Editor's preface

Physical Organic Chemistry is a mature discipline that is blessed with a rich history and a bright future. It is therefore fitting that the chapters in this 37th volume of *Advances in Physical Organic Chemistry* deal with investigations that can be traced back to the birth of the field, but which are continuing to produce results critical to our understanding of the stability of organic molecules and the mechanisms for their reactions.

Three chapters in this volume deal with various aspects of the stability of carbocations, and their role as putative intermediates in chemical and enzymatic reactions.

Abboud, Alkorta, Dávalos and Müller summarize the results of recent experimental work to determine the thermodynamic gas phase stabilities of carbocations relative to neutral precursors, the results of high level calculations of these stabilities and additional structural information that can only be easily obtained through such calculations. The chapter concludes with a description of the effect of solvent on the condensed-phase stability of carbocations, and the role of solvent in determining whether carbocations form as intermediates of solution reactions. This chapter is richly referenced and can be read with interest by anyone wishing to remain abreast of modern developments in a historically important subject.

The generation and characterization of vinyl carbocations remains a challenging problem because of the great instability of positive charge at sp hybridized carbon. Physical organic chemists have classically produced unstable carbocations through heterolytic cleavage of bonds to weakly basic atoms or molecules. The past success of this approach has prompted the experimental studies of vinyl(aryl)iodonium salts described in the chapter by Okuyama and Lodder. Although these salts are good electrophiles that carry an excellent iodoarene nucleofuge, photoexcitation to the excited state is required to drive heterolytic cleavage to form simple primary vinyl carbocations. This chapter describes the great diversity in the products obtained from the thermal and photochemical chemical reactions of vinyl(aryl)iodonium salts and the reasoning used in moving from these product yields to detailed conclusions about the mechanisms for their formation.

Oxocarbenium ions are commonly written as intermediates of organic reactions. However, the lifetimes of oxocarbenium ions in water approach the vibrational limit and their formation as reaction intermediates in this medium is sometimes avoided through a concerted mechanism. The determination of whether oxocarbenium ions form as intermediates in related enzymatic processes is a particularly challenging problem, because the protein catalyst will shield these ions from interactions with solvent and solutes which might provide evidence for their formation. Deuterium, tritium, and heavy atom kinetic isotope effects provide a wealth of information about reaction mechanism, but these are sometimes masked for enzymatic reactions by the high efficiency for turnover of enzyme-bound substrates. However, it is often possible for creative enzymologists to develop substrates or reaction conditions under which the rate constants for enzymatic reactions are limited by chemical bond cleavage, and are therefore subject to significant kinetic isotope effects. The design and interpretation of such multiple kinetic isotope effect studies to probe the changes in chemical bonding at sugar substrates that occur on proceeding to the transition states for enzyme-catalyzed cleavage of glycosides is described in a chapter by Berti and Tanaka.

Organometallic chemistry has largely been the domain of synthetic organic and inorganic chemists, and the mechanisms for organometallic transformations have not generally been subject to the same type of detailed experimental analyses developed in studies on the mechanism of multistep organic reactions. Claude Bernasconi, a leading figure in the study of organic reaction mechanisms, summarizes here the results of studies on the reactions of organometallic Fischer carbene complexes in aqueous solution. Finally, Drechsler and Rotello describe how the redox properties of flavins, quinones and related molecules can be varied through the rational design of molecules in which a given oxidation state is stabilized by electrostatic, hydrogen-bonding, π -stacking and other noncovalent interactions. The design and physical characterization of these finely tuned redox systems has potential applications in the development of a variety of molecular "devices".

We are pleased to note that the masthead lists a revamped and expanded Editorial Advisory Board. This board is assisting the coeditors in the planning of future volumes in order to ensure that *Advances in Physical Organic Chemistry* continues to highlight the most important applications of physical and theoretical methods to the characterization of the structure and stability of organic molecules and the mechanisms for their reactions.

J. P. Richard T. T. Tidwell

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Nucleophilic Vinylic Substitution and Vinyl Cation Intermediates in the Reactions of Vinyl Iodonium Salts

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1 Introduction

Nucleophilic vinylic substitution $(S_N V)$ has a wide variety of mechanistic possibilities, including more than 10 main routes which can be further sub-divided into over 30 variants, as reviewed by Rappoport.¹ Four main straightforward mechanisms are illustrated in Scheme 1.

Nucleophilic attack at the alkene carbon can in principle occur either toward the π^* or σ^* orbitals since the sp²-hybridized carbon has two types of vacant orbitals. That is, both out-of-plane (perpendicular) and in-plane attack are possible. The out-of-plane mode of attack is part of the addition–elimination pathway (Ad_N–E) with a



Scheme 1 Mechanisms of nucleophilic vinylic substitution.

carbanion as an intermediate. This associative mechanism is generally observed for activated unsaturated systems¹⁻³ and gives rise to products with stereochemistry ranging from retention to stereoconvergence depending on the (in)stability of the intermediate carbanions.

As the leaving group ability of Y^- increases, it ultimately departs concurrently with the nucleophilic attack and the reaction occurs via a "concerted addition– elimination" mechanism.^{1–3} This type of mechanism should result in retention of configuration and may be designated as $S_N V \pi$. In contrast, the in-plane σ^* attack of a nucleophile should lead to substitution with inversion of configuration and may be referred to as $S_N V \sigma$.

With further increasing leaving ability of Y^- , the reaction becomes dissociative and becomes a vinylic $S_N 1$ ($S_N V1$) reaction involving a vinyl cation as intermediate. $S_N V1$ reactions have been extensively studied, both with substrates giving stabilized vinyl cations and/or with substrates with a good nucleofuge such as triflate (trifluoromethanesulfonate, TfO⁻) and are the subjects of several reviews.⁴⁻⁶ Their stereochemical consequences are discussed in Section 2.

The $S_N V \sigma$ mechanism is a logical analogue of the $S_N 2$ reaction at saturated carbon that occurs via backside attack of the nucleophile, but it has long been rejected as a feasible pathway on the basis of steric considerations^{7,8} and of early theoretical calculations on a rather crude model system.⁹ However, quite recently definite examples of $S_N V \sigma$ reactions have been found,^{10,11} and recent theoretical studies^{12–16} show that the $S_N V \sigma$ as well as the $S_N V \pi$ mechanism is feasible. If imbalance of bond formation and bond cleavage occurs, the dissociative extreme of



the in-plane $S_N V \sigma$ mechanism is formation of a vinyl cation ($S_N V1$ mechanism) and the associative extreme of the out-of-plane $S_N V \pi$ mechanism is the Ad_N -E route. The reviews of Rappoport¹⁻³ mainly concern the associative to concerted part, Ad_N -E/ $S_N V \pi$, of the mechanistic spectrum of nucleophilic vinylic substitution. In this chapter we discuss the concerted-dissociative part ($S_N V \sigma / S_N V1$ dichotomy) of the mechanistic spectrum, mainly on the basis of our recent results obtained in the study of reactions of vinyl iodonium salts. Also, photochemical dissociative reactions generating vinyl cations are discussed and compared with the corresponding thermal reactions.

2 Vinylic S_N2 reactions

S_NV REACTIONS WITH INVERSION

The stereochemistry of nucleophilic vinylic substitution via the Ad_N–E mechanism ranges from retention of configuration to stereoconvergence. For S_NV1-type reactions, often partial inversion has been observed. The inversion has been attributed to nucleophilic attack on an ion pair intermediate.^{17–20} As illustrative examples, results of the acetolysis of the vinyl triflates **1a–c** in the presence of sodium acetate are given in Scheme 2.^{19,20} The fraction of inversion ranges from 40 to 80%, and is smaller the more stable the intermediate cation; β -alkyl substitution





Scheme 4

stabilizes vinyl cations, as discussed below. In less nucleophilic solvents such as trifluoroethanol and trifluoroacetic acid the fractions of inversion are also much smaller (<10%).^{18,20} More stabilized vinyl cations, with e.g., an α -aryl substituent, lead to products of stereoconvergence. These trends are consistent with a change of mechanism from ion pair to free ion with increasing stabilization of the cations.

The intramolecular processes outlined in Scheme 3 occur formally with inversion of configuration at the electrophilic carbon. Example (1) is participation of a nucleophilic neighboring group, where the nucleophile is constrained to attack inplane, while example (2) is the microscopic reverse of the first one, and the preferred path must again be in-plane attack. These reactions are postulated to occur via the $S_N V \sigma$ pathway to give the inverted product.^{21–23} Their consecutive occurrence results in an anchimerically assisted reaction, and the net stereochemical outcome is retention of configuration due to double inversion. The overall mechanism of this type of reaction can be classified as $S_N V1$, since the corresponding process in the solvolysis of saturated aliphatic derivatives is classified as an S_N1 reaction with neighboring group participation.

Participation of β -sulfur,²⁴ β -iodine,²⁵ and β -aryl groups²⁶ has been reported for the solvolysis of the vinylic substrates 2–4 (Scheme 4). Each of the reaction steps occurs with inversion, and the overall process results in retention of configuration. The cyclic 3-membered thiirenium ion 5 also gave exclusively inverted



nucleophilic substitution product (Scheme 5).^{27–29} However, retention of configuration was observed for the reaction of the neutral cyclic substrate, thiirene 1,1-dioxide **6** (Scheme 6).³⁰

The intramolecular transfer of the nucleophilic group in Scheme 7 is also claimed to be an $S_N V \sigma$ reaction.^{22,23} The anionotropic rearrangement of the di-*tert*-butylthiirenium ion 7 into the thietium ion 8 involves the stereospecific backside attack of the internal methide at the unsaturated carbon.

Examples of acyclic vinylic systems that undergo nucleophilic substitution with complete inversion of configuration are very rare. The reaction of 1,2-dibromo-1,2-difluoroethene (9) with *p*-toluenethiolate affords products of apparently inverted structure without loss of stereochemical purity (Scheme 8).³¹ However, the authors are not sure whether this is due to the stereospecificity of the reaction or to the thermodynamic stabilities of the products.

An unambiguous example is the nucleophilic substitution of 1-alkenyl(aryl)iodonium salts with halide ions (Scheme 9).³² 1-Decenyl(phenyl)iodonium tetra-



Scheme 9

fluoroborate (11) gave the completely inverted haloalkene 12 when it reacted with chloride, bromide, or iodide ions. Details of this reaction are discussed in a section below (see Scheme 14).

The nucleophilic substitution of the alkylidene carbenoid 1-bromo-1-lithioethene **13** with *tert*-butyllithium stereospecifically gives the inverted product (Scheme 10).^{33–35} This could be an $S_N V \sigma$ reaction.

An S_N 2-type substitution at the sp²-hybridized nitrogen atom of a C=N bond has recently been reported for the acid-catalyzed intramolecular reaction of oxime **14** (Scheme 11).³⁶ The results have been rationalized by theoretical calculations.

THEORETICAL CONSIDERATIONS

It has long been believed that in-plane nucleophilic substitution at vinylic carbon $(S_N V \sigma)$ is an unfavorable process, partly due to steric reasons.^{7,8} The nucleophilic attack occurs at the vacant orbital of the electrophilic substrate, and the LUMO of alkenes was generally thought to have π symmetry and not σ symmetry (σ^* orbitals are in general higher in energy than π^* orbitals). This would make perpendicular attack at the vinylic carbon (π^*) by a nucleophile more favorable than in-plane attack (σ^*). Early theoretical calculations for the system H₂C=CH₂ + H⁻, using the extended Hückel method, in fact showed a very high energy barrier (326 kJ mol⁻¹) for the in-plane S_NV σ reaction and a low barrier (75 kJ mol⁻¹) for the perpendicular π attack.⁹ Later theoretical studies undertaken in the 1970s and 1980s focused on the perpendicular and not on the in-plane reactions. Inspired by experimental indications that the in-plane S_NV process is feasible, more sophisticated MO calculations on such reactions were undertaken in the 1990s. The results thereof are at variance with the conclusions of the early calculations.

First, it was shown that the LUMO of vinylic substrates is not necessarily of π symmetry. For some classes of vinylic compounds the LUMO is a σ^* orbital (Table 1).¹³ While simple vinylic compounds such as vinyl chloride and triflate (entries 4 and 5) have an anticipated π^* orbital as LUMO, charged substrates (entries



| No. | | LUMO (symmetry) | LUMO + 1 (symmetry) |
|-----|---------------------|----------------------------------|---------------------|
| 1 | S+ H | -0.1046 (<i>σ</i>) | $-0.0954(\pi)$ |
| 2 | S O ₂ | 0.0819 (<i>π</i>) | 0.1878 (σ) |
| 3 | | -0.1395 (σ) | -0.0713 (π) |
| 4 | | 0.1454 (<i>π</i>) | $0.1824~(\sigma)$ |
| 5 | | 0.1328 (<i>π</i>) | $0.1811~(\sigma)$ |
| 6 | =∖+ I-Me | $-0.065 \left(\sigma\right)^{a}$ | $-0.029 (\pi)^{a}$ |
| 7 | | $-0.106 (\sigma)^{a}$ | $-0.035 (\pi)^{a}$ |
| 8 | Br F F Br | $0.1226 (\sigma)^{b}$ | $0.1421 (\pi)^b$ |
| 9 | F Br Br | $0.1355 (\sigma)^{b}$ | $0.1391 (\pi)^b$ |
| 10 | | 0.0476 (<i>o</i>) | 0.1195 (<i>m</i>) |

Table 1 Energies of the lowest vacant orbitals (in hartree) with π and σ symmetry for the vinylic portion of some model substrates

Calculated at 6-311G**//3-21G* (Ref. 13) unless noted otherwise. ^aCalculated at 6-31G* (MP2).¹⁴ ^bCalculated at 3-21G*//3-21G*.¹³

1, 3, 6, and 7) have a σ^* orbital as LUMO. Thus, frontier orbital considerations rationalize the observed inversion in the S_NV reactions of the charged substrates in Schemes 5, 7 and 9. In accordance with the experimental observation of substitution with retention in Scheme 6, the cyclic sulfone (entry 2) has a LUMO with π symmetry. Polyhaloethenes have lowest σ^* and π^* orbitals which are very close in energy to
each other, and the bromo analogues (8 and 9) have σ LUMO in accordance with the observed inversion in the nucleophilic substitution of this kind of substrate (Scheme 8). The carbenoid (entry 10) has a long and very weak C–Cl bond,³⁴ and this may be why it has a σ LUMO and in turn why it gives an S_NV σ reaction (Scheme 10).

Ab initio MO calculations at the G2(+) level of theory in the gas phase have shown that even the prototypical S_NV reaction of vinyl chloride with chloride ion can favorably take place via backside in-plane σ attack, in spite of its LUMO with π symmetry.¹² The energy profiles calculated for the in-plane and the out-of-plane attack are shown in Fig. 1. Attack by Cl⁻ on CH₂=CHCl first forms a hydrogen-



Reaction Coordinate

Fig. 1 Energy profile for the identity S_N^2 reaction of CH_2 =CHCl with Cl^- , calculated at the G2(+) level. Bond lengths are in angstroms. Taken from Ref. 12.

bonded complex in which the incoming Cl^- is located in the plane of the alkene. The reaction then proceeds via the in-plane σ ($S_N V \sigma$) pathway with a barrier of 137 kJ mol⁻¹ or via the out-of-plane π pathway ($S_N V \pi$) with a barrier of 179 kJ mol⁻¹. Thus, the $S_N V \sigma$ process is 42 kJ mol⁻¹ more favorable than the $S_N V \pi$ pathway in the gas phase. This conclusion has recently been confirmed at higher levels of theory.¹⁶ The negative charge in the transition state (TS) of the former pathway is less delocalized than that in the latter TS, and solvent effects stabilizing the TSs are expected to be larger for the former route. This simplistic consideration suggests that the gas-phase preference for the $S_N V \sigma$ pathway will be maintained in solution. No two-step pathway via a carbanion intermediate (Ad_N–E) was found for this unactivated vinyl substrate. The β , β -dichloroethyl anion, the potential intermediate in the Ad_N–E process, is unstable and collapses to the starting complex.

Quite recently, Lee and co-workers¹⁵ have theoretically examined the reactions of vinyl chloride with various nucleophiles in quest of the driving force of the unexpected preference for the S_NV σ process. Calculations were carried out at three levels, RHF/6-311 + G**(RHF), MP2/6-311 + G**(MP2), and G2(+)(MP2), with MP2/6-311 + G** geometries for the latter two levels. The preference for σ or π attack is dependent on the level of calculation and the nature of the nucleophile. The highest level results are summarized in Table 2 and typical structures of TSs for the S_NV σ route is energetically favored over the S_NV π route for Cl⁻ and Br⁻ as nucleophile, while the reverse is the case for OH⁻ and SH⁻.

The substitution reactions of CH₂=CHCl with OH⁻ and SH⁻ are exergic, whereas that with Br⁻ is endergic (ΔG^{0}). The S_NV σ route is not feasible with the strong base OH⁻ in the gas phase; the TS for the σ attack by OH⁻ could not be located, and the barrier for the S_NV π route is very low ($\Delta G^{\ddagger} = 9.0 \text{ kJ mol}^{-1}$). The barriers for the σ and π route of Br⁻ ($\Delta G^{\ddagger} = 134$ and 165 kJ mol⁻¹) are both higher than those for Cl⁻ (125 and 144 kJ mol⁻¹), largely due to the endergicity of the

| Mechanism | Nu ⁻ | $\Delta G_{	extsf{g}}^{\ddagger}$ | $\Delta G_{ m g}^0$ | $\Delta G_{ m AN}^{\ddagger}{}^a$ |
|---------------------------|--|-----------------------------------|--------------------------------|-----------------------------------|
| S _N V <i>σ</i> | Cl ⁻ | 124.5 | 0.0 | 192.9 |
| | OH^{-b} SH ⁻ | - 110.7 | - 79.7 | 194.8 - 171.6 |
| S _N V π | Cl ⁻ Br ⁻ OH ⁻ SH ⁻ | 144.5 165.3 9.0 101.9 | 0.0 38.2 -222.6 -79.7 | 210.1 222.2 72.6 160.8 |

Table 2 Energies $(kJ \text{ mol}^{-1})$ for the reactions of vinyl chloride with various nucleophiles Nu⁻, in the gas phase and in acetonitrile, calculated at the G2(+)(MP2) level¹⁵

^aValues in acetonitrile calculated by the isodensity polarizable continuum model.

^bThe transition state could not be located for the $S_N V \sigma$ -route.



Fig. 2 Transition states for the $S_N 2$ reaction of vinyl chloride with Br^- . Bond lengths are in angstroms. Adopted from Ref. 15.

reaction of Br⁻ ($\Delta G^0 = 38 \text{ kJ mol}^{-1}$). The reverse reaction, Cl⁻ + CH₂=CHBr, should be exergic and the barriers should be lower. The halide exchange reactions proceed preferably via the S_NV σ rather than the S_NV π pathway.

The TSs for the $S_N V \sigma$ route are quite loose with a small degree of bond formation and a large degree of bond cleavage. On the other hand, the TSs for the $S_N V \pi$ pathway are relatively tight with a large extent of bond formation and a small degree of bond cleavage. This contrast is clearly demonstrated by the calculated percentages of bond-order changes in going from the initial state to the TS, $\%\Delta n^{\ddagger}$, given in Table 3. The bond formation and cleavage percentages are 30–35 and 55–65 for the $S_N V \sigma$ route and 55–60 and 25–40 for the $S_N V \pi$ route. In the $S_N V \pi$ TS the π bond is partially broken (to a single bond in the limit), while in the $S_N V \sigma$ transition state a second π bond is partially formed (toward a triple bond). That is, triple-bond character has developed in the $S_N V \sigma$ TS.

