happened so far with aP vaccines. The use of wP-based vaccines makes the vaccines more affordable than their acellular pertussis counterparts, with significantly lower prices (see ► Chap. 18).

20.4.2 Hexavalent

DTaP-IPV-Hib- HepB (Infanrix Hexa®, Vaxelis®, Hexyon/Hexacima/Hexaxim®)

hINFANRIX Hexa® (GSK) ► http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Product_Information/human/000296/WC500032505.pdf

Hexyon®, Hexacima®, Hexaxim® (Sanofi Pasteur) ► http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Product_Information/human/002702/WC500145808.pdf

Vaxelis® Sanofi Pasteur and MSD ▶ http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Product_Information/human/003982/WC500202435.pdf

Four hexavalent vaccines have been licensed in Europe in the last 16 years and Europe has been the first region in the world to adopt hexavalent vaccines as part of the routine immunization program. As many as 20 out of 33 European countries routinely use vaccines combining antigens of six different diseases in children (■ Table 20.3).

Immune responses to the diphtheria, tetanus, and polio components of the different hexavalent combinations are non-inferior to those of the separate components. Although there is no serological correlate of protection against pertussis disease, the clinical efficacy of Infanrix® Hexa against pertussis has been demonstrated in household contact studies, and the more recent hexavalent vaccines have shown to achieve comparable seroprotective titers for the shared antigens. Hexyon®, Infanrix Hexa®, and Vaxelis® include 2, 3, and 5 pertussis antigens respectively, with pertussis toxoid and filamentous hemagglutinin common to the three formulations (see ▶ Chap. 18).

A fourth hexavalent vaccine, Hexavac®, was withdrawn in 2005 because of rapid waning of antibody titers against Hep B component. Currently available hexavalent vaccines induce comparable immune responses to Hep B Infanrix®. Hexa contains the same HepB component as used in Engerix®B- with a different dose compared with Hexavac®. The three hexavalent vaccines use recombinant DNA technology for B hepatitis antigen production in yeast: Infanrix Hexa® and Vaxelis® use *Saccharomyces cerevisiae* whereas Hexyon® produces it in *Hansenula polymorpha* cells.

While Vaxelis® and Hexyon® are fully liquid, ready-to-use vaccines, Infanrix® Hexa requires reconstitution before administration. Data regarding the long-term persistence of immune response, immune memory, and vaccine effectiveness of Vaxelis® and Hexyon® are still needed as compared with Infanrix® Hexa.

20.5 Practical Considerations

20.5.1 Concomitant Administration with Other Vaccines

DTaP combination vaccines may be given at the same time as pneumococcal conjugate, rotavirus, meningococcal conjugate and measles, mumps, rubella, and varicella vaccines. The potential for the interaction of DTaP-based penta- and hexavalent vaccines with these four vaccines has been studied in several clinical trials, and no important variations in the antibody titers were found.

20.5.2 Interchangeability

Monovalent vaccines and combination vaccines for the same diseases produced by the same manufacturer usually carry similar antigens, with no issues regarding interchange of vaccines. Questions arise, however, between hexavalent vaccines manufactured by different companies.

Several studies have addressed interchangeability and shown that vaccines containing diphtheria, tetanus, poliovirus, HepB, and Hib antigens are generally interchangeable. As there is no serological correlation of protection for pertussis, the interchangeability for those vaccines containing pertussis antigens has remained unclear for a long time. Owing to this, recommendations state that whenever possible, it would be preferable to use the same manufacturer's vaccine, at least for priming, but no contraindication has been stated against the opposite procedure. A number of studies have shown that combining aP-containing vaccines from different manufacturers regardless of the immunization schedule will provide similar seroprotective levels and immune memory as if they were the same vaccine.

In general, it is always preferable to use the same vaccine, at least in the priming schedule. Only if the timeliness of the immunization of the child can be affected, or if the vaccine administered previously is unknown, vaccination with vaccines containing similar antigens is not contraindicated.

Table 20.3 Use of pentavalent and hexavalent vaccines in immunization schemes in Europe

	Countries	DTPa, VPI, Hib		HepB		Hexavalent	Pentavalent
		Priming age (months)	Booster age (months)	Universal	Schedule (months)	Use (months)	DTPa, VPI, Hib (months)
2 + 1	Austria	3, 5	12	Yes	3, 5, 12	3, 5, 12	No
	Italy		11–13	Yes	3, 5–6, 11–13	3, 5–6, 11–13	3, 5–6, 11–13
	Iceland		12	No	–	No	3, 5, 12
	Denmark		12	No, RG only	–	3, 5, 12	3, 5, 12
	Finland		12	No, RG only	–	No	3, 5, 12
	Norway		12	No, RG only	–	No	3, 5, 12
	Sweden		12	No, RG only	–	3, 5, 12	3, 5, 12
	Slovakia	2, 4	10–11	Yes	2, 4, 10	2, 4, 10	No
	France		11	Yes	2, 4, 11	2, 4, 11	No
	Spain		11	Yes	2, 4, 11	2, 4, 11	No
3 + 1	Greece	2, 4, 6	15–18	Yes	2, 4, 6–18	No	No
	Ireland		13 (Hib)	Yes	2, 4, 6	2, 4, 6	No
	Portugal		18 (DTPa, Hib)	Yes	0, 2, 6	No	2, 4, 6
	Romania		12	Yes	0, 2, 6	2, 4, 11	No
	Lithuania		18	Yes	0, 1, 6	No	2, 4, 6, 18
	Latvia		12–15	Yes	2, 4, 6, 12–15	2, 4, 6, 12–15	2, 4, 6
	Cyprus		15–18	Yes	2, 4, 8–12	No	2, 4, 6, 15–18
	Croatia		12–23	Yes	0, 2, 6	2, 4, 6, 12	No
	Switzerland		15–24	No	1, 6, 15–24	No	2, 4, 6, 15–24
	Germany	2, 3, 4	11–14	Yes	2, 3, 4, 11–14	2, 3, 4, 11–14	2, 3, 4, 11–14
	Belgium		15	Yes	2, 3, 4, 15	2, 3, 4, 15	No
	Netherlands		11	Yes	2, 3, 4, 11	2, 3, 4, 11	No
	Luxembourg		13	Yes	2, 3, 13	2, 3, 13	4
	UK		12–13 (Hib)	No, RG only	–	No	2, 3, 4
	Malta		18	Yes	12, 13, 18	No	6 weeks, 3, 4, 18
	Hungary		18	Yes	Over 2 years	No	2, 3, 4, 18
	Czech Republic		10	Yes	2, 3, 4, 10	2, 3, 4, 18	No
	Bulgaria	16	Yes	0, 1, 6	2, 3, 4	2, 3, 4, 16	
	Estonia	3, 4–5, 6	24	Yes	0, 1, 6	No	3, 4–5, 6, 2 years
	Slovenia		12–24	No, RG only	Over 2 years	No	3, 4–5, 6, 18
Poland	16–18 (DTPw, VPI, Hib)		Yes	0, 2, 7	No	No	

Data compiled in January 2017
 RG risk groups

20.5.3 Vaccination schedules

In general, the schedules regarding pentavalent/hexavalent vaccines used in Europe can be summarized as either 2 + 1 or 3 + 1. Both schedules have proved to be effective for pentavalent and hexavalent vaccines. The specific schedules of the available hexavalent vaccines according to their label are summarized in [Table 20.4](#).

20.6 Concerns and Issues of a Lifetime with Combination Vaccines

20.6.1 Multiple Antigens and Immunity Overload

As the number of antigens administered to infants has kept growing, some parents and also

Table 20.4 Posology specified in the summary of product characteristics of the different hexavalent vaccines available

	Full-term infants		Preterm infants >24 weeks		HepB
	Primary vaccination (minimum 6 weeks old)	Booster vaccination	Primary vaccination	Booster vaccination	
Infanrix® Hexa	3-dose (at least 1-month intervals between doses)	At least 6 months after priming and preferably before 18 months ^a	3-dose (at least 1-month intervals between doses)	At least 6 months after priming and preferably before 18 months	In the absence of hepatitis B vaccination at birth, it is necessary to give a hepatitis B vaccine booster dose. Hexavalent vaccines can be considered for HepB booster dose. When a hepatitis B vaccine is given at birth, hexavalent vaccines can be used as replacement for supplementary HepB doses after week 6.
	2-dose (at least 2-month intervals between doses)	At least 6 months after priming and preferably before 11–13 months ^a			
Hexyon®	3-dose (at least 1-month intervals between doses)	At least 6 months after priming ^b	No data available		
	2-dose (at least 2-month intervals between doses)	At least 6 months after priming ^b			
Vaxelis®	3-dose (at least 1-month intervals between doses)	At least 6 months after priming ^c	Can be given	Can be given	
	2-dose (at least 1-month intervals between doses)	At least 6 months after priming ^c	Can be given	Can be given	

^aNot after 36 months old

^bNot after 24 months old

^cNot after 15 months old

healthcare professionals have expressed concerns about a possible overload of the immune system of children. This theory has been widely discussed and convincingly refuted, but misguided concerns still populate the internet. Children are commonly exposed to many more antigens in daily life than those injected in the vaccines, with no negative impact on the immune system.

20.6.2 Hexavalent Vaccine Safety and Their Relation to Sudden Unexpected Death

An association between hexavalent vaccination and the occurrence of sudden unexpected death (SUD) was suspected when a series of three SUDs were reported in Germany within 48 h of the administration of the booster dose of Hexavac® between 2000 and 2003. Standardized mortality ratios for SUD cases within 1 day of vaccination were 31.3 (95% CI 3.8–113.1; 2 cases observed; 0.06 cases expected), and 23.5 within 2 days of vaccination (95% CI 4.8–68.6; 3 cases observed; 0.13 cases expected), so even when these data did not prove a causal relationship, an alarm signal was raised and further investigation began. The Committee for Proprietary Medicinal Products (CPMP) issued a statement in 2003 after a statistical analysis based on the German data, and found no plausible biological cause for association between hexavalent vaccines and SUD in the 2nd year of life.

In Italy, a case series studying neonates born in the period 1999–2004 reported that the association between hexavalent vaccine administration and risk of SUD in the first 14 days after vaccine administration was significantly lower than that estimated in Germany; the authors claimed that this association was limited to the first vaccine dose only, at an age coinciding with the highest incidence of SUD. Relative risk in the first 2 days after vaccination was 0.7 and 2.3 for Hexavac® and Infanrix® Hexa respectively; the risk was 2.8 vs 1.4 and 1.6 vs 1.5 for the first week and for the 2 weeks after vaccine administration respectively. Based on these data, it was concluded that the limited increase in relative risk appeared to be confined to the first dose, and that it may be partially explained by the confounding effect of age.

Other studies performed so far have confirmed that none of the hexavalent vaccines used at the moment had any distinct effect on SUD.

20.6.3 Reduced Hib Response When Combined with DTaP

The most commonly reported example of immune interference in DTaP-based combination vaccines is the reduction in antibody titers to the Hib component of the vaccine polyribosylribitol phosphate (PRP) antigen. wP-based vaccines do not show this interference to the same extent, as the wP component may be acting as an adjuvant.

An interference between tetanus toxoid (TT) and Hib has been demonstrated. In Hib vaccines, TT acts as a carrier protein conjugated to the PRP. Several reasons for this interference have been mentioned: competition between TT-specific and PRP-specific B cells for the Hib conjugate antigen, suppression of PRP response by clonal expansion of TT-specific B cells, and physical prevention of the binding between the conjugate antigen and PRP-specific B cells by the TT carrier protein. FHA has also been proven to interact with PRP. Studies show that FHA is a suppressor of IL12 and IFN γ , suppressing immune responses to co-injected antigens. Lastly, aluminum hydroxide has been reported to be incompatible with Hib, with 5–11 times lower levels of PRP antibodies.

Whatever the case, this lower response does not have a clinical impact. It has been stated that the current seroprotective threshold against PRP is probably too high, and that antibody responses below this threshold are similarly protective. Furthermore, the newest hexavalent vaccine combines PRP with meningococcal outer membrane protein (PEP-OMPC), which is known to elicit a stronger early immunogenic response against Hib than the PRP-T antigen.

20.6.4 Combining with Neonatal Hepatitis B Immunization

In the case of hepatitis B, several countries administer the first dose at the time of birth, as recommended by the WHO. The other compo-

nents of the combination vaccine are not to be administered in the first days of life and a combination of HepB and DTaP still requires administration of monovalent HepB at birth followed by doses in combination with DTaP at 2, 4, and 6 months, resulting in an unnecessary fourth dose of HepB at the 6 month. A study comparing the DTaP–HepB combination administered at 2, 4, and 6 months with separate administration of HepB at birth, 1, and 6 months and DTaP at 2, 4, and 6 months showed significantly lower HepB antibody titers with the combination vaccine. However, antibody levels were still above serologically recognized levels of protection in 99% of the subjects. Furthermore, administration of a DTaP–HepB–IPV/Hib vaccine at 2, 4, and 6 months after a dose of HepB vaccine shortly after birth did not have an impact on protective anti-HBs titers and was not more reactogenic than the same combination given without the birth dose of HepB.

20.6.5 HepB Reduction in Long-Term Protection

Rapid waning of hepatitis B vaccine-induced antibodies was the reason for the withdrawal of the hexavalent combination vaccine, Hexavac®, by the EMEA in 2005. Although >95% of children vaccinated with Hexavac® had seroprotective antibody levels after primary vaccination, up to 20% of them were relatively low (≤ 100 IU/L) and these subjects had a lower response to the booster dose. This observation was also reflected in studies where Hexavac® was co-administered with pneumococcal vaccine or meningococcus C conjugate vaccine. It was assumed that these children might not have assured protection against hepatitis B during adolescence and adulthood. This theory notwithstanding, no increase in hepatitis B infection has been recorded in those countries where Hexavac® was widely used. In a subsequent study, Zanetti et al. showed that even though 60% of the 5- to 6-year-old children studied did not have seroprotective levels against HepB before the booster dose, a protective antibody response was induced in 92.1% of the participants. The authors concluded that Hexavac®-vaccinated children

maintained T-cell memory and were able to trigger anti-HB production by B cells when exposed to the viral antigen.

At the same time, it has been shown that vaccine dosage and the length of the gap between the last and preceding doses in the primary series are the main determinants of immune persistence in HepB vaccination. The new generation of hexavalent vaccines contain increased amounts of Hep B antigen to avoid this issue.

20.6.6 Shortage Acellular Pertussis Component

Starting in 2015, there was a shortage in the pertussis acellular component of the combination vaccines in Europe, owing to reduced production capacities. This situation affects not only acellular pertussis vaccines, but also all the combination vaccines containing this component.

Europe has issued some recommendations to modify the immunization calendars of those countries enduring the shortage. Priority should be given in the following order:

- The infant primary immunization series (first year of life).
- The first toddler booster (second year of life) dose.
- If applicable, the first toddler booster dose should be prioritized over the school-entry booster.
- Eventually, the use of a low-antigen-content pertussis vaccine as a pre-school booster, instead of a regular-dose vaccine, while vaccinating these cohorts at a later age.

In countries where vaccination during pregnancy is recommended and Tdap vaccine is in short supply, it is suggested that doses should be preserved for maternal immunization, instead of adolescent or pre-school booster doses, since maternal immunization directly benefits newborns.

As an example, Spain has had to adjust its immunization schedule as a result of the shortage. Following the rise in demand for Tdap vaccines resulting from the start of the vaccination program in pregnant women against pertussis, it has been decided to temporarily withdraw the booster dose indicated for 6-year-old children to

preserve these pertussis-containing vaccine doses for primary vaccination, pregnant women, the and toddler booster dose. In addition, hexavalent vaccine is now administered in a 2 + 1 schedule.

20.6.7 Pertussis Components and Immunity Waning

The main components of the aP pertussis vary between different vaccines and include PT, FHA, PRN, and Fimbriae type 2 and 3 (FIM). Of these, only the PT component is deemed essential for conferring protection against pertussis infection, as demonstrated for example in Denmark, where a monovalent pertussis vaccine containing only PT has been in use for more than 15 years, with no pertussis outbreak since 2002. Conversely, several published papers have shown that other components such as FHA or pertactin do not protect against pertussis infection. Regardless of the inclusion or exclusion of the different components, no inferiority in immune response, immune duration, efficacy or safety has been reported in any of the commercialized DTaP combination vaccines.

Further Reading

- Aristegui J, Dal-Re R, Diez-Delgado J, et al. Comparison of the reactogenicity and immunogenicity of a combination diphtheria, tetanus, acellular pertussis, hepatitis B, inactivated polio (DTPa-HBV-IPV) vaccine, mixed with the *Haemophilus influenzae* type b (Hib) conjugate vaccine and administered as a single injection, with the DTPa-IPV/Hib and hepatitis B vaccines administered in two simultaneous injections to infants at 2, 4 and 6 months of age. *Vaccine*. 2003;21:3593–600.
- Carlsson RM, Claesson BA, Selstam U, Fagerlund E, Granström M, Blondeau C, Hoffenbach A. Safety and immunogenicity of a combined diphtheria-tetanus-acellular pertussis-inactivated polio vaccine-Haemophilus influenzae type b vaccine administered at 2-4-6-13 or 3-5-12 months of age. *Pediatr Infect Dis J*. 1998;17(11):1026–33.
- Decker MD. Principles of pediatric combination vaccines and practical issues related to use in clinical practice. *Pediatr Infect Dis J*. 2001;20(11 Suppl):S10–8.
- ECDC. Shortage of acellular pertussis-containing vaccines and impact on immunisation programmes in the EU/EEA. [▶ http://ecdc.europa.eu/en/publications/Publications/RRA-shortage-of-aP-containing-vaccines.pdf](http://ecdc.europa.eu/en/publications/Publications/RRA-shortage-of-aP-containing-vaccines.pdf).
- Eskola J, Olander RM, Hovi T, Litmanen L, Peltola S, Käyhty H. Randomised trial of the effect of co-administration with acellular pertussis DTP vaccine on immunogenicity of Haemophilus influenzae type b conjugate vaccine. *Lancet*. 1996;348(9043):1688–92.
- Eskola J, Ward J, Dagan R, Goldblatt D, Zepp F, Siegrist CA. Combined vaccination of Haemophilus influenzae type b conjugate and diphtheria tetanus pertussis containing acellular pertussis. *Lancet*. 1999;354:2063–8.
- Eposito S, Tagliabue C, Bosis S, Ierardi V, Gambino M, Principi N. Hexavalent vaccines for immunization in paediatric age. *Clin Microbiol Infect*. 2014;20(Suppl 5):76–85.
- Lee AW, Jordanov E, Boissard F, Marshall GS. DTaP5-IPV-Hib-HepB, a hexavalent vaccine for infants and toddlers. *Expert Rev Vaccines*. 2017;16(2):85–92.
- Mallet E, Belohradsky BH, Lagos R, Gothefors L, Camier P, Carrière JP, Kanra G, Hoffenbach A, Langue J, Undreiner F, Roussel F, Reinert P, Flodmark CE, Stojanov S, Liese J, Levine MM, Muñoz A, Schödel F, Hessel L, Hexavalent Vaccine Trial Study Group. A liquid hexavalent combined vaccine against diphtheria, tetanus, pertussis, poliomyelitis, Haemophilus influenzae type B and hepatitis B: review of immunogenicity and safety. *Vaccine*. 2004;22(11–12):1343–57.
- Maman K, Zöllner Y, Greco D, Duru G, Sendyona S, Remy V. The value of childhood combination vaccines: from beliefs to evidence. *Hum Vaccin Immunother*. 2015;11(9):2132–41.
- Marshall GS, Adams GL, Leonardi ML, Petrecz M, Flores SA, Ngai AL, Xu J, Liu G, Stek JE, Foglia G, Lee AW. Immunogenicity, safety, and tolerability of a hexavalent vaccine in infants. *Pediatrics*. 2015;136(2):e323–32. doi:10.1542/peds.2014-4102.
- Martín-Torres F, Boissard F, Thomas S, Sadorge C, Borrow R. PRI02C study group. Immunogenicity and safety of a new hexavalent vaccine (DTaP5-IPV-HB-Hib) administered in a mixed primary series schedule with a pentavalent vaccine (DTaP5-IPV-Hib). *Vaccine*. 2017;35(30):3764–72. doi:10.1016/j.vaccine.2017.05.043. Epub 2017 Jun 2.
- Nunes MC, Madhi SA. Review of a new fully liquid, hexavalent vaccine: Hexaxim. *Expert Opin Biol Ther*. 2013;13(4):575–93. doi:10.1517/14712598.2013.774368.
- Obando-Pacheco P, Rivero-Calle I, Gómez-Rial J, Rodríguez-Tenreiro Sánchez C, Martín-Torres F. *Vaccine*. New perspectives for hexavalent vaccines. 2017; S0264-410X(17)30860-5. doi: 10.1016/j.vaccine.2017.06.063. [Epub ahead of print]
- Skibinski DA, Baudner BC, Singh M, O'Hagan DT. Combination vaccines. *J Global Infect Dis*. 2011;3(1):63–72.
- Syed YY. DTaP5-HB-IPV-Hib Vaccine (Vaxelis®): a review of its use in primary and booster vaccination. *Paediatr Drugs*. 2017;19(1):69–80. doi:10.1007/s40272-016-0208-y.
- Thollot F, Scheifele D, Pankow-Culot H, Cheuvart B, Leyssen M, Uliyanov L, Miller JM. A randomized study to evaluate the immunogenicity and safety of a heptavalent diphtheria, tetanus, pertussis, hepatitis B, poliomyelitis, haemophilus influenzae b, and meningococcal serogroup C combination vaccine administered to

- infants at 2, 4 and 12 months of age. *Pediatr Infect Dis J.* 2014;33(12):1246–54.
- Vennemann MM, Butterfass-Bahloul T, Jorch G, Brinkmann B, Findeisen M, Sauerland C, Bajanowski T, Mitchell EA, GeSID Group. Sudden infant death syndrome: no increased risk after immunisation. *Vaccine.* 2007; 25(2):336–40.
- Vesikari T, Becker T, Vertruyen AF, Poschet K, Flores SA, Pagnoni MF, Xu J, Liu GF, Stek JE, Boissnard F, Thomas S, Ziani E, Lee AW. A Phase III randomized, double-blind, clinical trial of an investigational hexavalent vaccine given at two, three, four and twelve months. *Pediatr Infect Dis J.* 2017a;36(2):209–15. doi:[10.1097/INF.0000000000001406](https://doi.org/10.1097/INF.0000000000001406).
- Vesikari T, Borrow R, Da Costa X, Richard P, Eymen C, Boissnard F, Lockhart S. Concomitant administration of a fully liquid, ready-to-use DTaP-IPV-HB-PRP-T hexavalent vaccine with a meningococcal serogroup C conjugate vaccine in infants. *Vaccine.* 2017b;35(3):452–8. doi:[10.1016/j.vaccine.2016.11.053](https://doi.org/10.1016/j.vaccine.2016.11.053).
- Zepp F, Schmitt HJ, Cleerhout J, Verstraeten T, Schuerman L, Jacquet JM. Review of 8 years of experience with Infanrix Hexa (DTPa-HBV-IPV/Hib hexavalent vaccine). *Expert Rev Vaccines.* 2009;8(6):663–78.

Pneumococcal Vaccines

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21.1 Burden of Pneumococcal Disease

Streptococcus pneumoniae (Pnc) is a major cause of morbidity and mortality in children and the elderly worldwide. When classified by its polysaccharide capsule, Pnc has >95 serotypes, each capable of causing disease. However, the invasiveness varies by serotypes. Diseases caused by pneumococcus include severe infections, such as meningitis and bacteremia (both regarded as invasive pneumococcal disease; IPD), pneumonia, and other milder mucosal diseases, such as middle ear infection (otitis media) and sinusitis.