The loose TS on the $S_N V \sigma$ path is associated with a large energy of deformation (ΔE_{def}) , which is the energy required to deform the reactant, vinyl chloride, to its geometry in the TS without interaction with the nucleophile). This should lead to a high activation barrier. The difference in the ΔE_{def} values for the two routes (Table 3, last column) must be closely related to the relative ease of the σ and π attack. The

| Mechanism | Nu ⁻ | C–Nu | C–Cl | C=C | $\Delta E_{\rm def} \ (\rm kJ \ mol^{-1})$ |
|---------------------------|-----------------|------|------|------|--|
| $\overline{S_N V \sigma}$ | Cl ⁻ | 34.5 | 65.5 | 14.2 | 247.4 |
| | Br ⁻ | 37.3 | 66.2 | 9.1 | 258.0 |
| | SH ⁻ | 30.8 | 57.4 | 2.1 | 204.3 |
| $S_N V \pi$ | C1 ⁻ | 62.3 | 37.7 | 20.5 | 164.9 |
| | Br ⁻ | 62.3 | 43.5 | 16.7 | 195.6 |
| | OH ⁻ | 22.5 | 3.5 | 13.5 | 11.9 |
| | SH ⁻ | 56.3 | 24.0 | 32.0 | 108.4 |

Table 3 Percentage changes in bond order, $\%\Delta n^{\ddagger}$, and deformation energies (ΔE_{def}) on going from reactants to the transition state¹⁵

larger energies of deformation for the $S_N V \sigma$ than for the $S_N V \pi$ pathway were considered to be responsible for the unfavorability of the in-plane σ attack compared with the out-of-plane π attack, as has long been accepted by organic chemists. What factors cause the reverse of this trend in ΔE_{def} values to overall preference for the $S_N V \sigma$ over the $S_N V \pi$ pathway?

As already pointed out the frontier MO levels do not always predict correctly the ease of the reaction pathways. The ground state LUMO symmetry at the α -carbon of the vinylic substrate is therefore not the dominant factor determining the mechanistic preference in the S_NV reactions. Another factor considered, intramolecular *geminal* and *vicinal* σ - σ * type charge-transfer interactions in the TS, is also not responsible for the energetic preference of the S_NV σ pathway in the substitution of unactivated vinylic substrate by halide ions. Examination of σ - σ * proximate delocalization in the TSs showed that such TS stabilizing interactions are larger in the S_NV σ pathway.

Another important energy term in TS interactions is the electrostatic one. In the loose TS of the $S_N V \sigma$ process, a considerable positive charge is developed at the α -carbon, which gives a strong electrostatic (Coulombic) interaction with the anionic nucleophile. The electrostatic interaction energies ΔE_{es} are quite large for the $S_N V \sigma$ reactions involving Cl⁻ and Br⁻ (Table 4). The stabilization due to ΔE_{es} exceeds the larger destabilization due to ΔE_{def} for the $S_N V \sigma$ routes and more than compensates for the large deformation energies in favor of the $S_N V \sigma$ over the $S_N V \pi$ process. Also solvent effects have been studied, using a continuum model, but the relative preference of the two pathways is barely affected by the solvent, acetonitrile (Table 2, ΔG_{AN}^{\ddagger}).

In conclusion, in the substitution reaction of unactivated vinylic substrates the out-of-plane $S_N 2$ pathway with retention $(S_N V \pi)$ is favored with chalcogenides, OH⁻ and SH⁻, while the in-plane $S_N 2$ pathway with inversion $(S_N V \sigma)$ is energetically preferred with halides, Cl⁻ and Br⁻. The $S_N V \pi$ TS is relatively tight and predominantly stabilized by proximate $\sigma - \sigma^*$ charge-transfer interactions. In contrast, the $S_N V \sigma$ TS is relatively loose and stabilized mainly by electrostatic interactions.

| Mechanism | Nu^{-} | C –Nu | C _C1 | H _Nu | H _Cl | Total |
|----------------|-----------------|-------------------|------------------|---------------------------------|--------------------|---------|
| Wieenamsin | Itu | c_{α} -Itu | c_{α} –cr | Π_{α} – Π_{α} | Π_{α} –Cl | Total |
| $S_N V \sigma$ | Cl^{-} | -74.1 | -74.1 | -118.0 | -118.0 | - 384.1 |
| | Br^- | 46.4 | -51.0 | -98.3 | -109.6 | - 305.4 |
| | SH^{-} | -20.5 | -18.8 | - 106.3 | -90.0 | -235.6 |
| $S_N V \pi$ | Cl^{-} | 0.4 | 0.4 | - 34.7 | - 34.7 | -68.6 |
| | Br^{-} | 7.1 | 9.6 | -29.3 | -40.2 | - 52.7 |
| | OH^- | -10.0 | -0.8 | - 165.3 | -10.0 | - 186.2 |
| | SH^{-} | 30.1 | 22.2 | - 33.5 | -23.0 | -4.2 |

Table 4 Calculated energies (ΔE_{es} , kJ mol⁻¹) for the major electrostatic interactions of C_{α} and H_{α} with Nu and leaving Cl¹⁵





The reactions of the vinyl iodonium ions 15a-e with chloride ion (Scheme 12) have been theoretically examined at the MP2/DZ + d level.¹⁴ The transition structures calculated for the reaction of divinyliodonium ion 15e with chloride are shown in Fig. 3. The Cl⁻ first coordinates to the positive iodine to form a λ^3 -iodane intermediate; next, the TSs for the in-plane (S_NV σ) and out-of-plane attack (S_NV π)



Fig. 3 Calculated structures of $(CH_2=CH)_2I^+$ (**15e**) and its λ^3 -chloroiodane and transition states for the $S_NV\sigma$ and $S_NV\pi$ reactions. Bond lengths are in angstroms. Adopted from Ref. 14.

| Substrate | $S_N V \sigma$ | | $ m S_N V \pi$ | | |
|-----------|----------------|----------|----------------|----------|--|
| | Gas phase | Solution | Gas phase | Solution | |
| 15a | 105.0 | 84.5 | 95.4 | 101.7 | |
| 15b | 106.3 | 111.3 | 102.1 | 117.6 | |
| 15c | 115.1 | 138.9 | 94.6 | 112.5 | |
| 15d | 130.5 | 136.8 | 98.7 | 107.5 | |
| 15e | 111.7 | 55.6 | 95.0 | 97.9 | |

Table 5 Energy barriers, ΔG^{\ddagger} (in kJ mol⁻¹) calculated for the reactions of vinylic iodonium ion (15) with Cl⁻ in the gas phase and in acetonitrile¹⁴

of chloride are reached. In the TS of the latter route chlorine interacts with iodine. Thus, this reaction is a quasi-intramolecular process within the λ^3 -iodane intermediate, and is called the ligand coupling reaction. The gas-phase activation energies summarized in Table 5 show that the $S_N V \sigma$ TS is higher than the $S_N V \pi$ TS in spite of the σ symmetry of the LUMO of all vinyl iodonium ions calculated. In the reaction of these charged substrates with an anionic nucleophile, the TSs are neutralized and the loose $S_N V \sigma$ TSs have a larger dipole moment than the $S_N V \pi$ TSs. Hence, solvent effects on the two reaction routes are different, as demonstrated by using a dielectric continuum model calculation (Table 5). This is contrary to the finding for the neutral substrates examined above. For the unsubstituted vinyl derivatives **15a** and **15e**, the solvent effects reverse the mechanistic preference in solution. However, the $S_N V \sigma$ pathways of the β -tert-butyl- and β -chlorovinyl iodonium ions **15c** and **15d** have considerably higher energy barriers than the $S_N V \pi$ or ligand coupling routes both in the gas phase and in solution. These trends are consistent with the experimental observations, presented in the next section.

IN-PLANE SUBSTITUTION $(S_N V \sigma)$ OF VINYL IODONIUM SALTS

Vinyl iodonium salts are good substrates for vinylic nucleophilic substitution owing to the excellent nucleofugality of the iodonio leaving group, which is evaluated to be about 10^{12} times a better leaving group than iodide itself and 10^6 times better than triflate.³⁷ General methods of preparation of this class of iodonium salts were developed only in the 1980s,^{38–43} and it was found that their S_NV reactions give products with stereochemistry ranging from exclusive inversion to complete retention, depending on their structure and on the reaction conditions.^{44,45}

Before discussing the S_NV reactions of vinyl iodonium salts, it is worthwhile mentioning their structures. The tetrafluoroborate salts are essentially dissociated or exist as an ion pair in solution. In the crystalline state the counterion BF_4^- is located in the apical position of a pseudo-trigonal bipyramidal structure with the iodine at the center.³⁹ However, the long atomic distance of I–F(BF₃) suggests that the apical bonding is ionic rather than hypervalent, even in the solid state.⁴⁴ In contrast, the halide salts of vinyl iodonium compounds in solution are in equilibrium with hypervalent λ^3 -haloiodanes (**17**), and also with dihaloiodates (**18**) at higher



Scheme 13

concentration of halide ions (Scheme 13).^{10,11,46–49} The solution of a tetrafluoroborate salt of a vinyl iodonium ion usually shows only a strong UV–Vis absorption band below 250 nm, but upon addition of halide ions a strong new band develops at longer wavelengths (Fig. 4). The new absorption is ascribed to λ^3 -haloiodane 17, and the association constants can be evaluated from the dependence of the absorbance on halide concentrations (Fig. 5). Their values are listed in Table 6 for the iodonium salts 11 and 20–22, given in Scheme 14. The associated forms, however, are less reactive than the free iodonium ions.^{10,11,47,19} Furthermore, there is little information about the association with most other anions. For these reasons, the substrates are simply designated as iodonium rather than iodane, although at higher concentrations of halide ions in solution the reactive species can actually be the λ^3 -haloiodane.

Ochiai and co-workers³² first discovered that the nucleophilic substitution of (E)-1-alkenyl(phenyl)iodonium tetrafluoroborates (e.g., **11**) with tetrabutylammonium halides (except with fluoride) in various solvents at room temperature occurs with complete inversion of configuration to give (Z)-1-haloalkenes (e.g., Z-**12**) (Scheme



Fig. 4 UV absorption spectra of vinyl iodonium tetrafluoroborate **11** in acetonitrile solution containing no additional salt (—) and 0.01 mol dm⁻³ of Bu₄NCl (– – –), Bu₄NBr (––––), and Bu₄NI (– – –) measured at 25°C. Taken from Ref. 11.



Fig. 5 The initial absorbance of **20** at 250 nm in Bu₄NCl solution in acetonitrile at $\mu = 0.10$ and 25°C. The solid curves are the theoretical ones calculated with data given in Table 6. Taken from Ref. 10.

14; also see Scheme 9). Various mechanistic examinations established that the reaction occurs via the $S_N V \sigma$ mechanism,^{10,11} a conclusion supported by theoretical calculations.¹⁴ The side reaction is elimination, yielding a 1-alkyne (e.g., **19**). The product distributions of the reactions of (*E*)-1-decenyl(phenyl)iodonium tetrafluor-oborate (**11**) with halide ions (X⁻) at [Bu₄NX] = 0.1 mol dm⁻³ and 25°C are summarized in Table 7.¹¹ The fraction of elimination product **19** decreases with concentration of halide and in the order, $Cl^- > Br^- > I^-$. Deuterium labeling of

 Table 6
 Product ratios and kinetic parameters for the reactions of 1-alkenyl(phenyl)iodonium salts 11 and 20–22 with chloride ion

| | 20 (R = Me) | $ \begin{array}{l} 11 \\ (R = C_8 H_{17}) \end{array} $ | $\begin{array}{l} 21 \\ (\mathbf{R} = i\text{-}\mathbf{Pr}) \end{array}$ | $\begin{array}{l} 22 \\ (\mathbf{R} = t - \mathbf{Bu}) \end{array}$ |
|---|-----------------------|---|--|---|
| s/e ^a | 72/28 | 54/46 | 41/59 | 0/100 |
| $K_1 ({\rm mol}^{-1}{\rm dm}^3)$ | 5900 | 7200 | 7600 | 5600 |
| $K_2 (\text{mol}^{-1} \text{dm}^3)$ | 16 | 15 | 25 | 11 |
| $k_1 + k'_2 K_1 \; (\text{mol}^{-1} \text{dm}^3 \text{s}^{-1})$ | 0.82 | 2.30 | 2.91 | 4.65 |
| $k_2K_1 + k'_3K_1K_2 \;(\text{mol}^{-2} \text{dm}^6 \text{s}^{-1})$ | 95.7 | 203 | 239 | 0.93 |

Equilibrium and rate constants are defined in Schemes 13 and 16. Values were determined at 25° C and the ionic strength of 0.1 in acetonitrile.¹⁰

^aProduct ratio of substitution/elimination obtained at $[Bu_4NCl] = 0.01 \text{ mol dm}^{-3}$ without adjusting the ionic strength.

Scheme 14

substrate **11** showed that these halides gave **19** with loss of the β -hydrogen (via β -elimination).^{11,32} In contrast, fluoride ion induces exclusively elimination affording **19** in all solvents. In this case, deuterium labeling experiments showed that product **19** was formed with loss of the α -hydrogen (via α -elimination). The kinetic isotope effects are also consistent with this mechanistic variation: α - or β -deuterium exhibits a primary kinetic isotope effect depending on the mechanism. This contrasting behavior of halides is due to the basicity of fluoride (p K_a 3.17), which induces abstraction of the highly acidic α -hydrogen within the λ ³-haloiodane intermediate (Scheme 15).

Solvent effects on the product distribution of the reactions of **11** at 50°C and $[Bu_4NX] = 0.05 \text{ mol dm}^{-3}$ including protic solvents are presented in Table 8.¹¹ The fraction of substitution tends to increase with increasing polarity of the solvent. The substitution products are exclusively in the *Z* form (inversion) in aprotic solvents and in methanol, trifluoroethanol (TFE), and acetic acid. In more highly ionizing solvents such as hexafluoro-2-propanol (HFIP), formic acid, and trifluoroacetic acid (TFA) a small amount of the *E* isomer (*E*-**12**) is also formed. The partial formation of retained product in the highly ionizing solvents at first seemed to be due to ionization of the substrate, i.e. the ion pair mechanism. However, at a lower concentration of chloride ([C1⁻] = 0.01 mol dm⁻³) the fraction of *E*-**12a** was as high as 85%, and as by-product (*E*)-iodoalkene **12c** was formed. From these

| - | | | |
|-----------------|---|-----------------------------------|-----|
| Ialide | Solvent | Z-12 | 19 |
| 21- | CHCl ₃ | 85 | 15 |
| 21- | MeCN | 90 | 10 |
| 21- | THF | 95 | 5 |
| 3r ⁻ | CHCl ₃ | 92 | 8 |
| 3r ⁻ | MeCN | 96 | 4 |
| - | CHCl ₃ | 91 | 9 |
| - | MeCN | 99.6 | 0.4 |
| ;- | MeCN | 0 | 100 |
|]] }r | THF CHCl ₃ MeCN CHCl ₃ MeCN MeCN | 95 92 96 91 99.6 0 | |

Table 7 Product distributions in the reactions of 1-decenyl(phenyl)iodonium salt 11 withvarious halide ions

Obtained with tetrabutylammonium halide at $[Bu_4NX] = 0.1 \text{ mol dm}^{-3}$ and $25^{\circ}C.^{11}$



observations, the retained product is concluded to be produced via the perpendicular attack ($S_N V \pi$), as discussed below in terms of the ligand coupling mechanism.

Kinetic results are also consistent with a mechanism involving mainly the $S_N V \sigma$ pathway. Solvent effects on the rate of the reaction of **11** with bromide at $[Bu_4NBr] = 0.05 \text{ mol dm}^{-3}$ are rather small and tend to decrease with increasing polarity of the solvents. The effect of substituents at the leaving phenyliodonio group is also small: ρ is about 0.75, which should be compared to the substituent effect with $\rho = 1.8$ on the solvolysis of 4-*t*-butyl-1-cyclohexenyl(aryl)iodonium tetrafluoroborates,³⁷ where a vinyl cation is produced in the rate-determining step (see Scheme

| Halide | Solvent ^{<i>a</i>} (Y_{OTs}^{b}) | 12 (Z/E) | 19 | Remarks |
|--------------------------|--|-----------------|----|---------------------|
| Cl- | CHCl ₃ | 61 (100/0) | 39 | |
| Cl^{-} | MeCN | 76 (100/0) | 24 | |
| Cl^{-} | MeOH (-0.92) | 80 (100/0) | 20 | Solvolysis products |
| Cl^{-} | TFE (1.80) | 94 (100/0) | 6 | Solvolysis products |
| Cl^{-} | 97HFIP (3.61) | 92 (98.4/1.6) | 8 | 2 1 |
| Cl^{-} | AcOH (-0.61) | 87 (100/0) | 13 | |
| Cl^{-} | HCO ₂ H (3.04) | 98 (96.9/3.1) | 2 | Solvolysis products |
| Cl^{-} | TFA (4.57) | 100 (85/15) | 0 | 0.7% 12c |
| Cl^{-c} | TFA (4.57) | 100 (15/85) | 0 | 3.5% 12c |
| Br^{-} | MeOH (-0.92) | 84 (100/0) | 16 | |
| Br^{-} | TFE (1.80) | 97 (100/0) | 3 | |
| Br^{-} | AcOH (-0.61) | 86 (100/0) | 14 | |
| Br^{-d} | HCO ₂ H (3.04) | 98 (98.4/1.6) | 2 | |
| Br^{-} | TFA (4.57) | 100 (98.6/1.4) | 0 | |
| Br^{-c} | TFA (4.57) | 100 (85/15) | 0 | 0.6% 12c |

Table 8Product distributions in the reactions of 1-decenyl(phenyl)iodonium salt 11 with
halide ions in various solvents

Obtained with tetrabutylammonium halide at $[Bu_4NX] = 0.05 \text{ mol dm}^{-3}$ and 50°C.¹¹

 a TFE = 2,2,2-trifluoroethanol, HFIP = 1,1,1,3,3,3-hexafluoro-2-propanol, TFA = trifluoroacetic acid.

^bSolvent ionizing power.⁵⁰

^cObtained at $[X^-] = 0.01 \text{ mol dm}^{-3}$.

^dNaBr was used as a salt.



Fig. 6 Rate constants for the reaction of **11** with chloride ion in acetonitrile at $\mu = 0.10$ and 25°C: (•) k_{obsd} ; (Δ) k_s (for substitution); (\Box) k_e (for elimination). The solid curves are calculated with data given in Table 6. Taken from Ref. 10.

25). The secondary kinetic deuterium isotope effects at the α - and β -position of **11** of about 1.1 and 1.1–1.2, respectively, on the rate of substitution are also consistent with the S_NV σ mechanism. Furthermore, the small positive apparent volume of activation obtained at [Bu₄NBr] = 0.05 mol dm⁻³ and 20°C, $\Delta V^{\ddagger} = 4.6$ cm³ mol⁻¹, is not incompatible with this mechanism.¹¹

The dependence of the observed rate constants on the concentration of halide ion $[X^-]$ is complicated due to formation of the iodane adduct in a pre-equilibrium. Kinetic analysis shows that the nucleophilic substitution is bimolecular. A typical rate dependence is shown in Fig. 6 for the reaction of **11** with chloride ion in acetonitrile at 25°C.¹⁰ The curvatures are theoretically simulated according to the reaction scheme given in Scheme 16 and parameters are given in Table 6. Because of the pre-equilibrium formation of the two adducts, iodane and iodate, the intramolecular nature of the β -elimination, and the lower reactivity of iodate, the curvature for the elimination reaction decreases after its initial increase with increasing [Cl⁻]. In contrast, the curvature for the substitution reactions of **11**, and of the iodane with chloride, and the accompanying formation of the unreactive adduct, iodate. The equilibrium constants for the formation of iodane and iodate from the kinetic analysis agree with those obtained from the changes in UV absorbance (Fig. 5).

The effects of the β -alkyl groups of the 1-alkenyliodonium salts 11 and 20–22 on the product ratios and rate constants in their reaction with chloride ion are



summarized in Table 6.¹⁰ The results are consistent with the proposed mechanisms involving in-plane substitution $(S_N V \sigma)$ and β -elimination (Scheme 16). The fraction of substitution decreases in the order $R = Me > n-C_8H_{17} > i-Pr \gg t$ -Bu, suggesting steric hindrance against the in-plane nucleophilic attack. The *t*-butyl derivative **22** only gives the elimination product and the kinetic curve is typical of the intramolecular reaction of the iodane adduct (Fig. 7).

A stereochemical investigation using substrates of opposite configuration (e.g., Z-11 next to *E*-11) is desirable to establish that the substitution reaction is stereospecifically inversion. However, Z-11 does not undergo nucleophilic substitution with halides but yields quantitatively the elimination product, 19. (Z)-1-Alkenyliodonium salts are known to be too labile to be isolated due to their facile *anti-β*-elimination (Scheme 17).⁵¹



Fig. 7 Observed rate constants for the reaction of **22** with chloride ion in acetonitrile at 25°C. The solid curve is calculated with data given in Table 6. Taken from Ref. 10.



Therefore, the reactions of geometrically isomeric iodonium salts of β , β -dialkylvinyl derivatives with halide ions have been investigated.⁴⁹ However, the reactions are not stereospecific and give significant amounts of rearranged products. The results are discussed in Section 4 as a borderline case. The mechanism is obviously different from that of the monoalkylvinyl derivatives.

Other nucleophiles, which give substitution with inversion in their reaction with (E)-1-alkenyliodonium salts, include dialkyl sulfide and selenide,⁴⁵ phosphoroselenoates,⁵² and sulfonates⁵³ (Scheme 18). Amides also react with (E)-1alkenyliodonium salts and give a Z enol formate (Scheme 19).⁵⁴ The mechanism of these reactions, involving very weakly basic or non-basic nucleophiles, may well be in-plane $S_N V \sigma$ attack. Moderately basic nucleophiles such as amines and carboxylates, even dichloroacetate, give α -elimination.

It was found that the tetrafluoroborate anion itself can act as nucleophile: inverted (Z)-1-fluoroalkenes were obtained upon thermolysis of (E)-1-alkenyl(phenyl)iodo-





Scheme 18



nium tetrafluoroborates at 60°C in chloroform (Scheme 20).⁵⁵ In this reaction, tetrafluoroborate and not fluoride is the nucleophile. If free fluoride ion F^- were formed by dissociation of BF_4^- , α -elimination would have occurred. It should be noted that even the *t*-butyl derivative **22** gives some inverted product.

The intramolecular Friedel–Crafts-type reaction of alkenyliodonium salts 23 proceeds efficiently at 40–60°C in various solvents including methanol, acetonitrile, and chloroform (Scheme 21).⁵⁶ The Z isomer of 23c does not undergo ring closure, thus the reaction does not involve the formation of a vinyl cation. The mechanism of the reaction may well be in-plane $S_N V \sigma$ attack by the intramolecular aromatic nucleophile.



Scheme 20



OUT-OF-PLANE SUBSTITUTION (S_NV π) OF VINYL IODONIUM SALTS

The reactions of the β -halogenovinyliodonium salts **24a,b** with halide ions are quite slow, but proceed at 60°C to give a substitution product, (*Z*)-1,2-dihaloalkene **25**, with complete retention of configuration (Scheme 22).^{46,57} No inverted products were detected. As shown in Fig. 8 the rate of the reaction changes with halide concentration in the same way as for the β -elimination. That is, the reaction must occur as an intramolecular process of the λ ³-haloiodane intermediate as does the β -elimination. Another characteristic of the reaction is the formation of the retained iodoalkene **26**. The results are best accounted for by the ligand coupling mechanism within the hypervalent iodine intermediate **27**.^{58–60} The trends have been explained using theoretical calculations on compound **15d** (Scheme 12, Table 5).¹⁴ The β -chloro substituent considerably raises the barrier for the S_NV σ reaction, but does not significantly influence the S_NV π barrier.

The iodane 27 is in equilibrium with its conformational isomer 27'. Coupling between the apical and equatorial ligand should result in 25 from 27 and 26 from 27'. The ligand coupling mechanism can be regarded as a pseudo-intramolecular (unimolecular) variant of the (bimolecular) $S_N V \pi$ pathway, as discussed in the theoretical section (Fig. 3).



Scheme 22



Fig. 8 Observed rate constants for the reactions of **11** (\bigcirc , $10^2 k_{obsd}$, 50° C) and **24b** (•, $10^5 k_{obsd}$, 60° C) with bromide ion in acetonitrile. Solid curves are theoretical ones. Taken from Ref. 46.

The reaction of 1-decenyliodonium salt **11** with halide ions in acidic solvents such as trifluoroacetic acid, formic acid, and HFIP, yields significant amounts of the retained products *E*-**12** (Table 8).¹¹ The reactions in these solvents were quite slow and product *E*-**12** was always accompanied by a small amount of iodoalkene *E*-**12c**. The retained substitution products must be produced via ligand coupling in the intermediate iodane. When the facile $S_N V \sigma$ reaction becomes sluggish, due to the reduced reactivity of the nucleophile (by hydrogen-bond solvation), a second possible $S_N V \pi$ reaction shows up. So, all substitution reactions of 1-alkenyliodonium salts can be interpreted solely in terms of $S_N V2$ mechanisms, mostly $S_N V \sigma$, but accompanied by ligand coupling ($S_N V \pi$) if the nucleophilicity of the halide is reduced by hydrogen-bond solvation.

3 Vinyl cations as S_NV1 intermediates

Vinyl cations are admittedly unstable species in both thermodynamic and kinetic terms, and were recognized as reactive intermediates only in the mid-1960s.⁶¹ Early progress in this field of chemistry was made in the 1970s through studies of solvolysis of vinylic substrates, especially by using the "super" leaving group triflate.^{4,62} This progress has been summarized in the monograph *Vinyl Cations* in 1979.⁵ In the next two decades, modern techniques of chemistry, such as NMR, laser



photolysis, and sophisticated computational methods, extended the scope of vinyl cation chemistry, as reviewed in the book *Dicoordinated Carbocations* in 1997.⁶ Most recently, a new family of compounds, vinyl iodonium salts, allowed the study of simpler, less stabilized vinylic cations thanks to the enormous nucleofugality of the iodonio group.

STABILITY OF VINYL CATIONS

The parent vinyl cation, $H_2C=C^+H$, and the primary vinyl cations RHC=C⁺H, are very unstable both thermodynamically and kinetically and can only be generated under forced conditions in solution. Lucchini and Modena⁶³ suggested the formation of the parent vinyl cation as an intermediate in the acid-catalyzed hydration of ethyne in strong sulfuric acid (>15 mol dm⁻³). Formation of a primary vinyl cation from 2-butyne was proposed on the basis of trapping a rearranged cation with carbon monoxide in the superacid FHSO₃/SbF₅ (Scheme 23).⁶⁴ Fornarini and Speranza^{65,66} used the nuclear decay of tritiated ethene to generate the parent vinyl cation (Scheme 24). Radioactive tritium (³H, half-life, 12.26 years) spontaneously decays to ³He with emission of a β^- particle to give vibrationally excited $[H_2C=CH-^3He]^+$, which converts to the free vinyl cation by heterolysis of the very weak C-³He bond. In benzene as solvent, the Friedel-Crafts product, styrene, is detected after 10–14 months. Photoexcitation of appropriate precursors also generates primary vinyl cations as discussed in the last part of Section 3 and in Section 5.

Scheme 24

In the gas phase, vinyl cations are easily produced by mass spectrometric techniques, which also allow measurement of their heats of formation.⁶⁷ Gas-phase spectroscopic studies show that the smallest vinylic cation, $C_2H_3^+$, has a structure of protonated acetylene (**28**_b) and not that of the classical vinyl cation (**28**_a).⁶⁷⁻⁶⁹



The results of recent *ab initio* MO studies are consistent with the experimental data and show that the bridged form (28_b) is 19 kJ mol⁻¹ more stable than the classical one (28_a) .⁶⁸ Thus, primary vinyl cations generated in solution may well also be bridged, though no solution-phase structural information is available. Theoretical calculations using dielectric continuum models showed that solvation has only a minor effect on the relative stability of the two forms of the vinylic cation 28.⁷⁰ The RISM SCF calculations also suggested similar stabilization of both forms of the cation by solvent.⁷¹ These results indicate that the hydrogen-bridged form of the vinyl cation is also in solution more stable than the open ion.

Table 9 summarizes the relative stabilities of related cations obtained from theoretical calculations and/or gas-phase thermochemical data. The values are based on the isodesmic reaction $RH + C_2H_3^+ \rightarrow R^+ + CH_2=CH_2$, and give an idea how unstable simple vinyl cations are. The stability of the parent vinyl cation is in between that of the methyl and ethyl cation. α -Alkyl substitution markedly stabilizes the cation, and stabilization by an α -phenyl group is still much larger. Such stabilized secondary vinyl cations are common intermediates in vinylic solvolysis and in electrophilic addition to alkynes. A substituent at the β -position also significantly affects the stability of the vinyl cation. A β -alkyl group stabilizes the classical vinyl cation while a β -phenyl group destabilizes it.⁷² 1,2-Participation by the β -substituent stabilizes the cation. The phenyl-bridged vinylenebenzenium ion **29** is quite stable,^{73,74} and is an intermediate in the solvolysis of β -aryl-substituted vinyl derivatives.^{26,75-77}



| $\overline{\mathbf{R}^+}$ | $\Delta E^a \ (\text{kJ mol}^{-1})$ | $-\Delta \text{HIA}^{b} (\text{kJ mol}^{-1})$ |
|--------------------------------------|-------------------------------------|---|
| CH ₃ ⁺ | 97 | 108 |
| н С=С ⁺ н н | 19 | - |
| л н−с́≡с−н | 0 | 0 |
| $CH_3 - CH_2^+$ | - 47 | - |
| ́́́́́́́́́́́́́́́́́́́́́́́́́́́́́́́́́́́́ | - 74 | - 73 |
| $C = C^+ CH_3$ | - 97 | - 96 |
| CH_3 $C=C-CH_3$ H | - 122 | |
| CH_3 $C=C-CH_3$ | - 151 | |
| $H_2C=CH-CH_2^+$ | - 167 | - 134 |
| $\dot{C} = C^{+}CH = CH_{2}$ | | - 138 |
| H C=C ⁺ -Ph H | | - 202 |
| H c=c ⁺ −< | | - 185 |
| -+ | | - 155 |

^{*a*}The energy of the isodesmic reaction of $RH + C_2H_3^+ \rightarrow R^+ + CH_2=CH_2$ calculated by the *ab initio* MO method at MP2(fu)/6-31G* (taken from compilations in Ref. 68).

^{*b*}The hydride ion affinity of the cation ($R^+ + H^- \rightarrow RH$) relative to that of the vinyl cation (taken from a list in Ref. 67).



Because of the preferred linearity of vinyl cations, the stability of cyclic vinyl cations depends on the ring strain or the ring size. Rates of solvolysis of triflates show that the 1-cyclohexenyl cation is 6×10^{-5} -fold less readily generated than the 1-methyl-1-propenyl cation.^{78,79} The 1-cyclopentenyl cation is not formed at all.⁸⁰ On the other hand, 1-cyclobutenyl triflate is 3700 times more reactive than 1-cyclohexenyl triflate.⁸¹ The stability of the cyclobutenyl cation is due to resonance involving cyclopropyl-stabilization of the positive charge.⁶⁸



LEAVING ABILITY OF THE IODONIO GROUP

The kinetic stability of vinyl(phenyl)iodonium salts is largely due to the instability of the corresponding vinyl cations formed on departure of the iodobenzene nucleofuge. Thus, α -phenyl- or α -alkyl-substituted (secondary) vinyl iodonium ions so far could not be prepared as stable salts.⁴³

1-Cyclohexenyliodonium salts are moderately stable due to a compromise between the stability of the secondary vinyl cation and the ring strain of the cyclic structure. Solvolysis of 4-*t*-butyl-1-cyclohexenyl(phenyl)iodonium tetrafluoroborate (**30**) was investigated in various alcoholic and aqueous solutions.³⁷ The main products are those expected for a cyclohexenyl cation intermediate (**31**), the enol ether (**32**) and/or cyclohexanone as well as iodobenzene (Scheme 25).

In addition to these expected solvolysis products, (4-t-butyl-1-cyclohexenyl)iodobenzenes (33) were obtained in yields of 13–15% in methanol and aqueoussolutions and of 30–40% in TFE. The*ortho*derivative*o*-33 is the predominantisomer of these products, and evidently is formed via the internal return of thecontact ion–molecule pair of cyclohexenyl cation (31) and iodobenzene. The rates



Scheme 25

of solvolysis are only slightly (the Winstein–Grunwald m = 0.12)⁸² affected by the solvents employed as shown in Fig. 9. This is compatible with a reaction of an ionic substrate giving an ionic intermediate in the rate-determining step. From comparison of the rate constant of **30** with that of 1-cyclohexenyl triflate,^{78,79} the phenyliodonio group was evaluated to be about 10⁶ times more effective as a nucleofuge than the triflate leaving group. Introduction of an electron-withdrawing substituent in the iodonio group further enhances the leaving ability. These values show that the iodonio group is the most efficient leaving group ever determined quantitatively.



Fig. 9 Solvent effects on the solvolysis rates of 30 at 50°C. Closed circles show data obtained in ethanol–water solvents. Taken from Ref. 37.



Intermediacy of the cyclohexenyl cation was further supported by the observation of carbocationic rearrangement during the solvolysis of 2-methyl-1-cyclohexenyl-(phenyl)iodonium tetrafluoroborate (**34**). The strained bent 2-methyl-1-cyclohexenyl cation (**35**) formed rearranges to the more stable linear secondary vinyl cation **36** (Scheme 26). Solvolysis of **34** in 60% aqueous ethanol at -20° C gave both a rearranged and an unrearranged ketone in the ratio of 14/86. Such a 1,2-rearrangement is characteristic of electron-deficient carbon species. The 2-methyl iodonium salt **34** decomposes quite readily and its rate of solvolysis in 60% aqueous ethanol shows that the 2-methyl group enhances the reactivity of **30** about 250-fold (provided the influence of the 4-alkyl group is small). This is in accordance with the prediction that a β -methyl group stabilizes vinyl cations.

Although no free-radical products were detected under the usual solvolysis conditions, careful examination of the products of **30** in the presence of a good hydrogen atom donor, 1,4-cyclohexadiene, or in 2-propanol, showed formation of about 1% of 4-*t*-butyl-1-cyclohexene. In principle, apparent heterolysis can occur via initial homolysis followed by single electron transfer (SET). However, this possibility is inconsistent with the rate effects of substituents at the leaving iodobenzene. Electron-withdrawing groups, which would destabilize the iodobenzene cation radical, accelerate the solvolysis with $\rho = +1.7$. The major reaction pathway must be rate-determining heterolysis yielding vinyl cation **31** (S_NV1). The free radical product may be formed via reverse SET (Scheme 27), since the ionization potentials of the cyclohexenyl radical and iodobenzene are quite similar.



Scheme 27



VINYLENEBENZENIUM ION INTERMEDIATES

β-Phenylvinyliodonium salts are not expected to afford the primary classical βphenylvinyl cation since this species is thermodynamically less stable than the parent vinyl cation (p. 27). Solvolysis of (*E*)-styryl(phenyl)iodonium tetrafluoroborate (**37**), however, takes place at a reasonable rate at $50-70^{\circ}$ C.^{75,76} Substitution as well as elimination products, **38** and **39**, are formed (Scheme 28). The ratios of substitution/elimination (**38/39**) and the *Z/E* isomeric ratios of **38** depend on the solvent used (Table 10). The only product of trifluoroethanolysis is the (*E*)-styryl ether *E*-**38c**, formed via exclusive substitution with retention. With increasing nucleophilicity/basicity of the solvent, the percentage of inversion in the substitution products increases and so does the percentage of elimination: in methanol and ethanol more than 90% is elimination and one-third of the substitution occurs with inversion. The elimination product **39** is formed via α-elimination due to the basicity of the solvent. But what is the mechanism of the substitution?

In TFE, the α -deuterated substrate **37**(D) gives a 1:1 mixture of α - and β -deuterated *E*-**38c** (Scheme 29). These stereochemical and H/D scrambling results are best explained by a symmetric 1,1-vinylenebenzenium ion intermediate **29**(D).

In acetic acid and methanol, 37(D) affords both stereoisomers of 38, and the deuterium distributions are different in the two isomers: complete scrambling of deuterium between the α - and β -positions in *E*-38 as was the case in TFE, and complete retention of the isotope at the original position in *Z*-38 (Scheme 30, results in methanol). Obviously *E*-38 and *Z*-38 are formed via different mechanisms. The inverted products *Z*-38 are formed presumably via the in-plane S_NV σ pathway. If

| Solvent | Temperature (°C) | 38 (Z/E) | 39 | |
|-------------------|------------------|-----------------|----|--|
| TFE | 50 | 100 (0/100) | 0 | |
| AcOH | 70 | 85 (14/86) | 15 | |
| $MeOH^a$ | 60 | 6 (34/66) | 94 | |
| EtOH ^a | 60 | 7 (30/70) | 93 | |

 Table 10
 Product distribution in the solvolysis of 37 in some solvents⁷⁶

^aContained trifluoroacetic acid (1 mmol dm⁻³).



they were formed via a primary vinyl cation intermediate, the deuterium distribution between the α and β position of *E*-**38** would not have been equal.

Solvent effects on the rate of solvolysis of **37** are also consistent with these conclusions. As Fig. 10 shows, a large scattering is obvious in the plot against the solvent ionizing power parameter, Y_{OTs} ,^{50,83} but the rates correlate with the solvent nucleophilicity parameter, $N_{T.}$.^{84,85} The solvolysis of the ionic substrate via a benzenium ion should not be very dependent on the solvent polarity or nucleophilicity, but the competing pathways of bimolecular substitution (S_NV σ) and α -elimination will be very dependent on the solvent nucleophilicity/basicity. Large amounts of the latter products were formed in the more nucleophilic solvents, consistent with the kinetic results.

The benzenium ion intermediate **29** is formed by participation of the β -phenyl group of **37**. For stereoelectronic reasons the participation should occur from the *trans* position with respect to the nucleofuge. This can be confirmed by comparing the product distributions of the *E* and *Z* isomers of the substrate.

The Z isomer of **37** is too unstable to be isolated as a salt due to its facile β elimination.⁵¹ Therefore, the solvolysis of (*E*)- and (*Z*)-2-phenyl-1-propenyl(phenyl)iodonium tetrafluoroborate (**40**) was examined.⁷⁷ As anticipated, the solvolysis
of the *E* isomer *E*-**40** is much faster than that of *Z*-**40**, e.g., 4000 times in TFE at 60°C.
The distributions of the products from *E*-**40** and *Z*-**40** are quite different (Table 11).







Fig. 10 Solvent effects on the solvolysis rates of **37** obtained at 50°C as plotted against solvent ionizing power Y_{OTs} (a) and nucleophilicity N_{T} (b). Taken from Ref. 76.

The *E* substrate *E*-40 gives mainly the phenyl-migrated substitution product 42 and two elimination products, 44 and 45, which are derived from the phenyl-migrated cation 47 (and/or 46) (Scheme 31). In the less nucleophilic solvents the methyl-migrated product 43 is also formed. In contrast, *Z*-40 gives unrearranged 41 and the methyl-migrated products, 43 and 44. *Z*-40 does not yield any phenyl-migrated products. The results are consistent with participation of the *trans*



Scheme 31

| | | | 2 | | | | | |
|--------------|---------|-------------------|-----------------------|---------------------|-----------------------|-----|-----|-----|
| Substrate | Solvent | Reaction time (h) | 41 $(E/Z)^{a}$ | 42 $(E/Z)^b$ | 43 $(E/Z)^{c}$ | 44 | 45 | PhI |
| E- 40 | MeOH | 1 | 0 | 29 (15/0) | 0 | 28 | 30 | 97 |
| Z-40 | MeOH | 95 | 5.8 (5.8/0) | 0 | 4.9 | 81 | 0 | 87 |
| E- 40 | AcOH | 1 | 0 | 42 (35/7.1) | 8.1 (3.1/5.0) | 35 | 7.3 | 97 |
| Z-40 | AcOH | 122 | 12.8 (5.2/7.6) | 0 | 56 (20/36) | 30 | 0 | 92 |
| E- 40 | TFE | 0.5 | 0 | 20 | 11 | 17 | 6.4 | 85 |
| Z-40 | TFE | 170 | 0 | 0 | 14 | 6.7 | 0 | 22 |

Table 11 Product distributions (in percent yields) in the solvolysis of **40** at $60^{\circ}C^{77}$

For structures of the products, see Scheme 31.