21.2 IPD: Bacteremia, Bacteremic Pneumonia, Meningitis, and Other IPD

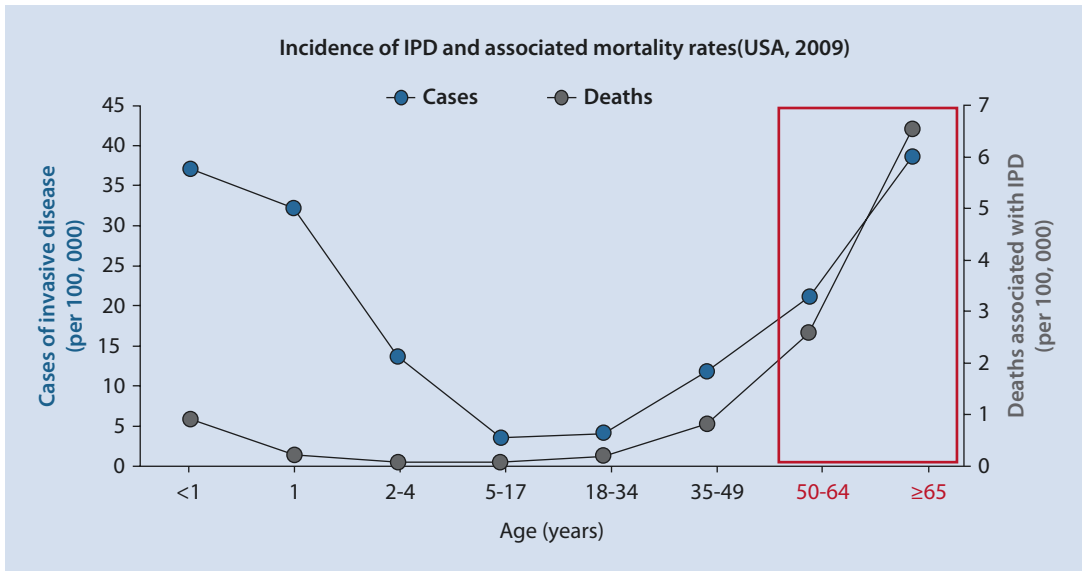
The highest IPD incidence occurs in children <2 years old. National pneumococcal surveillance programs are conducted in a number of countries in Europe, mainly in western countries, but also in Central and Eastern Europe (such as Croatia, Czech Republic, Hungary, Poland, Romania, and Slovakia) and in Israel. It was stated that in Europe, before widespread pneumococcal conjugate vaccine (PCV) immunization, the overall mean annual incidence of IPD in children aged <2 years was 44.4/100,000, and the mean case fatality rate for IPD was 3.5%. It is clear that figures vary between countries and populations and are largely dependent, beyond true differences, on epidemiological methods and reporting. Thus, for example, the rates reported for children <2 years, were approximately 15 cases per 100,000 in Germany and the Netherlands, but >90 and 104 per 100,000 in Spain and in Belgium respectively. Furthermore, considerable differences in IPD rates were noted comparing different studies conducted even in the same country (Table 21.1). Overall, in Europe, age-specific IPD rates were highest in those aged 65 years and over (13.8 cases per 100,000 population), followed by infants under 1 year of age (11.3 cases per 100,000 of the population). ▶ http://ecdc.europa.eu/en/health-topics/pneumococcal_infection/Pages/Annual-epidemiological-report-2016.aspx#sthash.53mozZJl.dpuf

Table 21.1 Annual incidence per 100,000 of invasive pneumococcal disease (IPD) in children <2 years old in Europe, pre-pneumococcal conjugate vaccine (PCV)

Country	Age (years)	Mean IPD incidence	Year
Austria	<2	14.5	2001–2003
Belgium	<2	104.4	2002–2003
Denmark	<2	43	1995–1999
Denmark	<2	50.9	2000–2005
Finland	<2	45.3	1985–1989
Germany	<2	16	1997–1998
Germany	<2	16.3	1997–2000
Hungary	2	12.5	2002–2004
Israel	<2	68.3	1993–1997
Israel	<2	77.4	1998–2002
Israel	<2	92.0	2003–2007
Italy	<2	11.3	2001–2002
Norway	<2	18.6	2001
Norway	<2	50	2000–2005
Poland	<2	19	2003–2004
Portugal	1	11.5	1999–2001
Slovenia	0–1	56.9	1993–2001
Spain	<1	110.2	1998–2001
Spain	<2	32.4	1997–2001
Spain	<2	48.4	1999–2001
UK	<2	17.2	1995–1997
UK	<2	37.8	1980–1999

Adapted from Isaacman et al. (2010)

Globally, pneumococcal infections cause ~11% of all deaths in children aged <5 years, mainly from pneumonia, reaching ~500,000 deaths annually. The pneumococcal vaccine could have the potential to reduce deaths from pneumonia and the impact on mortality could potentially be greater than that from the prevention of IPD in developed countries (Fig. 21.1), where hospitalization for pneumonia and the use of medical services for otitis media (OM) in young children constitute a considerable eco-



■ Fig. 21.1 Invasive pneumococcal disease (IPD) burden. IPD invasive pneumococcal disease, CDC ► <http://www.cdc.gov/abcs/reports-finding/survreports/spneu09.html>

nomic burden, particularly among the very young population (<5 years old).

Pneumococcal nasopharyngeal (NP) carriage precedes disease and is the source of pneumococcal spread in the community. Carriage rates are highest during early childhood, and thus, not only pneumococcal disease rates peak in young children, but these children are also the main source of Pnc spread.

Carriage rates vary considerably across Europe, and can be influenced by several factors, including the age of the population sampled, concomitant diseases, daycare center attendance, number of siblings, antibiotic usage, and the introduction and uptake of vaccines. In general, studies conducted in European crowded populations or in daycare centers show higher carriage rates.

The likelihood of *S. pneumoniae* causing disease depends upon several factors, including the invasiveness of the strains, the host susceptibility, and the existence of preceding or concurrent viral infection. Transmission of pneumococcus occurs mainly through direct and indirect contacts with respiratory secretions from patients and healthy carriers. In most cases, the individual is transiently and asymptotically colonized. However, occasionally, pneumococci can spread from the nasopharynx to cause mucosal disease, such as otitis media (by aspiration to the middle ear fluid through the Eustachian tube), sinusitis, and pneu-

monia (by *S. pneumoniae* aspiration to the lungs), or by direct invasion to the bloodstream, resulting in IPD, i.e., bacteremia (in some cases, sepsis), bacteremic pneumonia, and meningitis. (■ Fig. 21.2).

High-risk groups for the development of pneumococcal disease (both mucosal and IPD) include mostly either the very young or the elderly, children suffering from malnutrition, and immunocompromised populations (HIV, asplenia, immunosuppressive therapy, etc.).

21.3 *Streptococcus pneumoniae* Epidemiology

21.3.1 Pneumonia

Estimating the burden of childhood pneumonia is difficult, mainly because of the differences in case definitions and variations in trial end-points assessing this burden. The diagnosis of pneumonia usually derives from the clinical presentation: cough, fever, increased respiratory rate, crackles, and decreased respiration sounds. In young children, some of these clinical signs and symptoms can be absent. Radiography remains the best available tool for diagnosing pneumonia, although inter-observer variations are frequent. There is usually no confirmation of etiology in pneumonia cases (except in uncommon cases of bacteremic

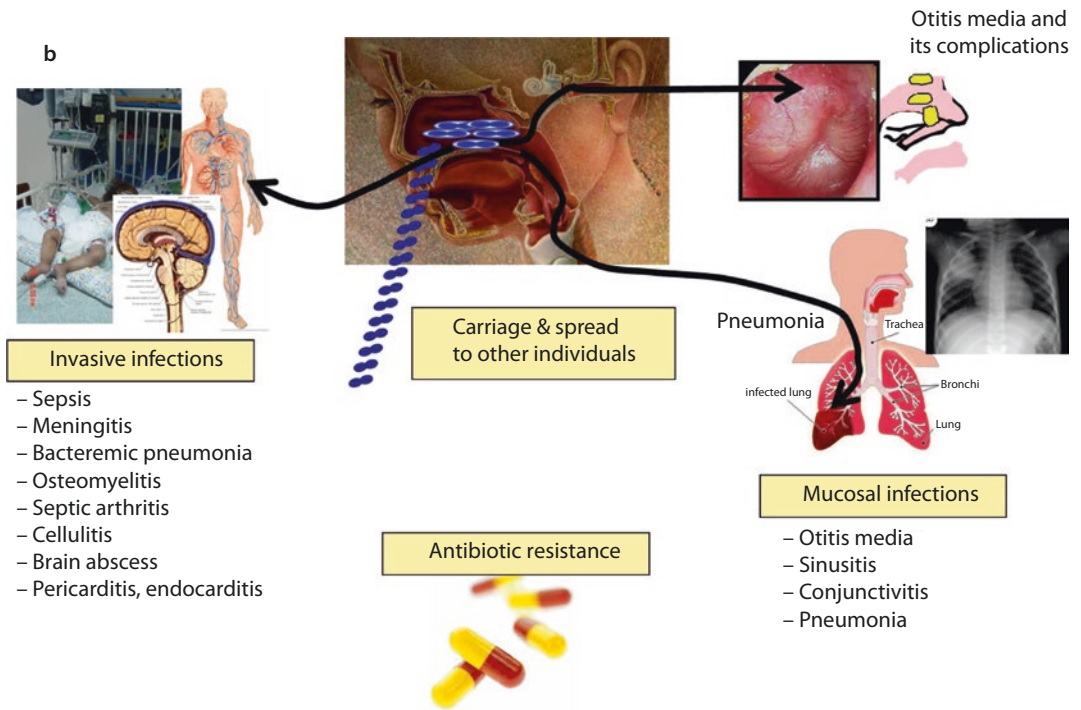
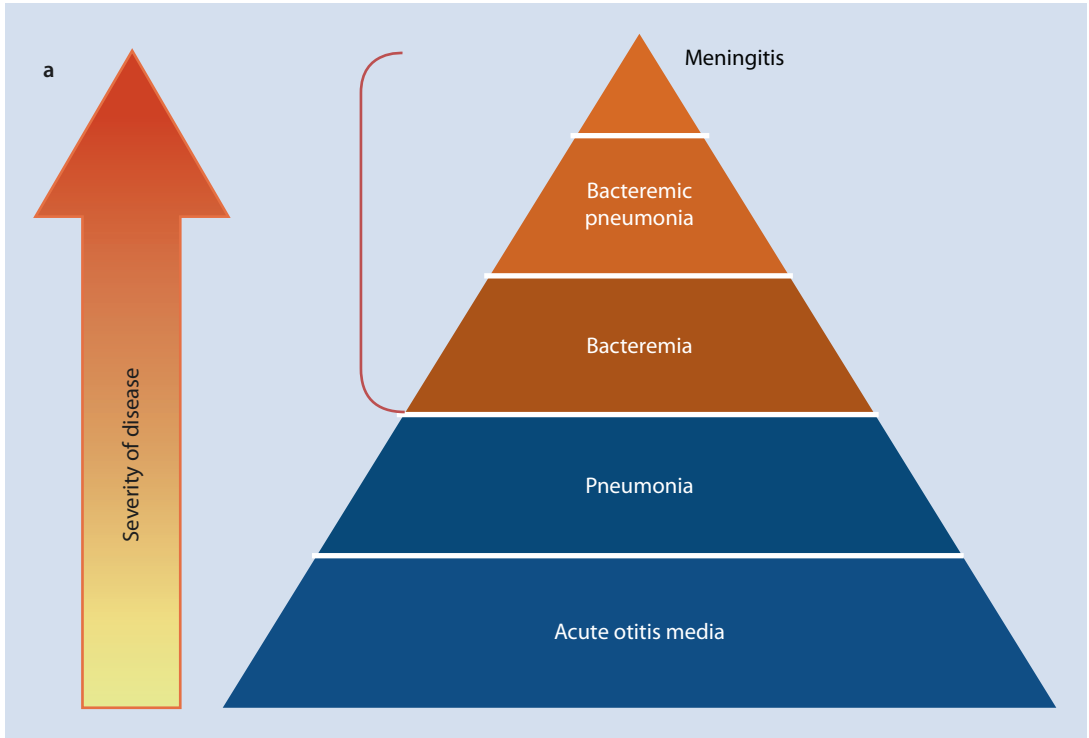
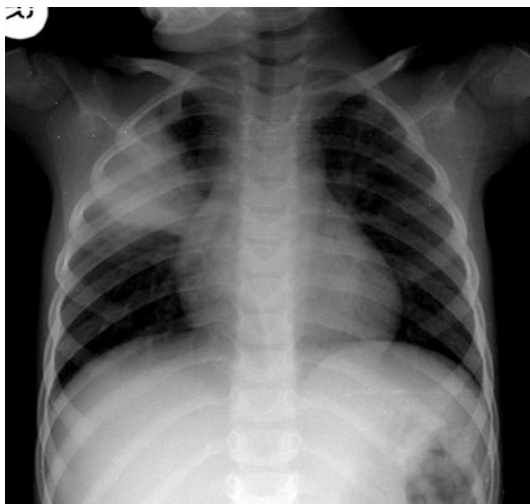


Fig. 21.2 a Non-invasive and IPD burden (Atkinson et al. 2009). b Mucosal and IPD burden



■ Fig. 21.3 Alveolar pneumonia on a chest X-ray

pneumonia, mechanically ventilated children with pneumonia, and pneumonia with pleural effusion).

Pneumococcal vaccination itself offers a method of estimation of the role of pneumococci in pneumonia. Reduction in all-cause pneumonia after vaccination is likely to reflect the etiological share of pneumococci.

Although alveolar infiltrates are considered mainly compatible with bacterial pneumonia, they are not pathognomonic and are also present in viral infections or viral–bacterial co-infections (■ Fig. 21.3). Furthermore, the WHO guidelines for the interpretation of chest radiographs resulted in a relatively high level of agreement between readers for the definition of “alveolar pneumonia” and “no pneumonia,” but poor agreement for non-alveolar pneumonia. This demonstrates the difficulties involved in reaching a consensus on the diagnosis of pneumonia.

Definitive measures such as positive blood cultures are only positive in 1–10% of all alveolar pneumonia cases. Sputum cultures, routinely used in adults, have a very low yield in children, as children cannot produce deep sputum, reflecting lower respiratory tract secretions.

21.3.2 Otitis Media

Otitis media is a major public health problem in early childhood worldwide; it is estimated that most children will suffer at least once from OM

and ~20% will suffer from recurrent or chronic OM (complex OM). The OM burden is huge in terms of the number of sick children, primary physician visits and antibiotic prescriptions. The disease peaks between the ages of 6 and 24 months. Before PCV introduction, *S. pneumoniae* accounted for approximately 30–60% of cases, and serotypes included in PCV7 and PCV13, constituted approximately 65% and 90% respectively of all pneumococcal cases. It is increasingly clear now that early OM is mainly caused by *S. pneumoniae*, especially by the more invasive serotypes, a high proportion of which are vaccine serotypes. Such early acute infections may be often missed clinically, as they may be asymptomatic or only mildly symptomatic during viral infections. Recurrent, nonresponsive, spontaneously draining, and chronic OM (termed together complex OM), are the sequelae of the first infections. In contrast to the first acute OM cases, in complex OM cases, the role of nontypeable *Haemophilus influenzae* (NTHi) is increasingly important, because, as with other chronic or recurrent respiratory tract infections, this organism recognizes damage and starts a process of prolonged infections, often involving multiple organisms and biofilm formation.

21.3.3 Mastoiditis

Acute mastoiditis is the inflammation of the mastoid process of the temporal bone that follows as a suppurative, relatively rare, complication of acute otitis media. *Streptococcus pneumoniae* is regarded as one of the major bacterial pathogens causing mastoiditis.

21.4 Pneumococcal Vaccines

Two types of pneumococcal vaccines are currently available: the nonconjugated, polysaccharide vaccine (PPV23) and the 10 and 13 valent pneumococcal conjugated vaccines (PCV10 and PCV13). The conjugated vaccines (PCVs) offer several advantages over PPV23. First, PCVs are licensed for use in infants 6 weeks of age and older, whereas PPV23 is only licensed for children >2 year old. This is because PCVs already offer protection from early infancy. Second,

PCVs elicit T-dependent immune response and thus also memory, which are not elicited by PPV23.

PPV23 was introduced in 1983 and is available in Europe for immunization against pneumococcal diseases caused by the 23 serotypes contained in the vaccine (1, 2, 3, 4, 5, 6B, 7F, 8, 9N, 9V, 10A, 11A, 12F, 14, 15B, 17F, 18C, 19F, 19A, 20, 22F, 23F, and 33F) in adults and children aged ≥ 2 years.

In 2000, PCV7 (serotypes 4, 6B, 9V, 14, 18C, 19F, 23F conjugate to CRM₁₉₇) was first licensed, and has increasingly been used globally. Currently, two more extended-serotype PCVs are licensed (whereas PCV7 is no longer manufactured): PCV10 (PCV7 serotypes + serotypes 1, 5, and 7F) and PCV13 (PCV10 serotypes + serotypes 3, 6A and 19A). In PCV10, eight serotypes are conjugated to NTHi protein-D, serotype 19F to diphtheria toxoid, and serotype 18C to tetanus toxoid. In PCV13, all serotypes are conjugated to CRM₁₉₇.

Description of PNEUMOVAX®23™ According to SPC

► http://www.merck.com/product/usa/pi_circulars/p/pneumovax_23/pneumovax_pi.pdf

PNEUMOVAX 23 is approved for use in persons 50 years of age or older and persons aged ≥ 2 years who are at an increased risk for pneumococcal disease. PPV23 is not approved for use in children younger than 2 years of age because children in this age group do not develop an effective immune response to capsular types contained in the polysaccharide vaccine.

Description of Prevnar 13® According to SPC

► http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Product_Information/human/001104/WC500057247.pdf

Therapeutic indications

Active immunization for the prevention of invasive disease, pneumonia, and acute otitis media caused by *Streptococcus pneumoniae* in infants, children, and adolescents from 6 weeks to 17 years of age.

Active immunization for the prevention of invasive disease and pneumonia caused by *Streptococcus pneumoniae* in adults ≥ 18 years of age and the elderly.

Three-dose primary series

The recommended immunization series consists of four doses, each of 0.5 ml. The primary infant series consists of three doses, with the first dose usually given at 2 months of age and with an interval of at least 1 month between doses. The first dose may be given as early as 6 weeks of age. The fourth (booster) dose is recommended between 11 and 15 months of age.

Two-dose primary series

Alternatively, when Prevnar 13 is given as part of a routine infant immunization program, a series consisting of three doses, each of 0.5 ml, may be given. The first dose may be administered from the age of 2 months, with a second dose 2 months later. The third (booster) dose is recommended between 11 and 15 months of age.

Description of PHiD-CV10 (Synflorix®)

According to SPC

► http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Product_Information/human/000973/WC500054346.pdf

Therapeutic indications

Active immunization against invasive disease, pneumonia, and acute otitis media caused by *Streptococcus pneumoniae* in infants and children from 6 weeks up to 5 years of age.

Three-dose primary series

The recommended immunization series to ensure optimal protection consists of four doses, each of 0.5 ml. The primary infant series consists of three doses with the first dose usually given at 2 months of age and with an interval of at least 1 month between doses. The first dose may be given as early as 6 weeks of age. A booster (fourth) dose is recommended at least 6 months after the last priming dose and preferably between 12 and 15 months of age.

Two-dose primary series

Alternatively, when Synflorix is given as part of a routine infant immunization program, a series consisting of three doses, each of 0.5 ml may be given. The first dose may be administered from the age of 2 months, with a second dose 2 months later. A booster (third) dose is recommended at least 6 months after the last primary dose.

21.5 Pneumococcal Polysaccharide Vaccine PNEUMOVAX®23™

Pneumococcal polysaccharide vaccine (PPV23) is not included in pediatric National Immunization Programs (NIPs), as it is not approved for use in children younger than 2 years of age. It is recommended for usage in high-risk individuals ≥ 2 years of age, including (but not limited to): asplenia (anatomical, functional); chronic renal insufficiency; cochlear implant; complement and properdin deficiency; CSF leak; hematopoietic organ disorder; HIV; hypogammaglobulinemia; immunodeficiency (congenital, acquired); malignancy; nephrotic syndrome; sickle-cell anemia; and transplantation (organ, subsequent to stem cell transplantation). However, there is a considerable variability in vaccine recommendations among different European countries.

A second dose of PPSV23 is recommended 5 years after the first dose of PPSV23 for children who have anatomical or functional asplenia, including sickle-cell diseases, HIV infection, or other immunocompromising conditions.

Whenever PPV23 is recommended in children, it should be administered at least 8 weeks after the last PCV dose. If PPV23 was administered before PCV, administration of the latter should be delayed for at least a year.

Pneumococcal polysaccharide vaccine 23 is also recommended for adults at a high risk and all adults aged 65 years and older. The vaccine has been shown to be moderately effective in preventing invasive pneumococcal disease among the general elderly population. However, its effectiveness against IPD in the high-risk elderly may be lower. The vaccine has not been clearly demonstrated to prevent pneumonia in any age group, and it does not prevent nasopharyngeal carriage at any age.

21.6 Introduction of Pneumococcal Conjugate Vaccines and Vaccine Uptake

Pneumococcal conjugate vaccine7 was added to the US infant immunization schedule in 2000. In Europe, however, PCV7 introduction varied considerably among countries, with Spain, Ireland,

and Luxembourg introducing PCV7, at least partially, in the years 2001 through 2003; Austria, Belgium, Italy, and Slovenia in 2004 and 2005; Greece, Slovakia, France, Netherlands, Germany, Norway, UK, Iceland, Malta, and Denmark in 2006 and 2007; and Poland, Cyprus, Hungary, Finland, Sweden, Czech Republic, Latvia, Bulgaria, Portugal, and Israel only during 2008 through 2010. Furthermore, vaccine uptake and recommendations regarding immunization schedule also varied considerably among countries (■ Table 21.2).

In contrast to the late introduction of PCV7 in Europe, PCV13 and PCV10 were introduced into European countries (mostly in Western Europe), shortly after their licensure (2010 and 2011 respectively). Several countries replaced PCV7 with PCV13, including Belgium (but there was a return to PCV10 in 2015), Denmark, France, Ireland, Norway, Spain (Madrid), Switzerland, UK, Italy, and Israel.

In the Netherlands and Austria, PCV10 replaced PCV7 in 2011 and 2012 respectively, whereas in Finland and Iceland, PCV10 was introduced as the first PCV in the National Vaccination Program in September 2010 and April 2011 respectively.

Other countries, including Spain (Catalonia, Navarra), Portugal, Slovakia, the Czech Republic, and Sweden used both PCV13 and PCV10. In Germany, PCV7 was introduced to the NIP in July 2006, and was replaced by PCV10 in April 2009 and PCV13 December 2009, with PCV13 predominantly used (>90% market share). The number of European countries introducing PCV10 and PCV13 to their NIPs has been constantly increasing (■ Fig. 21.4).

21.7 Different Vaccine Schedules

Most schedules in European countries include two primary PCV doses in the 1st year of life, with a booster dose in the 2nd year of life (2 + 1 schedule). However, several European countries have a 3 + 1 schedule, with the first three doses given in the 1st year of life and a booster dose at the age of 1 year or older (■ Fig. 21.5). Some differences also exist in the time intervals between doses and the timing of the booster.

Table 21.2 Characteristics of national pneumococcal vaccination programs in EU countries in 2010

Country	Date PCV7 introduction	Scope of PCV7 vaccination program	Immunization schedule (dose)	Vaccine coverage ^e
Austria	July 2004	Universal	3 + 1	–
Belgium	January 2005	Universal	2 + 1	97
Bulgaria	April 2010	Universal	3 + 1/2 + 1	–
Cyprus	August 2008	Universal	3 + 1	–
Czech Republic	January 2010	Risk-based	3 + 1	86.3
Denmark	October 2007	Universal	2 + 1	85
Estonia	–	–	Not decided	–
Finland	January 2009	Risk-based	2 + 1	–
France	June 2006	Universal	2 + 1	81
Germany	July 2006	Universal	3 + 1	52.9
Greece	January 2006	Universal	3 + 1	–
Hungary	October 2008	Universal	2 + 1	81.1
Iceland	December 2006	Risk-based	2 + 1	–
Ireland	October 2002	Universal	2 + 1	89
Israel ^a	July 2009	Universal	2 + 1	90
Italy	May 2005	Universal/risk-based	2 + 1	55
Latvia	January 2010	Universal	3 + 1	51
Lithuania	–	–	3 + 1	–
Luxembourg	February 2003	Universal	3 + 1	86
Malta	January 2007	Risk-based	3 + 1	–
Netherlands	June 2006	Universal	3 + 1	94
Norway	July 2006	Universal	2 + 1	90
Poland	May 2008	Risk-based	3 + 1/2 + 1	1.7
Portugal	June 2010	Risk-based	2 + 1	52
Romania ^b			3 + 1	
Slovakia ^c	January 2006	Risk-based	2 + 1	99.2
Slovenia	September 2005	Risk-based	3 + 1	–
Spain ^d	June 2001	Risk-based	3 + 1	–
Sweden	January 2009	Universal	2 + 1	–
United Kingdom	September 2006	Universal	2 + 1	90

Navarro Torné et al. (2014)

^aData not included in the original table

^bPCV7 was registered in September 2007 for voluntary use on a private basis

^cUniversal as of April 2008

^dUniversal introduction in the autonomous region of Madrid in November 2006

^eSources: VENICE II and WHO estimates of PCV7 coverage



Fig. 21.4 European pneumococcal conjugate vaccine (PCV) National Immunization Programs. Belgium mixed situation: PCV10 in Flanders region (since July 2015) and Prevenar13 in the Brussels/Wallonia region (until February 2016). Greece: in the National Immunization Program (NIP), the only pneumococcal vaccine included for children is PCV13. However, it is merely a recommendation; thus, a physician could also prescribe PCV10, but in that case, it should not be fully reimbursed. Spain: PCV13 NIP in 11 out of 17 regions. Six regions continue with

PCV13, 95% through the private market; * Spain: 14 out of 17 regions have started PCV13 national immunization program. ** Both PCVs are available/reimbursed in the NIP or the NIP consists of different PCVs by region PCV13 is a pneumococcal polysaccharide conjugate vaccine (13-valent, adsorbed). Please refer to the Summary of Product Characteristics and official recommendations. 1. Gavi Alliance Progress Report 2013. ► <http://www.gavialliance.org/results/gavi-progress-reports/> 2. Data on file, Pfizer

21.8 General Comments on PCV Impact and Impact Studies

When considering the effects of a vaccine, one must understand the difference among efficacy, effectiveness, and impact. Efficacy is measuring the potential of a vaccine to protect against a specific end-point, compared with placebo or a control vaccine, in randomized control trials.

Effectiveness measures a similar effect, but in real life, and is therefore affected by other factors beyond those of efficacy (i.e., refrigerator conditions, vaccination errors). Hence, effectiveness is usually assessed retrospectively and is measured by using the case-control methodology.

In contrast to efficacy and effectiveness, when measuring impact, the overall reduction (or increase) and the dynamics of rates following vaccine implementation are measured. When assessing impact, it may be more difficult to appreciate

the true vaccine effect, differentiating it from potential other factors. However, these are the only studies that show the actual vaccine effect following vaccine introduction.