^aTotal yields of **41** and **41C** with the isomer yields of *E*-**41**/*Z*-**41** in parentheses.

^bTotal yields of 42 and 42C with the isomer yields of E-42/Z-42 in parentheses.

^cTotal yields of 43 and 43C with the isomer yields of E-43/Z-43 in parentheses.



 β -substituent (phenyl or methyl) in the departure of the iodobenzene nucleofuge. In the less nucleophilic solvents a further 1,2-hydride shift across the double bond of vinyl cation 47, formed from *E*-40, occurs to give the more stable cation 48, but the reverse rearrangement does not take place. The phenyl group has a higher migratory aptitude than the methyl group. This may be the reason why *Z*-40 affords the unrearranged product 41, and *E*-40 only gives rearranged products. The results indicate that the primary vinyl cation 49 is not formed during the thermal solvolysis of *E*-40 or *Z*-40.

Photosolvolysis of **40** gives quite different results, which suggest formation of the primary cation **49**, as discussed in Section 5.

β -ALKYL GROUP PARTICIPATION

 β , β -Dialkylvinyliodonium salts have a better chance to afford primary vinyl cations than their β -phenyl counterparts due to the stabilizing effect of β -alkyl substituents discussed above. Solvolyis of (*E*)- and (*Z*)-2-methyl-5-phenyl-1-pentenyl(phenyl)iodonium tetrafluoroborate (**50**) gave a variety of extensively rearranged products, as illustrated in Scheme 32 and summarized in Table 12.^{86,87}

The unsymmetrically substituted substrate gave preferentially the products derived from migration of the *trans*-alkyl group, but the products of the *cis*-alkyl migrations were also observed (last column in Table 12). The amount of the latter products was very small in methanol, but substantial in less nucleophilic solvents. If the migration of the alkyl group was solely due to participation in the heterolysis reaction, only the *trans* group could have migrated, and the formation of products from the migration of the *cis*- β -alkyl group could be considered as evidence for the formation of the classical primary vinyl cation **59**. However, the *cis*-alkyl-migrated

| Structure of 50 | Solvent | Time (h) | Solvent Time (h) | Time (h) | Time (h) | PhI | | Yield (%) | | | | | | % Rearr. | Ratio ^c |
|-----------------|---------|----------|------------------|-----------------|-----------------------|-------|-----|-----------|-----------|-----|----|-------|--|----------|--------------------|
| | | | | 51 | 51 $(Z/E)^{a}$ | 52 | 53 | 54 | 55 | 56 | 57 | | | | |
| E | TFE | 30 | 85 | 0.9 (nd) | 15 | 8.9 | 30 | 0.54 | 0.17 | 0 | 98 | 63/37 | | | |
| Ζ | TFE | 80 | 62 | 2.9 (nd) | 11 | 17 | 34 | 0.34 | 0.39 | 0 | 96 | 39/61 | | | |
| Ε | AcOH | 40 | 63 | 2.0 (trace/0.4) | 21 | 3.4 | 50 | 2.7 | $(1)^{b}$ | 0 | 98 | 84/16 | | | |
| Ζ | AcOH | 80 | 70 | 9.0 (1.0/3.9) | 7.7 | 22 | 47 | 0.5 | $(3)^{b}$ | 0 | 90 | 25/75 | | | |
| Ε | EtOH | 14 | 98 | 27 (5.8/12) | 16 | 0.8 | 38 | 12 | 0.9 | 4.6 | 73 | 94/6 | | | |
| Ζ | EtOH | 40 | 89 | 62 (13/38) | 0.8 | 5.6 | 15 | Trace | 3.9 | 11 | 37 | 8/92 | | | |
| Ε | MeOH | 16 | 80 | 26 (6.2/16) | 12 | Trace | 25 | 7.0 | 0.3 | 2.8 | 63 | Large | | | |
| Ζ | MeOH | 46 | 68 | 61 (10.3/31) | Trace | 3.7 | 12 | Trace | 2.7 | 5.4 | 23 | Small | | | |
| E(Z) | MeOH | 1 | 79 | 80 (17/47) | 0 | 0 | 1.7 | 0 | 0 | 9.3 | | | | | |

Table 12Product distribution in the solvoly is of 50 at $60^{\circ}C^{87}$

^aCombined yields of **51** and the aldehyde **51**C obtained after acid treatments are given with percent yields of the Z and E isomers of **51** in parentheses before acid hydrolysis.

^bPartially overlapped with another peak.

^cRatio of **52** + **55** to **53** + **56**



Fig. 11 Energy barrier for the degenerate hydride transfer of 1,2-dimethylvinyl cation calculated at MP2/6-31G*. Data are taken from Ref. 87.

products can also be formed via the very facile interconversion of the secondary vinyl cations, **57** and **58**, by 1,2-hydride shift.

Model calculations of the dimethylvinyl cation system at the MP2/6-31G* level show that the hydrogen-bridged cation is more stable than the secondary cation and that the barrier for the degenerate hydride transfer is very low (<1 kcal mol⁻¹, Fig. 11).⁸⁷ The isomeric allylic cation, which could be generated by a 1,2-hydride shift across the single bond of the secondary vinyl cation, is much more stable than the vinyl cation, but the barrier for this 1,2-hydride shift is very high since allylic conjugation cannot contribute to the stabilization of the transition state for the hydride shift.

Also, the unrearranged enol ether products **51** were obtained, without stereoselectivity. This seems to suggest the intermediacy of the primary cation **59**. However, products **51** in methanol and ethanol were concluded to be derived mostly from the alkylidenecarbene **60** formed by α -elimination (Scheme 33). Products **51** in more polar solvents amount to only a few percent and we now think that their formation is no definite support for the intermediacy of **59**. Alternative direct routes (S_NV σ and S_NV π) to give these products seem to be more reasonable in view of the results obtained with other systems (see the next section and Section 4).

Hinkle and co-workers^{88–90} have studied the thermal decomposition of a number of β , β -dialkylvinyl iodonium triflates, using an iodobenzene leaving group of enhanced nucleofugality, i.e. 4-trifluoromethyl and 3,5-bis(trifluoromethyl) deriva-



Scheme 33



Scheme 34

tives. The 3,5-bis(trifluoromethyl)phenyl iodonium salts are 28 times more reactive than the phenyl salts, and were expected to generate primary vinyl cations. Typical results of the thermolysis of the *E/Z* isomeric triflate salts **61** and **62** in chloroform are given in Scheme 34. As in the solvolysis of **50**, extensive rearrangements and formation of both isomers of the unrearranged substitution products were observed. The results were initially regarded as evidence for the involvement of primary vinyl cation intermediates,⁸⁹ but are now concluded to indicate participation of the *trans* β -alkyl group in the C–I bond heterolysis, followed by a 1,2-hydride shift to give the isomeric rearranged product.⁹⁰ The S_N2 and ligand coupling pathways afford the unrearranged *E* and *Z* products. These conclusions are in full agreement with our views summarized above.

CHIRALITY PROBE APPROACH

The product analysis of solvolysis of unsymmetrically substituted β , β -dialkylvinyl substrates does not provide definitive information about the intermediacy of primary vinyl cations, mainly due to the facile 1,2-hydride shift of the secondary vinyl cations involved. Cyclic vinyl cations are expected to have a high barrier for this rearrangement, since hydrogen-bridged vinylic cations have a structure similar to that of a protonated acetylene and small-ring acetylenes are very strained. In fact, *ab initio* MO calculations (MP2/6-31G*) of the cycloheptenyl cationic system show the barrier for the 1,2-hydride shift to be quite high (23 kcal mol⁻¹) (Fig. 12).⁹¹ So, solvolysis of optically active (*R*)-4-methylcyclohexylidenemethyl(phenyl)iodonium tetrafluoroborate (**63**) should give the optically active (*R*)-4-methylcycloheptenone (**64**) with complete retention of enantiomeric purity if the rearrangement occurs with β -C-C bond participation, producing the chiral secondary cation **65** (Scheme 35).⁹¹ On the other hand, if the primary vinyl cation **66** was generated



Fig. 12 Energy barrier for the hydride transfer of cycloheptenyl cation calculated at MP2/6-31G*. Taken from Ref. 91.

during the reaction, extensive racemization should occur due to the loss of chirality: the linear vinyl cation is achiral. A further advantage of the chiral system **63** is that the two β -substituent groups are identical and the complications arising from different migratory aptitudes of the β -alkyl groups of unsymmetrical substrates such as **50**, **61**, and **62** are avoided.



Scheme 35



Cycloheptanone **64** produced in the solvolysis of (R)-**63** of 69% ee (enantiomeric

excess) in methanol, aqueous methanol, TFE, and HFIP (1,1,1,3,3,3)-hexafluoro-2propanol) in all cases was the *R* isomer of 68–69% ee, the ee being unchanged within experimental error. So, the primary vinyl cation **66** is not formed under the reaction conditions, not even in a highly ionizing non-nucleophilic solvent. Ion **66** is also not produced upon using a better leaving group: the 3-trifluoromethyl derivative of **63** provided the same stereochemical results.

In view of the results reported by Hinkle and co-workers for the reactions of vinyl iodonium triflates in chloroform,^{88–90} the reactions of **63** with weakly nucleophilic sulfonates in chloroform have also been examined.⁹² Reaction of **63** with triflate and mesylate in chloroform gave both unrearranged and rearranged sulfonates, **67S** and **68S** (Scheme 36). Unexpectedly, the products are racemized to various extents: the 7-membered-ring mesylate, **68Ms**, is largely racemized, but the triflate products only moderately. In the framework of a mechanism similar to that given in Scheme 35, there are two possibilities for racemization of **68Ms**. One is formation of the primary cation **66** and the other is a 1,2-hydride shift of the rearranged cation **65**.

The two possibilities can be distinguished by using substrate **63** labeled with ${}^{13}C$ at the exocyclic position (${}^{13}C$ -**63**, Scheme 37). The label will be at the 2-position of



Scheme 37



Scheme 38

product **68Ms**, irrespective of intervention of **66**, but at the 1- and 2-position if interconversion between the enantiomeric ions **65** occurs. NMR analysis of the reaction products shows extensive scrambling of the ¹³C between the 1- and 2-position of **68Ms**. The extent of the scrambling agrees well with the degree of racemization of the product obtained from optically active (*R*)-**63**. That is, the primary cation **66** is obviously not involved in the reaction, and the racemization occurs due to scrambling of the exocyclic carbon atom of the substrate. What is the cause of this scrambling? Can the 1,2-hydride shift in **65**, which was not observed during the solvolysis reaction, be facilitated by added mesylate ion?

In the presence of a small amount of CH_3OD , the reaction of 63 with mesylate afforded **68Ms** containing a considerable amount of deuterium at the 2-position. This deuterium incorporation suggests an elimination-addition mechanism of interconversion of the isomeric cations 65 via cycloheptyne 69 as intermediate. Cycloalkyne 69 could in fact be trapped by tetraphenylcyclopentadienone giving 70 during the reaction (Scheme 38). In this trapping experiment, some adduct 68Ms was still isolated, now with the same optical purity as substrate (R)-63. These results are best accommodated within the mechanism outlined in Scheme 38. Mesylate ion acts on **65** both as a base and a nucleophile to give **69** and (R)-**68Ms**, respectively. The highly strained cycloalkyne then undergoes nucleophilic attack by the mesylate ion affording racemic 68Ms. In contrast, triflate reacts with 65 mostly as a nucleophile to give (R)-68Tf, and 69 is essentially not involved. This was confirmed by performing the reaction of 63 in the presence of both nucleophiles. The stereochemical purity of each of the products was not affected by the presence of the other sulfonate. Triflate cannot react with 69. The cycloalkyne 69 is captured by the nucleophile mesylate, but not reprotonated to give back 65: this would lead to interconversion of (S)- and (R)-65, which does not occur.



Scheme 39

OTHER ATTEMPTS TO GENERATE PRIMARY VINYL CATIONS

Hanack and co-workers⁹³ have suggested the formation of a primary vinyl cation during the solvolysis of cyclohexylidenemethyl triflate (**71**) in 50% aqueous methanol at 140°C. This was based on their observation that the solvolysis gave the rearranged ketone, cycloheptanone as well as cyclohexanecarboxaldehyde in spite of the lack of reactivity of **71** in less polar pure methanol (Scheme 39). A chirality probe approach has now also been applied to this system. Solvolysis of the optically active (*R*)-4-methylcyclohexylidenemethyl triflate (**72**, 73% ee)⁹⁴ was carried out under the same conditions as employed by Hanack *et al.*⁹³ The main product is (*R*)-4-methylcycloheptanone with 71% ee (Scheme 40). That is, the rearranged product retains the optical purity of the substrate, which is definitive evidence against involvement of the primary vinyl cation **66**. The rearrangement occurs via β -C–C bond participation, as in the solvolysis of the iodonium salt **63**. The aldehyde product may be formed via a nucleophilic reaction at the sulfonic sulfur yielding an enol.

Moloney and co-workers⁹⁵ have studied the decomposition of vinyllead triacetates prepared by the reactions of vinylmercury or vinyltin compounds with lead tetraacetate. Vinyllead triacetates generated in chloroform undergo reductive elimination of lead(II) acetate to afford vinyl cations (Scheme 41). The products are those of substitution (acetates) or elimination (alkynes), depending on the reaction







Scheme 41

conditions. However, the α -hydrogen-bearing vinylic derivatives only gave rearranged products or products of β -elimination (Scheme 42). No indication of the formation of primary vinyl cations was found.

The extreme leaving group He generated by nuclear decay of tritium has been demonstrated to give a vinyl cation from tritioethene^{65,66} (see Scheme 24). However, the super leaving groups of triflate and iodonio do not afford primary vinyl cations as described above. Diazonium salts with N₂ as leaving group, which has a leaving ability in between that of He and the super leaving groups, are promising precursors to unstable carbocations. The phenyl cation has been generated from its diazonium precursor.⁹⁶ Several attempts to generate primary vinyl cations by dediazoniation of alkenediazonium salts have been reported, and usually rearranged products were obtained.⁹⁷ One example is given in Scheme 43.⁹⁸ Rearranged products are expected if primary cations are formed, but the possibility of their direct formation via neighboring group participation has never been excluded. More detailed studies of alkenediazonium salts are required to reach a conclusion about the involvement of primary vinyl cations in their chemistry.



Scheme 42





The leaving group ability of halides in electronically excited vinyl halides is greatly enhanced compared to the ground state molecules. This makes photolysis of vinyl halides an excellent entry to vinyl cations.⁹⁹ For a dozen compounds the method has been used to produce cations from α -hydrogen-bearing vinylic bromides and iodides. In all cases studied but one, rearrangement of the initially formed primary vinyl cations to more stable secondary species is much faster than reaction with nucleophiles or bases. For example, iodomethylenecyclohexane (73) exclusively yields a nucleophilic photosubstitution and a photoelimination product derived from the ring-expanded, cycloheptenyl, cation 74 (Scheme 44).¹⁰⁰

In the exceptional case, the reaction of the cation with the nucleophile occurs intramolecularly. Irradiation of the β , β -bis(*o*-methoxyphenyl)vinyl bromide **75** yields a benzofuran derived from **76**, next to a benzofuran and an acetylene derived from **77** (Scheme 45).¹⁰¹ Here, the reaction of the primary vinyl cation **76** with the internal nucleophile can compete with the 1,2-shift across the C=C double bond, yielding the α -aryl-substituted vinyl cation **77**.

4 Borderline mechanisms

Thus far, we have discussed typical vinylic $S_N 2$ and $S_N 1$ reactions. The former involve reactions of good nucleophiles with simple 1-alkenyliodonium salts ($S_N V \sigma$) and unactivated β -halo derivatives ($S_N V \pi$), while the latter reactions occur under poor nucleophilic conditions (solvolysis) if the substrates can give stabilized vinylic cations of secondary or bridged structure. In this section, reactions which have characteristics of both categories are presented.



Scheme 44


SOLVOLYSIS OF β -ALKYLVINYL IODONIUM SALTS

The typical primary vinylic substrate (E)-1-decenyl(phenyl)iodonium tetrafluoroborate (11) displays unexpected reactivity under solvolytic conditions.¹⁰² Its solvolysis was expected to be sluggish due to the instability of the possible primary vinyl cation intermediate, but it proceeded smoothly and as rapidly as that of the cyclohexenyl derivative **30** in alcoholic and aqueous solvents. However, the rates of solvolysis of **11** depend on the nucleophilicity of the solvent and not on the ionizing power. Both substitution (enol ether or aldehyde) and elimination products (**19**) are formed, in ratios which depend on the basicity of the medium (Scheme 46).

The enol ether products are essentially only formed with inversion of configuration and are concluded to be produced via an S_NV2 reaction, and not via an intermediate primary vinyl cation. Both the in-plane $S_NV\sigma$ and the out-of-plane $S_NV\pi$ route are theoretically feasible as S_NV2 pathways, leading to inversion and retention of configuration, respectively (see Scheme 1), and the major substitution reaction of **11** follows the first route, and the minor reaction the second route.



Scheme 46



No indications for the formation of the primary 1-alkenyl cation were detected. If it were formed, its rearrangement via a facile 1,2-hydride shift to the more stable secondary cation should have been observed (Scheme 47). Furthermore, the possibility of the formation of an alkyl-bridged cation and the alkyl-participation mechanism were ruled out on the basis of the lack of isotope scrambling in the product from the deuterium-labeled substrate.⁷⁵

The elimination product **19** in Scheme 46 is formed via α - or β -elimination, depending on the reaction conditions. Fluoride ion induces the α -elimination pathway, while chloride and bromide ions induce β -elimination (see Scheme 15). In neutral methanol, the ratio of α - to β -elimination is about 3:1, but in the presence of weakly basic additives such as dichloroacetate (p K_a 1.35) and dihydrogen phosphate (p K_a 2.12) in methanol the main route is α -elimination.¹⁰²

REACTIONS OF β , β -DIALKYLVINYL IODONIUM SALTS WITH HALIDE IONS

Comparative investigations of geometrically isomeric β -monoalkylvinyl iodonium salts such as **11** are not possible due to the unavailability of the *Z* isomer of 1-alkenyliodonium salts.⁵¹ Therefore, the reactions of the *E*- and *Z*-isomer of β , β -dialkylvinyliodonium salt **50** with halide ions were studied.⁴⁹ Although the main products are those of substitution with inversion, significant amounts of retained products are also formed, as well as small amounts of rearranged products (Scheme 48, Table 13). These results seem to suggest formation of a primary vinyl cation,



Scheme 48

| Structure of 50 | X^{-a} (conc./mol dm ⁻³) | Solvent (μ^b) | Temperature, °C (time, h) | Product yield (%) | | | |
|-----------------|--|--------------------------|---------------------------|-------------------|-----|-------|-------|
| | | | | 78 (Z/E) | 54 | 55 | 56 |
| E | Cl ⁻ (0.04) | MeCN (0.2) | 50 (40) | 70 (75/25) | 11 | 0.5 | 0 |
| Ζ | $Cl^{-}(0.04)$ | MeCN (0.2) | 50 (30) | 87 (9/91) | 2.9 | 0 | Trace |
| Ε | $Cl^{-}(0.001)$ | MeCN (0.2) | 50 (40) | 25 (75/25) | 30 | 0.8 | 0 |
| Ε | $Cl^{-}(0.001)$ | MeCN (0.001) | 50 (40) | 47 (72/28) | 18 | 0.3 | 0 |
| Ζ | $Cl^{-}(0.001)$ | MeCN (0.2) | 50 (45) | 65 (10/90) | 11 | 0 | 0.7 |
| Ζ | $Cl^{-}(0.001)$ | MeCN (0.001) | 50 (40) | 72 (13/87) | 4.0 | 0 | Trace |
| Ε | $Cl^{-}(0.04)$ | CHCl ₃ (0.2) | 50 (20) | 72 (77/23) | 5 | Trace | 0 |
| Ζ | $Cl^{-}(0.04)$ | CHCl ₃ (0.04) | 50 (24) | 61 (77/23) | 0 | 0 | 0 |
| Ε | $Br^{-}(0.04)$ | MeCN (0.1) | 50 (20) | 84 (77/23) | 5.2 | 0.3 | 0.1 |
| Ζ | $Br^{-}(0.04)$ | MeCN (0.1) | 50 (20) | 98 (8/92) | 0.9 | Trace | 0 |
| Ε | $I^{-}(0.04)$ | MeCN (0.04) | 50 (17) | 90 (71/29) | 1.2 | 0 | 0 |
| Ζ | $I^{-}(0.04)$ | MeCN (0.04) | 50 (16) | 98 (8/92) | 0 | 0 | 0 |
| E^{c} | $Br^{-}(0.05)$ | MeOH (0.05) | 60 (25) | 18 (80/20) | 17 | 4.2 | 0.2 |
| Z^{c} | $Br^{-}(0.05)$ | MeOH (0.05) | 60 (25) | 45 (4.4/95.6) | 8.5 | 0 | 2.1 |
| E^{c} | $Br^{-}(0.05)$ | TFE (0.05) | 60 (25) | 11 (86/14) | 39 | 2.4 | 0.5 |
| Z^c | $Br^{-}(0.05)$ | TFE (0.05) | 60 (25) | 37 (2.6/97.4) | 15 | Trace | 0.6 |

 Table 13
 Product distribution in the reactions of 50 with halide ions⁴⁹

^aAdded as Bu₄NX.