Several components influence the impact observed after PCV introduction. The impact of PCV on the pneumococcal carriage of vaccine serotypes (VTs) is of utmost importance. This effect is the key point in the prevention of both pneumococcal diseases among the vaccine recipients on the one hand, and the prevention of spread and early exposure to vaccine-type strains in unvaccinated individuals on the other hand, resulting in indirect (herd) protection. Other important components determining PCV impact include vaccine uptake (affecting both direct and indirect impact), serotype coverage of the vaccine (PCV7, -10, -13), time elapsed since vaccine introduction (affecting the indirect impact), vaccine efficacy against different disease end points

	Months												
	2	3	4	5	6	10	11	12	13	14	15	18	23
Austria		PCV		PCV				PCV ¹					
Bulgium	PCV		PCV					PCV					
Bulgaria	PCV	PCV	PCV					PCV					
Croatia													
Cyprus	PCV		PCV					PCV ⁶					
Czech Republic	PCV10 ⁸	PCV10 ⁸	PCV10 ⁸					PCV10 ⁸					
Denmark		PCV13		PCV13				PCV13					
Estonia													
Finland		PCV10		PCV10				PCV10					
France	PCV		PCV				PCV						
Germany	PCV		PCV					PCV				PCV ¹³	
Greece	PCV		PCV		PCV		PCV						
Hungary	PCV13 ¹⁸		PCV13 ¹⁸					PCV13 ¹⁸					
Iceland		PCV10		PCV10				PCV10					
Ireland	PCV				PCV			PCV					
Italy		PCV		PCV				PCV					
Latvia	PCV		PCV					PCV					
Liechtenstein	PCV13 ²¹		PCV13 ²¹					PCV13 ²¹					
Lithuania	PCV		PCV					PCV					
Luxembourg	PCV		PCV					PCV					
Malta													
Netherlands	PCV		PCV				PCV						
Norway		PCV13		PCV13				PCV13					
Poland			PCV ²⁴										
Portugal	PCV13		PCV13					PCV13					
Romania	PCV ²⁴		PCV ²⁴									PCV ²⁴	
Slovakia	PCV		PCV				PCV						
Slovenia		PCV	PCV					PCV					
Spain	PCV ²⁷		PCV ²⁷					PCV ²⁷					
Sweden		PCV		PCV				PCV					
United Kingdom	PCV13		PCV13					PCV13					

Fig. 21.5 Recommended immunizations for pneumococcal disease in European children aged <2 years. ► <http://vaccine-schedule.ecdc.europa.eu/Pages/Scheduler.aspx>

(i.e., IPD vs mucosal), and local epidemiological characteristics, including serotype distribution before PCV introduction and immunodeficient population (i.e., HIV prevalence).

Impact studies are also important in advancing our understanding of the role of vaccine-type pneumococcal serotype in the etiology of mucosal syndromes, such as pneumonia and OM. In the case of OM, the introduction of PCV7 resulted in a moderate effect of up to ~25% reduction in OM rates. Further studies conducted following PCV13 introduction observed substantial (up to ~70%) reduction of overall OM, with the near elimination of vaccine-type OM, mainly complex OM. These findings hint at a new paradigm, suggesting that early OM episodes might mainly be caused by PCV13 serotypes, and that preventing these episodes might result in preventing acute OM sequelae, including complex OM. Similarly, the introduction of PCV13 resulted in a substantial reduction of pneumonia episodes, to the magnitude of 50%, suggesting the major role of vaccine serotypes in the etiology of pneumonia. Thus, impact figures, depending on multiple factors and endpoints acting in concert, are much greater than those calculated for efficacy. The observations with regard to PCV impact on disease, elucidating the role of vaccine-type strains in disease etiology are termed “vaccine probe” studies.

21.9 Implementation of PCV and Post-PCV Impact

True appreciation of the impact of a vaccine depends on a reliable, long-standing, pre-vaccine surveillance system. In this regard, there is a real gap of knowledge, as for the pre-PCV pneumococcal disease rates, especially beyond IPD. Although IPD rates are relatively easy to estimate, this is not the case with pneumonia and OM, as disease rates are highly variable because of differences in case definitions and the lack of national surveillance systems in most countries.

In general, all PCVs lead to a rapid and profound reduction in pneumococcal disease rates in vaccinated infants and children if widely introduced, and most studies also showed an indirect effect (herd protection) in older individuals who were not vaccinated.

The first seven-valent pneumococcal conjugated vaccine (PCV7) was developed based on

data demonstrating that within the USA and several other developed countries, the PCV7 serotypes were responsible for >80% of IPD in young children. Subsequent studies showed the important global role of additional serotypes, especially 1, 3, 5, 7F, 6A, and 19A. For one vaccine (PCV10, also termed PHiD-CV), efforts were made to add serotypes 1, 3, 5, and 7F to form an 11-valent vaccine, but following the failure to demonstrate protection against serotype 3 in an otitis media efficacy study, the final product has added only three additional serotypes (1, 5, and 7F) to the initial seven.

For the formulation of both PCV7 and PCV10, it was assumed that serotypes 6B and 19F present in these vaccines could protect against the prevalent and important (including often antibiotic-resistant) serotypes 6A and 19A respectively. For serotype 6A, cross-protection by serotype 6B was seen, at least for IPD, in fully vaccinated children. For 19A, no cross-protection was shown using PCV7. Limited cross-protection was observed for 19A in fully vaccinated infants with PCV10. However, probably because of the short duration of protection against IPD and the absence of efficacy against carriage, the overall picture post-implementation in the community regarding serotype 19A resembled that of PCV7, with an overall increase in disease in all ages in most countries using PCV10, which have been conducting appropriate epidemiological surveys. The prolonged use of PCV7 in some countries resulted in reduced disease caused by serotype 6A in all ages. Similarly, in countries using PCV10, a reduction in serotype 6A IPD in children aged <5 is usually observed. However, beyond this age group, the effect is dependent upon indirect protection derived from the impact on carriage, and thus has been more variable. In most countries using PCV10, rates of serotype 6A IPD in adults either did not decrease or even increase, meaning that the impact of PCV10 on 6A carriage in vaccine recipients was often insufficient.

Pneumococcal conjugate vaccine 13 was licensed in 2010. Implementation of this vaccine in several countries with well-conducted epidemiological studies and high vaccination coverage has shown a rapid reduction of the additional serotypes in all ages and for all endpoints. The one exception is serotype 3, where contradictory data were generated regarding its impact after the first 5–6 years post-PCV13 implementation. The final

verdict concerning its impact on serotype 3 disease has not yet been reached.

In contrast to PCV7 and PCV10, the introduction of PCV13 resulted in a rapid and profound decrease in all endpoints of disease and carriage by serotypes 6A and 19A in all ages. Furthermore, the presence of serotype 6A antigen in PCV13 resulted in its impact on disease from the carriage of cross-reactive serotype 6C, one of the most important replacing serotypes after the implementation of PCV7 or PCV10.

21.10 PCV Schedules

Post-implementation, the impact of the two different schedules were not directly compared, except in a double-blind, randomized controlled Finnish trial designed to document the effectiveness of the PCV10 vaccine against invasive pneumococcal disease, where vaccine effectiveness estimates of both 3 + 1 and 2 + 1 schedules were similar. However, the differences between the two regimens could not be fully assessed for all outcomes because of the paucity of outcome cases. Furthermore, whether data for comparison by one vaccine (PCV10) can directly be extrapolated to another vaccine (PCV13) is not clear.

Some data exist, though, to compare the impact of the various regimens on carriage. In VT carriage, antibody concentrations post-PCV administration may be related to efficacy. Thus, efficacy against carriage after two infant doses may be reduced compared with after three doses. PCV10 studies in Finland suggested that for PCV10, even after a booster, the 2 + 1 regimen is inferior to the 3 + 1 regimen.

In any case, even in countries with a 2 + 1 regimen, it is recommended that immunodeficient individuals (including those born prematurely) receive an additional PCV dose (i.e., a 3 + 1 schedule).

21.11 Impact of PCV on IPD in Young Children

Although IPD constitutes only a small proportion of all pneumococcal diseases, it is extremely important, as some of the IPD manifestations (i.e., sepsis, meningitis) are the most severe pneumococcal disease manifestations and result in the highest mortality rates.

The introduction of PCV7, PCV10, and PCV13 was associated with a rapid and profound reaction in IPD caused by the respective vaccine serotypes in children <5 years old. In countries introducing first PCV7, its replacement by PCV10 or PCV13 further reduced IPD caused by the additional serotypes, showing a two-step reduction pattern (■ Fig. 21.6). As discussed above, for the cross-reacting serotype 6A, all three PCVs showed a similar impact in young children. However, no apparent impact on serotype 19A was observed in countries using PCV10 and in several countries (i.e., Finland, Chile, and New Zealand), IPD caused by serotype 19A even increased in young children. IPD caused by some non-VTs increased in young children after the introduction of PCV, the most commonly observed serotypes in countries using PCV10 or PCV13 being 8, 12F, 15A, 15B/C, 22F, 24F, and 33F. In addition, following the introduction of PCV10, disease caused by serotype 3 also frequently increased.

As most of the non-PCV serotypes are less invasive than most PCV serotypes, it is not surprising that post-PCV implementation, the proportions of compromised patients increased within cases of IPD.

In general, the overall impact of PCV7/PCV13 in children was less prominent in meningitis than in non-meningitis IPD, probably attributable to the younger age of children with meningitis and some underlying conditions resulting in differences in causative serotypes between the two groups, as the decline of VT meningitis and nonmeningitis IPD was similar.

21.12 Impact of PCV on Pneumonia

Estimating the impact of PCV on pneumonia rates is difficult, for two main reasons: (1) the definition of pneumonia is not clear, as will be discussed later; and (2) the microbiological diagnosis of pneumonia is complex and unclear. As expected, the highest reductions were observed in studies evaluating bacteremic pneumococcal pneumonia (accounts for ~25–35% of all IPD cases), where disease rates declined in a similar manner to those of other nonpneumonia IPD.

Microbiological studies in cases of empyema or pleural effusion (pleuropneumonia) suggest that

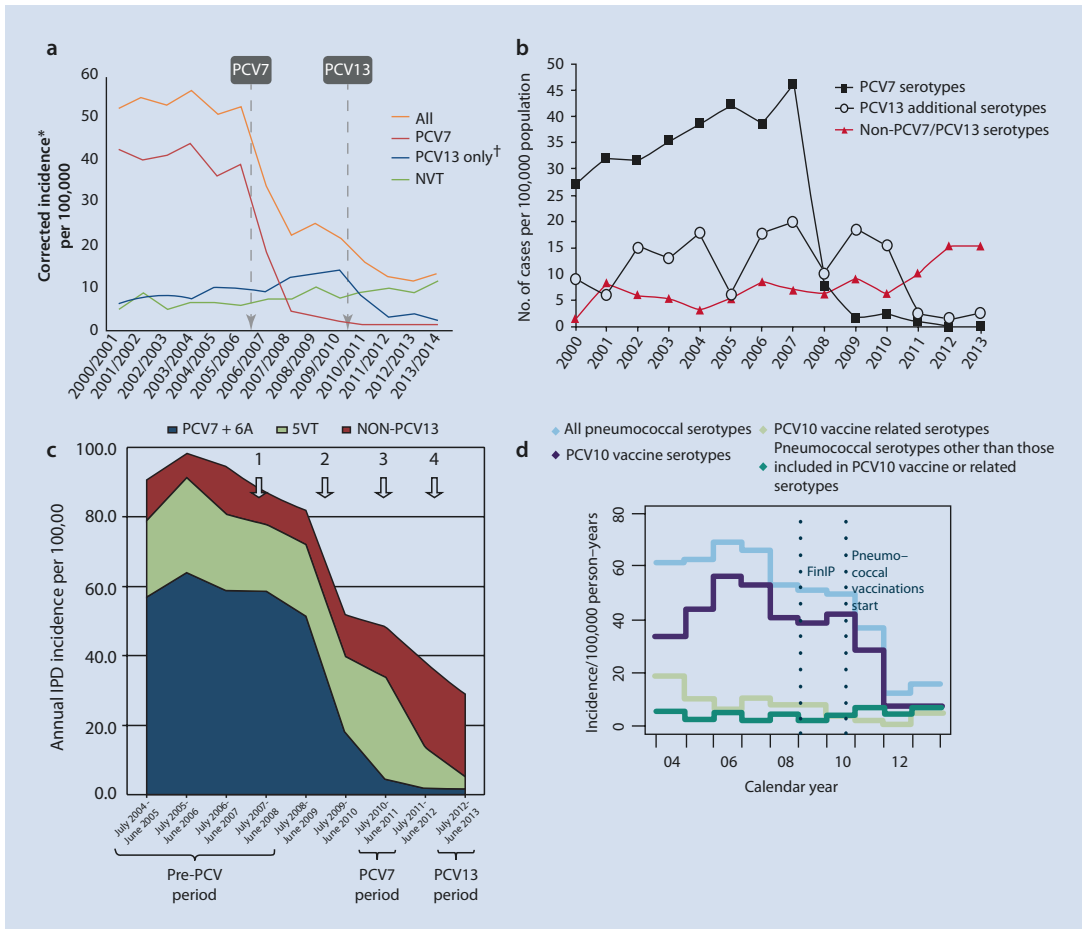


Fig. 21.6 Incidence of IPD in children <2 years before and after PCV introduction in England and Wales, Denmark, Israel, and Finland. **a** England and Wales, PCV13; **b** Denmark, PCV13; **c** Israel, PCV13; **d** Finland, PCV10

the most common serotypes, accounting for >50% of cases, might be serotype 14 (a PCV7 serotype) and the additional PCV10/PCV13 serotypes 1, 3, 5, 7F, and 19A. Thus, it is not surprising that post PCV7 implementation, pleural pneumonia did not decrease and even increased. However, post-PCV10 and PCV13 implementation, the incidence declined. As pleuropneumonia and bacteremic pneumonia constitute only a minority of pneumonia cases, in most other pneumonia cases, only partial information on the causative serotype exists.

When alveolar pneumonia (also termed lobar or segmental pneumonia) was examined, post-implementation reduction of up to 50% or more was seen, especially after PCV10 and PCV13 implementation (emphasizing the importance of serotype 1, 5, 7F, and 19A in pneumonia). This type of pneumonia is usually considered to be of bacterial origin, mainly pneumococcal.

These observations are consistent with the finding that the highest efficacy against pneumonia in randomized clinical studies with PCV7 and PCV10 was observed for alveolar pneumonia. However, much more common, but less obviously of pneumococcal origin, were all-cause pneumonia cases (a term that includes all end-points of pneumonia, such as non-alveolar, chest radiology-negative pneumonia, or even clinical-only pneumonia). For these more inclusive but less-specifically defined cases, overall reduction, as expected, was more variable, ranging from <20% to ~70%. In any case, the repeated findings of reduced rates of all-cause pneumonia emphasize not only the pneumococcal role in pneumonia, but probably the important role of vaccine serotypes as causative agents in all-cause pneumonia. These “vaccine probe” studies, in fact, showed clearly that the impact on pneumonia was much

greater than initially expected, with the number of prevented cases higher by orders of magnitude than the IPD figures.

21.13 PCV Impact on Otitis Media

Pre-licensure efficacy studies showed an efficacy of 57–67% against PCV7 serotype OM. PCV7 also showed a similar reduction for the cross-reacting serotype 6A. In contrast, rates of OM caused by non-PCV7 serotype did not decrease in these efficacy studies and in some cases even increased, along with nonpneumococcal cases. This resulted in an only modest efficacy against all-cause OM that did not reach statistical significance in most studies. However, measuring the overall OM incidence does not appropriately reflect the real OM burden, which is better reflected by measuring the impact of OM sequelae, such as recurrent, nonresponsive, and chronic OM (collectively termed complex OM). Although *S. pneumoniae* is not the only major pathogen in OM, it is found mostly in early OM, and becomes less prominent later, when the frequency of complex otitis increases. In complex OM, a high frequency of NTHi is observed, sometimes with *Moxarella catarrhalis* and other organisms, and frequent findings of biofilm formation. Thus, it is plausible that preventing early, acute pneumococcal OM might reduce the burden imposed by its sequelae. Indeed, pre-PCV7 licensure randomized controlled studies and post-introduction impact studies showed a significant reduction in OM-associated burden due to complex OM, with a reduction of recurrent otitis or ventilation tube insertions, despite the paucity of the presence of VT pneumococci at these end-points. Despite the increasing evidence for such an impact, the lack of post-PCV microbiological data raised some skepticism regarding the actual extent of OM burden reduction by PCVs. This was mainly because measuring pathogen-specific impact is particularly problematic, as it depends on obtaining middle-ear fluid cultures, usually performed selectively.

In Israel, the impact of PCV13 on OM cases necessitating middle-ear fluid cultures (mainly complex OM cases) was documented in a population-based, active surveillance system, in children <3 years old. Following the sequential introduction of PCV7/PCV13, a decline of 95% in the incidence

for the PCV7 + 6A serotypes was observed with a decline of 89% in the incidence of the additional PCV13 serotypes (1, 3, 5, 7F, and 19A) disease. Overall, complex OM-enriched pneumococcal OM incidence declined by 78%. Furthermore, non-pneumococcal OM episodes were also reduced, as expected. In this regard, it is important to remember that it has been long recognized that early OM cases in young infants are most likely to be associated with complex OM cases in large studies.

The prevention of early OM post-PCV implementation is an excellent example of dual protection provided by PCV. On the one hand, it reduces VT carriage (see in later paragraphs) to an extent where very young infants rarely encounter any VT in the community, and on the other hand, once the infants encounter one of the VTs, the vaccine provides additional direct protection against disease. Thus, the prevention of early encounters with vaccine-type *S. pneumoniae* results in a marked reduction of early acute OM episodes, and therefore, the subsequent sequelae.

With regard to PCV10, one hoped to see a direct effect of the vaccine on NTHi OM, as most serotypes in PCV10 (or its precursor PCV11) are conjugated to NTHi-derived protein-D. However, even though protein-D was immunogenic, PCV10 did not show direct protection against any NTHi outcome. One study suggested an exception. The POET study was conducted in the Czech Republic and Slovakia using PCV11 (the precursor of PCV10) against OM. This placebo-controlled study showed a significant reduction against NTHi-OM. However, in this study, the trigger to enroll children was for children visiting an otolaryngologist office, thus enriching the population with complex cases. Thus, the reduction of NTHi OM by protein-D conjugated PCV, documented only in the POET study, could be explained again by the prevention of early pneumococcal OM with a secondary prevention of NTHi otitis as part of the sequelae.

Another study on the efficacy of PCV10 against OM (the COMPAS study), conducted in Latin America, failed to show any effect of PCV10 on NTHi OM.

Post-PCV10 impact data on OM are scarce, but recent data from Iceland and Brazil suggest trends toward reductions of OM and recurrent OM. However, whether the extent of the impact will be similar for the PCV10 and PCV13 remains to be clarified.

21.14 PCV Impact on Carriage and the Resulting Indirect (Herd) Protection

The widespread introduction of PCV7 resulted in a rapid and substantial indirect (herd) protection. Herd protection is achieved through a reduction in the carriage of vaccine serotype pneumococci in vaccinated children, and thus a reduction of their spread in the community. On the other hand, the near elimination of the NP carriage of VT following PCV introduction led to replacement of the carriage by non-VTs, often less invasive. Because non-VT strains were less invasiveness overall, partial or no replacement disease was observed in most studies. Therefore, the disease replacement phenomenon was limited and was mainly observed in compromised patients. As elderly people can often be considered immunocompromised, it is not surprising that this population was most affected by the increase in non-VT strains in the community, following PCV introduction. However, a longer follow-up is needed to ascertain the continuous net positive effect of PCVs regarding replacement disease.

As all PCVs reduce the nasopharyngeal carriage of VT pneumococci, widespread vaccination resulted in reduced circulation of these serotypes in the community, hence the reduced encounters of both vaccinated and unvaccinated individuals with vaccine serotypes. As discussed above, the reduction in nasopharyngeal carriage is the most important factor determining impact, along with vaccination coverage. The reduced carriage protects both vaccinated and unvaccinated individuals. As an example, if PCVs have ~60% efficacy against VT OM, and if at the same time there is a 60% reduction in VT carriage, the vaccinated infants encounter only 40% of what he or she would have encountered in the pre-PCV era. In this given example, the dual protection results in ~85% protection against VT pneumococcal OM.

Three main groups have herd protection:

1. Those who are too young to be vaccinated (i.e., infants aged <4 months who usually by this age have only ≤ 1 doses); this early protection against VT disease may be the most important means of preventing complex OM, as very early OM (before reaching the age of full vaccination) is the most important risk factor for complex OM (beyond genetics).

2. The vaccinated individual (as specified above), as efficacy never reaches 100%.
3. Individuals too old to be vaccinated (practically all individuals >5 years of age).

We do not know how long the immunity afforded by PCVs lasts, especially in terms of mucosal immunity, but the indirect protection also ensures that those immunized in the past can be protected, even if they had already lost the vaccine-acquired immunity.

As discussed previously in this chapter, not all PCVs are equally efficacious against carriage in general, and some possess unique serotypes that others do not have. However, in general, in all countries where PCVs were introduced, an impressive reduction of IPD caused by vaccine-serotype pneumococci was recorded at all ages, because of the combined direct and herd protection. However, in compromised patients (including the elderly), replacement diseases caused by non-VTs is common. Current epidemiology data strongly suggest that PCV13 might provide a more rapid and profound herd protection, especially because of the reduction of the carriage of serotypes 6A, 19A, and the cross-reacting serotype 6C, compared with PCV10. A longer period of follow-up is needed to confirm these findings.

21.15 PCV Impact on Antibiotic Resistance

In the field of pneumococci, the general term for antibiotic nonsusceptibility is often preferred over the term “resistance,” as at times, especially for β -lactams antibiotics, the minimal inhibitory concentration (MIC) increases, meaning that the organism is less susceptible to the drug, but no full resistance has yet been reached. It is well-established that antibiotic nonsusceptibility among pneumococci (like most bacteria) can rarely occur by mutation, but rather widespread antibiotic use is the main contributor to the promotion of carriage and the circulation of antibiotic nonsusceptible *S. pneumoniae* (ANSP). The main antibiotics responsible for ANSP promotion and spread are the long-acting macrolides (in particular, azithromycin) and oral cephalosporins, whereas the least powerful promoter is high-dose amoxicillin (with or without clavulanate). However, any antibiotic drug can promote ANSP, and thus indiscriminate use of

antibiotics, which has often been practiced since the 1980s in many societies, is responsible for increasing ANSP prevalence. Since ANSP resides in the nasopharynx, antibiotic drugs given for any reason, will select these strains over susceptible ones, resulting in their promotion and spread in the community.

Among pneumococci, the most successful colonizers in young children are serotypes 6A, 6B, 9V, 14, 19A, 19F and 23F. These serotypes are also the main strains that express multidrug resistance and high-level resistance. They are also responsible for most disease (both IPD and mucosal diseases) in children and adults. Therefore, it is not surprising that the most important ANSP serotypes are included in the vaccines. Of these, serotypes 6B, 9V, 14, 19F, and 23F are included in PCV7, which also confers some cross-protection against serotype 6A (although as reviewed above, not complete in the case of carriage). PCV10, which adds the important serotypes 1, 5, and 7F beyond PCV7, does not significantly improve the impact on ANSP prevalence, as these three additional serotypes are rarely carried and rarely non-susceptible. However, the addition of serotypes 6A and 19A in PCV13 made an important contribution, as these two serotypes are often multidrug-resistant with a high level of resistance. This is the basis for the potential reduction of ANSP disease and circulation by PCVs.

All PCVs were shown to reduce antibiotic nonsusceptibility by three main mechanisms. First, they reduce VT disease (efficacy against pneumococcal diseases), including disease caused by the VT ANSP; second, they reduce the carriage and thus the spread of ANSP; third, the reduction of disease incidence results in a reduction of antibiotic use and thus a reduction in the antibiotic pressure on strains carried in the nasopharynx or other sites of the flora microbiota. These positive forces by the vaccine, are necessarily accompanied by a marked (although many times not complete) replacement in the carriage by non-VT. Nonsusceptibility, especially high-level and multidrug resistance, was remarkably less common among non-VTs before the introduction of PCVs. However, post-vaccination, by occupying the nasopharynx more frequently and for longer periods because of replacement, the non-VTs are

now under increased antibiotic pressure. Indeed, ANSP and even multidrug resistance among non-VTs are increasing at an alarming rate. However, because in general, the overall invasiveness among non-VTs is lower compared with VTs (with a few exceptions, i.e., serotypes 12F, 24F, 8, and 22F), the net effect is usually reduced disease caused by ANSP. It is not surprising that in adults, ANSP disease is influenced by childhood widespread PCV vaccination, through the major change in nasopharyngeal carriage. Thus, in many respects, ANSP IPD in adults follows that of childhood.

As discussed before, several serotypes (i.e., serotypes 8, 10A, 11A, 12F, 15A, 15B/C, 22F, 33F, and 35B) are generally the most important replacing serotypes, meaning that most of these are successful colonizers in the absence of competition with VTs. Therefore, it is only natural that increasing resistance and multidrug resistance are found in some of these serotypes. This scheme is especially worrisome in compromised patients, in whom replacement disease is most frequent.

21.16 Future Vaccines

All currently licensed pneumococcal vaccines have limitations due to their capsular serotype specificity.

Potential approaches to addressing current PCV limitations include higher valency PCVs. Attempts to extend the number of serotypes in PCV beyond the 13 serotypes in PCV13 are being made, but expanding the serotype spectrum in any PCV (higher valency PCV) is technically difficult. One alternative possibility is to have additional PCVs with some of the common replacement serotypes to be administered sequentially after PCV10/PCV13, or to adults only. Another alternative is to use pure protein of *S. pneumoniae* or polypeptide derivatives to develop protein-based vaccines. Protein vaccine candidates are ideally highly conserved by all pneumococcal strains, and exhibit high immunogenicity. However, so far, all attempts to develop such vaccines were not successful. Thus, it seems that in the next 5–10 years, no protein vaccine will emerge and be licensed for general use. A further possible approach is the use of whole killed cell vaccines, currently in human trials.

Further Reading

- Atkinson W, et al., editors. Epidemiology and prevention of vaccine-preventable diseases. CDC Pink Book. 11th ed. Washington DC: Public Health Foundation; 2009. p. 217–30.
- Ben-Shimol S, Givon-Lavi N, Leibovitz E, Raiz S, Greenberg D, Dagan R. Near-elimination of otitis media caused by 13-valent pneumococcal conjugate vaccine (PCV) serotypes in southern Israel shortly after sequential introduction of 7-valent/13-valent PCV. *Clin Infect Dis*. 2014;59(12):1724–32.
- Dagan R. Impact of pneumococcal conjugate vaccine on infections caused by antibiotic-resistant *Streptococcus pneumoniae*. *Clin Microbiol Infect*. 2009;15(Suppl 3):16–20.
- Isaacman DJ, McIntosh ED, Reinert RR. Burden of invasive pneumococcal disease and serotype distribution among *Streptococcus pneumoniae* isolates in young children in Europe: impact of the 7-valent pneumococcal conjugate vaccine and considerations for future conjugate vaccines. *Int J Infect Dis*. 2010;14(3):e197–209.
- Navarro Torné A, et al. European enhanced surveillance of invasive pneumococcal disease in 2010: data from 26 European countries in the post-heptavalent conjugate vaccine era. *Vaccine*. 2014;32(29):3644–50.
- Nuorti JP, Whitney CG, Centers for Disease Control and Prevention (CDC). Prevention of pneumococcal disease among infants and children – use of 13-valent pneumococcal conjugate vaccine and 23-valent pneumococcal polysaccharide vaccine – recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep*. 2010;59(RR-11):1–18. ► http://ecdc.europa.eu/en/publications/Publications/0701_TER_Use_of_pneumococcal_polysaccharide_vaccine.pdf
- O'Brien KL, Wolfson LJ, Watt JP, Henkle E, Deloria-Knoll M, McCall N, et al. Burden of disease caused by *Streptococcus pneumoniae* in children younger than 5 years: global estimates. *Lancet*. 2009;374:893–902.
- Palmu AA, Jokinen J, Borys D, Nieminen H, Ruokokoski E, Siira L, et al. Effectiveness of the ten-valent pneumococcal Haemophilus influenzae protein D conjugate vaccine (PHiD-CV10) against invasive pneumococcal disease: a cluster randomised trial. *Lancet*. 2013; 381(9862):214–22.

Meningococcal Vaccines

Andrew J. Pollard, Matthew D. Snape, and Manish Sadarangani

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22.1 Introduction

Meningococcal disease was first described in Europe as a characteristic outbreak in Geneva in 1805. *Neisseria meningitidis* (the meningococcus) is a Gram-negative diplococcus, divided into capsular groups determined by the polysaccharide capsule. Six of the twelve groups (A, B, C, W, X and Y) are responsible for almost all invasive diseases worldwide. While asymptomatic nasopharyngeal infection (colonisation or carriage) occurs in approximately 10% of the population, bacteria occasionally enter the bloodstream to cause devastating invasive diseases such as meningitis and septicaemia. In Europe it is typically a rare endemic disease, but hyperendemic and epidemic disease patterns also occur. Disease onset may be rapid and has a high case fatality rate, especially in those with septic shock. Many survivors suffer long-term neurological and non-neurological sequelae. Prevention of disease through vaccination is the only realistic prospect for disease control.

22.2 The Clinical Spectrum of Meningococcal Disease

Meningococcal infection ranges from asymptomatic nasopharyngeal carriage to fulminant septic shock, which can cause death within a few hours. Septicaemia and acute meningitis are the commonest manifestations of invasive disease. Meningococcal sepsis is classically described as a syndrome of fever and widespread purpura, with or without shock. Occult bacteraemia and chronic meningococcaemia can also occur. Occasionally the disease manifests as a focal infection such as pneumonia, septic arthritis, osteomyelitis, myocarditis, pericarditis, peritonitis, conjunctivitis, endophthalmitis, sinusitis or otitis media.

Invasive disease is often rapidly progressing from a non-specific febrile illness, indistinguishable from minor viral infections, to fulminant septicaemia and/or severe meningitis. In children who ultimately develop septicaemia, fever, nausea and vomiting and lethargy are the most frequent early symptoms. A blanching, salmon-coloured, maculopapular rash, similar to viral exanthems, may also be present. As disease progresses, signs of shock become more apparent. A rash occurs in 70–80% of meningococcal bacter-

aemia cases at hospital presentation and is usually non-blanching (i.e. petechial or purpuric). Most affected patients have only non-specific symptoms in the first 4–6 h of symptom onset, with the petechial/haemorrhagic rash, meningism and impaired consciousness developing later at a median of 13–22 h. Meningitis has more non-specific clinical features in infants and young children, when disease incidence is highest, compared with older children. Initial symptoms usually include fever, nausea and vomiting, photophobia and severe headache. Seizures can occur early or later in disease. Irritability, delirium and altered level of consciousness develop as central nervous system (CNS) inflammation progresses. The most specific signs are neck stiffness, associated with Kernig and Brudzinski signs, but these are often absent in children. Focal neurological abnormalities and signs of raised intracranial pressure may also occur. Where septicaemia and meningitis coexist, neurological features are due to cerebral ischaemia and/or meningeal inflammation.

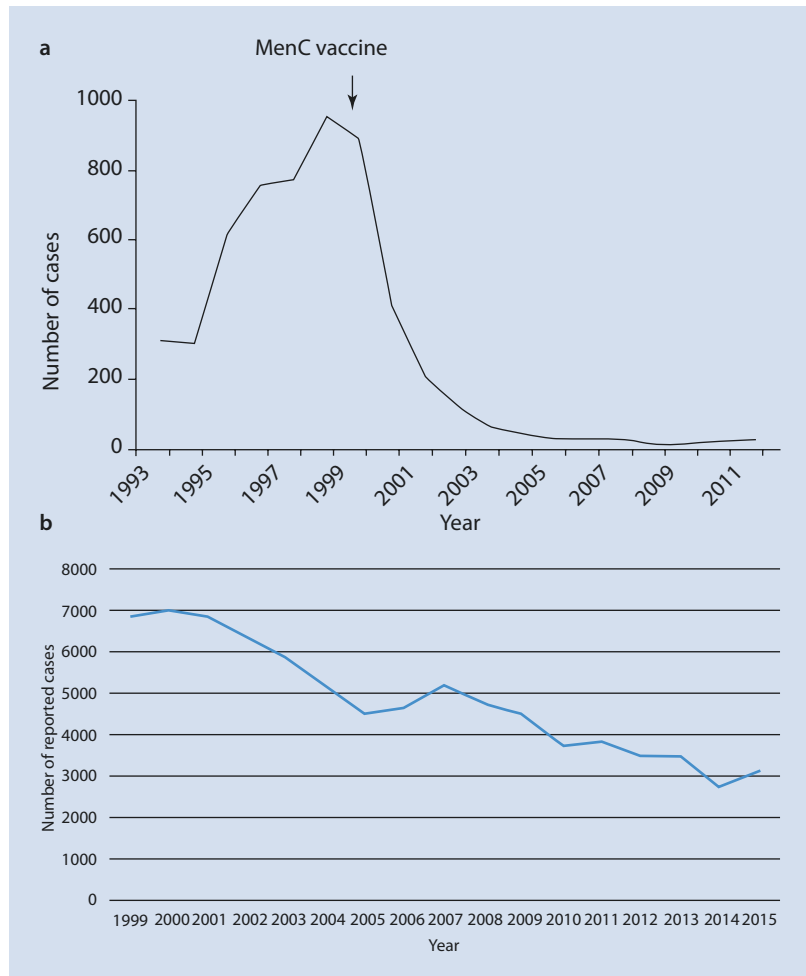
Despite medical advances, the case fatality rate in industrialised countries has remained around 5–15% since the 1950s, although some specialist centres have reported a case fatality rate of 5% with early aggressive circulatory support. Early neurological complications include seizures, syndrome of inappropriate antidiuretic hormone (SIADH), subdural effusions and empyema, hydrocephalus, raised intracranial pressure, focal neurological abnormalities, venous sinus thrombosis and cerebral infarction. Sequelae secondary to severe shock occur due to tissue hypoperfusion and include skin necrosis and subsequent scarring (which may need skin grafting) and gangrene of parts or entire limbs, requiring amputation. Growth plate damage may require multiple surgical procedures until growth is complete. There are very high rates of significant sequelae in survivors (up to 20–30% in most studies), leading to long-term disability. These include sensorineural hearing loss, epilepsy, learning difficulties, and motor/cognitive impairment. Arthritis can lead to permanent joint damage. Studies of longer-term outcomes, up to 15 years, after disease have described sequelae in up to 50–60%, including physical and neuropsychiatric problems. Significant emotional problems in close family members have also been found, highlighting the societal impact.

22.3 Epidemiology of Meningococcal Disease in Europe

Invasive meningococcal disease is rare in Europe, with rates of 0.09–6.6 cases per 100,000 population, depending on the country, between 2010 and 2014. Highest rates occur in Malta, Lithuania, Ireland and the United Kingdom. Infants (under 1 year of age) have the highest disease incidence rates (>10 per 100,000 per year), followed by 1–4-year-olds and adolescents/young adults aged 15–24 years. Most cases in 2012 were caused by group B organisms (68%), followed by group C (17%), Y (8%) and W (4%). Between 2008 and 2012, disease caused by group B and C organisms decreased, partly due to introduction of capsular group C (MenC) conjugate vaccines in several countries (■ Fig. 22.1a, b). The MenC conjugate

vaccine resulted in a tenfold drop in incidence of group C disease in those countries which introduced it, and most European countries include 1–3 doses of the monovalent MenC conjugate vaccine in the routine childhood immunisation schedule. In Austria, Greece and the United Kingdom, an adolescent booster of the quadrivalent MenACWY conjugate vaccine is used, and in the Czech Republic, the quadrivalent vaccine is used for all three doses (■ Fig. 22.2). In the United Kingdom, there was an increase in capsular group W (MenW) disease due to expansion of a hypervirulent strain, from 22 cases in 2009 to almost 180 in 2014–2015. Molecular characterisation of recent isolates has confirmed that the recent increase is due to a strain distinct from the 2000 Hajj-associated outbreak of MenW, which spread worldwide and lasted for several years. In 2013–2014, MenW was responsible for 15% of all cases

■ Fig. 22.1 a Impact of meningococcal capsular group C (MenC) conjugate vaccines on MenC disease in the United Kingdom. Number of laboratory confirmed cases of invasive capsular group C meningococcal disease in England and Wales between July 1993 and June 2012, before and after introduction of MenC vaccine into the UK routine immunisation schedule in 1999. b Meningococcal disease in Europe. Number of cases of meningococcal disease reported in the European Union/European Economic Area 1999–2015



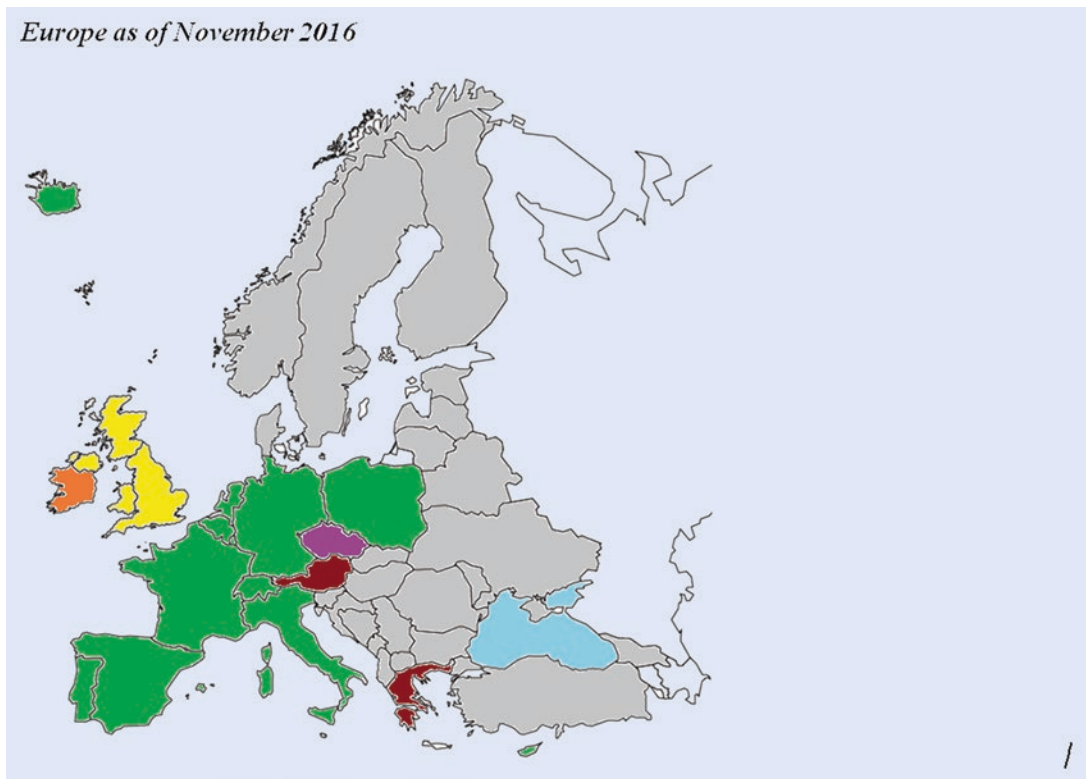


Fig. 22.2 Use of meningococcal vaccines in routine immunisation schedules across Europe as of November 2016. *Green* MenC vaccine only, *brown* MenC and MenACWY vaccines only, *orange* MenB and MenC vaccines only, *purple* MenB and MenACWY vaccines only, *yellow* MenB, MenC and MenACWY vaccines, *grey* no meningococcal vaccine. This image only depicts vaccines in routine

use; additional vaccines may be recommended in some countries in high-risk groups and/or travellers (Data from ► <http://vaccine-schedule.ecdc.europa.eu/Pages/Scheduler.aspx> and ► http://apps.who.int/immunization_monitoring/globalsummary/schedules (Accessed 2 November 2016)

of invasive disease, compared to historical levels of 1–2%. This increase prompted the change in the UK adolescent booster from the monovalent MenC vaccine to quadrivalent MenACWY vaccine in September 2015.

22.4 Polysaccharide Vaccines

The first meningococcal vaccines in regular use were plain polysaccharide vaccines developed in the 1960s. These vaccines are based on the capsule which surrounds the organism and is used for grouping and were produced to target disease caused by groups A, C, W and Y with bivalent MenAC and quadrivalent MenACWY vaccines produced. In clinical trials, the capsular groups A and C components of these vaccines had over 90% effectiveness in the short term against disease caused by these organisms, but protection

waned over time, especially among children. An intervention study in Quebec, Canada, showed that effectiveness of the group C component was 95% in children ≥ 6 years during the first 2 years post-vaccination, but was not effective in younger children. The group A component is immunogenic from a few months of age, therefore making it unlike other polysaccharide vaccines which do not induce protective immunity before 2 years of age. There are no protection data currently available for capsular group W or Y polysaccharides.

While these polysaccharide vaccines are effective in protecting older children and adults against disease, they are inadequate for young children with the highest disease incidence.

The immune response does not involve recruitment of helper T cells, so immunological memory does not occur; the vaccines induce short-term protection only, are associated with immunological hypo-responsiveness (reduced responses after

administration of booster doses), and do not elicit a response in children under 2 years of age. The lack of a response in young children is thought to be due to immaturity of the marginal zone B cells. Antibody responses to these vaccines are thought to be induced by cross-linking of the B-cell receptor by the repeating polysaccharide moieties, which results in differentiation of antigen-specific B cells into antibody-secreting cells without germinal centre formation in the draining lymph node. These vaccines have now almost entirely been replaced by protein-polysaccharide conjugate vaccines.

22.5 MenC Conjugate Vaccines

In the 1990s conjugate vaccines, which are able to overcome the problems of plain polysaccharide vaccines, were successfully developed. In these products the polysaccharides are conjugated to protein carriers CRM₁₉₇ (a non-toxic genetic variant of diphtheria toxin) (Meningitec, Nuron Biotech and Menjugate, GSK) or tetanus toxoid (NeisVac-C, Pfizer). The first monovalent MenC conjugate vaccines were licensed in the United Kingdom in 1999 and subsequently in the rest of Europe. The MenC conjugate vaccine was introduced into the UK routine immunisation programme from November 1999, and between 1999 and 2001 there was a reduction in MenC cases of 87% among the vaccinated groups. The MenC conjugate vaccine induces high levels of bactericidal antibodies in all age groups, and vaccine effectiveness correlates with the induction of these functional antibodies with a titre $\geq 1:8$ in the population.

In contrast to the polysaccharide vaccines, the MenC conjugate vaccines also induce immunological memory (eliciting an augmented response to subsequent doses of vaccine and/or the presence of MenC-specific memory B cells in the peripheral blood), which allow rapid (about 4 days) and high-magnitude responses to occur when a vaccinated individual is exposed to serogroup C meningococci. In unimmunised infants the response takes about 10 days and is of lower magnitude following their first dose of MenC vaccine. With the rapid onset of disease, however, this memory response within 4 days would not be sufficient to protect an individual, and maintenance of high levels of serum bactericidal antibody is likely to be necessary to preserve vaccine effectiveness.

22.6 Herd Immunity Induced by Conjugate Vaccines

The reduction in nasopharyngeal meningococcal carriage by conjugate vaccines has been a vital contribution to their remarkable success. The MenC conjugate vaccine in the United Kingdom reduced transmission of group C *N. meningitidis*, thereby providing herd protection – indirectly protecting unvaccinated individuals. After the vaccine was introduced, the number of cases among unvaccinated age groups fell by 67%, corresponding to a reduction in MenC carriage rates in vaccinated young adults. The highest rates of meningococcal carriage occur in adolescents and young adults, so many countries include an adolescent booster dose of MenC or MenACWY conjugate vaccine to maintain herd protection in the population, which will remain highly effective at maintaining protection of the population if high vaccine uptake rates can be maintained.

22.7 MenACWY Conjugate Vaccines

Three meningococcal ACWY (MenACWY) conjugate vaccines have been developed to provide broader protection against meningococcal disease (MenACWY polysaccharides conjugated to diphtheria toxoid, Menactra; tetanus toxoid, Nimenrix; or CRM197, Menveo), though only two are currently licensed in Europe (tetanus and CRM197 conjugate). Licensure trials undertaken with each of these products found them to be non-inferior in induction of bactericidal antibody when compared with either the previously licensed MenACWY polysaccharide vaccines or the first-licensed MenACWY diphtheria conjugate vaccine. These vaccines, like the MenC conjugate vaccines, induce immunological memory, and the responses to the vaccines can be boosted. However, although some of the clinical development studies were conducted in infants, these vaccines are not currently licensed for use in infants in Europe (tetanus conjugate is licensed from 12 months of age; CRM197 conjugate is licensed from 2 years of age).

Potential scheduling of MenACWY vaccines could include a toddler dose as a replacement for the toddler MenC dose use in a number of countries and/or an adolescent dose, to act as a booster

Table 22.1 Licensed schedules of MenACYW in Europe

Population	Age	Dose series	Interval	Comments
Children in high-risk groups	0–12 months	2	≥1 months	Unlicensed at this age but used off label in some countries where recommended
Unvaccinated children and high-risk groups	12 months to adulthood	1	N/A	MenACYW-TT licensed from 12 months of age and MenACYW-CRM from 24 months. Used for immunisation of toddlers and/or adolescents/adults in some countries. No data over 65 years of age

for earlier MenC doses (see [Table 22.1](#)) and to reduce nasopharyngeal carriage of meningococci among adolescents and disease caused by A, C, W and Y meningococci in these individuals and more widely through herd immunity. Serological evidence of protection has been shown to persist in most adolescents for at least 2–3 years after immunisation, but longer-term follow-up data are still needed. Antibody is not so well maintained after immunisation of younger children.

No prelicensure vaccine efficacy studies were undertaken, but one study estimated vaccine effectiveness over 6 years of use of the MenACWY diphtheria-conjugate in the United States to be 69% (95% CI, 50–81%). There was an indication that the vaccine effectiveness declined from a high of 82% (54–923%) after the first year from immunisation, in keeping with observations of a decline in bactericidal antibody, leading the US policy to use a two-dose schedule of the vaccine. The MenACWY-CRM-197 conjugate vaccine was assessed in a study evaluating effectiveness of meningococcal vaccines against nasopharyngeal carriage in almost 3000 university students, and the vaccine was found to reduce carriage of C, W and Y strains by 36.2% (15.6–51.7), suggesting the potential for the vaccine to induce herd immunity.

Preliminary trials showed the MenACWY conjugate vaccines to have a similar local and systemic reaction safety profile to that described for other conjugate vaccines and the licensed polysaccharide vaccines. Early reports of an association of the MenACWY-diphtheria conjugate vaccine with Guillain-Barre syndrome have not been confirmed in subsequent observations.

As a result of spread of a hyperinvasive clone of capsular group W meningococcus in the United Kingdom, immunisation of adolescents with

MenACWY conjugate vaccine from age 13/14 to 18 years of age commenced in 2015, and data are expected to emerge on vaccine effectiveness of the programme in the near future. 4CMenB, described below, is also likely to provide protection against some non-B strains.

22.8 Capsular Group B Meningococcal Vaccines

The poor immunogenicity of the group B polysaccharide made the development of vaccines against MenB disease particularly challenging; however, the use of subcapsular proteins as alternative vaccine targets has enabled the recent development of two vaccines that offer the potential to overcome this gap in meningococcal disease prevention.

One of these, 4CMenB (Bexsero, GSK), has been included in the routine infant immunisation schedule in the United Kingdom and Ireland, and effectiveness data from the first 10 months of the UK campaign has provided the first direct evidence that this vaccine is able to provide broad protection against capsular group B meningococcal disease in infants. The other, rLP2086 (Trumenba, Pfizer), is licensed for use in adolescents in the United States, and has recently been approved in Europe.

22.9 4CMenB

This vaccine was licensed in Europe in 2013 and has subsequently been licensed in more than 35 countries. In Europe 4CMenB is licensed from 2 months of age, with schedules differing according to age ([Table 22.2](#)).

Table 22.2 Licensed schedules of 4CMenB in Europe

Population	Age	Dose series	Interval	Booster recommended
Infants	2–5 months	3	≥1 month	One dose at 12–23 months
Unvaccinated infants	6–11 months	2	≥2 months	One dose at 12–23 months; ≥2 months from primary series
Unvaccinated children	12–23 months	2	≥2 months	One dose 12–23 months after the primary series
Unvaccinated children	2–10 years	2	≥2 months	
Adolescents and adults	11 years and older	2	1–2 months	

4CMenB contains four key immunogenic components:

- Detoxified outer membrane vesicles (OMVs) from strain 44/76, within which the immunodominant antigen is porin A (PorA)
- Factor H binding protein (fHbp)
- Neisserial adhesin A (NadA)
- Neisserial heparin-binding antigen (NHBA)

This multicomponent approach was taken to broaden the immunity against MenB provided by vaccines based on OMVs alone, which had been given in phase 3 effectiveness studies in Norway or in population-based interventions in Latin America, Normandy and New Zealand. These vaccines were effective against disease due to the strain from which the OMV was derived, but not against strains bearing variants of the immunodominant PorA protein (especially in infants). Their use was therefore confined to epidemics of MenB disease due to restricted lineages, rather than endemic disease. The use of the OMV in 4CMenB not only allowed inclusion of the PorA antigen, but may also non-specifically enhance the immune response to the additional vaccine antigens.

Of the “additional” proteins, fHbp and NHBA are nearly universal on pathogenic *N. meningitidis*, while genes for NadA were present in 23% of a European strain panel. Clinical trials in which 6427 participants from 2 months to adulthood received 4CMenB have shown these proteins induce bactericidal antibodies against MenB strains expressing closely matched antigens. However, pathogenic meningococci differ in the surface expression of these proteins, and like PorA, there is phenotypic variability that potentially restricts the breadth of cross protection

afforded by the antibodies induced by each individual vaccine component. Determining the likely breadth of direct protection afforded by immunisation with 4CMenB in any given population has to take into account all these factors, even before considering the potential for synergistic (or antagonistic) interactions between different vaccine-induced antibodies acting on the target bacteria at the same time.

Given these challenges, various methods have been used to predict the proportion of MenB disease potentially preventable by immunisation with 4CMenB. One of these, “MATS”, predicts that the potential coverage of 4CMenB in Europe varies by country between 73% and 85%. In England coverage by MATS was predicted to be 67.2% in 2014/2015, a fall from 73% in 2007/2008, whereas 88% of common disease-causing strains appeared susceptible to pooled post-immunisation sera. While still awaiting formal validation by comparison with the emerging “real-life” effectiveness data, these estimates have provided a starting point for consideration of the potential benefits and cost-effectiveness of the vaccine’s introduction.