^bIonic strength adjusted with Bu₄NClO₄. ^cSignificant amounts of solvolysis products were also obtained.



which would give mainly inversion product via nucleophilic trapping in the ion pair intermediate (ion pair mechanism, Scheme 2), and would afford the rearranged products via 1,2-alkyl shift. However, various lines of evidence are incompatible with this suggestion. The stereoselectivity for inversion is larger with the Z isomer than with the E isomer, and it tends to be larger in more polar media, that is, at higher ionic strength and in polar protic solvents. The halide reactions of **50** were concluded to be mainly $S_N V \sigma$ accompanied by minor pathways of $S_N V \pi$ and ligand coupling (intramolecular $S_N V \pi$) as well as heterolysis with β -alkyl participation leading to the rearranged secondary vinyl cations (see Scheme 32). Formation of 1iodoalkene **78** with retained configuration is considered evidence for the ligand coupling mechanism. However, the very small dependence of the stereoselectivity on the halide concentration indicates similarity in molecularity of the reactions which give the inverted and the retained products. As the inverted product is formed by the bimolecular $S_N V \sigma$ reaction, the bimolecular $S_N V \pi$ pathway to give the retained product must also be involved.

REACTIONS OF β -PHENYLVINYL IODONIUM SALTS WITH HALIDE IONS

The reactions of (*E*)-styryl(phenyl)iodonium salt **37** with halide ions give results similar to those obtained with **50** (Scheme 49, Table 14).⁴⁷ In this case the main product is that of elimination. In aprotic solvents the substitution products (**79**) are predominantly inverted but in TFE a significant amount of retained **79** was also

| Solvent | Time (h) | 79 (Z/E) | 39 | 79/39 |
|-------------------|--|---|---|---|
| MeCN | 8 | 14 (99.8/0.2) | 71 | 16/84 |
| CHCl ₃ | 5 | 6.7 (98.5/1.5) | 73 | 8.4/91.6 |
| MeCN | 3 | 48 (99.4/0.6) | 43 | 53/47 |
| CHCl ₃ | 3 | 26 (98.3/1.7) | 72 | 27/73 |
| Dioxane | 1.5 | 45 (99.6/0.4) | 40 | 53/47 |
| THF | 0.25 | 57 (99.8/0.2) | 49 | 59/41 |
| DMSO | 5 | 19 (98.6/1.4) | 17 | 53/47 |
| TFE | 120 | 17 (70/30) | 12 | b |
| MeCN | 2 | 77 (98.8/1.2) | 10 | 88/12 |
| | Solvent MeCN CHCl ₃ MeCN CHCl ₃ Dioxane THF DMSO TFE MeCN | Solvent Time (h) MeCN 8 CHCl ₃ 5 MeCN 3 CHCl ₃ 3 Dioxane 1.5 THF 0.25 DMSO 5 TFE 120 MeCN 2 | SolventTime (h) 79 (Z/E)MeCN814 (99.8/0.2)CHCl35 $6.7 (98.5/1.5)$ MeCN348 (99.4/0.6)CHCl3326 (98.3/1.7)Dioxane1.545 (99.6/0.4)THF0.2557 (99.8/0.2)DMSO519 (98.6/1.4)TFE12017 (70/30)MeCN277 (98.8/1.2) | SolventTime (h) 79 (Z/E) 39 MeCN814 (99.8/0.2)71CHCl35 6.7 (98.5/1.5)73MeCN348 (99.4/0.6)43CHCl3326 (98.3/1.7)72Dioxane1.545 (99.6/0.4)40THF0.2557 (99.8/0.2)49DMSO519 (98.6/1.4)17TFE12017 (70/30)12MeCN277 (98.8/1.2)10 |

 Table 14
 Product yields in the reactions of 37 with halide ions

Obtained at $[Bu_4NX^-] = 0.1 \text{ mol dm}^{-3}$ and 50°C.⁴⁷

^aTetrabutyl ammonium halide, Bu₄NX.

^bSolvolysis products were also obtained.



obtained. Products from the reaction of the α -deuterated substrate with bromide nucleophile in acetonitrile and TFE were examined by ¹H NMR (Scheme 50). In acetonitrile, no scrambling or loss of deuterium was observed. However, in TFE, the stereochemically retained product showed scrambling of the deuterium between the α - and β -positions while the inverted product did not. These results show that substitution with retention of configuration takes place via different mechanisms in aprotic solvent and in TFE: the S_NV π mechanism gives the normal product of retention but the D/H scrambled product is formed by trapping of the vinylenebenzenium ion intermediate in TFE as observed for trifluoroethanolysis (see Scheme 29).

5 Photochemical reactions

As discussed in the previous sections, the thermal solvolysis reactions of alkenyl iodonium salts usually occur via *trans* β -phenyl- or *trans* β -alkyl-assisted departure of the iodobenzene leaving group, by in-plane vinylic S_N2 substitution or by ligand



Scheme 51

| Solvent | Z-38 | E- 38 | 39 | 80 | 81 | PhI |
|-------------|---------------|---------------|---------------|------|------|--------------|
| MeOH TFE | 0.12 0.007 | 0.39 0.007 | 0.32 <0.02 | 0.04 | 0.01 | 0.75 0.46 |

Table 15 Quantum yields of formation of products in the photosolvolysis of **37** in MeOH and TFE at $20^{\circ}C^{103}$

coupling. The only unambiguous examples of spontaneous dissociative formation of vinyl cations (vinylic S_N1 reactions) seem to be the generation of secondary cations **31** from **30** (Scheme 25) and **35** from **34** (see Scheme 26).

The situation is quite different for the photochemical solvolysis reactions of vinyl iodonium salts. In all cases studied thus far these reactions involve direct, unassisted heterolytic cleavage of the vinylic C–I bond, yielding primary and endocyclic secondary vinyl cations. For example, photosolvolysis of (*E*)-styryl(phenyl)iodonium tetrafluoroborate (**37**) very efficiently yields the products resulting from heterolytic cleavage of the vinylic C–I bond, depicted in Scheme 51.¹⁰³ (Also smaller amounts of products resulting from heterolytic cleavage of the vinylic C–I bond are formed, but they are not discussed here.)

As in the corresponding thermal solvolysis (see Scheme 28) two unrearranged enol ethers, *E*- and *Z*-**38**, are formed but, in marked contrast, also a rearranged enol ether **80** (and its hydrolysis product **81**). The ratios of formation of unrearranged/ rearranged substitution product (**38/80** + **81**) of *E/Z* isomeric substitution product (*E*-**38/***Z*-**38**), and of substitution/elimination (**38** + **80** + **81/39**) depend on the solvent used (Table 15).

The photochemical results are best accounted for by heterolytic cleavage of the vinylic C–I bond, yielding iodobenzene and the open primary styryl cation 82 (Scheme 52). This species can undergo nucleophilic attack by the solvent yielding E/Z-38, or loss of a β -hydrogen yielding 39. Alternatively, 82 can rearrange by a 1,2-hydrogen shift to the α -phenylvinyl cation 83, which upon nucleophilic reaction with the solvent gives 80, and upon β -proton loss 39.

In methanol, the main enol ether products are the unrearranged **38**; in TFE by far the major product is the rearranged enol ether **80**. This difference is due to the difference in lifetime of the primary styryl cation (**82**) in the two solvents. Its



Scheme 52 Mechanism of formation of photoproducts from 37.



Scheme 53

lifetime will be longer in the good cation-stabilizing TFE than in the nucleophilic methanol. This gives **82** more time to rearrange to the more stable **83**.

In methanol the *E*/*Z* ratio of the unrearranged enol ethers (**38**) is 3.3 (i.e. mainly retention of configuration), while in TFE the *E*/*Z* ratio is 1. This difference is ascribed to the coordination of the nucleophilic solvent methanol with the hypervalent iodine (8-I-2) compound **37** to give the corresponding tricoordinated 10-I-3 (λ^3 -iodane) species. This coordination is less important for the less nucleophilic TFE. Excitation of the free iodonium yields **82** in an S_N1 mechanism, which gives substitution products **38** in ~ 1:1 *E*/*Z* ratio (Scheme 53). Excitation of the λ^3 -iodane results in the formation of **82**, iodobenzene and a solvent molecule within a solvent cage. Nucleophilic attack of the solvent molecule within the cage will give preferentially *E*-**38**, in an S_N*i* mechanism.

In methanol the efficiency of formation of elimination product **39** is higher than in TFE. In the cation-molecule pair formed upon photolysis of **37** in methanol (Scheme 53), the methanol not only acts as a nucleophile but also as a base. In TFE the proton has to be abstracted by the less basic agents iodobenzene, the leaving group, or external TFE.

The photochemical solvolysis of (E)- and (Z)-2-phenyl-1-propenyl(phenyl)iodonium tetrafluoroborate (**40**) in methanol and TFE yields the heterolytic vinylic C– I bond cleavage products shown in Scheme 54 and summarized in Table 16.⁷⁷



Scheme 54

| Substrate | Solvent | E- 41 | Z- 4 1 | E- 42 | Z- 42 | E- 43 | Z- 43 | 44 | PhI |
|--------------|---------|--------------|---------------|-------------------|--------------|--------------|--------------|--------|------|
| E- 40 | MeOH | 34.1 | 13.4 | 6.5 | 0 | 8.2 | 8.6 | 15.3 | 66.3 |
| Z-40 | MeOH | 14.8 | 38.5 | 7.5 | 0 | 10.8 | 11.5 | 16.7 | 71.5 |
| E- 40 | TFE | 0 | 0 | 32.6 ^a | 4.4^{a} | 36 | $.6^b$ | $<\!2$ | 81 |
| Z-40 | TFE | 0 | 0 | 11.0 ^c | 2.2^{c} | 54 | $.1^d$ | < 2 | 70 |

Table 16 Product distributions (in percentages) in the photosolvolysis of *E*- and *Z*-40 in MeOH and TFE at $20^{\circ}C^{77}$

^aIn addition 9.9% of ketone 42C is formed.

^bIn addition 10.2% ketone **43C**.

^cIn addition 2% 42C.

^dIn addition 10.4% **43C**.

Five enol ethers are produced, two unrearranged (41), one 1,2-phenyl-shifted (42) and two 1,2-methyl-shifted (43). Unlike the corresponding thermal reaction (see Scheme 31 and Table 11), the product patterns are barely affected by the geometrical structure of 40: the ratio 41/42 + 43 = 2.0 for *E*-40 and 1.8 for *Z*-40. This indicates formation of a common intermediate, the primary vinyl cation 49, from both stereoisomers (Scheme 55). This cation gives 41 and rearranges to the more stable vinylic cations 46-48, which in turn give 42 and 43.

As in the case of 37, the amount of rearranged product is much smaller in methanol than in TFE. Also, preferential retention of configuration in the unrearranged enol ethers 41 was observed in methanol but not in TFE, and



Scheme 55



elimination was found to be far more important in methanol than in TFE. As discussed for **37**, lifetime differences of the initially formed vinyl cation, ion nucleophile reaction inside or outside a solvent cage, and ion base reaction inside or outside a solvent cage (see Scheme 53), account for the solvent effects.

The product mixtures of the photochemical reactions of both **37** and *E*- and *Z*-**40** are strikingly different from those of the corresponding thermal reactions (Scheme 51/Table 15 versus Scheme 28/Table 10; and Scheme 54/Table 16 versus Scheme 31/Table 11). Photolysis of **37** gives a rearranged substitution product, while its thermolysis does not. Photolysis of *E*- and *Z*-**40** gives quite similar product mixtures, while their thermolysis gives quite different mixtures. These marked contrasts mean that the thermal and photochemical reactions of **37** and **40** do proceed through different product-forming reactive intermediates. Whereas primary vinyl cations are involved in the photolysis, they are not in the thermolysis.

A dichotomy in the thermal versus photochemical behavior of vinyl iodonium salts has also been observed in the reactions of **37** in chlorinated alkanes (Scheme 56).^{55,103}

In the thermolysis the *E* isomers of fluoro- and chlorostyrene (*E*-84-F and *E*-84-Cl) are formed exclusively, which indicates the intermediacy of the vinylenebenzenium ion **29** (see Scheme 29) as the species abstracting a fluoride from the counterion BF_4^- , or a chloride from the solvent. Photochemically both *E*- and *Z*-84 are formed, indicating the intermediacy of the primary styryl cation **82** (see Scheme 52). Preferentially the *Z*-isomer of **84-F** is formed. The fluoride-abstracting species must not be **82** itself, but a tight ion-molecule pair of **82** and iodobenzene (see Scheme 53), which hampers the approach of the fluoride donor to the side of **82** where this group leaves.

6 Summary

Vinyl(aryl)iodonium salts are good electrophiles carrying an excellent nucleofuge, an iodoarene, which makes them good substrates for nucleophilic substitution $(S_N 2)$

as well as versatile precursors for vinyl cations (S_N 1). For the vinylic S_N 2 reactions, two pathways are possible: $S_N V \sigma$ and $S_N V \pi$ leading to inversion and retention of configuration, respectively. Primary vinyl cations are unstable both thermodynamically and kinetically, and their thermal generation from vinyl iodonium salts is avoided by participation of the *trans* β -phenyl or β -alkyl groups or an external nucleophile. In contrast, photoexcited vinyl iodonium ions readily undergo heterolysis to give primary cations.

Acknowledgments

The authors are grateful to Professor Masahito Ochiai (Tokushima University) for the fruitful collaboration on the topics summarized in this chapter. They also appreciate the support of their cooperative research by the Yamada Science Foundation.

References

- 1. Rappoport, Z. (1985). Rec. Chim. Pays-Bas 104, 309-349
- 2. Rappoport, Z. (1981). Acc. Chem. Res. 14, 7-15
- 3. Rappoport, Z. (1992). Acc. Chem. Res. 25, 474-479
- 4. Stang, P.J. (1978). Acc. Chem. Res. 11, 107-114
- 5. Stang, P.J., Rappoport, Z., Hanack, M. and Subramanian, L.R. (1979). *Vinyl Cations*. Academic Press, New York
- 6. Rappoport, Z. and Stang, P.J. (eds) (1997). *Dicoordinated Carbocations*. Wiley, Chichester
- 7. Miller, S.I. and Yonan, P.K. (1957). J. Am. Chem. Soc. 79, 5931-5937
- 8. Miller, S.I. (1968). Adv. Phys. Org. Chem. 6, 185-332
- 9. Kelsey, D.R. and Bergman, R.G. (1971). J. Am. Chem. Soc. 93, 1953-1961
- Okuyama, T., Takino, T., Sato, K. and Ochiai, M. (1998). J. Am. Chem. Soc. 120, 2275–2282
- Okuyama, T., Takino, T., Sato, K., Oshima, K., Imamura, S., Yamataka, H., Asano, T. and Ochiai, M. (1998). *Bull. Chem. Soc. Jpn* **71**, 243–257
- 12. Glukhovtsev, M.N., Pross, A. and Radom, L. (1994). J. Am. Chem. Soc. 116, 5961-5962
- 13. Lucchini, V., Modena, G. and Pasquato, L. (1995). J. Am. Chem. Soc. 117, 2297-2300
- 14. Okuyama, T. and Yamataka, H. (1999). Can. J. Chem. 77, 577-583
- 15. Kim, C.K., Hyun, K.H., Kim, C.K. and Lee, I. (2000). J. Am. Chem. Soc. 122, 2294–2299
- 16. Bach, R.D., Baboul, A.G. and Schlegel, H.B. (2001). J. Am. Chem. Soc. 123, 5787-5793
- 17. Clarke, T.C., Kelsey, D.R. and Bergman, R.G. (1972). J. Am. Chem. Soc. 94, 3626–3627
- 18. Clarke, T.C. and Bergman, R.G. (1974). J. Am. Chem. Soc. 96, 7934-7944
- 19. Summerville, R.H. and Schleyer, P.v.R. (1972). J. Am. Chem. Soc. 94, 3629-3631
- 20. Summerville, R.H. and Schleyer, P.v.R. (1974). J. Am. Chem. Soc. 96, 1110-1120
- 21. Rappoport, Z. (1978). Tetrahedron Lett. 1073-1076
- 22. Lucchini, V., Modena, G. and Pasquato, L. (1993). J. Am. Chem. Soc. 115, 4527-4531