22.9.1 Experience of Use

To date 4CMenB has been used in outbreaks of MenB disease in the Saguenay-Lac-Saint-Jean region of Quebec, Canada, in educational institutions in the United States and Canada and has been introduced into the routine immunisation schedules in the United Kingdom and Ireland. Many countries have also recommended the use of 4CMenB for children with complement deficiencies and splenic dysfunction/asplenia. In the

United Kingdom, routine immunisation at 2, 4 and 12 months commenced in September 2015 (for infants born after 1 May 2015). Ten months into this immunisation campaign, 95.5% and 88.6% of eligible infants had received their first and second dose (respectively) by 6 months of age. Lower 4CMenB immunisation rates in infants experiencing disease compared with their age-matched population cohort suggested a vaccine effectiveness of 82.9% (95% C.I. 24.1–95.2), and the number of MenB disease cases in the vaccine-eligible age group was 50% lower in the period following vaccine introduction than the average of the previous 4 years. Of note is that this “2 + 1” schedule differs from the licensed schedule for 2 month olds; the same reduced schedule is being used in Ireland.

22.9.2 Reactogenicity/Safety

The most significant adverse events after immunisation of infants are fever and irritability, which are observed in approximately 60% and 75% of 2-month-old infants when 4CMenB is given with routine infant vaccines (DTaP-IPV-HepB and 7-valent pneumococcal vaccine). These relatively high rates of fever may be due to the inclusion of OMVs in this vaccine. Rates of fever were reduced by the use of prophylactic paracetamol at the time of immunisation (from 71% to 52% in 2-month-olds) without impacting on the vaccine's immunogenicity. Other reported reactions include tenderness at the injection site, which is reported as severe in 12–16% of infants (crying when moving leg) and 17% of adolescents (unable to perform unusual duties).

Concerns regarding the rates of post-immunisation fever in infants led to the recommendation in the United Kingdom that prophylactic paracetamol be administered at the time of 4CMenB administration for 2- and 4-month-olds, with two further doses given in the next 24 h. Despite this provisional data suggests an increase in emergency department attendances for post-immunisation reactions.

As with all new vaccines, extensive post-marketing surveillance is being conducted to detect any less common, but serious, adverse events following immunisation. Particular attention is being paid to Kawasaki disease and febrile seizures following cases of both conditions in

4CMenB clinical trials that were considered possibly related to 4CMenB.

22.9.3 Areas of Uncertainty

Two key determinants of the impact of 4CMenB immunisation campaigns that are as yet unknown are the duration of vaccine-induced immunity and whether such campaigns can induce herd immunity by reducing rates of nasopharyngeal carriage of potentially invasive MenB strains.

Following immunisation in infancy with three priming doses and boosting at 12 months of age, over 97% of children have bactericidal antibodies above the accepted correlate of protection for three key MenB strains, and it is from this presumed peak concentration that estimates of vaccine coverage have been made. By 4 years of age, these proportions had fallen to 9, 12 and 93%, depending on which strain was tested. Although a good response to a booster dose was observed, an additional booster dose is not included in the licensed 4CMenB schedule. Whether this waning of antibodies against some vaccine antigens will be of clinical relevance as children proceed through their school years and into adolescence will only be apparent from ongoing disease surveillance in an immunised population.

The potential for immunisation with 4CMenB to reduce nasopharyngeal carriage of MenB is also uncertain. A randomised clinical trial of UK university students demonstrated an 18% reduction in rates of nasopharyngeal carriage of all meningococci in 4CMenB recipients compared with controls; however, this was primarily in non-B capsular groups. No impact on circulation of meningococci is expected from the UK immunisation campaign as this does not include adolescents or young adults (the ages at which carriage is most common); however, studies are planned to assess further the potential of an adolescent immunisation campaign with 4CMenB to induce herd immunity.

The finding of an impact of 4CMenB immunisation on non-B meningococci in the carriage study highlights that the antigens contained in 4CMenB are not specific to the B capsular group, and indeed immunisation with 4CMenB has been shown to raise bactericidal antibodies to capsular group W. Whether this will translate into cross-capsular group protection against invasive disease is yet to be determined.

22.10 Bivalent rLP2086

This vaccine is licensed for use in adolescents in the United States in a three-dose (0, 1–2, 6 months) or two-dose (0, 6 months) schedule, the former being more appropriate in outbreak settings. As with 4CMenB, this vaccine has a “category B” recommendation for use in 16- to 23-year-olds in that country (i.e. may be administered to provide short-term protection against most strains of serogroup B meningococcal disease). An application for licensure in the European Union in those 10 years and was recently approved.

This vaccine is based on two variants of the fHbp protein that have had a lipid tail attached. As with 4CMenB, immunogenicity against a broad range of MenB strains has been demonstrated, and the vaccine has been licensed on this basis rather than on direct evidence of effectiveness.

Bivalent rlp2086 has been used in the context of college outbreaks in the United States; however, formal effectiveness studies have not been possible during these campaigns due to the low number of cases and brief duration of the outbreaks. No impact of vaccination on nasopharyngeal carriage was observed during an outbreak at Providence College, Rhode Island in 2015, however the relatively small sample size did not allow a definitive assessment of this vaccine’s potential to induce herd immunity.

Observed side effects following bivalent-rlp2086 administration to adolescents include injection site pain (severe in 8.2%), headache (56.9%, severe in 1.4%) and pyrexia (8.3%). Serious adverse events in clinical trials were no more common following this MenB vaccine than comparator, licensed, vaccines.

22.11 Conclusion

The availability of vaccines against MenC, MenACWY and MenB disease represents an important advance in the prevention of meningococcal disease. The dramatic changes in meningococcal epidemiology observed over the last two decades in Europe emphasise the need to have vaccines available to deal with existing and emerging threats in a timely manner to save lives. Further developments such as the recent emergence of serogroup X in sub-Saharan Africa, for which there is currently no licensed vaccine,

demonstrate that vaccine prevention of meningococcal disease is an ongoing and evolving challenge.

Further Reading

- Agnememel A, Hong E, Giorgini D, Nunez-Samudio V, Deghmane AE, Taha MK. Neisseria meningitidis Serogroup X in Sub-Saharan Africa. *Emerg Infect Dis*. 2016;22(4):698–702.
- Artenstein MS, Gold R, Zimmerly JG, Wyle FA, Schneider H, Harkins C. Prevention of meningococcal disease by group C polysaccharide vaccine. *N Engl J Med*. 1970;282(8):417–20.
- Bjune G, Hoiby EA, Gronnesby JK, Arnesen O, Fredriksen JH, Halstensen A, Holten E, Lindbak AK, Nokleby H, Rosenqvist E, et al. Effect of outer membrane vesicle vaccine against group B meningococcal disease in Norway. *Lancet*. 1991;338(8775):1093–6.
- Borrow R, Balmer P, Miller E. Meningococcal surrogates of protection—serum bactericidal antibody activity. *Vaccine*. 2005;23(17–18):2222–7.
- Christensen H, May M, Bowen L, Hickman M, Trotter CL. Meningococcal carriage by age: a systematic review and meta-analysis. *Lancet Infect Dis*. 2010;10(12):853–61.
- Cohn AC, MacNeil JR, Clark TA, Ortega-Sanchez IR, Briere EZ, Meissner HC, Baker CJ, Messonnier NE, Centers for Disease and Prevention. Prevention and control of meningococcal disease: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep*. 2013;62(RR-2):1–28.
- European Centre for Disease Prevention and Control. Surveillance atlas of infectious disease. Accessed 6th Sept 2016.
- Frosi G, Biolchi A, Lo Sapio M, Rigat F, Gilchrist S, Lucidarme J, Findlow J, Borrow R, Pizza M, Giuliani MM, Medini D. Bactericidal antibody against a representative epidemiological meningococcal serogroup B panel confirms that MATS underestimates 4CMenB vaccine strain coverage. *Vaccine*. 2013;31(43):4968–74.
- Jafri RZ, Ali A, Messonnier NE, Tevi-Benissan C, Durrheim D, Eskola J, Fermon F, Klugman KP, Ramsay M, Sow S, Zhujun S, Bhutta ZA, Abramson J. Global epidemiology of invasive meningococcal disease. *Popul Health Metr*. 2013;11(1):17.
- Kelly C, Arnold R, Galloway Y, O’Hallahan J. A prospective study of the effectiveness of the New Zealand meningococcal B vaccine. *Am J Epidemiol*. 2007;166(7):817–23.
- Kelly DF, Snape MD, Clutterbuck EA, Green S, Snowden C, Diggle L, Yu LM, Borkowski A, Moxon ER, Pollard AJ. CRM197-conjugated serogroup C meningococcal capsular polysaccharide, but not the native polysaccharide, induces persistent antigen-specific memory B cells. *Blood*. 2006;108(8):2642–7.
- Ladhani SN, Giuliani MM, Biolchi A, Pizza M, Beebeejaun K, Lucidarme J, Findlow J, Ramsay ME, Borrow R. Effectiveness of Meningococcal B Vaccine against Endemic Hypervirulent Neisseria meningitidis W Strain, England. *Emerg Infect Dis*. 2016;22(2):309–11.

- Maiden MC, Stuart JM, U. K. M. C. Group. Carriage of serogroup C meningococci 1 year after meningococcal C conjugate polysaccharide vaccination. *Lancet*. 2002;359(9320):1829–31.
- Peltola H, Makela H, Kayhty H, Jousimies H, Herva E, Hallstrom K, Sivonen A, Renkonen OV, Pettay O, Karanko V, Ahvonen P, Sarna S. Clinical efficacy of meningococcus group A capsular polysaccharide vaccine in children three months to five years of age. *N Engl J Med*. 1977;297(13):686–91.
- Read RC, Baxter D, Chadwick DR, Faust SN, Finn A, Gordon SB, Heath PT, Lewis DJ, Pollard AJ, Turner DP, Bazaz R, Ganguli A, Havelock T, Neal KR, Okike IO, Morales-Aza B, Patel K, Snape MD, Williams J, Gilchrist S, Gray SJ, Maiden MC, Toneatto D, Wang H, McCarthy M, Dull PM, Borrow R. Effect of a quadrivalent meningococcal ACWY glycoconjugate or a serogroup B meningococcal vaccine on meningococcal carriage: an observer-blind, phase 3 randomised clinical trial. *Lancet*. 2014;384(9960):2123–31.
- Sierra GV, Campa HC, Varcacel NM, Garcia IL, Izquierdo PL, Sotolongo PF, Casanueva GV, Rico CO, Rodriguez CR, Terry MH. Vaccine against group B Neisseria meningitidis: protection trial and mass vaccination results in Cuba. *NIPH Ann*. 1991;14(2):195–207; discussion 208–10.
- Snape MD, Dawson T, Oster P, Evans A, John TM, Ohene-Kena B, Findlow J, Yu LM, Borrow R, Ypma E, Toneatto D, Pollard AJ. Immunogenicity of two investigational serogroup B meningococcal vaccines in the first year of life: a randomized comparative trial. *Pediatr Infect Dis J*. 2010;29(11):e71–9.
- Snape MD, Perrett KP, Ford KJ, John TM, Pace D, Yu LM, Langley JM, McNeil S, Dull PM, Ceddia F, Anemona A, Halperin SA, Dobson S, Pollard AJ. Immunogenicity of a tetravalent meningococcal glycoconjugate vaccine in infants: a randomized controlled trial. *JAMA*. 2008;299(2):173–84.
- Trotter CL, Andrews NJ, Kaczmarek EB, Miller E, Ramsay ME. Effectiveness of meningococcal serogroup C conjugate vaccine 4 years after introduction. *Lancet*. 2004;364(9431):365–7.
- Vieusseux M. Memoire sur le maladie qui a regne a Geneva au printemps de 1805. *J Med Clin Pharm*. 1805;11:163–82.

Pediatric Vaccines for Travel Outside Europe

R.H. Behrens and N. Prevatt

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23.1 Introduction

Pediatric travellers face most of the infectious risks that adults face during travel, but their different immunological maturity and response to infection make them more vulnerable to disease.

An individualised risk assessment is essential for every Pediatric traveller. This should involve all aspects of the child's health including pre-existing health issues that may affect their ability to receive certain vaccinations safely, medical needs if any, and the time the parents have allowed to prepare for their journey.

23.2 Background to Vaccination for Travel

A wide range of travel vaccines exist, but it is important to balance the risk of disease against the risk associated with the vaccine.

Prior to travel the routine childhood immunisation schedule must be up to date and vaccines can be brought forward if necessary, so that essential vaccines and boosters are not missed during travel. This is particularly important for MMR and DTP since the risk of contracting these infections is much higher in the tropics than in many parts of Europe.

Hepatitis A virus is transmitted by ingestion of food and water contaminated by faeces and occasionally by close contact between children. As such the distribution of Hepatitis A is closely mapped to poor economic and sanitary conditions, with the highest incidence in the Indian subcontinent.

Children who have had hepatitis A, or are likely to suffer only mild illness (≤ 5 years of age), do not need to receive this vaccine. Pediatric formulations of the hepatitis A vaccine are available for use from 1 year of age should parents decide to have a younger child immunised. Vaccination against Hepatitis A should ideally be given 14 days prior to travel to become fully effective (see ► Chap. 12 for more details).

23.2.1 Yellow Fever

The yellow fever (YF) virus is an arthropod-borne flavivirus, which circulates between monkeys and

humans, and between humans, via *Aedes* species of mosquitoes.

Yellow fever (YF) ranges from a moderate flu-like illness with nausea and vomiting to a severe, multisystem haemorrhagic disease, with jaundice (hence the name) and circulatory shock. Approximately one quarter of patients die within 7–10 days of onset. Those patients who survive will have acquired lifelong immunity.

Global numbers estimate approximately 51,000–380,000 cases, with between 19,000 and 180,000 fatalities per annum. The number of cases reported to WHO are not indicative of the risk faced by travellers as very few travellers have ever been infected with yellow fever, with only six reported travel-acquired infections since 1990. The risk of YF is present in parts of South America and sub-Saharan Africa (see ■ Maps A and B).

Vaccine Requirements for Pediatric Travellers

The requirement for YF vaccine by destination is outlined in the International Health Regulations (IHRs), and infants and children aged over 9–12 months should receive the vaccine based on this indication, after a risk-benefit analysis which takes into account their journey and likely exposure.

The IHRs are designed to prevent the spread of the virus from endemic to non-endemic regions through travel. An international certificate of vaccination may, therefore, be required by endemic and non-endemic countries under the World Health Organization (WHO) regulations. Thus, the requirement for a YF vaccine certificate may not reflect the risk of exposure in that country. This is an important factor to consider where relative contraindications to the vaccination apply.

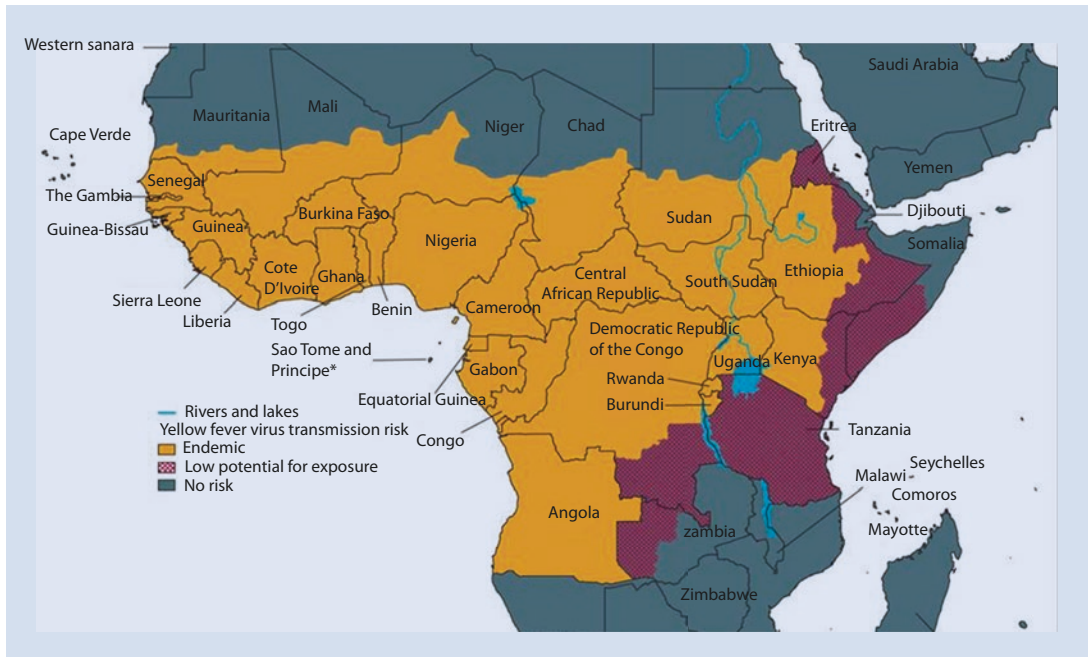
The following table explains when children may need to receive the vaccine

Travelling to countries with YF transmission	Travelling to countries requiring mandatory YF vaccination for travellers from all countries	Travelling to countries requiring YF vaccination for travellers arriving from countries with risk of YF transmission
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■ **Map A** Yellow fever distribution in Central and South America. Jentes ES, Pomeroy G Fau – Gershman MD, Gershman Md Fau – Hill DR, Hill Dr. Fau – Lemarchand J, Lemarchand J Fau – Lewis RF, Lewis Rf Fau – Staples JE, et al. The revised global yellow fever risk map and recommendations for vaccination, 2010: consensus of the Informal WHO Working Group on Geographic Risk for Yellow Fever. (1474–4457)

- ▶ <http://www.who.int/ith/en/>
- ▶ http://gamapserver.who.int/mapLibrary/Files/Maps/ITH_YF_vaccination_americas.png?ua=1
- Yellow fever vaccine recommendations in the Americas 2013
- WHO map of YF in the Americas and Africa
- ▶ <http://apps.who.int/ithmap/>



Map B Yellow fever distribution in Africa. Jentes ES, Pomeroy G Fau – Gershman MD, Gershman Md Fau – Hill DR, Hill Dr. Fau – Lemarchand J, Lemarchand J Fau – Lewis RF, Lewis Rf Fau – Staples JE, et al. The revised global yellow fever risk map and recommendations for vaccination,

2010: consensus of the Informal WHO Working Group on Geographic Risk for Yellow Fever. (1474–4457)

► http://gamapserver.who.int/mapLibrary/Files/Maps/ITH_YF_vaccination_africa.png?ua=1

Yellow fever vaccination recommendations in Africa, 2015

The list of country requirements for the YF vaccination certificate (or exemption certificate) can be found online on the WHO website.

23.2.2 Pediatric Yellow Fever Vaccination

Two vaccines, 17DD and 17D-204, were developed simultaneously in the 1930s and are thought to have similar immunogenicity and safety profiles. Currently the only licenced YF vaccine used in Europe is Stamaril[®], made by Sanofi Pasteur. It is a live attenuated 17D-204 strain of yellow fever virus, grown in embryonated chick eggs. Every 0.5 ml dose contains at least 1000 IU, which should cause a subclinical infection in healthy individuals.

The vaccine is administered as an intramuscular (IM) or deep subcutaneous injection. The dose is of 0.5 ml regardless of age. There is new evidence that the vaccine administered as 0.1 ml IM produces equivalent neutralising antibody titres, but this is not currently used outside of epidemic scenarios and WHO does not allow certification at this dose.

This vaccine is highly effective. At 10 days after vaccination, 90% have seroconverted; at 1 month nearly 100% of adults are protected. Pediatric studies show a minimum seroconversion rate of 86% after single-dose vaccination, with the lowest seroconversion rates in children 9–36 months of age.

A neutralising antibody titre of 1:10 is considered protective against YF. Yet neutralising antibody testing may underestimate protection because T-cell immunity may also be present.

The vaccine must be given more than 10 days prior to entry to comply with the International Health Regulations (IHRs), as the neutralising antibody response will have been achieved in this timescale.

Vaccine recipients maintain detectable levels of neutralising antibody for more than 10 years post vaccination, and boosters are not required unless the recipient was immunocompromised or below 2 years at the time of vaccination. In 2016, the World Health Organisation (WHO) amended the 10-year booster rule within the IHRs dictating that a single dose of vaccine should now be considered to confer lifelong protection and no further boosters are required.

Infants aged less than 6 months should not be given YF vaccine because of the risk of encephalitis. The vaccine can be considered between 6–9 months where there is a risk in outbreak settings. The vaccine should similarly only be given to pregnant or breastfeeding mothers under such high-exposure circumstances as visiting a region with an outbreak.

Vaccine Side Effect Profile

A range of temporary adverse events have been reported following the administration of YF vaccine. The most frequent reactions include headache, myalgia and injection site swelling, which occur in 10–15% of recipients. In infants and young children, the most frequently reported reactions are irritability, crying and appetite loss, and these are reported in approximately one third of children in the first few days. Pyrexia can develop up to 14 days afterwards. Reports of generalised allergic reactions indicate an incidence of 1 in 131,000.

However, YF is a vaccine with known rare serious adverse events: aside from hypersensitivity reactions, there are two distinct forms of important and severe adverse events associated with receipt of this vaccine. Thus far, these have mostly been described in people receiving a primary dose of the vaccine. They are yellow fever vaccine-associated viscerotropic disease (YEL-AVD) and yellow fever-associated neurologic disease (YEL-AND).

YEL-AVD usually occurs within 10 days of vaccination.

Features resemble fulminant infection by wild-type virus and thus may include fever, fatigue, myalgia, headache and jaundice. This may

progress to hypotension, metabolic acidosis, muscle and liver cytolysis, cytopenia and renal and respiratory failure. In these cases YF vaccine can be detected in serum and tissue PCR. The mortality rate has been around 60%. The risk of YEL-AVD in travellers is estimated to be 1 per 250,000 primary vaccines, with the highest risk in those over the age of 60 years. Cases of YEL-AND in children appear to be extremely scarce.

YEL-AND usually occurs within a month of vaccination.

Clinical features include high fever with headache that may progress to one or more of confusion, focal neurological deficit, encephalitis, meningitis or Guillain-Barré syndrome. Approximately one third of cases have been fatal, but the neurological sequelae may also be long-standing and disabling. Encephalitis is a particular risk in those under 9 months of age, with multiple cases described in infants less than 7 months old, prior to the establishment of a minimum age for vaccination. The incidence among infants aged below 6 months is highest and has been estimated as more than 0.5 cases per 1000. Cases of probable YEL-AND have also been reported following transmission from nursing mothers to infants.

Contraindications to YF Vaccine

YF vaccine can only be administered by designated clinics licensed by the health administration for the territory. Designated YF centre status is contingent on meeting the safety standards necessary to give this vaccine, one of which is staff training. ■ Table 23.1 denotes absolute and relative contraindications to the vaccine.

■ Table 23.1 Absolute and relative contraindications to YF vaccination

Contraindications in children	Precautions in children
Allergy to vaccine components (including anaphylaxis to egg) Age < 6 months Symptomatic HIV infection or CD4 < 200 (or <15% of the total in children <6 years) Thymus disorders (including thymoma/thymectomy/DiGeorge syndrome) Primary immunodeficiency Malignancy Transplantation Immunosuppressive therapy	Current febrile illness Age 6–9 months Asymptomatic HIV infection and (or CD4 15–24% of total in children <6 years)

Children with any of the contraindications listed are at risk of vaccine-associated disease and should be provided a letter of exemption.

Summary of Recommendations

Children should be given this vaccine for travel to areas of YF to meet certification for entry, unless they have contraindications or precautions. Where there are contraindications to the vaccine an exemption certificate should be issued to meet certification requirements.

23.2.3 Rabies

Rabies is caused by a *Lyssavirus*, which is transmitted to humans by the bite or scratch of infected mammals such as dogs, cats, monkeys and bats. Dog saliva in particular is the source of most rabies exposure.

The incubation period from bite to disease varies from weeks to years. First symptoms are fever, pain and paraesthesia around the wound. As the rabies virus spreads through the nervous system, it results in progressive paralysis, encephalitis and then certain death. Other infamous symptoms of furious rabies are aggression, aerophobia and hydrophobia. Children are symptomatic earlier as they are more likely to be bitten near to the head, and thus the central nervous system (CNS), on account of their height. Once symptoms occur rabies is invariably fatal.

Rabies is widespread across Africa, Asia, America and Europe, with the highest incidence in Asia. It causes an estimated 59,000 deaths per annum globally. Cases in travellers are very infrequent however an estimated 0.4% of adult travellers experience a potential animal exposure per months stay in an endemic country and appropriate post-exposure prophylaxis (PEP) is very difficult to obtain in many of the places where exposures occur. Since rabies is invariably fatal, it is an important disease to protect travellers against. The highest burden of exposures is among children, with 40% of PEP being accessed for child travellers.

Vaccine Requirements for Pediatric Travellers

A course of pre-exposure rabies vaccine is often considered for children who take multiple visits to or have prolonged stays in countries where rabies is endemic. Children put themselves at particular

risk by handling animals. A 'prolonged stay' is often considered to be over 1 month. WHO affirms that the risk assessment for this vaccine should not be based solely on the length of stay.

Parents should be encouraged to make this choice by balancing the cost of the vaccination with the cost of disrupting the trip in the case of a bite. Rabies vaccine may be viewed as insurance, as accessing PEP during travel can prove costly, time-consuming and difficult to obtain in some countries.

It is not routine practice to recommend vaccination for infants who are not ambulant as they are unlikely to come into contact with animals.

Pediatric Rabies Vaccination

There are two licenced rabies vaccines available in Europe:

- Human diploid cell vaccine
- Purified chick embryo cell vaccine

The cell culture vaccine is a Wistar rabies virus strain (PM/WI 38–1503-3 M) grown in human diploid cells, ultrafiltrated and inactivated. The chick embryo vaccine is prepared from a Flury LEP strain grown in chicken embryoblasts, centrifuged and inactivated.

The pre-exposure rabies schedule (the primary course) consists of three doses of vaccine at day 0, 7 and 28, intramuscularly. The third dose can be given early at day 21 if there is insufficient time before travel, and this has no significant effect on immunogenicity. Approximately 95% of adults respond to a full course of rabies pre-exposure vaccine, and the antibodies are long-lived. There is a paucity of Pediatric data. Intradermal vaccination is also effective in producing seroconversion and may be cheaper to administer to children.

Restarting an interrupted vaccine course is not necessary. The protective antibody level for rabies is estimated to be >0.5 IU/ml; however, there is no rationale for serological testing in a traveller (unless to confirm seroconversion in an immunosuppressed individual).

Post-exposure Prophylaxis (PEP)

Individuals who have been appropriately vaccinated prior to travel still require further booster doses when a significant bite occurs, but having a pre-exposure vaccination course eliminates the need for rabies immunoglobulin (RIG), which is in short global supply.

Table 23.2 Classifying rabies risk for the use of PEP

WHO category of exposure	
Category I	Licks to intact skin whilst feeding/touching (i.e. no exposure): <i>No PEP required</i>
Category II	Nibbling of skin, minor scratches or abrasions without bleeding: <i>Immediate PEP required</i>
Category III	Transdermal bites or scratches, licks on broken skin, contamination of mucous membranes with saliva from licks and exposure to bats: <i>Immediate PEP required and PLUS administration of rabies immunoglobulin (RIG) required for unvaccinated children</i>

The number of vaccine doses required post-exposure is dependent on the pre-exposure vaccine status. Travellers who have had a full course of pre-exposure vaccine will require two booster doses. Travellers who have not had any pre-exposure vaccination will require five to six doses of vaccine (depending on the regimen) (Table 23.2).

Children with category III exposures will require passive immunisation with rabies immunoglobulin (RIG) in addition to a post-exposure vaccine course if they have not received a full course of pre-exposure vaccinations. Rabies immunoglobulin (RIG) is an IgG prepared from the plasma of hyperimmunised individuals. Aside from the risks associated with blood-borne products, RIG is difficult to access and costly in endemic countries, and shortage of RIG is an important part of the risk analysis. For those who chose not to have their children vaccinated pretravel, it is wise to investigate the availability of RIG at the intended destination. In Southeast Asia an equine immunoglobulin is also available but is associated with significant risk of serum sickness and hypersensitivity reactions.

The decision to give PEP after rabies exposure in vaccine allergic individuals should be weighed carefully and the vaccine given with preparation for anaphylaxis being made.

23.3 Japanese Encephalitis

Japanese encephalitis (JE) is a mosquito-borne infection caused by the eponymous *Flavivirus*. It is transmitted in an enzootic cycle between mos-

quitoes and vertebrate hosts, usually pigs and birds. This transmission is by *Culex* sp. mosquitoes, which are evening and night-time biting mosquitoes. The main *Culex* vector for JE is *Culex tritaeniorhynchus*, which commonly breed in flooded rice fields and ground pools, and so the greatest transmission is in rural agricultural areas of Asia where there are rice paddy fields and pig farms. Some urban transmission does occur.

The incubation period is around 15 days, and symptoms include fever, flu-like symptoms and headache. Most infections are asymptomatic, but signs of encephalitis, such as altered level of consciousness and convulsions, occur in approximately 1 in 300 infections and are more common in the Pediatric population. Approximately 20–30% of symptomatic patients die, and of those that recover, 20–30% (21) are left with residual neurological problems including profound neurodisability, tremor, poor memory and psychological problems.

23.3.1 Epidemiology of JE in Travellers

JE was first described in Japan in the late 1800s, but it is now recognised throughout most of East and Southeast Asia, where it is a leading cause of viral encephalitis. China (excluding Taiwan) accounts for approximately 50 percent of cases. JE is also present in the Pacific Rim (see Map C).

Recent estimates are that around 68,000 cases occur annually in endemic countries (an annual incidence of approximately 1.8 per 100,000 popu-



Map C The global distribution of JE. (Source: CDC ► <https://www.cdc.gov/japaneseencephalitis/maps/index.html>, ► <http://apps.who.int/ithmap/>)

lation). Seventy-five percent of cases occur in children (annual incidence of approximately 5.4 per 100,000 population) with a higher frequency in those over 3 years.

Peak transmission of the virus occurs between *May and September* for the temperate regions of Asia such as Korea and Japan; between *March and October* for the more tropical countries of Southeast Asia such as Thailand, Cambodia and Vietnam; *September to December* for Nepal and Northern India; and *year-round* in countries with year-round rainfall such as Malaysia, Indonesia and the Philippines.

The risk to most travellers to both Asia and the Pacific is very low with an estimated incidence of less than one case per million travellers. The risk is presumed to be highest for travellers staying in endemic areas for more than a few months, particularly during transmission seasons. The mainstay of prevention remains bite avoidance. There is no real indication to vaccinate the majority of travellers to East and Southeast Asia. Children travelling to endemic areas for long periods during the transmission season may be vaccinated.

23.3.2 JE Vaccination

The mouse brain-derived Beijing-1 and Nakayama strain vaccines (Green Cross Vaccine and JE-

VAX[®]) are still used in some endemic countries but most are moving towards either the live attenuated recombinant vaccine (ChimeriVax-JE (IMOJEV[®])) or the Vero cell-derived vaccine that is used in Europe. Children from Australia or S.E. Asia may have received Chimerivax-JE, which contains a live attenuated YFV-17D with the prM/E genes replaced with the corresponding JE virus SA14-142 strain genes. Whilst it is worth being aware of the vaccines a child may have received overseas, IXIARO[®], by Valneva, is the only JE vaccine licenced for Pediatric administration in Europe. IXIARO[®] contains the SA14-14-2 strain of JE virus, produced in Vero cells and inactivated. It is licenced from 2 months of age.

The IXIARO[®] course consists of two doses given 1 month apart. The primary course should be completed at least 7 days before exposure. Children aged 2 months to 3 years receive 0.25 ml doses, and children aged above 3 years receive the adult doses of 0.5 ml. The manufacturer reports seroconversion in 85–100% of Pediatric recipients at 6 months. No long-term seroprotection data has been generated for children, but adults demonstrate continued protection for up to 3 years. As such, a booster dose is recommended within the second year if continued protection is required.

There is no data on interrupted schedules in children, but Japanese encephalitis vaccine is

highly immunogenic, and evidence from adults suggests that it is unnecessary to repeat the first dose after a schedule delay.

There was a relatively high risk of anaphylaxis with the mouse brain-derived JE vaccine and several contraindications, including that it was not to be used in children with neurological conditions. The Vero cell-derived, inactivated vaccine is usually well tolerated.

23.4 Cholera

Cholera is a bacterial disease caused by infection with toxigenic *Vibrio cholerae*. The main features of cholera are the result of the release of cholera toxin, which binds to the intestinal cells and causes the efflux of ions and water into the bowel lumen that leads to watery diarrhoea.

Only cholera serogroups 01 or 0139 produce toxin and thus cause epidemic disease. There are two biotypes of serogroup 01 – Classical and El Tor (which is further divided into Inaba, Ogawa and Hikojima).

Cholera is characterised by the sudden onset of profuse, watery stools with occasional vomiting. The incubation period is usually between 2 and 5 days but may be only a few hours. In severe cases, dehydration, metabolic acidosis and circulatory collapse may follow rapidly. Untreated, more than 50% of severe cases die within a few hours of onset. However, with prompt, correct treatment, mortality is less than 1%.

Cholera is caused by consuming cholera-contaminated food or water, typically present in countries with poor sanitation and food hygiene, worldwide.

Cholera outbreaks still occur in many low-income countries and particularly during humanitarian crises. Children in the 2–4 year age group are particularly affected.

23.4.1 Cholera Vaccination Requirements

The mainstay of cholera prevention is food and water hygiene. Vaccination is rarely necessary for travellers. Children travelling to remote areas with epidemic cholera and limited access to basic medical care can be considered for vaccine.

Cholera is similar in structure to some strains of *E.coli*. The cholera vaccine also has limited pro-

tective effect against heat-labile enterotoxin-producing *E.coli*, one of the many causes of travellers' diarrhoea. The WC/rCBt oral vaccine is licenced in some countries for preventing ETEC diarrhoea, but it should generally not be used for this purpose in travellers. The duration of protection against ETEC is <3 months.

The licenced vaccine in Europe is Dukoral®. It is an inactivated oral vaccine. The vaccine is supplied as granules, and a separate bicarbonate buffer suspension, which protects the vaccine from destruction by gastric acid. The vaccine contains recombinant cholera toxin B subunits plus the following strains of inactivated bacteria:

- *Vibrio cholerae* Inaba 01 classical biotype
- *Vibrio cholerae* 01 Inaba El Tor biotype
- *Vibrio cholerae* 01 Ogawa classical biotype

As such it confers protection against serogroup 01 only. It does not protect against serogroup 0139 or any other vibrio species.

The Dukoral® vaccine is licenced in Europe for children aged over 2 years. Since immunity is mediated by intestinal mucosal IgA, serological tests may not fully reflect immunity. The reported protective efficacy against serogroup 01 cholera is around 68%, but this begins to wane quickly, after around 6 months (28). The wane is faster in infants. For continuous protection, therefore, a single booster dose is recommended 2 years after completing the primary course for children over 6 years of age and 6 months after completing the primary course for children aged 2–6 years.

The primary course must be given 1–6 weeks apart. If more than 6 weeks elapse between doses, the primary course should be recommended.

There are a number of vaccines developed for immunisation against cholera, and most are oral. The least costly vaccine, used in endemic regions, is the bivalent 01 and 0139 whole-cell oral vaccine by Shanchol. This vaccine does not contain the cholera toxin B subunit and has an overall efficacy of about 52% during the first year and 62% in the second year, associated with minimal side effects.

23.4.2 Cholera Vaccine Side Effect Profile and Contraindications

Mild gastrointestinal symptoms (abdominal pain, cramping, diarrhoea and nausea) are commonly reported adverse effects associated with the vaccine.

Vaccine administration should be delayed in the event of an acute gastrointestinal or febrile illness.

The vaccine contains approximately 1.1 g of sodium per dose, and this can make it unsuitable for children with nephrotic syndrome or those who take a low-sodium diet for other reasons.

23.5 Typhoid

Salmonella typhi and *paratyphi* A/B/C are serotypes of the gram-negative bacteria *Salmonella enterica*. Travellers are infected by ingestion of contaminated food and water or by direct faeco-oral transmission in areas of poor sanitation.

The signs of enteric fever range from headache, myalgia, nausea and abdominal pain with constipation or diarrhoea to fever and sepsis with intestinal perforation and GI haemorrhage. Children may experience severe disease including meningitis and encephalopathy, presenting as seizures. Generally, severe disease (typhoid fever) is associated with *S. typhi* infection.

The majority of cases occur in Asia. The disease is endemic throughout Africa and South America but is rarely diagnosed in travellers from these two continents. The areas of highest incidence are India, Bangladesh and Pakistan. Seventy-eight percent of infections are in those who return to their birth countries to visit friends and relatives (VFRs). This highlights the importance of vaccinating travellers to these countries, targeting children of VFR parents in particular.

The attack rates are up to 478/100,000 annually in school-age children and 358/100,000 annually for 2–4-year-olds in high-risk areas. In travellers the estimated infection rates are 1–10 per 100,000 travellers to the high typhoid-burden countries such as India, Bangladesh and Pakistan.

23.5.1 Typhoid Vaccination

Typhoid vaccination is often recommended for travellers to areas of South Asia. For low- and moderate-risk areas such as sub-Saharan Africa and South America, vaccination is not recommended.

The current licenced vaccines do not offer significant protection against *S. paratyphi*, and none of the licenced vaccines are suitable for infants and children under 2 years of age. All travellers should be advised on personal and food hygiene to help reduce infection risk.

23.5.2 Typhoid Vaccines

The inactivated whole-cell typhoid vaccine provided around 65–70% protection but caused strong adverse reactions, and its use has long been discontinued in Europe.

There is now a live oral vaccine, and two Vi capsular polysaccharide vaccines licenced for *Salmonella typhi*. They offer some protection but they have multiple shortcomings (■ Table 23.3).

The oral vaccines contain a live attenuated lyophilized TY21a strain that may be more immunogenic, particularly enhancing mucosal immunity. The oral vaccine administered with bicarbonate buffer in three doses over several days has an efficacy of up to 50% in the first 2 years after vaccination. Since oral TY21a is a live vaccine, it is avoided in immunocompromised children, and the use of any concomitant antibiotics will affect its efficacy.

Vi is the virulence factor and protective antigen in *Salmonella typhi*. The polysaccharide vaccines are made of purified Vi polysaccharide from the Ty2 *Salmonella typhi* strain. They are single-dose vaccines. Neither elicits protective responses

■ Table 23.3 Typhoid vaccines currently in use

Vaccine type	Immunogenic constituents	Minimum age	Vaccine trade names	Duration of protection
Live attenuated oral vaccine	Ty21a	6 years	Vivotif® (Crucell) ZeroTyph (Boryung)	3 years (after 3 doses)
Parenteral vaccine	Vi polysaccharide	2 years (1 year if benefit outweighs risk)	Typhim Vi® (Sanofi Pasteur) Typherix® (GlaxoSmithKline)	3 years

in children under 2, as expected for a polysaccharide vaccine in this age group. In older children it provides protection of approximately 60%. It is recommended for three yearly boosting, but anti-Vi IgG titres have been shown to decline well before that time period.

Vi conjugate vaccines are in development. One VI-TT conjugate vaccine, coupled to tetanus toxoid, has been used in India, but there is little data on efficacy. New conjugate vaccines such as the Vi-CRM 197 and the Vi-rEPA have demonstrated significantly increased immunogenicity with good safety profiles. This new generation of typhoid conjugate vaccine may provide practitioners with a vaccine that is suitable for travellers, particularly for the high-risk group of pre-school children.

23.6 Vaccines with No Current Indications for Travellers

23.6.1 Dengue

Dengue is one of the worlds' most common infectious diseases, endemic to more than 110 countries (see [Map D](#)). The dengue virus, which belongs to the *Flaviviridae* family, is the etiologic

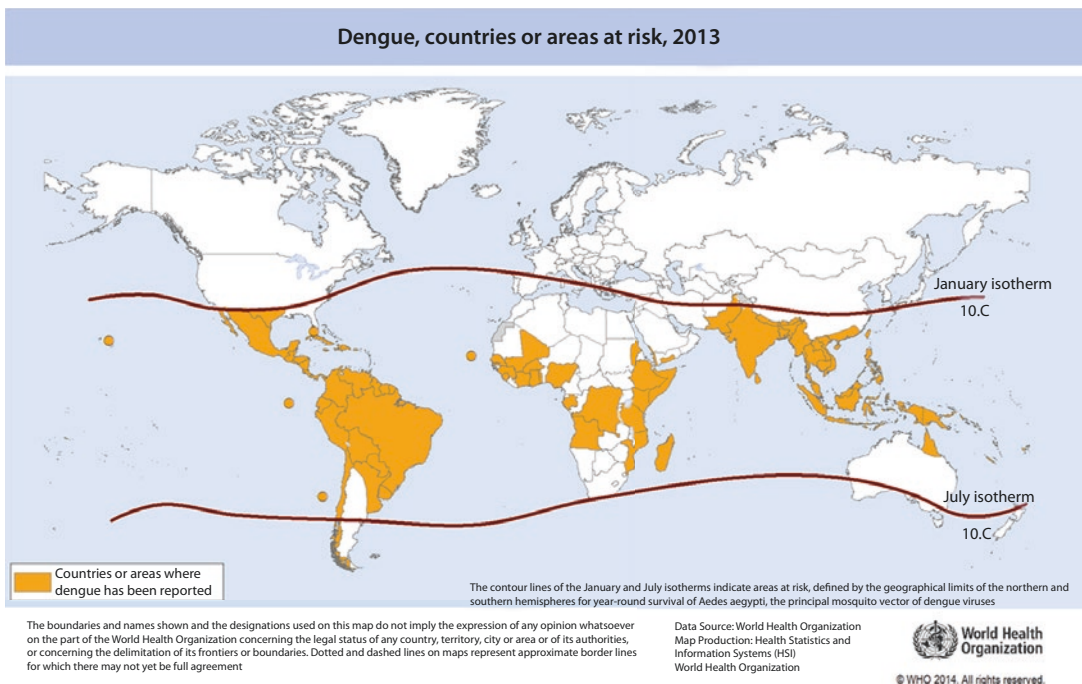
agent responsible for the broad spectrum of dengue symptoms and signs that range from mild fever (DF) to dengue haemorrhagic fever and dengue shock syndrome.

There are four dengue serotypes, DEN-1, DEN-2, DEN-3 and DEN-4, and any useful vaccine would need to provide protection to all four serotypes.

Dengue is a single-stranded RNA virus that encodes three structural proteins (capsid protein C, pre-membrane protein (prM) and envelope protein E), plus seven nonstructural proteins (NS1, NS2a, NS2b, NS3, NS4a, NS4b and NS5). The envelope protein (protein E) has the main epitopes for the production of neutralising antibody and is therefore considered the best target for vaccine development.

Dengue Vaccines

Dengvaxia[®], a live, attenuated, tetravalent vaccine developed by Sanofi Pasteur, is the first licenced vaccine for dengue prevention. It is available in several endemic countries where dengue is a leading cause of child mortality. Dengvaxia is based on a genetically engineered live attenuated YF virus, whereby the prM and E genes from each of the four dengue serotypes are substituted into the



backbone of the yellow fever virus, 17D vaccine strain.

Two large phase III efficacy trials conducted in endemic areas of Latin America and Asia showed the efficacy of the vaccine to vary from 77.7% for serotype 4, 74.0% for serotypes 2 and 3 to 42.3% for serotype 1. In the Pacific region, the overall efficacy is just 56.5%, with the greatest impact being in the prevention of severe dengue and hospitalisation.

A long-term follow-up study of children between 2 and 16 years of age in the Asia-Pacific and Latin American regions demonstrated increased and unexplained hospitalisations for severe dengue among children under 9 years.

Several other dengue vaccine candidates are in clinical trials, and multiple strategies have been exploited for vaccine development

Tetavalent inactivated vaccines appear to be safer but may have low immunogenicity across the four serotypes. A tetavalent dengue virus purified inactivated vaccine (TDEN PIV) strengthened by adjuvants is currently being evaluated by GlaxoSmithKline in a phase I clinical study.

A DNA vaccine candidate using prM and E protein DNA of all four serotypes can generate protective antibodies against all four dengue serotypes. Early trials are ongoing.

The NS1 surface protein is the other potential vaccine candidate. Passive immunisation with anti-NS1 antibodies prevented lethal dengue disease in a mouse model. As such, several strategies for NS1-based vaccines are under investigation.

Previous concerns regarding antibody-dependent enhancement effects and activity of cross-reactive T cells during repeat dengue infection have somewhat slowed the development of a safe and effective dengue vaccine.

23.6.2 Malaria

Malaria causes over 450,000 deaths per annum in sub-Saharan Africa. *Plasmodium falciparum* accounts for more than 90% of these deaths. Severe malaria and mortality peaks in the under-fives, particularly under twos.

Infants and children in Africa typically suffer multiple episodes of severe clinical malaria before developing a degree of immunity. It is unclear whether sterile immunity ever occurs in adults, as opposed to immune tolerance.

A vaccine is needed, primarily for the use in children living in endemic countries.

Antibodies to antigens present on malaria sporozoites such as the circumsporozoite protein (CSP) have been demonstrated to prevent sporozoites migrating to liver cells, and thus a vaccine targeted to CSP could theoretically stop a pre-erythrocytic infection from developing in the liver. Alternatively merozoite surface antigens or gametocyte surface antigens could be targeted to reduce multiplication in the blood stage. Combination vaccines acting on more than one stage of the parasites life cycle could induce broader immune responses.

Malaria Vaccines

There are currently more than 30 malaria vaccines in preclinical or clinical trials and we discuss two vaccines, one of which is licenced and the other shows a mechanism for complete protection.

RTS,S/AS01 Vaccine

RTS,S/AS01 (Mosquirix[®]) is the most advanced malaria vaccine candidate.

It has been developed by GSK and collaborators, within a public private partnership, to produce a vaccine for Africa. RTS,S acts on the pre-erythrocytic stage of malaria. It is a recombinant hybrid where portions of the CSP protein are fused to hepatitis B surface antigen and co-expressed in yeast. RTS,S virus-like particles are formed when the fusion protein is expressed within yeast cells. AS01 is the adjuvant, made of immunomodulatory molecules and liposomes. The vaccine is given by IM injection and has been evaluated in trials using a 0-, 1-, 2-month schedule, with a booster at 18 months after the third dose.

Clinical trials have demonstrated safety and immunogenicity. They also importantly demonstrated non-inferiority of hepatitis B immunity compared to Engerix-B vaccination and no deleterious effect on other co-administered vaccines responses. A double-blind, randomised controlled trial, conducted in seven African countries (Burkina Faso, Gabon, Ghana, Kenya, Malawi, Mozambique and Tanzania) from 2009–2011, showed that the three-dose primary schedule of RTS,S reduced clinical malaria cases by 28% in children and 18% in infants, over at least 3 years after vaccination. A booster dose of RTS,S administered 18 months after the primary schedule further reduced the number of cases of clinical

malaria in children (aged 5–17 months at first vaccination) by 36% and in infants (aged 6–12 weeks at first vaccination) by 26% to the end of the study, with average follow-up of 48 and 38 months, respectively.

Although very promising, protection decreased over time in both age groups. There was also an increased risk of severe malaria and malaria mortality and a significant disproportionate increase in overall mortality in children who did not receive the booster dose. There is also concern that the RTS,S/AS01 vaccination may prevent the normal development of malaria immunity, and malaria interventions in young children might lead to rebound morbidity and mortality in older age groups.

The European Medicines Agency gave a positive scientific opinion for RTS,S, but the vaccine is not being studied in travellers at present. The question of whether children who received a malaria vaccine would be at higher risk after a period of non-exposure, or without boosters, than malaria-naïve children is still unanswered. It is unlikely ever to be used in travellers, as the efficacy would be suboptimal.

Sporozoite Vaccine

Vaccines against the sporozoite are very attractive as the infecting dose is small (~6 sporozoites), and there is little chance of selecting escape mutants. However, the sporozoite is a complex organism that develops in the mosquito, and the in vitro production of the sporozoite for the use as an antigen has yet to be achieved. The vaccine appears safe and trials continue. Whilst it is likely to be an unattractive and expensive choice for short-stay travellers, it could appeal to long-term expatriate travellers.

Further Reading

- Bhutta Z, et al. Immunogenicity and safety of the v-CRM197 conjugate vaccine against typhoid fever in adults, children, and infants in south and southeast Asia: results from two randomised, observer-blind, age de-escalation, phase 2 trials. *Lancet Infect Dis.* 2014;14(2):119–29.
- CDC. Grading of recommendations, assessment, development, and evaluation (GRADE) for yellow fever vaccine booster doses. Advisory Committee on Immunization Practices. CDC. 2015.
- de Menezes MR, Fernandes Leal Mda L, Homma A. Serious adverse events associated with yellow fever vaccine. *Hum Vaccin Immunother.* 2015;11(9):2183–7.
- Gautret P, Parola P. Rabies vaccination in travelers: a global perspective. *J Travel Med.* 2012;19(6):395–6.
- Gautret P, Shaw M, Gazin P, Soula G, Delmont J, Parola P, et al. Rabies postexposure prophylaxis in returned injured travelers from France, Australia, and New Zealand: a retrospective study. *J Travel Med.* 2008;15(1):25–30.
- Gotuzzo E, Yactayo S, Cordova E. Efficacy and duration of immunity after yellow fever vaccination: systematic review on the need for a booster every 10 years. *Am J Trop Med Hyg.* 2013;89(3):434–44.
- Hills SL, Griggs AC, Fischer M. Japanese encephalitis in travelers from non-endemic countries, 1973–2008. *Am J Trop Med Hyg.* 2010;82(5):930–6.
- Japanese Encephalitis Vaccines: WHO position paper – February 2015. WHO; February 2015.
- Kossaczka Z, Lin FY, Ho VA, NTT T, Bay PV, Thanh TC, et al. Safety and immunogenicity of vi conjugate vaccines for typhoid fever in adults, teenagers, and 2- to 4-year-old children in Vietnam. *Infect Immun.* 1999;67(11):5806–10.
- Liu Y, Liu J, Cheng G. Vaccines and immunization strategies for dengue prevention. *Emerg Microbes Infect.* 2016;5(7):e77.
- Malerczyk C, Vakil HB, Bender W. Rabies pre-exposure vaccination of children with purified chick embryo cell vaccine (PCECV). *Hum Vaccin Immunother.* 2013;9(7):1454–9.
- Monath TP. Review of the risks and benefits of yellow fever vaccination including some new analyses. *Expert Rev Vaccines.* 2012;11(4):427–48.
- Morbidity and Mortality Weekly Report: Human Rabies Prevention – United States, 2008. Recommendations of the Advisory Committee on Immunization Practices [press release]. CDC: 2008.
- Roukens AH, Vossen AC, van Dissel JT, Visser LG. Reduced intradermal test dose of yellow fever vaccine induces protective immunity in individuals with egg allergy. *Vaccine.* 2009;27(18):2408–9.
- RTS,S Clinical Trials Partnership. Efficacy and safety of RTS,S/AS01 malaria vaccine with or without a booster dose in infants and children in Africa: final results of a phase 3, individually randomised, controlled trial. *Lancet.* 2015;368(9988):31–45.
- Seder RA, et al. Protection against malaria by intravenous immunization with a nonreplicating sporozoite vaccine. *Science.* 2013;341(6152):1359–65.
- Shlim DR, Solomon T, Ericsson CD, Steffen R. Japanese encephalitis vaccine for travelers: exploring the limits of risk. *Clin Infect Dis.* 2002;35(2):183–8.
- Steinberg EB, Bishop R, Haber P, Dempsey F, Hoekstra M, Nelson JM, et al. Typhoid fever in travelers: who should be targeted for prevention? *Clin Infect Dis.* 2004;39:186–91.
- Sur D, et al. Efficacy and safety of a modified killed-whole-cell oral cholera vaccine in India: an interim analysis of a cluster-randomised, double-blind, placebo-controlled trial. *Lancet.* 2009;374(9702):1694–702.
- Szu S. Development of vi conjugate – a new generation of typhoid vaccine. *Expert Rev. Vaccines.* 2013;12(11):1273–86.
- WHO. World Health Organisation, Rabies Fact Sheet. <http://www.who.int/rabies/en/>. WHO; 2014 (updated 2016).
- Winge A, Ritter V, Coelho-Amaral P, Traiber C. Infant meningoencephalitis probably caused by yellow fever vaccine virus transmitted via breastmilk. *J Pediatr.* 2011;87(3):269–72.