- Destro, R., Lucchini, V., Modena, G. and Pasquato, L. (2000). J. Org. Chem. 65, 3367–3370
- 24. Modena, G. and Tonellato, U. (1971). J. Chem. Soc (B) 374
- 25. Bassi, P. and Tonellato, U. (1974). J. Chem. Soc. Perkin Trans. 2 1283
- 26. Stang, P.J. and Dueber, T.E. (1977). J. Am. Chem. Soc. 99, 2602-2610
- 27. Capozzi, G., Lucchini, V., Modena, G. and Scrimin, P. (1977). Tetrahedron Lett. 911-912
- 28. Lucchini, V., Modena, G. and Zaupa, T. (1982). J. Org. Chem. 47, 590-592
- 29. Lucchini, V., Modena, G., Vale, G. and Cappozi, G. (1981). J. Org. Chem. 46, 4720-4724
- 30. Yoshida, Y., Komatsu, M., Ohshiro, Y. and Agawa, T. (1979). J. Org. Chem. 44, 830
- 31. Shainyan, B.A. and Rappoport, Z. (1993). J. Org. Chem. 58, 3421-3428
- 32. Ochiai, M., Oshima, K. and Masaki, Y. (1991). J. Am. Chem. Soc. 113, 7059-7061
- 33. Duraisamy, M. and Walborsky, H.M. (1984). J. Am. Chem. Soc. 106, 5035-5037
- Boche, G., Marsch, M., Müller, A. and Harms, K. (1993). Angew. Chem. Int. Ed. Engl. 32, 1032–1033
- Topolski, M., Duraisamy, M., Rachon, J., Gawronski, J., Gawronska, K., Goedken, V. and Walborsky, H.M. (1993). J. Org. Chem. 58, 546
- 36. Mori, S., Uchiyama, K., Hayashi, Y., Narasaka, K. and Nakamura, E. (1998). Chem. Lett. 111–112
- Okuyama, T., Takino, T., Sueda, T. and Ochiai, M. (1995). J. Am. Chem. Soc. 117, 3360–3367
- 38. Ochiai, M., Sumi, K., Nagao, Y. and Fujita, E. (1985). *Tetrahedron Lett.* 26, 2351–2354
- Ochiai, M., Sumi, K., Takaoka, Y., Kunishima, M., Nagao, Y., Shiro, M. and Fujita, E. (1988). *Tetrahedron* 44, 4095–4112
- Ochiai, M., Toyonari, M., Nagaoka, T., Chen, D.-W. and Kida, M. (1997). *Tetrahedron Lett.* 38, 6709–6712
- 41. Hinkle, R.J., Poulter, G.T. and Stang, P.J. (1993). J. Am. Chem. Soc. 115, 11626-11627
- 42. Hinkle, R.J. and Stang, P.J. (1994). Synthesis 313-316
- 43. Stang, P.J. and Zhdankin, V.V. (1996). Chem. Rev. 96, 1123-1178
- 44. Okuyama, T. (1999). Rev. Heteroatom Chem. 21, 257–275
- 45. Ochiai, M. (2000). J. Organomet. Chem. 611, 494-508
- 46. Okuyama, T., Takino, T., Sato, K. and Ochiai, M. (1997). Chem. Lett. 955-956
- 47. Okuyama, T., Oka, H. and Ochiai, M. (1998). Bull. Chem. Soc. Jpn 71, 1915-1921
- Ochiai, M., Kida, M., Sato, K., Takino, T., Goto, S., Donkai, N. and Okuyama, T. (1999). *Tetrahedron Lett.* 40, 1559–1562
- 49. Okuyama, T., Sato, K. and Ochiai, M. (2000). Bull. Chem. Soc. Jpn 73, 2341-2349
- 50. Schadt, F.L., Bentley, T.W. and Schleyer, P.v.R. (1976). J. Am. Chem. Soc. 98, 7667-7674
- 51. Ochiai, M., Oshima, K. and Masaki, Y. (1991). J. Chem. Soc. Chem. Commun. 869-870
- 52. Yan, J. and Chen, Z.-C. (1999). Tetrahedron Lett. 40, 5757-5758
- 53. Fujita, M., Sakanishi, Y., Furutani, M. and Okuyama, T. (2001). Unpublished results
- 54. Ochiai, M., Yamamoto, Y. and Sato, K. (1999). J. Chem. Soc. Chem. Commun. 1363–1364
- 55. Okuyama, T., Fujita, M., Gronheid, R. and Lodder, G. (2000). *Tetrahedron Lett.* **41**, 5125–5129
- Ochiai, M., Takaoka, Y., Sumi, K. and Nagao, Y. (1986). J. Chem. Soc. Chem. Commun. 1382–1384
- 57. Ochiai, M., Oshima, K. and Masaki, Y. (1994). Chem. Lett. 871-872
- 58. Oae, S. (1986). Croatia Chem. Acta 59, 129-151
- 59. Oae, S. (1996). Pure Appl. Chem. 68, 805-812

- 60. Finet, J.-P. (1998). Ligand Coupling Reactions with Heteroatomic Compounds, pp. 122-129. Pergamon, Oxford
- 61. Grob, C.A. and Cseh, G. (1964). Helv. Chim. Acta 47, 194
- 62. Stang, P.J., Hanack, M. and Subramanian, L.R. (1982). Synthesis 85-126
- 63. Lucchini, V. and Modena, G. (1990). J. Am. Chem. Soc. 112, 6291-6296
- 64. Hogeveen, H. and Roobeek, C.F. (1971). Tetrahedron Lett. 3343-3346
- 65. Fornarini, S. and Speranza, M. (1984). Terahedron Lett. 25, 869-872
- 66. Fornarini, S. and Speranza, M. (1989). J. Am. Chem. Soc. 111, 7402-7407
- Aue, D.H. (1997). In *Dicoordinated Carbocations*, Rappoport, Z. and Stang, P.J. (eds), pp. 105–156. Wiley, Chichester
- 68. Apeloig, Y. and Müller, T. (1997). In *Dicoordinated Carbocations*, Rappoport, Z. and Stang, P.J. (eds), pp. 9–104. Wiley, Chichester
- 69. van Alem, K., Lodder, G. and Zuilhof, H. (2000). J. Phys. Chem. A 104, 2780-2787
- 70. Bagno, A. and Modena, G. (1999). Eur. J. Org. Chem. 2893-2897
- 71. Sato, H. (2001). Unpublished results
- Kobayashi, S., Hori, Y., Hasako, T., Koga, K. and Yamataka, H. (1996). J. Org. Chem. 61, 5274–5279
- 73. Rappoport, Z., Kobayashi, S., Stanger, A. and Bese, R. (1999). J. Org. Chem. 64, 4370-4375
- 74. Gronheid, R. (2001). PhD Thesis, Leiden University, The Netherlands
- 75. Okuyama, T. and Ochiai, M. (1997). J. Am. Chem. Soc. 119, 4785-4786
- 76. Okuyama, T., Ishida, Y. and Ochiai, M. (1999). Bull. Chem. Soc. Jpn 72, 163-170
- 77. Gronheid, R., Lodder, G., Ochiai, M., Sueda, T. and Okuyama, T. (2001). J. Am. Chem. Soc. 123, 8760–8765
- Pfeifer, W.D., Bahn, C.A., Schleyer, P.v.R., Bocher, S., Harding, C.E., Hummel, K., Hanack, M. and Stang, P.J. (1971). J. Am. Chem. Soc. 93, 1513–1515
- 79. Hargrove, R.J. and Stang, P.J. (1976). Tetrahedron 32, 37-41
- 80. Hanack, M., Bentz, H., Märkl, R. and Subramanian, L.R. (1978). Ann. Chem. 1894
- 81. Hanack, M., Carnahan, E.J., Krowczyniski, A., Schoberth, W., Subramanian, L.R. and Subramanian, K. (1979). J. Am. Chem. Soc. 101, 100–108
- 82. Grunwald, E. and Winstein, S. (1948). J. Am. Chem. Soc. 70, 846-858
- 83. Bentley, T.W. and Llewellyn, G. (1990). Prog. Phys. Org. Chem. 17, 121-158
- 84. Kevill, D.N. and Anderson, S.W. (1991). J. Org. Chem. 56, 1845-1850
- Kevill, D.N. (1996). In Advances in Quantitative Structure-Property Relationship, Charton, M. (ed.), vol. 1, pp. 61–115. JAI Press, Greenwich, CT
- 86. Okuyama, T., Sato, K. and Ochiai, M. (1998). Chem. Lett. 1177-1178
- Okuyama, T., Yamataka, H. and Ochiai, M. (1999). Bull. Chem. Soc. Jpn 72, 2761–2769
- 88. Hinkle, R.J. and Thomas, D.B. (1997). J. Org. Chem. 72, 7534-7535
- Hinkle, R.J., McNeil, A.J., Thomas, Q.A. and Andrews, M.N. (1999). J. Am. Chem. Soc. 121, 7437–7438
- McNeil, A.J., Hinkle, R.J., Rouse, E.A., Thomas, Q.A. and Thomas, D.B. (2001). J. Org. Chem. 66, 5556–5565
- (a) Fujita, M., Sakanishi, Y. and Okuyama, T. (2000). J. Am. Chem. Soc. 122, 8787– 8788; (b) Fujita, M., Sakanishi, Y., Nishii, M., Ymataka, H. and Okumaya, T. (2002). J. Org. Chem. (in press)
- (a) Fujita, M., Sakanishi, Y. and Okuyama, T. (2001). J. Am. Chem. Soc. 123, 9190– 9191; (b) Fujita, M., Sakanishi, Y., Nishii, M. and Okumaya, T. (2002). J. Org. Chem. (in press)
- 93. Hanack, M., Märkl, R. and Martinez, A.G. (1982). Chem. Ber. 115, 772-782
- 94. Fujita, M., Yamamoto, A., Sugimura, T. and Okuyama, T. (2001). Chem. Lett. 806-807

- 95. Moloney, M.G., Pinhey, J.T. and Stoermer, M.J. (1990). J. Chem. Soc. Perkin Trans. 1 2645–2655
- 96. Zollinger, H. (1994). Diazo Chemistry. I. Aromatic Heteroaromatic Compounds, pp. 162–177. VCH, Weinheim
- Bott, K. (1983). In The Chemistry of Functional Groups, Supplement C. The Chemistry of Triple-Bonded Functional Groups, Patai, S. and Rappoport, Z. (eds), pp. 671–697. Wiley, Chichester
- 98. Szele, I., Tencer, M. and Zollinger, H. (1983). Helv. Chim. Acta 66, 1704-1709
- 99. Lodder, G. (1997). In *Dicoordinated Carbocations*, Rappoport, Z. and Stang, P.J. (eds), pp. 377–431. Wiley, Chichester
- 100. Kropp, P.J., McNeely, S.A. and Davis, R.D. (1983). J. Am. Chem Soc. 105, 6907-6915
- 101. Suzuki, T., Kitamura, T., Sonoda, T., Kobayashi, S. and Taniguchi, H. (1981). J. Org. Chem. 46, 5324–5328
- 102. Okuyama, T., Imamura, S. and Ishida, Y. (2001). Bull. Chem. Soc. Jpn 74, 543-548
- 103. Gronheid, R., Lodder, G. and Okuyama, T. (2002). J. Org. Chem. 67, 693-702

The Interplay between Redox and Recognition Processes: Models and Devices

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1 Introduction

Understanding the fundamental principles controlling the interdependence of molecular recognition and redox processes is of crucial importance both in the context of biochemistry as well as in the field of materials science,¹ where biological systems can be used as models for pragmatic devices.^{2–5} Potential applications include, among others, molecular shuttles,⁶ switches,⁷ wires,⁸ logical gates,⁹ and biosensors¹⁰ (Fig. 1).

In biological systems, the interdependence between electron-transfer processes and specific non-covalent interactions is a common phenomenon. Enzymes containing redox-active cofactors, such as flavins,¹¹ quinones,¹² nicotinamides,¹³ or pterins¹⁴ use specific interactions between these small molecules and the enzyme to regulate the reactivity of the cofactor. Through hydrogen bonding, aromatic stacking, donor atom– π interaction, and other electrostatic interactions, these systems selectively stabilize oxidation and protonation states by controlling the redox behavior of the cofactor. In addition, enzyme–cofactor interactions provide control over the molecular orbital distribution, and finally, the protein matrix plays a crucial role in both intra- and interprotein electron-transfer processes.^{15,16} In consequence, redox enzymes can be seen as powerful molecular devices that utilize molecular recognition to control all aspects of electron-transfer processes.

In biological systems, the effects of the various electrostatic interactions are difficult to isolate and quantify individually, weak interactions may even go unnoticed. In order to acquire insight into these processes and to apply them to



Fig. 1 Schematic illustration of redox-based recognition-mediated molecular switches discussed within this chapter.

the design of practical devices, model systems utilizing individual effects have to be developed.

This chapter will discuss our investigations into the fundamental non-covalent enzyme–cofactor interactions controlling the redox behavior and physical properties of bio-organic cofactors, as well as studies on supramolecular model systems designed to decouple the effects of the different electrostatic and other interactions. Research towards the design and fabrication of molecular devices based on these same principles will be presented.

2 Non-covalent interactions and redox potentials

Redox-active cofactors are important species in biological systems, playing vital roles in redox and electron-transfer processes. Among the structurally and functionally diverse redox enzymes, flavoproteins containing the flavin cofactors flavin adenine dinucleotide (FAD) or flavin mononucleotide (FMN) are involved in many different biochemical processes serving as a highly versatile redox system.^{11,17–20} These enzymes display considerable variations in their redox behavior using a single electrochemically active unit.²¹ The oxidized flavin (Fl_{ox}) can undergo either one- or two-electron reduction to a flavosemiquinone in an anionic (Fl_{rad}–) or neutral (Fl_{rad}H) form, or the fully reduced flavohydroquinone anion (Fl_{red}H⁻), respectively (Fig. 2a). Similar one- or two-electron reductions are observed in quinone-based systems (Fig. 2b).

The one-electron reduction potential of flavin in flavoproteins varies over a wide (>500 mV) potential range, corresponding to a free energy difference of $\Delta\Delta G > 10$ kcal/mol.¹⁷ Since the redox-active isoalloxazine nucleus is the same in all systems, these extensive variations must arise from apoenzyme–cofactor non-



Fig. 2 Common oxidation and protonation states of (a) flavin and (b) quinone cofactors.

covalent interactions. These interactions include charge attraction and repulsion, dipolar interactions such as hydrogen bonding, aromatic stacking, as well as less prominent donor atom – π interactions. As will be demonstrated within this chapter, all these interactions modulate the redox potentials of the cofactor. Given the essentially electrostatic nature of recognition events,^{22,23} they should be particularly sensitive to changes in the charge density distribution within the host–guest complex. Obviously, significant changes occur upon changing the oxidation state in redox-active molecules. Using well-established B3LYP-DFT calculations, which have been shown to reproduce experimentally available data accurately,^{24,25} these





Fig. 3 Electrostatic potential maps derived from B3LYP-DFT calculations for lumiflavin and ubiquinone in the respective oxidized (Fl_{ox} and Q_{ox}) and radical anion forms (Fl_{rad^-} and Q_{rad^-}). Contour lines are at 60 kcal/mol intervals.



path C

Scheme 1 Proton- and electron-transfer pathways.

changes can be visualized in electrostatic potential maps for both the oxidized and reduced forms. Figure 3 shows the electrostatic potentials for the flavin and ubiquinone systems. As can be seen from the calculations for flavin, the one-electron reduction substantially increases the negative potentials of the carbonyl oxygens O(2) and O(4): the O(2) potential rises from -45 kcal/mol (Fl_{ox}) to -116 kcal/mol (Fl_{rad}-) at the van der Waals radius, making these oxygens more basic, and thus stronger hydrogen bond acceptors.

This reduction also converts the electron-deficient aromatic framework of Fl_{ox} into the highly electron-rich Fl_{rad} . In the oxidized form the flavin nucleus, particularly the central ring, is capable of forming favorable π -stacking interactions with electron-rich aromatic systems, as well as interacting with electron-rich functional groups. The reduced Fl_{rad} on the other hand, exhibits mostly negative electrostatic potentials over the entire framework, hence making interactions with neighboring electron-rich moieties highly unfavorable. The relationship between selective binding and redox potentials can be quantified by the equation

$$\frac{K_{\rm a}(\rm red)}{K_{\rm a}(\rm ox)} = e^{nF/RT(E_{1/2}(\rm bound) - E_{1/2}(\rm unbound))}$$
(1)

where K_a (red) and K_a (ox) are the respective association constants in the reduced and oxidized form, and $E_{1/2}$ (bound) and $E_{1/2}$ (unbound) the standard reduction potentials in the host–guest complex and the free ligand. According to this equation, increased binding in the reduced form (K_a (red) > K_a (ox)) inherently renders the reduction potential in the bound form more positive ($E_{1/2}$ (bound) – $E_{1/2}$ (unbound) > 1).²⁶

Another important feature in redox processes is electron transfer, including proton-coupled electron transfer, a prevalently observed phenomenon. These events follow three basic mechanisms (Scheme 1): proton transfer can occur after (path A), prior to (path B), or simultaneous with (path C) electron transfer.

The first mechanism is often found in the electrochemical reduction and oxidation of quinones and flavins in aprotic media.²⁷ Using cyclic voltammetry (CV) and simultaneous electrochemistry and electron paramagnetic resonance (SEEPR),



Scheme 2 Schematic representation of the ECE pathway for the two-electron reduction of flavin in aprotic media.

studies on flavin in solvents of low dielectric constants, which effectively mimic the hydrophobic environment inside a protein, were carried out. These investigations have established that intermolecular transfer of the acidic imide proton of Fl_{ox} to an already reduced Fl_{rad-} causes the cascade of reactions shown in Scheme 2. Proton transfer occurs as the second step in an electrochemical/chemical/electrochemical (ECE) two-electron reduction to the fully reduced state. Since electron transfer rates.

A similar ECE two-electron reduction was also observed for a ubiquinone– thiourea model system, where the complexing component not only serves as a proton source, but also enables specific recognition, which in turn provides direct control of the switching between one- and two-electron reactions. In these host– guest complexes, hydrogen bonding to the quinone allows direct two-electron reduction to occur via a facilitated proton transfer.²⁸ In this system, however, it appears to be difficult to determine whether proton transfer follows or precedes the second electron transfer. Preliminary studies suggest a pathway between a pre- or post-reduction proton transfer.

In contrast to protonation of the already reduced species, transfer of the proton prior to electron transfer causes significant changes in both the electron-transfer rates and the redox potentials. Simplified, the process can be seen as "acid-catalyzed" electron transfer.²⁹ Upon protonation of the redox-active unit, the reduction potential becomes substantially less negative, an effect analogous to hydrogen bonding. Although the conditions required for pre-protonation of the cofactor are highly acidic (pH < 0), and therefore it would seem unlikely to occur in biological systems, there are systems where kinetic data indicate proton transfer occurs prior to or at least simultaneous with electron transfer.³⁰

The third pathway (see Scheme 1, path C), consisting of a directly coupled electron and proton transfer (i.e., formal hydrogen atom transfer), requires both



Fig. 4 Hydrogen-bonded complex between naphthylimide radical anion (N_{rad}) and a diacyl diaminopyridine receptor (1), and the negative control *N*-methyl naphthylimide (2), in which the methyl group blocks the binding site.

electron and proton tunneling.³¹ While electron tunneling is an accepted and wellunderstood process, proton tunneling is far less common. However, proton tunneling can be readily understood considering the following: since the de Broglie wavelength of a proton possessing an energy of 2 kcal/mol is ~ 0.5 Å, and the distance required to transfer a proton from a hydrogen bond donor to an acceptor is 0.7 Å, proton tunneling coupled to protein–cofactor motion is a reasonable possibility. As can be seen from these considerations, donor and acceptor must be in the close proximity provided by, e.g., hydrogen bonding, in order to enable efficient proton tunneling. This close spatial alignment is a natural feature in enzyme– cofactor complexes and has been replicated in hydrogen-bonding-based model systems.²⁸ In biomolecular systems, evidence for proton tunneling has been obtained from kinetic isotope effect experiments.³² In a study on photoinduced electron transfer in metalloporphyrin model systems, it has been determined that the electron-transfer rate in a hydrogen-bound structure was twice that of a covalently linked analogue.³³

Non-covalent interactions not only modulate the reduction potentials, but also have a significant effect on the orbital geometries of redox-active cofactors.^{34,35} Changing the shape of the highest occupied molecular orbital (HOMO) will affect the oxidation behavior of a reduced species, while altering the geometry of the lowest unoccupied molecular orbital (LUMO) will have an impact on the reduction of an oxidized cofactor. While correlations between orbital geometries and redox behavior could provide further insight into redox processes, direct observations of the structures of HOMO and LUMO are not experimentally feasible. The singly occupied molecular orbital (SOMO) of a radical, on the other hand, can be monitored by EPR spectroscopy, where experimentally accessible hyperfine coupling constants (HFCs) are correlated to theoretical values. As a consequence,



Fig. 5 (a) Acceptor–donor–acceptor three-point hydrogen bonding in flavin. (b) Hydrogenbond complexes between oxidized flavin (Fl_{ox}) and 2,6-diacyl diaminopyridine (1) and 2,6diaminotriazine (3).

cofactors possessing stable radical forms are excellent tools for studying recognition-modulated effects on molecular orbitals.

In a recent study, the effects of hydrogen bonding on the spin density and thus the shape of the SOMO of a radical anion were examined.³⁶ The system studied consists of a complex between the naphthylimide radical anion (N_{rad^-}) and a diacylated 2,6-diaminopyridine (**1** in Fig. 4). This system was chosen because of its resemblance to natural host–cofactor complexes and its relative structural simplicity, which provides readily interpretable EPR spectra and also allows application of complex computational methods. The SEEPR spectrum of N_{rad^-} is significantly altered upon addition of receptor **1**. Control experiments with the non-binding *N*-methyl- N_{rad^-} (**2**) confirmed that the change in the spectra is indeed a result of specific recognition. Spectrum simulation and curve fitting finally provided experimental HFCs.