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24.1 Group B Streptococcus Vaccines

24.1.1 Burden of Disease

Group B streptococcus (GBS) is well recognized as a cause of early neonatal infection in high-income countries (HICs), with long-term adverse neurodevelopmental outcomes in up to 50% of survivors of GBS meningitis. A recent meta-analysis reported an overall global estimate of GBS incidence of 0.53 per 1000 live births and a mean case fatality ratio of 9.6% (95% CI 7.5–11.8). There is, however, significant variation in the estimated incidence of GBS disease between and within global regions.

Many HICs have introduced intrapartum antibiotic prophylaxis (IAP) strategies in order to reduce the burden of neonatal early-onset GBS disease (EOD). Since the introduction of IAP policies in the USA, rates of EOD have declined from 1.7 per 1000 live births in the 1990s to 0.34–0.37 per 1000 live births in 2014.

24.1.2 Epidemiology

Group B streptococcus can colonize the vagina and gastrointestinal tract of pregnant women and be transmitted vertically to their babies during delivery. Up to 30% of women carry GBS in the vagina or rectum without it causing symptoms. Vertical transmission occurs in 15–50% of infants born to a colonized mother. Although the majority of such babies will not go on to develop invasive disease, maternal colonization is a prerequisite for EOD (days 0–6 of life) and a significant risk factor for late-onset disease (LOD) (days 7 to 89). Overall, EOD accounts for approximately 60–70% of all neonatal GBS disease, depending on the use of IAP in the population. LOD also results from vertical transmission from a colonized mother, but nosocomial transmission, breast milk, and community sources are also recognized.

24.1.3 GBS Vaccines

Given the very early onset of neonatal GBS disease and the shortcomings of IAP-based prevention strategies, there is considerable interest in developing an effective antenatal vaccine. The major-

ity of work in this field over the last 30 years has focused on the development of a vaccine based on capsular polysaccharide (CPS), in part reflecting the success of this approach for other encapsulated bacteria such as *Streptococcus pneumoniae*, but also by the demonstration, initially by Baker et al. in 1976, of the association between GBS ST-specific capsular antibody concentrations and invasive GBS disease in newborns. In the USA and Europe, GBS STs causing invasive disease are predominantly STs Ia, Ib, II, III, and V.

CPS-protein conjugate vaccines against all relevant ST have been assessed in healthy, non-pregnant women and demonstrated satisfactory immunogenicity and safety. More recently, conjugate vaccines have been developed based on tetanus toxoid and CRM197 as the carrier proteins. Studies in pregnant women have also established the immunogenicity and safety of these candidates. A recent study of a trivalent CRM-conjugate vaccine involving 470 pregnant women demonstrated a satisfactory reactogenicity profile, immunogenicity, and antibody transfer to the infant. A pentavalent conjugate vaccine is now proposed, with estimated coverage of >90% of global disease-causing serotypes.

The use of CPS conjugate vaccines is not without its drawbacks, including cost, limited strain coverage, and, potentially, serotype replacement. One way of overcoming these limitations is to develop a vaccine based on highly conserved surface proteins. A phase I trial of a protein vaccine incorporating Rib and Alpha C surface proteins is currently underway (► [Clinicaltrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT02459262): NCT02459262). The use of “reverse vaccinology” has also identified four proteins that could be the basis of a “universal” vaccine: the pilus proteins and the Sip protein.

Several obstacles exist in moving the most advanced vaccine into phase III clinical trials. Given the relative rarity of GBS disease in Europe and the USA, large numbers of infants would need to be recruited to determine vaccine efficacy. Efficacy trials are likely to be needed because it is not currently known what concentration of antibody is required to protect infants. However, generation of robust data supporting serological correlates of protection could facilitate the licensure of a GBS vaccine without the need for large-scale prelicensure efficacy trials in pregnant women.

24.2 Cytomegalovirus (CMV)

24.2.1 CMV Vaccines

Cytomegalovirus (CMV) is the most common cause of congenital infection globally and can be associated with significant sequelae in affected infants. Congenital CMV infection is the leading nongenetic cause of sensorineural hearing loss (SNHL), the only potentially treatable cause, and is associated with neurodevelopmental delay.

CMV infection is usually asymptomatic or associated with mild, transient symptoms in immunocompetent children and adults; however, in immunocompromised individuals, primary or reactivated virus can cause substantial morbidity.

Transmission of CMV to the fetus can occur following primary maternal infection, reactivation, or reinfection with a different strain. The rate of transmission to infants born to women with primary CMV infection is substantially higher, 32%, compared to those infants born to women with reactivation, 1.4%. The global birth prevalence of cCMV is 0.64%, with significant variation. The total prevalence represents the sum of transmission following primary infection and reactivation during pregnancy.

Around 10–15% of congenitally infected infants (cCMV) will have symptoms at birth. Clinical features of cCMV seen in the majority (>50%) of symptomatic infants include petechiae, jaundice, hepatosplenomegaly, microcephaly, intrauterine growth retardation, elevated ALT, and low platelets. Features observed less frequently include chorioretinitis, optic atrophy, purpura, and seizures. The most common finding on neuroimaging is intracranial calcification, with some infants also demonstrating ventricular dilatation, cysts, and lenticulostriate vasculopathy. Most symptomatic infants (40–60%) will experience adverse neurodevelopmental outcomes, such as cerebral palsy, cognitive impairment, and SNHL.

About 10–15% of infants with cCMV who are not symptomatic at birth will still develop SNHL, which in some is progressive.

Prevention of congenital CMV (cCMV) infection is the main driver of CMV vaccine development and therefore is the focus here.

24.2.2 Vaccines

CMV vaccine development has a relatively long history, starting in the 1970s with live-attenuated vaccines. The live-attenuated CMV vaccines were associated with only mild injection site reactions and no systemic reactions and induced antibodies at similar concentrations to natural infection, and no excretion of virus was detected. The laboratory strains of CMV lost the ULb' region of the genome during the multiple cell culture passages, therefore losing genes permitting entry into epithelial cells. Therefore, these vaccines did not elicit high concentrations of antibody that prevent viral entry into cells, and a clinical trial of women with young children in childcare showed that vaccination did not prevent primary or secondary infection.

In the 1980s and 1990s, recombinant subunit vaccines incorporating CMV surface glycoprotein B (gB), adjuvanted with MF59, were first developed and tested. The vaccine-induced gB antibody is thought to be important for prevention of viral entry into fibroblasts. The vaccine was well tolerated and immunogenic, more so in infants than in adults, and induced antibody responses of higher magnitude than natural infection; however, immunity quickly waned. The protection afforded to adolescent girls was at best modest, up to 45%. Similar results were also observed in CMV seronegative women with vaccine efficacy of 50%.

Other candidates in the vaccine pipeline include CMV DNA vaccines that contain both gB and pp65, another surface protein. The pp65 protein is an abundant protein in CMV virions and is a major target of the T-cell responses to CMV. One such vaccine, CyMVectin, is in late preclinical development and is intended to induce immunity prior to pregnancy in order to prevent cCMV.

Most recently, there has been interest in vaccines containing more immunogens. CMV has a pentameric gH/gL/UL128-UL130-UL131 complex on its surface that is critical to viral entry and is an important target of the neutralizing antibody response in seropositive individuals. A recent study has shown an association between antibodies to this pentamer and prevention of transmission of primary CMV from mother to fetus. With these recent findings, there is renewed hope for a vaccine candidate that will have clinical efficacy.

24.2.3 Other Issues

The immune correlates of protection against cCMV have not fully been elucidated, including the contribution of humoral and cellular immunity to maternal-fetal transmission. Determining relevant endpoints in clinical trials to support vaccine licensure is critical, given that immune correlates remain elusive and clinical endpoints are required, such as cCMV infection. Such trials necessitate very large sample sizes and long follow-up to achieve sufficient statistical power and will be costly. Optimizing the protective efficacy of CMV vaccines in both seronegative and seropositive individuals is critical, since a significant number of infants with cCMV are born to women with preexisting CMV antibody.

A further significant issue is the timing of vaccination. A vaccine should be administered prior to pregnancy to ensure immunity before the first trimester; however, many pregnancies are not necessarily planned and women do not necessarily seek preconception healthcare. Vaccinating adolescents is an alternative; however, persistence of immunity into reproductive years may be challenging, and a vaccine would need to be effective in both seronegative and seropositive females. Another possibility is to vaccinate in early childhood. Vaccinating as part of the routine infant immunization program would ensure high coverage; could prevent infection prior to first encounter, thereby overcoming the problems of immunity in seropositive individuals; and would interrupt viral circulation by preventing prolonged shedding of CMV in the urine and saliva of infected toddlers. This age group is the most common source of infection to pregnant women and therefore would afford protection to the mothers or caregivers of young children. Modeling suggests that a combination strategy may be preferable.

24.3 Respiratory Syncytial Virus (RSV)

24.3.1 RSV Vaccines

Globally, respiratory syncytial virus (RSV) is a major cause of infant morbidity and mortality. Although natural infection offers limited protection against reinfection, passive immunization

with RSV-specific antibody can protect infants against severe lower respiratory tract disease. This suggests that vaccine-induced antibody could benefit infants and therefore supports maternal vaccination as a potential strategy for protecting young infants. The duration of such protection will dictate the need for additional infant vaccines. For high-risk groups a strategy of passive immunization with monoclonal antibodies continues to be an important approach.

24.3.2 RSV Disease

RSV in some infants causes bronchiolitis (inflammation of the lower airways). This can be a significant respiratory illness which causes hospitalizations, respiratory failure, and death; around 99% of such deaths occur in low- and middle-income countries (LMIC). There is an estimated 34 million cases of RSV-associated acute lower respiratory tract infection in infants under 5 years of age every year (22% of ALRI episodes), 3.4 million of which are severe, and RSV is estimated to account for 3–9% of all fatal lower respiratory tract infections in infants.

RSV infection is seasonal in most countries; outbreaks occur most frequently in the cold season in areas with temperate and Mediterranean climates and in the wet season in tropical countries with seasonal rainfall. Young age is the major risk factor for RSV disease, peaking at about 8–12 weeks of age and becoming lower after 6 months of age. A number of genetic and environmental factors combine with the age of the infant to increase risk of severe RSV disease. Certain groups are known to be at high risk from more severe RSV infections: prematurity, bronchopulmonary dysplasia, congenital heart disease, immunodeficiency, cerebral palsy, and Down's syndrome. Nevertheless, the majority of emergency admissions occur in otherwise healthy infants born at term.

24.3.3 Vaccines

Support for a vaccine strategy comes from studies using monoclonal RSV antibody preparations (palivizumab, motavizumab) which are able to protect infants against severe lower respiratory disease. Indeed these are currently the only means of protecting high-risk infants against

RSV and are recommended in many countries for specific groups of infants, albeit with different strategies.

Vaccine development for RSV stalled for several decades as a result of a trial in the 1960s with a formalin-inactivated RSV vaccine. This resulted in enhanced, including fatal, cases of RSV disease in vaccine recipients. However the field is now flourishing; a number of candidates are under development, and several have reached phase 3 trials. Vaccine strategies being considered for protecting infants include infant vaccination, maternal vaccination, and vaccinating contacts of infants in order to prevent transmission. Maternal immunization, which aims to provide protection to the infant by boosting the levels of transplacental antibody, is the leading strategy.

The most advanced candidates are subunit vaccines containing purified RSV fusion (F) protein. One pregnancy trial has been completed; vaccinees and their infants had a fourfold rise in serum RSV IgG concentrations and breast milk RSV-specific IgA and IgG concentrations were also boosted. A phase II clinical trial in women of childbearing age with an F protein subunit vaccine recently showed that both neutralizing and palivizumab-competing antibodies could be induced in this population. Furthermore, a significant reduction in RSV infections was demonstrated.

Vaccines designed for infants need to overcome the difficulties of generating a protective response at this age and the theoretical risks associated with generating an inappropriate response. Current candidates include gene-based vector vaccines (adenovirus) and particle-based and live-attenuated vaccines. For infants in LMIC, where children up to 5 years of age continue to suffer severe RSV disease, boosting maternal antibody alone may not be the most effective strategy to fully protect infants. The development of an effective pediatric vaccine will therefore become a necessary part of a complete prevention strategy.

Further Reading

Adler SP, Starr SE, Plotkin SA, Hempfling SH, Buis J, Lou Manning M, et al. Immunity induced by primary human cytomegalovirus infection protects against secondary infection among women of childbearing age. *J Infect Dis.* 1995;171:26–32. doi:10.1093/infdis/171.1.26.

- Baker CJ, Kasper DL. Correlation of maternal antibody deficiency with susceptibility to neonatal group B streptococcal infection. *N Engl J Med.* 1976;294(14):753–6.
- Baker CJ, Rench MA, McInnes P. Immunization of pregnant women with group B streptococcal type III capsular polysaccharide-tetanus toxoid conjugate vaccine. *Vaccine.* 2003;21(24):3468–72.
- Bernstein DI, Munoz FM, Callahan ST, Rupp R, Wootton SH, Edwards KM, et al. Safety and efficacy of a cytomegalovirus glycoprotein B (gB) vaccine in adolescent girls: a randomized clinical trial. *Vaccine.* 2016;34:313–9. doi:10.1016/j.vaccine.2015.11.056.
- Brodeur BR, Boyer M, Charlebois I, Hamel J, Couture F, Rioux CR, et al. Identification of group B streptococcal sip protein, which elicits cross-protective immunity. *Infect Immun.* 2000;68(10):5610–8.
- Dollard SC, Grosse SD, Ross DS. New estimates of the prevalence of neurological and sensory sequelae and mortality associated with congenital cytomegalovirus infection. *Rev Med Virol.* 2007;17:355–63. doi:10.1002/rmv.544.
- Fouts AE, Chan P, Stephan JP, Vandlen R, Feierbach B. Antibodies against the gH/gL/UL128/UL130/UL131 complex comprise the majority of the anti-cytomegalovirus (anti-CMV) neutralizing antibody response in CMV hyperimmune globulin. *J Virol.* 2012;86:7444–7. doi:10.1128/JVI.00467-12.
- Glenn GM, Fries LF, Thomas DN, Smith G, Kpamegan E, Lu H, et al. A randomized, blinded, controlled, dose-ranging study of a respiratory syncytial virus recombinant fusion (F) nanoparticle vaccine in healthy women of childbearing age. *J Infect Dis.* 2016;213(3):411–22.
- Hall CB, Weinberg GA, Iwane MK, Blumkin AK, Edwards KM, Staat MA, et al. The burden of respiratory syncytial virus infection in young children. *N Engl J Med.* 2009;360(6):588–98.
- Heath PT, Schuchat A. Perinatal group B streptococcal disease. *Best Pract Res Clin Obstet Gynaecol.* 2007;21(3):411–24.
- Heyderman RS, Madhi SA, French N, Cutland C, Ngwira B, Kayambo D, et al. Group B streptococcus vaccination in pregnant women with or without HIV in Africa: a non-randomised phase 2, open-label, multicentre trial. *Lancet Infect Dis.* 2016;16(5):546–55.
- Kapikian AZ, Mitchell RH, Chanock RM, Shvedoff RA, Stewart CE. An epidemiologic study of altered clinical reactivity to respiratory syncytial (RS) virus infection in children previously vaccinated with an inactivated RS virus vaccine. *Am J Epidemiol.* 1969;89(4):405–21.
- Lambert L, Sagfors AM, Openshaw PJ, Culley FJ. Immunity to RSV in early-life. *Front Immunol.* 2014;5:466.
- Lanzieri TM, Bialek SR, Ortega-Sanchez IR, Gambhir M. Modeling the potential impact of vaccination on the epidemiology of congenital cytomegalovirus infection. *Vaccine.* 2014;32:3780–6. doi:10.1016/j.vaccine.2014.05.014.
- Le Doare K, Heath PT. An overview of global GBS epidemiology. *Vaccine.* 2013;31(Suppl 4):D7–12.
- Lillier D, Kabanova A, Revello MG, Percivalle E, Sarasini A, Genini E, et al. Fetal human cytomegalovirus transmission correlates with delayed maternal antibodies to gH/gL/pUL128-130-131 complex during primary

- infection. *PLoS One*. 2013;8:e59863–13. doi:[10.1371/journal.pone.0059863](https://doi.org/10.1371/journal.pone.0059863).
- Maione D, Margarit I, Rinaudo CD, Massignani V, Mora M, Scarselli M, et al. Identification of a universal group B streptococcus vaccine by multiple genome screen. *Science*. 2005;309(5731):148–50.
- Margarit I, Rinaudo CD, Galeotti CL, Maione D, Ghezzi C, Buttazzoni E, et al. Preventing bacterial infections with pilus-based vaccines: the group B streptococcus paradigm. *J Infect Dis*. 2009;199(1):108–15.
- Munoz FM. Respiratory syncytial virus in infants: is maternal vaccination a realistic strategy? *Curr Opin Infect Dis*. 2015;28(3):221–4.
- Munoz FM, Piedra PA, Glezen WP. Safety and immunogenicity of respiratory syncytial virus purified fusion protein-2 vaccine in pregnant women. *Vaccine*. 2003;21(24):3465–7.
- Murray J, Bottle A, Sharland M, Modi N, Aylin P, Majeed A, et al. Risk factors for hospital admission with RSV bronchiolitis in England: a population-based birth cohort study. *PLoS One*. 2014;9(2):e89186.
- Nair H, Nokes DJ, Gessner BD, Dherani M, Madhi SA, Singleton RJ, et al. Global burden of acute lower respiratory infections due to respiratory syncytial virus in young children: a systematic review and meta-analysis. *Lancet*. 2010;375(9725):1545–55.
- Neostrep. Neostrep – development of group B streptococcal vaccine. 2016 [cited 2016 .24/01/16]; Available from: ► <http://www.neostrep.eu/index.html>
- Palivizumab, a humanized respiratory syncytial virus monoclonal antibody, reduces hospitalization from respiratory syncytial virus infection in high-risk infants. The IMPact-RSV Study Group. *Pediatrics*. 1998;102(3 Pt 1):531–7.
- Pass RF, Zhang C, Evans A, Simpson T, Andrews W, Huang M-L, et al. Vaccine prevention of maternal cytomegalovirus infection. *N Engl J Med*. 2009;360:1191–9. doi:[10.1056/NEJMoa0804749](https://doi.org/10.1056/NEJMoa0804749).
- PATH. Respiratory syncytial virus. Vaccine development against a major cause of childhood respiratory illness 2016 [13/9/16]. Available from: ► <http://sites.path.org/vaccinedevelopment/respiratory-syncytial-virus-rsv/>.
- Plotkin S. The history of vaccination against cytomegalovirus. *Med Microbiol Immunol*. 2015;204:247–54. doi:[10.1007/s00430-015-0388-z](https://doi.org/10.1007/s00430-015-0388-z).
- Schrag SJ, Verani JR. Intrapartum antibiotic prophylaxis for the prevention of perinatal group B streptococcal disease: experience in the United States and implications for a potential group B streptococcal vaccine. *Vaccine*. 2013;31(Suppl 4):D20–6.

Norovirus Vaccine

Timo Vesikari

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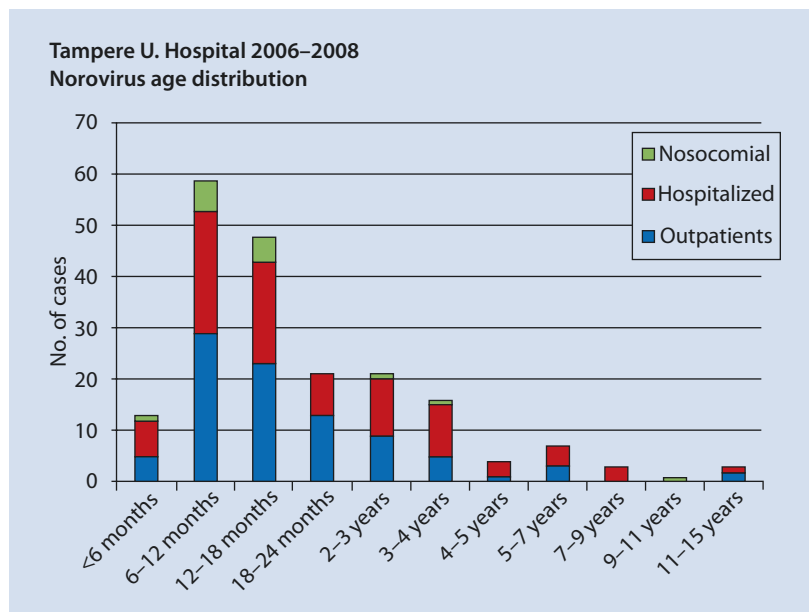
Noroviruses (NoVs) are, after rotaviruses (RV), the second most common causative agents of acute gastroenteritis (GE) in young children. Globally, NoV GE carries significant mortality in children under 5 years of age; one estimate puts the annual death toll at 71,000, which is about one third of that associated with rotavirus (see ► Chap. 11). In industrialized countries deaths in children are rare, but there is significant mortality from NoV GE in the elderly. Therefore, NoV vaccine development is targeted at both children and adults, often with greater emphasis on the latter. In industrialized countries, the burden of NoV disease in children, as measured by severe cases seen in hospital, is about one third to one half of that of RV disease.

Noroviruses were discovered in 1972 by AZ Kapikian of NIH using electron microscopy on stool samples collected from a 1968 outbreak in Norwalk, Ohio. The new virus was called Norwalk virus and later the name was given to all noroviruses. The Norwalk virus is a genogroup I NoV (GI.1). GI NoVs are common in foodborne and waterborne outbreaks of GE, which may occur in any age group. Conceivably, such outbreaks are more difficult to target by vaccination, although some special groups such as cruise ship passengers and military recruits could be targeted.

In contrast, the epidemic NoV GE in children has a predictable seasonal pattern and occurs every winter. The age distribution is similar to that of RV GE (■ Fig. 25.1), and a vaccination approach should be targeted at infants or toddlers at the latest. In epidemic NoV GE, the predominant viruses are of genogroup II, particularly genotype GII.4, which is also the prime candidate for NoV vaccine development. There are at least 22 genotypes of NoVs within genogroup II and 9 genotypes within genogroup I. Repeated NoV infections and episodes of GE occur in young children. The resulting immunity is serotype-specific and not long-lasting. A NoV vaccine could not possibly contain all genotypes, and a changing composition (such as influenza vaccine) would also be difficult. Therefore, it is commonly held that a NoV vaccine should contain both genogroups, but induce cross-protective immunity within the genogroup.

Noroviruses do not grow in normal cell culture and only poorly in explants of gut tissue. Therefore, a live virus is not regarded as an option. Most probable candidate vaccines are NoV virus-like particles (VLPs), which can be produced in baculovirus insect cell system or in plants. VLPs are highly antigenic and may be administered either by injection or mucosally (e.g., intranasally). Only one NoV VLP vaccine has progressed

■ Fig. 25.1 Age distribution of NoV gastroenteritis in Tampere University Hospital 2006–2008 ((Reproduced with permission. Räsänen S, PhD thesis. University of Tampere 2016))



to clinical trials. This vaccine is produced by Takeda (by acquiring LigoCyte). It is a bivalent NoV GI.1 + GII.4 VLP vaccine combined with aluminum adjuvant. The GII.4 component is based on a “consensus” sequence and is not any variant that occurs in nature. The idea is that such a consensus VLP induces a cross-reacting immune response and elicits protection against other GII.4 variants and, possibly, more broadly, against other GII NoVs.

The results of a proof-of-concept challenge trial in adult volunteers are shown in [Table 25.1](#). In this study, the subjects received two doses of Takeda’s candidate NoV VLP vaccine and were challenged on day 42 with a naturally occurring wild-type GII.4 NoV Farmington strain. The results indicate that the vaccine induced partial protection against heterologous challenge. Specifically, there was high-level protection against severe NoV GE, less against mild NoV GE, and no protection against NoV infection.

The NoV challenge study experience resembles the performance of the RV vaccine in that a NoV VLP vaccine seems to prevent severe disease and not NoV infection. This should be seen as a realistic target for a future NoV vaccine in children. A parenteral vaccine given in two or three

doses to infants or toddlers would induce broadly reactive cross-protection against severe NoV, but would not fully prevent NoV infection with mild symptoms. Even so, a successful NoV vaccine would do better than nature.

A new concept is the combination of NoV VLP vaccine with rotavirus VP6. In this combination, RV VP6 would not only protect against RV GE, but also enhance the immune response to NoV, like an adjuvant. Such a combination might become a universal vaccine against childhood GE.