Using UHF and DFT-B3LYP calculations, theoretical isotropic HFCs were calculated according to the equation

$$a(N) = \frac{8\pi}{3} g_e \beta_e g_N \beta_N \rho(r_N)$$
⁽²⁾

where the HFC of nucleus N, a(N) is directly proportional to the corresponding Fermi contact integral (i.e., spin density) at the nucleus; g_e and g_N denote the respective electronic and nuclear g-factors, while β_e and β_N represent the Bohr and nuclear magneton, respectively. Theoretically obtained HFCs were in good overall quantitative agreement with the experimental results, and can therefore be used to predict properties not experimentally accessible, such as electrostatic potential, electron density, and spin-density distribution. According to the calculations,



Fig. 6 Plot of the binding enthalpy for the flavin–DMF complex as a function of N(3)–O distance in the oxidized and radical anion form, based on B3LYP calculations.

complexation by the receptor shifts spin density away from the carbonyl oxygens involved in the recognition process, and towards carbons 3 and 8 within the naphthalene framework. Considering that hydrogen bonding is a two-electron process, lower, filled orbitals are distorted towards the binding site, which leads to an increase in charge density at the carbonyl oxygens. To maintain orthogonality between wavefunctions, the SOMO is distorted away from the binding site.

3 Redox modulation through hydrogen bonding

Since hydrogen bonding is a directed interaction,³⁷ a distinct array of hydrogen bond donor–acceptor functionalities provides greatly increased specificity to a recognition event. Single-, or even more efficiently, multiple-point hydrogen-bonding interactions can selectively stabilize or destabilize individual oxidation states, thereby modulating the redox potential between these states. An example for three-point hydrogen bonding between enzyme and cofactor can be seen in many flavoproteins, where the two carbonyl oxygens O(2) and O(4) serve as hydrogen

bond acceptors, and the imide N(3)–H function provides a donor site (Fig. 5a).³⁸ As pointed out in the previous section, reduction of the flavin cofactor to Fl_{rad-} is expected to increase charge density on the carbonyl oxygens, making them stronger acceptors. Consequently, hydrogen bonding to O(2) and O(4) selectively stabilizes the Fl_{rad-} oxidation state, thereby facilitating the reduction from Fl_{ox} to Fl_{rad-} . To quantify this prediction, a number of studies have been carried out employing diaminopyridine model receptors mimicking the three-point hydrogen bonding. Using nuclear magnetic resonance (NMR) and CV measurements, it has been determined that complexation of flavin to diacyl diaminopyridine renders the reduction potential of $Fl_{ox} \rightarrow Fl_{rad-}$ 155 mV (3.6 kcal/mol) less negative.³⁹

Binding involving the imide N(3)-H function, however, is expected to show the opposite effect on the redox behavior. Reduction of Flox to Fl_{rad}- increases the charge density at this position, which in turn makes it a weaker donor compared to the oxidized form. Therefore, interactions at this position will make the reduction of Flox to Flrad- more unfavorable. In a recent study the role of N(3)-H has been investigated. Insight into recognition at this site was obtained through calculations on the lumiflavin-dimethylformamide (lumiflavin-DMF) complex providing a model for an isolated imide-carbonyl interaction. Based on B3LYP calculations, the binding enthalpy for Flox and Flrad- as a function of N(3)-O distance was examined. As can be seen in Fig. 6, the Flox-DMF complex shows a behavior expected for a normal hydrogen bond: an optimal N(3)-O distance of 2.87 Å with a maximum binding energy of -5.3 kcal/mol. In contrast, no defined optimum was found for the Fl_{rad}-DMF interaction. In a series of experiments the redox potential of flavin in solvents with and without hydrogen-bond-accepting functionality were determined, demonstrating that hydrogen bonding at N(3)-H makes the reduction from Flox to Flrad- up to 80 mV (1.8 kcal/mol) more difficult. Through the separation of the N(3)-H interaction from the competing set of the two hydrogen-bondaccepting interactions, it is possible to make quantitative predictions on the effect of binding to the carbonyl oxygens: subtraction of the effect of N(3)-H from the potential shifts observed with the diacyl diaminopyridine receptor indicates that hydrogen bonding to O(2) and O(4) makes the reduction potential 227 mV (5.2 kcal/ mol) less negative.40

Simplified, it can be deduced that each individual hydrogen bond donated to the redox-active unit contributes more than 100 mV to the facilitation of the reduction. In contrast, ligands that accept hydrogen bonds from the redox-active unit make the reduction substantially more difficult. Since the number of experimental studies available is very limited, this quantification is obviously quite preliminary. Changing the receptor from diacyl diaminopyridine (1) to a corresponding 2,6-diaminotriazine system (3 in Fig. 5b) leads to a similar binding (K_a (pyridine) = 537 M⁻¹ versus K_a (triazine) = 480 M⁻¹), with the impact on the reduction potential, however, being much weaker ($\Delta E_{1/2}$ (pyridine) = 155 mV versus $\Delta E_{1/2}$ (triazine) = 18 mV).⁴¹ From these results, it becomes clear that the structure–function relationship in these systems is not yet fully understood, and careful parametric studies are required to shed more light on this aspect.



Fig. 7 Structures of polymeric host (4), electroactive guest (5), and control receptor (6); and schematic representation of encapsulation of guest (5) by host (4).

With respect to the applicability of the hydrogen bonding-redox modulation interplay to functional devices, it is equally, if not more important to examine the inverse effect, i.e., the modulation of the specific binding induced by an oxidation/reduction event. In the previously discussed electrochemical studies on the flavin-diaminopyridine system, we observed that near-maximal shifts of the redox potentials occurred at receptor concentrations at which only 22% of the Flox would be bound. This indicates that binding to the receptor is dramatically increased upon reduction of Flox to Flrad-. As already pointed out, this reduction leads to an increased hydrogen-bond-accepting ability of the carbonyl oxygens. The resulting recognition enhancement can be quantified using experimentally accessible thermodynamic variables to determine the free energy ΔG and the association constant K_a for the binding of complex Fl_{rad} receptor. Using these considerations, a 500-fold increase in binding $(K_a(\text{red}) = 250,000 \text{ M}^{-1} \text{ versus } K_a(\text{ox}) = 537 \text{ M}^{-1})^{39}$ was observed, corresponding to a free energy of $\Delta G = -7.3$ versus -3.7 kcal/mol in the oxidized form. This dramatic increase in binding makes the system particularly attractive as a potential molecular device, as will be discussed later in this chapter.

The above examples clearly demonstrate the profound effect of specific recognition on the redox potentials and vice versa. Specific recognition via hydrogen bonding can also be used to provide site-isolation of an electroactive center, preventing unwanted interactions between individual electroactive molecules. In a recent set of experiments, the effect of recognition-driven encapsulation



Fig. 8 Cyclic voltammograms for guest (5) alone (trace 1), complex between 5 and control receptor (6) (trace 2), and complex between polymer (4) and 5 (trace 3).

of a redox-active guest (5) within a globular polymer (4) bearing complementary diaminotriazine functionalities was examined (Fig. 7).⁴² Using NMR measurements, it was established that the flexible polymer "wraps around" an individual receptor molecule, thereby providing efficient encapsulation. This isolation has a significant effect on the redox behavior of the ferrocene, as can be seen from CV experiments (Fig. 8). The CV of a solution of **5** exhibits a sharp reduction wave that arises from precipitation of the oxidized ferrocenium species. Addition of the monomeric control receptor (**6**) had only a small effect on the electrochemical behavior (Fig. 8, trace 2), whereas addition of an equimolar amount of the polymer led to almost complete reversibility of the redox couple (Fig. 8, trace 3). These results clearly demonstrate that encapsulation of **5** within the polymeric host effectively prevents aggregation of the oxidized species.

4 Redox modulation through π -stacking and donor atom- π interactions

In addition to hydrogen bonding, aromatic stacking^{43,44} and interactions of the cofactor π system with electron-rich donor atoms such as oxygen^{45,46} or sulfur³⁸ are among the fundamental forces in molecular recognition. In biological systems, it has been shown that interactions of the electron-deficient π systems of cofactors such as flavins or quinones with electron-rich sidechain residues and substrates have a considerable influence on the redox behavior of the cofactor.⁴⁵ In a study on the *Desulfovibrio vulgaris* flavodoxin, an electron-transfer protein,^{47,48} selective mutations of aromatic residues near the flavin binding site were induced. It was



Fig. 9 Molecular modeling structure (AMBER) of the flavin-host (7) complex and thermodynamic constants at 298 K.

found that the replacement of Tyr98 by Ala made the reduction potential of the $Fl_{rad}H/Fl_{red}H^-$ couple (i.e., the second reduction) 140 mV less negative,⁴⁹ a range comparable to the effect of hydrogen bonding.

To determine directly the effects of interactions involving the π system on the redox behavior of flavin and related compounds, we developed a series of bidentate model receptors. These receptors contain a xanthene-based scaffold that provides for the arrangement of recognition elements in a well-defined geometry (Fig. 9). Using three-point hydrogen bonding, the diaminotriazine moiety at the top orients the flavin over the bottom aromatic surface. The phenyl receptor (7a) is expected to show no other interactions besides hydrogen bonding, and can therefore be used as a reference in order to isolate any other effects. In a first set of experiments,⁵⁰ the aromatic-stacking interactions between receptors 7a-c and flavin were verified independently by fluorescence quenching and NMR titration studies. It was shown that with increasing π -overlap the fluorescence decreased progressively, to an almost complete quenching in the presence of the anthracene system (7c). Through NMR titration experiments, an enhanced association energy with receptors 7b and 7c relative to 7a could be quantified. Due to the additional $\pi - \pi$ interaction in system Fl_{0x} – 7c the association constant shows a 44-fold increase, corresponding to an increase in the free energy of 2.1 kcal/mol over the phenyl control system. Insight into the modulation of the redox potentials upon aromatic stacking was gained using CV experiments. While complexation of flavin with phenyl receptor 7a resulted in

an 18 mV less negative reduction potential compared to the free flavin due to stabilization of Fl_{rad^-} by hydrogen bonding, increased aromatic-stacking interactions resulted in a shift to more negative values making the reduction more difficult. Using the phenyl receptor (**7a**) as a control, it was shown that the stacking interactions shift the reduction potential 81 mV more negative. This shift is indicative of unfavorable interactions between the electron-rich Fl_{rad^-} and the likewise electron-rich anthracene unit, resulting in a more-or-less prominent electrostatic repulsion.

Due to its modular architecture, the same xanthene-based receptor type can be used to study the less obvious interactions between a π -electron-containing redoxactive moiety and electron-rich groups or atoms in close proximity.⁵¹ The rigid scaffold places donor atoms in direct contact with the electron-deficient region of the flavin nucleus, analogous to their flavoenzyme counterparts (Fig. 9). Again, parametric quantification of interactions can be achieved through reference to an electrostatically neutral control receptor. NMR titration and fluorescence quenching studies revealed up to fivefold increases in flavin binding upon incorporation of a donor atom, corresponding to a free energy change of -0.95 kcal/mol relative to the control receptor (7d). In these studies, it has been observed that the increase of association can be correlated with the size and polarizability of the donor atom. However, it is not yet clear which of these factors, size or polarizability contributes more to the interaction.⁵² As expected, CV experiments showed that donor atom $-\pi$ interactions shift the reduction potential to more negative values, indicating unfavorable interactions between two electron-rich systems upon reduction.⁵¹

In addition to these effects, other electrostatic interactions will affect the redox behavior of redox-active species. A study on flavodoxins for instance, has shown that anionic residues as far away as 13 Å have a measurable influence on the FMN cofactor.⁵³ A different study considering the influence of metal cations on the redox modulation established that Ca^{2+} binding to a pyrroloquinolinequinone (PQQ) cofactor made the reduction of the oxidized cofactor 570 mV (13 kcal/mol) less negative.⁵⁴

5 Redox modulation and specific binding applied to the design of molecular devices

As pointed out in the previous sections, biological systems use the interplay of redox events and specific molecular recognition to regulate a wide variety of processes and transformations. We have also shown that the effects of hydrogen bonding, aromatic stacking and other electrostatic interactions on the redox potentials of electroactive molecules can be probed individually, opening the possibility of utilizing biochemical systems as prototypes for pragmatic molecular devices providing the ultimate goal in the miniaturization of mechanical systems.

One common requirement for the functioning and thus utilization of molecular devices is the ability to access them. Hence, the development of suitable interfaces



Fig. 10 Structure of the flavin-receptor (8) complex and association constants at 298 K.

will be crucial. In recent studies, chemical and photochemical stimuli have been used to control the operation of a variety of devices.^{55–59} Application of these devices, however, is complicated by the need to convert electronic input from a computer to either photonic or, even less feasible, chemical stimuli and vice versa. The use of electrochemical activation, on the other hand, provides an opportunity to greatly simplify the application of molecular devices.⁶⁰ Such systems allow direct interfacing to computer systems through the use of electronic input.⁶¹ This simplicity makes redox-controlled systems very attractive candidates as the basis for molecular devices. The implementation of molecular recognition into the control of redox processes should enhance even further the viability of these systems.

Another equally important consideration is the communication of stimuli arising from molecular recognition events as well as the modification of molecular recognition over extended distances. This communication of information is central to the regulation of biological processes and the creation of functional devices. Based on a study of a series of diaminotriazine receptors for flavins which demonstrated that spatially remote substituents alter the hydrogen bond properties,⁴¹ a more recent study extended this approach to even greater distances.⁶² Using a family of chemosensors (**8a**–**e**) for flavins featuring extended conjugation (Fig. 10), the effects of variation of spatially remote (~ 11 Å away) substituents on the efficiency of the hydrogen-bonding-based recognition process were examined.

NMR titration experiments revealed that the strength of the receptor-flavin recognition is highly dependent upon the nature of the substituents present. Binding constants K_a varied from 37 M⁻¹ for the dimethylamino group to 82 M⁻¹ for the chloro substituent, resulting from alterations in the electrostatic potential and the polarizability of the hydrogen-bonding surface. Receptors carrying a donor substituent display a diminished acidity of the amide protons, and thus a weaker hydrogen-bonding capacity. Although this effect is in part compensated by an increased basicity of the triazine ring nitrogen, which would improve its hydrogen bond acceptor strength, the overall effect is a weakened recognition.



Fig. 11 Reversible conformational rearrangement upon reduction of vicinal tricarbonyl (9).

Electron-accepting remote substituents cause the opposite effect and thus result in stronger binding.

To gain insight into the converse effect of binding events on the electronics of the receptor system, effects that are not experimentally accessible, a series of B3LYP-DFT calculations has been carried out. From these calculations, it can be seen that, upon binding, dramatic increases in electron density in the distal ring occur for the chloro receptor. This effect arises from an electron release in the amide protons upon binding. In strong contrast, for the dimethylamino-substituted receptor practically no change in the atomic charges upon binding to flavin is observed, an effect due to the electron-releasing nature of this group, which compensates for the changes in the electron-deficient triazine nucleus. From these results, it can be seen that information is communicated over a nanometer distance, a feature that makes these systems prototypes for molecular wires.^{8,63–65} Another completely different approach to enable communication over extended distances is the utilization of molecular shuttles,^{55,57} where a molecular probe "commutes" between two distinct locations.

A fundamental requirement for the creation of devices is the ability to switch between individual states; considerable interest has been devoted to the design and fabrication of organic-based systems possessing the potential to function as information storage or switching devices. Switching action can be provided, for example, by a conformational change in a molecule⁶⁶ or a considerable alteration of the binding behavior of a host–guest system upon external stimulation, as well as combinations of both. As shown in a recent study, vicinal tricarbonyls can undergo an electrochemically induced conformational switch.⁶⁷ SEEPR and CV experiments demonstrated that vicinal tricarbonyl (9 in Fig. 11) can readily be reduced to the corresponding radical anion. This reduction was also shown to possess a very large peak potential difference ($\Delta E_p = \sim 400$ mV), a diagnostic of dramatic structural reorganization upon reduction.

To understand the nature of this reorganization, a series of B3LYP-DFT calculations was performed. Geometry optimization for 9_{ox} showed that the carbonyl groups are arranged in a helical fashion, due to electrostatic repulsion between the oxygen atoms. Upon reduction, however, the central O(2) carbonyl unit rearranges into an antiperiplanar position (Fig. 11). The calculations suggest that this particular conformation of 9_{rad} is stabilized by 20 kcal/mol over the helical conformation observed in 9_{ox} . This effect is due to an enhanced electron delocalization, as



Fig. 12 Structure of the naphthylimide-anthracyl receptor (7c) complex.

supported by the SEEPR spectrum, where greatly increased spin density in the phenyl ring is observed.

In the previous section, we have demonstrated that reduction of one component in a π -stacked system causes dramatic changes in the stacking behavior; two aromatic systems being attracted to each other in the oxidized form will repel each other upon reduction. This behavior can be utilized in the design of a conformational switch. In order to examine the feasibility of such a system, a series of experiments on a xanthene–naphthylimide system was carried out (Fig. 12).⁵⁰ Similar to the analogous flavin system, binding of the oxidized naphthylimide to the anthracene receptor (**7c**) was greatly increased relative to the phenyl control (**7a**), which proves a strong aromatic-stacking interaction.

SEEPR experiments were used to gain insight into the binding behavior of the reduced naphthylimide radical anion. Practically no change in the EPR spectra of both the anthracene and the phenyl system was observed, indicating a total lack of interaction between the anthracene moiety and N_{rad^-} . This behavior supports a structure for the complex with divergent aromatic surfaces; repulsion of the aromatic moieties causes a conformational change, where one aromatic system is "flipped away" (Fig. 13). Since reduction of the naphthylimide, on the other hand, enhances hydrogen bonding, the loss of aromatic-stacking interactions is in part compensated: the binding constant, as determined through NMR titrations, drops from 1840 M^{-1} in the oxidized form to 1700 M^{-1} upon reduction.



Fig. 13 Schematic illustration of the redox-based conformational switching in the naphthylimide-7c complex.



Fig. 14 Schematic representation of two-pole, three-component switching based on redox-modulated recognition.

In essence, it can be seen that hydrogen bonding and aromatic stacking show complementary modulation of the redox potentials, which in turn leads to redox-state-dependent molecular recognition. This selective recognition behavior can be employed to enhance the efficiency of a molecular switch, where the binding preference of a redox-active guest to each of the two competing hosts is regulated by the oxidation state of the guest. Using the bidentate receptor (**7c**) and a diacyl diaminopyride (**1**), capable of only hydrogen binding, as the two hosts and naphthylimide as a redox-active guest, oxidation-state-mediated binding prefer-



Fig. 15 Predicted structures (AMBER) of the azobenzene receptor (10)–naphthylimide (11) complexes and schematic illustration of redox (11_{ox} , 11_{rad^-} , $11_{red^{2-}}$, left to right) and photochemical switching.

ences, and thus switching capabilities, were examined.⁶⁸ As already discussed, binding of the naphthylimide to the bidentate receptor (**7c**) decreases slightly upon reduction to N_{rad} . Binding of the naphthylimide to diaminopyridine, on the other hand, increases drastically upon reduction to N_{rad} . NMR titration experiments on a mixture of the hosts and the oxidized guest showed a clear preference of the guest for **7c**, due to favorable aromatic stacking. The opposite binding preference in the radical anion state has been demonstrated using SEEPR experiments. Using experimentally obtained association constants and $E_{1/2}$ values, these preferences can be quantified. From thermodynamic considerations, it is shown that N_{ox} has a preference of 1.5 kcal/mol for the π -stacking receptor, whereas N_{rad} - binds 1.9 kcal/mol more strongly to the hydrogen-bonding receptor (Fig. 14). This change in binding preference caused by a change of the oxidation state makes the system an efficient three-component, two-pole molecular switch.