Further Reading

- Atmar RL, Bernstein DI, Harro CD, et al. Norovirus vaccine against experimental human Norwalk virus illness. *N Engl J Med.* 2011;365:2178–87.
- Bernstein DI, Atmar RL, Lyon GM, et al. Norovirus vaccine against experimental human GII.4 virus illness: a challenge study in healthy adults. *J Infect Dis.* 2015;95:2734–47.
- Blazevic V, Lappalainen S, Nurminen K, Huhti L, Vesikari T. Norovirus VLPs and rotavirus VP6 protein as combined vaccine for childhood gastroenteritis. *Vaccine.* 2011;29(45):8126–33.
- Blazevic V, Malm M, Arinobu D, Lappalainen S, Vesikari T. Rotavirus capsid VP6 protein acts as an adjuvant in vivo for norovirus virus-like particles in a combination vaccine. *Hum Vaccin Immunother.* 2016;12(3):740–8.
- Kapikian AZ, Wyatt RG, Dolin R, Thornhill TS, Kalica AR, Chanock RM. Visualization by immune electron microscopy of a 27-nm particle associated with acute infectious nonbacterial gastroenteritis. *J Virol.* 1972;10:1075–81.
- Lopman BA, Hall AJ, Currs AT, Parashar UD. Increasing rates of gastroenteritis hospital discharges in US adults and the contribution of norovirus, 1996–2007. *Clin Infect Dis.* 2011;52:466–72.
- Pang XL, Joensuu J, Vesikari T. Human calicivirus-associated sporadic gastroenteritis in Finnish children less than two years of age followed prospectively during a rotavirus vaccine trial. *Pediatr Infect Dis J.* 1999;18:420–6.
- Prasad BV, Rothnagel R, Jiang X, Estes MK. Three dimensional structure of baculovirus-expressed Norwalk virus capsids. *J Virol.* 1994;68:5117–25.
- Räsänen S, Lappalainen S, Salminen M, Huhti L, Vesikari T. Noroviruses in children seen in a hospital for acute gastroenteritis in Finland. *Eur J Pediatr.* 2011;170(11):1413–8.
- Vesikari T, Blazevic V. Norovirus vaccine: one step closer. *J Infect Dis.* 2015;211:853–5.

Table 25.1 Results of a norovirus GI/GII VLP vaccine (Takeda) challenge study

Outcome	Protection	<i>p</i>
Severe vomiting/diarrhea	100%	0.054
Moderate to severe vomiting/diarrhea	68%	0.068
Any vomiting/diarrhea	47%	0.074
Infection	14%	0.420

Adapted from Bernstein et al. (2015) 50 vaccinees and 48 control subjects. A NoV vaccine containing a consensus sequence of GII.4 was given intramuscularly in two doses to healthy adult volunteers followed by challenge on day 42 with GII.4 Farmington strain NoV

Registration of Vaccines, Safety Follow-Up, and Pediatric Investigation Plan

Carlo Giaquinto and Francesca Rocchi

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Before a new vaccine is approved for release onto the market, a rigorous regulatory procedure to assess the quality, efficacy, and safety of the product must be undertaken. In Europe, the vaccine manufacturer has to obtain a marketing authorization (MA) or product license. The MA is granted after an evaluation that relates to a number of vaccine properties such as quality, safety, and efficacy in addition to compliance with good practices in the areas of manufacturing and clinical or laboratory testing.

In the European Union (EU), there are two types of MA for vaccines: the national and the European. The national MA is issued by the competent authorities of the individual Member States. In this case, the vaccine may be put on the market in all Member States that have granted an authorization for it. If a company is seeking a national MA in more than one Member State, the *mutual recognition* or *decentralized procedure* is available to facilitate the granting of harmonized national authorizations across Member States. For the authorization of traditional non-recombinant vaccines in the EU, the developer can submit the Marketing Authorization Application (MAA) for review to one or more national competent authorities for medicines. On the other side, the Community marketing authorization is a single authorization that allows the medicinal product to be put on the market in all Member States and is granted by the European Commission, following a positive opinion from the European Medicines Agency (EMA). The EMA is a decentralized agency responsible for the scientific evaluation, supervision, and safety monitoring of medicines developed by pharmaceutical companies for use in Europe. The EMA is primarily involved in the centralized procedure for obtaining an EU MA. The Agency also gives scientific advice to companies on the development of new vaccines and develops guidelines on quality, safety, and efficacy testing requirements. Innovative vaccines, and in particular, recombinant vaccines (recombinant protein-based vaccines and recombinant viral-vectored vaccines), must be evaluated and approved in the EU via the centralized procedure. Other novel vaccines can also be approved centrally if justified by the applicant (eligibility to the centralized procedure under the “optional scope,” as outlined in Article 3 of Regulation (EC) No. 726/2004). The central-

ized procedure is mandatory for certain types of medicinal products and optional for others. Medicinal products made of recombinant proteins, advanced therapy medicinal products (ATMPs) for human use, human medicinal products containing a new active substance for the treatment of acquired immune deficiency syndrome, cancer, neurodegenerative disorders, diabetes, viral diseases, auto-immune diseases/other immune dysfunctions, and designated orphan medicinal products fall within the mandatory scope and must be filed centrally at the EMA. Although the European pharmaceutical legislation does not provide a formal definition, vaccines are typically considered medicinal products containing one or more immunogenic antigens intended for the prevention of disease from infective agents. Medicinal products containing one or more immunogenic antigens for the treatment of disease, e.g., chronic HIV infection, chronic hepatitis B or C infection, cancer, or Alzheimer’s disease, are typically referred to as therapeutic vaccines or active immunotherapy. The same scientific principles for their product development as for prophylactic vaccines against infectious diseases apply. Vaccines against infectious diseases that are based on viral (or other) vectors or on DNA plasmids are specifically excluded from the definition of a gene therapy medicinal product (GTMP).

The scientific evaluation of the application is carried out by the Committee for Medicinal Products for Human Use (CHMP) of the EMA and a scientific opinion is prepared in co-operation with other EMA committees, as applicable, together with many expert groups and working groups, e.g., the Vaccine Working Party, that contribute to the review of applications. The opinion is sent to the European Commission, which drafts a decision and, having consulted the Member States through the relevant standing committee, adopts the decision and grants a MA. In Europe, the vaccines authorized via a centralized procedure have one invented name (trade name), one common labelling, translated into 23 languages and comprises Summary of medicinal Product Characteristics (SmPC) and the user package leaflet and package labelling. Approved conditions of use are laid down in the summary of product characteristics, the SmPC, (prescribing information for health professionals), the labelling, and the package leaflet for users.

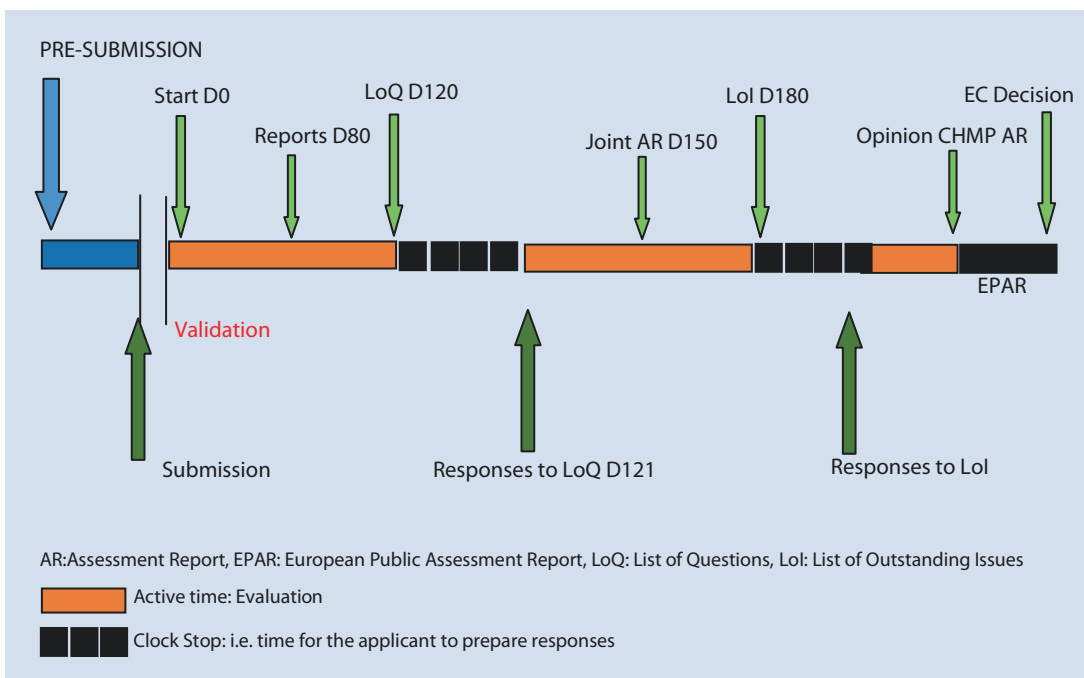
Once the evaluation is completed within 210 days, the CHMP adopts a favorable or unfavorable opinion on whether to grant the authorization. Once the Community MA is granted, the EMA publishes the CHMP assessment report on the vaccine, which includes the reasons for its opinion. This document is called the European Public Assessment Report (EPAR). The EPAR includes a summary, in all EU languages, written in a manner that is understandable to the public. EPARs and their summaries are published on the EMA website (■ Fig. 26.1).

To meet unmet medical needs of patients and in the interests of public health, the CHMP can recommend the granting of MAs based on less complete data than is normally required. In such cases, the granting of an MA is subject to certain specific obligations to be reviewed annually (“conditional marketing authorization”). For example, in 2010, two pandemic influenza vaccines (H1N1), Arepanrix[®] and Humeza[®], received conditional marketing approval. In exceptional circumstances, a MA can be granted subject to a requirement for the applicant to introduce specific procedures (safety procedures, program of studies, prescription or administration conditions, product information), in particular, con-

cerning the safety of the product (“marketing authorization under exceptional circumstances”). Continuation of the authorization is linked to the annual reassessment of these procedures. Imvanex[®], a vaccine against smallpox, was approved in 2013 under exceptional circumstances because it had not been possible to obtain complete information about Imvanex[®] because of the absence of the disease.

In the area of vaccines, it is worthwhile mentioning Article 58 of Regulation (EC) No. 726/2004, which allows the CHMP to give opinions, in co-operation with the World Health Organization (WHO), on medicinal products for human use that are intended exclusively for markets outside of the EU. Medicines eligible for this procedure are used to prevent or treat diseases of major public health interest. This includes vaccines used in the WHO Expanded Program on Immunization for protection against a public health priority disease. The CHMP carries out a scientific assessment of applications submitted under Article 58, and, after consultation with the WHO, adopts a scientific opinion.

The requirements for the structure and content of the MAA are laid down in the EU Common Technical Document (CTD) and provide for a



■ Fig. 26.1 Centralized assessment procedure (CAP)

harmonized structure and format for MAAs in Europe, Japan, and the USA. Data generated from pharmaceutical tests, nonclinical and clinical tests and trials with the vaccine concerned, in addition to other information required by the EU legislation, need to be submitted to the EMA and all CHMP members for evaluation. The application dossier for the vaccine must be presented in accordance with the EU-CTD. The CTD is an internationally agreed format for the preparation of a well-structured application to be submitted to regulatory authorities in the three International Conference on Harmonization (ICH) regions of Europe, USA, and Japan.

26.1 Paediatric Investigation Plans

As for all medicinal products, since 26 January 2007, vaccine developers are obliged to submit the results of studies conducted in compliance with an agreed Paediatric Investigation Plan (PIP) to have a valid application for a new MA. The Paediatric Regulation (EC) No. 1901/2006 requires the PIP to be submitted to the EMA as early as possible (ideally soon after the phase I–II clinical trial conducted in adult populations).

The PIP describes planned clinical studies in children, including the proposed timing of the studies, formulation adaptations to make it suitable for children and nonclinical studies in Juvenile animals if required. This is to ensure that the necessary data are generated determining the conditions under which the vaccine may be authorized to treat the paediatric population; in other words, a PIP should provide the data to enable the assessment of the quality, safety, and efficacy in children, and consequently the benefit/risk profile in the paediatric population. The PIP must be agreed by the EMA Paediatric Committee (PDCO) before applying for MA for any age group. The key element of the Paediatric Regulation is the early involvement of vaccine developers in the research and development program of a medicinal product through the requirement to reach an agreement with the PDCO on the PIP. Decision on a waiver may be issued by the EMA when such paediatric development is not needed or not appropriate. Deferrals may also be granted.

During the period from 2007 to 2016, a total of 52 PIPs/waivers on vaccines were agreed by the PDCO out of a total of 808; 12

vaccine PIPs were completed, leading to the authorization of 12 paediatric vaccine indications (■ Table 26.1).

The PDCO has developed a number of standard PIPs. These are documents that provide recommendations for the key binding elements to be included in the PIP opinion with the aim of assisting applicants with the agreement of PIPs on specific types or classes of medicines. A particularly challenging project was the drafting of the standard PIP for the tetanus-diphtheria pertussis (DTaP) vaccines, owing to the complexity of vaccination programs and differences across Member States. The PDCO has defined, in collaboration with the European Centre for Disease Prevention and Control (ECDC) and European public health vaccinology experts, the schedule that should be evaluated during clinical trials in children when developing a new DTaP-containing combination vaccine. The proposed schedule has been defined as the one producing data that can cover the various vaccination schedules in the individual European Member States, through extrapolation of results to immunologically less challenging schedules.

26.2 Vaccine Development – Requirements for the MAA

26.2.1 Quality

The development of vaccines is addressed in a variety of guidelines on vaccines. There is considerable interest in developing new adjuvants for both existing and novel vaccines. The area is quite complicated and the nature and mode of action of novel adjuvants is quite wide. EU guidance on the development and regulatory approval for an adjuvant is available. An important aspect in developing a novel adjuvant is to show that it does enhance the immune response to the antigen with associated clinical benefit. The safety of novel adjuvants is also an important factor.

The quality section of an MAA requires a detailed characterization of the vaccine, a detailed description of the manufacturing process, a description of all raw materials and components used in the manufacturing process, and a description and validation of all quality control tests applied during the manufacturing process and to the vaccine itself. This section should also consider the consistency of vaccine production and

Table 26.1 New medicines (CAPs, initial marketing authorizations) including a pediatric indication by year of authorization

Year	Active substance	Trade name	Indication pediatric-only or mixed ^a
2007	Human papillomavirus vaccine (types 16, 18)	Cervarix	Mixed
2009	Pneumococcal polysaccharide conjugate vaccine	Synflorix	Pediatric only
2009	Pneumococcal polysaccharide conjugate vaccine (13-valent, adsorbed)	Prevenar 13 (PIP not yet completed)	Pediatric only
2010	Meningococcal group A, C, W135, and Y conjugate vaccine	Menveo (PIP completed)	Mixed
2011	Influenza vaccine (live attenuated, nasal)	Fluenz (waiver)	Pediatric only
2012	Prepandemic influenza vaccine (H5N1) (whole virion, inactivated, prepared in cell culture)	Vepacel	Mixed
2012	Meningococcal group A, C, W135, and Y conjugate vaccine	Nimenrix	Mixed
2013	Meningococcal group B vaccine (RDNA, component, adsorbed)	Bexsero	Mixed
2013	Diphtheria (D), tetanus (T), pertussis (acellular, component) (PA), hepatitis B (RDNA) (HBV), poliomyelitis (inactivated) (IPV) and <i>Haemophilus influenzae</i> type b (Hib) conjugate vaccine (adsorbed)	Hexacima	Pediatric only
2013	Diphtheria (D), tetanus (T), pertussis (acellular, component) (PA), hepatitis Bb (RDNA) (HBV), poliomyelitis (inactivated) (IPV), and <i>Haemophilus influenzae</i> type b (Hib) conjugate vaccine (adsorbed)	Hexyon	Pediatric only
2013	Influenza vaccine (live attenuated, nasal)	Fluenz tetra	Mixed
2015	Human papillomavirus vaccine (types 6, 11, 16, 18, 31, 33, 45, 52, 58) (recombinant, adsorbed)	Gardasil 9	Mixed

^aAdult and pediatric

the stability of the vaccine and describe an appropriate and validated potency assay for the vaccine.

There is no generic EU guideline addressing the quality requirements of vaccines; however, the requirements are similar for most biological medicinal products. Guidance is available for some specific vaccines including smallpox vaccine, influenza vaccine, recombinant viral-vec-tored vaccines¹⁶, and DNA vaccines.

26.2.2 Nonclinical

The “Note for Guidance on preclinical pharmacological and toxicological testing of vaccines”

focuses on the preclinical evaluation of new vaccine products (those containing antigens not yet described in the European Pharmacopoeia monographs or in WHO requirements, or using a new conjugate for a known antigen, or any combination of known and/or new antigens). As the vaccine represents a heterogeneous class of agents, preclinical pharmacological and toxicological testing of the vaccine may be adapted for the product in question. Single-dose toxicity data from at least one animal species should be performed, with a dose providing an adequate safety margin in relation to the human dose; a study on repeated dose toxicity in one animal species for vaccines that will require multiple

doses in a clinical setting is normally required. Data on reproductive function (fertility) are not usually needed, but this depends on the vaccine indication, and embryo/fetal, perinatal toxicity data, and carcinogenicity/mutagenic studies are usually not needed either. However, there are exceptions for vaccines with new adjuvants where special considerations are needed. Local tolerance should be evaluated, as vaccines are in most cases administered intramuscularly, subcutaneously or intradermally. Immunogenicity studies look at humoral and cell-mediated response in appropriate animal models for the disease to indicate dose, schedule, and route of administration in future clinical studies. Protection studies basically establish the proof of concept of protection from disease and are established by challenge studies if feasible (e.g., ferrets challenge studies for influenza pandemic vaccines).

Secondary pharmacodynamics include safety pharmacology for the potential evaluation of undesirable pharmacological activities (the circulatory and respiratory system) and should be considered depending on the new vaccine. For vaccines protecting against infectious diseases, not all aspects of a classical nonclinical development program need to be covered, e.g., pharmacokinetics is generally not required for vaccines. More specific nonclinical guidance is available for vaccines containing adjuvants, for smallpox vaccines, and for live recombinant viral-vectored vaccines.

26.2.3 Clinical

The EU Guideline on the clinical evaluation of vaccines provides a comprehensive explanation of the design of clinical development programs for new vaccines that are intended to provide pre- and post-exposure prophylaxis. In the development of any new vaccine, adequate data on immunogenicity should be assembled during the clinical development program. Aspects that should usually be covered include characterization of the immune response, investigation of an appropriate dose and primary schedule, assessment of the persistence of detectable immunity, and consideration of the need for and response to booster doses. Additionally, for vectored vaccines, the determination and characterization of the pre-existing immunity to the vector should be

addressed. Pharmacokinetic studies might be required for MA when new delivery systems are used or when the vaccine contains novel adjuvants or excipients.

Ideally, protective efficacy should be performed before licensing a new vaccine. However, it is recognized that there are situations where such studies are not necessary and/or not feasible before licensing for all types of vaccines; for example, when there are established immunological correlates of protection against a specific infection such as diphtheria or tetanus or hepatitis B, immunogenicity studies may be considered sufficient. In addition, when the disease does not occur, e.g., smallpox or pandemic influenza, estimating protective efficacy is not feasible.

Vaccine effectiveness reflects direct (vaccine-induced) and indirect (population-related) protection during routine use. Whether or not protective efficacy is assessed in the pre-authorization period, attempts should be made to estimate vaccine effectiveness in the post-authorization period. With the increasing complexity of vaccines and the frequent need for co-administration of multiple vaccines, immune interference has become a very important consideration.

Concerning clinical safety as pre-authorization requirements, unless otherwise justified, the recommended minimum sample size would be at least 3,000 subjects for a new vaccine; the total data for pre-authorization studies should usually be sufficient to reliably determine the frequency of uncommon local and systemic adverse events, i.e., frequency 1/100 to 1/1,000.

By the time a MA is granted, a risk specification should have been finalized that includes a description of possible safety issues related to the intrinsic character of the vaccine; a risk management plan (RPM) should have been agreed with the EMA; and a pharmacovigilance system (as defined in the current EU legislation) and procedures should have been put in place. The RMP defines a set of pharmacovigilance activities and interventions that identify, characterize, prevent, or minimize risks relating to the medicinal product, including the assessment of the effectiveness of those interventions. New pharmacovigilance legislation came into operation in 2012, and new provisions for Periodic Safety Update Reports (PSURs), RMPs, safety signals and Post Authorization Safety Studies (PASS)

were introduced. In addition, literature monitoring and several tools for product safety reviews at the EU level are part of this legislation. A Pharmacovigilance Risk Assessment Committee (PRAC) has been established at the EMA, and as one of its tasks, the PRAC assesses the RMP.

Considering that vaccines are almost always administered to healthy persons, the continued re-assessment of the overall risk–benefit profile has great implications.

26.3 Conclusion

Many new vaccines will become available in the very near future, which poses important challenges to the regulatory process in Europe and the USA. Large clinical trials have been carried out in the past to evaluate the efficacy and safety of vaccines, which in some cases delayed the global introduction of an important vaccine. New technologies for vaccine manufacturing have been developed that are not fully known in terms of safety and long-term efficacy. Therapeutic vaccines are becoming available for chronic diseases and it is not clear how these should be evaluated, especially in the long term. These issues require the development of a strong regulatory environment that will be able to guarantee the “overall quality” of the new vaccines and the need for a fast and efficient process.

Further Reading

- 10-year Report to the European Commission General report on the experience acquired as a result of the application of the Paediatric regulation. European Medicines Agency and its Paediatric Committee November 2016. ► http://ec.europa.eu/health/sites/health/files/files/paediatrics/2016_pc_report_2017/ema_10_year_report_for_consultation.pdf.
- COMMISSION REGULATION (EC) No 507/2006, 29 Mar 2006, Conditional marketing authorisation for medicinal products for human use falling within the scope of Regulation (EC) No 726/2004 of the European Parliament and of the Council.
- Commission Regulation (EU) No 1235/2010 of the European Parliament and of the Council of 15 Dec 2010 amending, as regards pharmacovigilance of medicinal products for human use, Regulation (EC) No 726/2004 laying down Community procedures for the authorisation and supervision of medicinal products for human and veterinary use and establishing a European Medicines Agency, and Regulation (EC) No 1394/2007 on advanced therapy medicinal products. ► http://ec.europa.eu/health/files/eudralex/vol-1/reg_2010_1235/reg_2010_1235_en.pdf.
- DIRECTIVE 2001/83/EC OF THE EUROPEAN PARLIAMENT AND OF THE COUNCIL 6 Nov 2001 on the Community code relating to medicinal products for human use.
- Directive 2010/84/EU of the European Parliament and of the Council of 15 Dec 2010 amending, as regards pharmacovigilance, Directive 2001/83/EC on the Community code relating to medicinal products for human use. ► http://ec.europa.eu/health/files/eudralex/vol-1/dir_2010_84/dir_2010_84_en.pdf and corrigendum. ► http://ec.europa.eu/health/files/eudralex/vol-1/dir_2010_84_cor/dir_2010_84_cor_en.pdf.
- Explanatory note on immunomodulators for the guideline on adjuvants in vaccines for human use. CHMP/VWP/244894/2006. ► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003810.pdf.
- Final paediatric investigation plan: Expected key elements and requirements for a new DTaP-containing combination vaccine for primary and booster vaccination in infants and toddlers. EMA/82701/2015. ► http://www.ema.europa.eu/docs/en_GB/document_library/Regulatory_and_procedural_guideline/2015/11/WC500196478.pdf.
- Guideline on adjuvants in vaccines for human use. EMEA/CHMP/VEG/134716/2004. ► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003809.pdf.
- Guideline on clinical evaluation of new vaccines. EMEA/CHMP/VWP/164653/2005. ► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003870.pdf.
- Guideline on quality, non-clinical and clinical aspects of live recombinant viral vectored vaccines. EMA/CHMP/VWP/141697/2009. ► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2010/08/WC500095721.pdf.
- http://ec.europa.eu/health/files/eudralex/vol-2/a/chap-4rev200604_en.pdf.
- <http://www.who.int/immunization/en/>.
- Klug B, Celis P, Ruepp R, Robertson J. Vaccines: EU regulatory requirements. *Mol Vaccines*. 2014;2:845–50.
- Note for guidance on cell-culture-inactivated influenza vaccines. Annex to the note for guidance on the harmonisation of requirements for influenza vaccines 2002.CPMP/BWP/2490/00. ► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003877.pdf.
- Note for guidance on pre-clinical safety evaluation of biotechnology-derived pharmaceuticals (ICH S6). CPMP/SWP/465/95. ► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/10/WC500004004.pdf.
- Note for guidance on the development of vaccinia virus based vaccines against smallpox. CPMP/1100/02 2009. ► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003900.pdf.
- Note for guidance on the harmonisation of requirements for influenza vaccines. CPMP/BWP/214/96 1997.

► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003945.pdf.

Notice to Applicants Volume 2 B – presentation and format of the dossier – Common Technical Document (CTD) May 2008. ► http://ec.europa.eu/health/files/eudralex/vol--2/b/update_200805/ctd_05-2008_en.pdf.

Notice to Applicants Volume 2A – procedures for marketing authorisation. Chapter 4: Centralised procedure. 2016.

Points to consider on the development of live attenuated influenza vaccines 2019. EMEA/CPMP/BWP/1765/99.

► http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003899.pdf.

REGULATION (EC) No 1901/2006 OF THE EUROPEAN PARLIAMENT AND OF THE COUNCIL of 12 December 2006 on medicinal products for paediatric use.

Regulation (EC) No 726/2004 of the European Parliament and of Council, 31 Mar 2004, Community procedures for the authorisation and supervision of medicinal products for human and veterinary use and establishing a European medicines Agency. ► <http://eurlex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2004:136:0001:0033:en:PDF>.

Supplementary Information

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