As seen from the previous examples, molecular switches generally rely on twostate (off–on) switching. One way to further enhance the versatility of such devices is to incorporate multi-state switching. A recent study, for instance, used TTF in its three stable oxidation states to achieve three-pole switching.⁶⁹ Diversity of function can be even further increased through a set of orthogonal input stimuli.^{70,71} In a recent study, a system capable of processing orthogonal inputs has been developed.⁷² As shown in Fig. 15, a host–guest complex in which the host (**10**) possesses two discrete states that can be switched photochemically (*trans–cis*), together with a guest (**11**) featuring three stable oxidation states (ox, rad⁻, red²⁻)⁷³ provides six possible discrete states.

Again, the system utilizes the complementarity of hydrogen bonding and aromatic-stacking effects. Initial studies using UV–Vis and NMR spectroscopy demonstrated the rapid and efficient reversible photoisomerization as well as an overall enhanced binding of the guest in the *trans* form, due to aromatic-stacking interactions. Stepwise reduction of the guest, on the other hand, resulted in a gradual decrease of binding strength, due to unfavorable electrostatic interactions between the electron-rich azobenzene moiety and the increasingly charged guest. Using electrochemically determined changes in free energy, the association constants of the receptor in both the *trans*- and *cis*-forms with the guest in all three oxidation states could be quantified. As can be seen in Fig. 15, the six possible binding states possess five distinct affinities, since the 10_{cis} · 11_{ox} and 10_{trans} · 11_{red^2-} complexes exhibit virtually identical binding constants. These five affinities are almost equally spaced exponentially over a 70-fold range, a feature extremely useful to fine-tune recognition properties.

So far, we have shown that redox-modulated recognition, a prevalent feature in biological systems, can be employed in the design of functional devices. As mentioned earlier, one challenge to the efficient application of such systems is the ability to immobilize, order and thus individually address them. One way to provide the desired anisotropy is through the use of colloids functionalized with self-assembled monolayers (SAMs).^{74,75} In a recent model study, a diacyl diaminopyridine-functionalized gold colloid (DAP-Au), capable of binding flavin, has been



Fig. 16 Self-assembly of the hydrogen bonding–aromatic stacking binding site on a colloid (**12**).

prepared.⁷⁶ Using NMR titration experiments, the association constant for the DAP-Au·Fl_{ox} complex was found to be 196 M⁻¹, consistent with previously observed values ($K_a = 193 - 537 \text{ M}^{-1}$). CV measurements showed a 81 mV (1.85 kcal/mol) stabilization of the bound Fl_{rad}. Using equation (1), it has been established that the binding of flavin increases by greater than 20-fold upon reduction of Fl_{ox} to Fl_{rad}. An additional feature of these SAM-functionalized colloids, the mobility of thiols on the SAM surface,⁷⁷ led to an extension of this approach to create multivalent receptors providing templated guest recognition.⁷⁸ To provide a suitable system, thiol-functionalized diacyl diaminopyridine moieties were place-exchanged onto a colloid functionalized with pyrene units capable of aromatic stacking, to provide the bifunctional colloid (**12**). NMR titration experiments demonstrated the expected enhanced binding of this system to Fl_{ox} due to a multivalent interaction. An association constant of $K_a = 323 \text{ M}^{-1}$ was obtained, nearly twice that observed for the DAP-Au·Fl_{ox} system, where only hydrogen bonding can occur.

To explore the use of colloid–flavin binding to self-assemble pre-organized binding sites on the nanoparticle surface, a trifunctional system with dilute hydrogen bonding and aromatic-stacking elements has been employed (Fig. 16). The templation of colloid binding sites was demonstrated by time-course NMR experiments. Enhanced binding over the course of 70 h was observed by monitoring the gradual downfield shift of the flavin N(3)–H proton. The association constant calculated from this shift increased from 168 to 235 M^{-1} , demonstrating the capability of this colloid–SAM system to reorganize to an imprinted system through the use of binding enthalpy.

6 Conclusion and outlook

The interplay between redox and recognition events is of crucial importance in biochemical regulation processes. Specific recognition controls various aspects of the redox process such as redox potentials, electron-transfer pathways, protonation states, and orbital geometries. Series of parametric studies using synthetic model receptors have provided considerable insight into the nature of these regulation processes, probing the individual effects of hydrogen bonding, aromatic stacking, donor atom $-\pi$ and other interactions on the redox properties of electroactive cofactors.

The results obtained from our model studies were found to correlate with effects observed in naturally occurring enzymes, and are furthermore directly applicable to the creation of redox-based pragmatic molecular devices. However, many aspects remain to be explored in both biomolecular and synthetic model systems; one particular challenge will be the understanding of how the individual processes are linked together in biological systems. This understanding will ultimately provide a comprehensive model for the design of highly complex pragmatic devices on the molecular level.

References

- 1. Kaifer, A.E. (1999). Acc. Chem. Res. 32, 62-71
- Roque, A., Pina, F., Alves, S., Ballardini, R., Maestri, M. and Balzani, V. (1999). J. Mater. Chem. 9, 2265–2269
- Diederich, F., Echegoyen, L., Gomez-Lopez, M., Kessinger, R. and Stoddart, J.F. (1999). J. Chem. Soc., Perkin Trans. 2 1577–1586
- 4. Mattei, P. and Diederich, F. (1997). Helv. Chim. Acta 80, 1555-1588
- 5. Lehn, J.-M. (1995). Supramolecular Chemistry: Concepts and Perspectives. VCH, Weinheim
- 6. Anelli, P.L., Spencer, N. and Stoddart, J.F. (1991). J. Am. Chem. Soc. 113, 5131-5133
- Otsuki, J., Tsujino, M., Iizaki, T., Araki, K., Seno, M., Takatera, K. and Watanabe, T. (1997). J. Am. Chem. Soc. 119, 7895–7896
- 8. Zhou, Q. and Swager, T.M. (1995). J. Am. Chem. Soc. 117, 12593-12602
- 9. De Silva, A.P., Gunaratne, H.Q.N. and McCoy, C.P. (1997). J. Am. Chem. Soc. 119, 7891-7892
- 10. Crooks, R.M. and Ricco, A. (1998). J. Acc. Chem. Res. 31, 219-227
- 11. Ghisla, S. and Massey, V. (1989). Eur. J. Biochem. 181, 1-17
- 12. Anthony, C. (1996). Biochem. J. 320, 697-711
- 13. Popov, V. and Lamzin, V.S. (1994). Biochem. J. 301, 625-643
- 14. Blakley, R.L. and Benkovic, S.J. (1985). *Chemistry and Biochemistry of Pterins*. Wiley, New York
- 15. Sharp, R.E., Chapman, S.K. and Reid, G.A. (1996). Biochemistry 35, 891-899
- 16. Tollin, G., Hurley, J., Hazzard, J. and Meyer, T. (1993). Biophys. Chem. 48, 259-279
- 17. For a comprehensive review, see Mueller, F. (ed.) (1990). *Chemistry and Biochemistry of Flavoenzymes*, vols. 1–3. CRC Press, Boca Raton
- 18. Yano, Y. (2000). Rev. Heteroatom Chem. 22, 151–179
- 19. Hasford, J.J., Kemnitzer, W. and Rizzo, C.J. (1997). J. Org. Chem. 62, 5244-5245
- 20. Li, W.S. and Sayre, L.M. (2001). Tetrahedron 57, 4523-4536
- 21. Mueller, F. (1983). In *Topics in Current Chemistry*, Boschke, F.L. (ed.), vol. 108, pp. 71–108. Springer, Berlin
- Klarner, F.G., Burkert, U., Kamieth, M., Boese, R. and Benet-Buchholz, J. (1999). *Chem. Eur. J.* 5, 1700–1707

- 23. Mecozzi, S., West, A.P. and Dougherty, D.A. (1996). Proc. Natl Acad. Sci., USA 93, 10566-10571
- 24. Lozynski, M., Rusinska-Roszak, D. and Mack, H.-G. (1998). J. Phys. Chem. A 102(17), 2899–2903
- 25. Alem, K.V., Sudholter, E.J.R. and Zuilhof, H. (1998). J. Phys. Chem. A 102(52), 10860-10868
- 26. Niemz, A. and Rotello, V.M. (1999). Acc. Chem. Res. 32, 44-52
- 27. Niemz, A., Imbriglio, J. and Rotello, V.M. (1997). J. Am. Chem. Soc. 119, 887-892
- 28. Greaves, M.D., Niemz, A. and Rotello, V.M. (1999). J. Am. Chem. Soc. 121, 266–267
- 29. Fukuzumi, S. (1997). Bull. Chem. Soc. Jpn 70, 1-28
- Graige, M.S., Paddock, M.L., Bruce, J.M., Feher, G. and Okamura, M.Y. (1996). J. Am. Chem. Soc. 118, 9005–9016
- 31. Cukier, R. and Nocera, D. (1999). Annu. Rev. Phys. Chem. 49, 337-369
- 32. Kohen, A. and Klinman, J. (1998). Acc. Chem. Res. 31, 397–404
- 33. de Rege, P.J., Williams, S. and Therien, M. (1995). Science 269, 1409-1413
- 34. Takahashi, M., Matsuo, M. and Udagawa, Y. (1999). Chem. Phys. Lett. 308, 195-198
- 35. Turi, L. (1999). Chem. Phys. 110, 10364-10369
- 36. Niemz, A. and Rotello, V.M. (1997). J. Am. Chem. Soc. 119, 6833-6836
- 37. Scheiner, S. (1997). Hydrogen Bonding. Oxford University Press, Oxford
- Mattevi, A., Obmolova, G., Kalk, K.H., van den Berkel, W.J.H. and Hol, W.G.J. (1993). J. Mol. Biol. 230, 1200–1215
- 39. Breinlinger, E., Niemz, A. and Rotello, V.M. (1995). J. Am. Chem. Soc. 117, 5379-5380
- 40. Cuello, A.O., McIntosh, C.M. and Rotello, V.M. (2000). J. Am. Chem. Soc. 122, 3517-3521
- 41. Deans, R., Cooke, G. and Rotello, V.M. (1997). J. Org. Chem. 62, 836-839
- 42. Galow, T.H., Ilhan, F., Cooke, G. and Rotello, V.M. (2000). J. Am. Chem. Soc. 122, 3595–3598
- 43. Hunter, C.A. (1994). Chem. Soc. Rev. 23(2), 101-109
- 44. Chen, Y., Kampf, J.W. and Lawton, R.G. (1997). Tetrahedron Lett. 38, 5781-5784
- 45. Watt, W., Tulinski, A., Swenson, R. and Wattenpugh, K. (1991). J. Mol. Biol. 218, 195-208
- 46. Fukuyama, K., Matsubara, H. and Rogers, L.J. (1992). J. Mol. Biol. 225, 775-789
- 47. Swenson, R.P., Krey, G.D. and Eren, M. (1987). In *Flavins and Flavoproteins*, Edmondson, D. and McKormick, D. (eds), pp. 98–107. de Gruyter, Berlin
- 48. Zhou, Z. and Swenson, R.P. (1996). Biochemistry 35, 15980-15988
- 49. Swenson, R.P. and Krey, G.D. (1994). Biochemistry 33, 8505-8514
- 50. Breinlinger, E. and Rotello, V.M. (1997). J. Am. Chem. Soc. 119, 1165-1166
- 51. Breinlinger, E., Keenan, C. and Rotello, V.M. (1998). J. Am. Chem. Soc. 120, 8606-8609
- 52. Rotello, V.M. (1998). Heteroatom Chem. 605-606
- 53. Zhou, Z. and Swenson, R.P. (1996). Biochemistry 35, 12443-12454
- 54. Itoh, S., Kawakami, H. and Fukuzumi, S. (1998). J. Am. Chem. Soc. 120, 7271-7277
- 55. Bisell, R.A., Cordova, E., Kaifer, A.E. and Stoddart, J.F. (1994). *Nature* **369**, 133–137
- Schoevaars, A.M., Kruizinga, W., Zijlstra, R.W.J., Veldman, N., Spek, A.L. and Feringa, B.L. (1997). J. Org. Chem. 62, 4943–4948
- 57. Balzani, V., Credi, A., Langford, S.J., Raymo, F.M., Stoddart, J.F. and Venturi, M. (2000). J. Am. Chem. Soc. **122**, 3542–3543
- 58. Gobbi, L., Seiler, P. and Diederich, F. (1999). Angew. Chem., Int. Ed. Engl. 38, 674-678
- Kern, J.M., Raehm, L., Sauvage, J.-P., Divisia-Blohorn, B. and Vidal, P.L. (2000). *Inorg. Chem.* 39, 1555–1560
- Abel, E., Castro, R., McRobbie, I.M., Barbour, L., Atwood, J.L., Kaifer, A.E. and Gokel, G.W. (1998). Supramol. Chem. 9, 199–202

- Collier, C.P., Wong, E.W., Belohradski, M., Raymo, F.M., Stoddart, J.F., Kuekes, P.J., Williams, R.S. and Heath, J.R. (1999). *Science* 285, 391–394
- 62. Deans, R., Cuello, A.O., Galow, T.H., Ober, M. and Rotello, V.M. (2000). J. Chem. Soc., Perkin Trans. 2 1309–1313
- 63. Swager, T.M. (1998). Acc. Chem. Res. 31, 201-207
- Jestin, I., Frere, P., Blanchard, P. and Roncali, J. (1998). Angew. Chem., Int. Ed. Engl. 37, 942–945
- Bumm, L.A., Arnold, J.J., Cygan, M.T., Dunbar, T.D., Burgin, T.P., Jones, L., Allara, D.L., Tour, J.M. and Weiss, P.S. (1996). *Science* 271, 1705–1707
- 66. Monahan, C., Bien, J.T. and Smith, B.D. (1998). J. Chem. Soc., Chem. Commun. 431-432
- 67. Galow, T.H., Cuello, A.O. and Rotello, V.M. (2000). Tetrahedron Lett. 41, 9489–9492
- Deans, R., Niemz, A., Breinlinger, E. and Rotello, V.M. (1997). J. Am. Chem. Soc. 119, 10863–10864
- Ashton, P.R., Balzani, V., Becher, J., Credi, A., Fyfe, M.C.T., Mattersteig, G., Menzer, S., Nielsen, M.B., Raymo, F.M., Stoddart, J.F., Venturi, M. and Williams, D.J. (1999). J. Am. Chem. Soc. 121, 3951–3957
- 70. Doron, A., Katz, E., Tao, G.L. and Willner, I. (1997). Langmuir 13, 1783-1790
- 71. Gilat, S.L., Kawai, S.H. and Lehn, J.-M. (1995). Chem. Eur. J. 1, 275-284
- 72. Goodman, A., Breinlinger, E., Ober, M. and Rotello, V.M. (2001). J. Am. Chem. Soc. 123, 6213–6214
- Osuka, A., Yoneshima, R., Shiratori, H., Okada, T., Taniguchi, S. and Mataga, N. (1998). J. Chem. Soc., Chem. Commun. 1567–1568
- Andres, R.P., Bein, T., Dorogi, M., Feng, S., Henderson, J.I., Kubiak, C.P., Mahoney, W., Osifchin, R.G. and Reifenberger, R. (1996). *Science* 272, 1323–1325
- 75. Liu, J., Xu, R. and Kaifer, A.E. (1998). Langmuir 14, 7337-7339
- 76. Boal, A.K. and Rotello, V.M. (1999). J. Am. Chem. Soc. 121, 4914-4915
- Badia, A., Cuccia, L., Demers, L., Morin, F. and Lennox, R.B. (1997). J. Am. Chem. Soc. 117, 2682–2692
- 78. Boal, A.K. and Rotello, V.M. (2000). J. Am. Chem. Soc. 122, 734-735
The Physical Organic Chemistry of Fischer Carbene Complexes

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1 Introduction

WHAT ARE FISCHER CARBENE COMPLEXES?

Fischer carbene complexes belong to a class of compounds commonly referred to as transition metal carbene complexes. The 1983 book by Dötz *et al.*¹ provides an excellent introduction to many of the most important general aspects of these

complexes. A common feature of transition metal carbene complexes is that they contain a formal carbon-to-metal double bond as schematically represented below:



The electronic structure of these complexes can be visualized as arising from a coupling of the carbene fragment, CR_1R_2 , to the metal fragment, L_nM , in the following way.²⁻⁴ The carbon of the carbene fragment is sp² hybridized. The two electrons that are not involved in the C–R bonds are assumed to be in the lower-lying sp² orbital, leaving the p orbital unoccupied (Fig. 1a). Depending on the identity and the oxidation state of the metal, the ligands and the R-groups attached to the carbene carbon, two kinds of bonding situations arise. One leads to Fischer-type complexes, the other to Schrock-type complexes.

The Fischer carbene complexes are characterized by late-transition metals of low oxidation state, e.g., Cr(0), W(0), Fe(0), Re(0), etc.; they have π -acceptor ligands such as CO or NO at the metal and at least one π -donor group such as OMe, SMe, or NMe₂, etc. at the carbene carbon. The bonding situation is shown in Fig. 1b. The σ -bond is formed by donation of the lone pair on the carbene fragment to the metal. The π -bond may be understood as the result of back donation from a metal d_{π} orbital to the carbon p orbital. Because the energy level of the metal d_{π} orbital is lower than that of the carbene p orbital, the π -electrons remain largely on the metal fragment (Fig. 1b). As a consequence, the carbene carbon becomes highly electron deficient; this explains the requirement for a π -donor group that can partially compensate for this electron deficiency, as shown in **2** for a methoxy carbene complex. That the zwitterionic structure contributes extensively to the resonance hybrid is evidenced by the fact that, at low temperature, the complex **3** shows two separate ¹H NMR signals for the methoxy protons.^{1,5}





Fig. 1 The relative energies of the $M(d_{\pi})$ and the $C(p_z)$ orbitals control the electrophilic or nucleophilic character of the carbene. (a) The orbitals of free CH₂. (b) If the $M(d_{\pi})$ levels are lower in energy, a Fischer carbene will result. (c) If the $M(d_{\pi})$ levels are higher in energy, a Schrock carbene will result. Shading represents occupied orbitals. (Figure taken from Crabtree, Ref. 4, by permission from Wiley & Sons.)



This situation is due to the relatively high barrier to rotation of the C–O bond, making the interconversion between the two isomers (**3a** and **3b**) slow on the NMR time scale. Additional evidence for the π -donor effect of alkoxy as well as other groups (mainly alkyl amino) comes from ⁵³Cr NMR studies⁶ and theoretical calculations.⁷ Nevertheless, the carbene carbon remains electrophilic, a central feature of Fischer carbenes.

Schrock carbene complexes are characterized by early transition metals of higher oxidation state, e.g., Ta(V), W(VI); they have non- π -acceptor ligands such as alkyl, η -C₅H₅, Cl, etc. and mainly alkyl groups or hydrogen but no π -donors attached to the carbene carbon. Just as for the Fischer carbenes, the σ -bond is the result of donation of the carbene lone pair to the metal. However, the polarization of the π -bond is such that most of the electron density is on the carbene carbon because here the metal d $_{\pi}$ orbital is higher in energy than the carbene p orbital (Fig. 1c). Crabtree's⁴ description is particularly illuminating: "One way of looking at this is to say that the two electrons originally in M(d $_{\pi}$) transfer to the more stable C(p_z) orbital, oxidizing the metal by two units and giving a CR₁R₂²⁻ ligand. The system can, therefore, be seen as a metal-stabilized carbanion acting as a σ and π donor to the metal translates into the Schrock carbene carbon acting as an X₂ ligand, just as the oxo group acts as O²⁻ in a complex such as Re(=O)Cl₃(PPh₃)₂ or Re(=O)Me₄."

The extensive and ever increasing activity in the area of transition metal carbene complexes has been mainly driven by their usefulness in synthesis. To this date, the chemistry of the Fischer-type complexes has enjoyed the most extensive development and the present chapter will deal exclusively with this type. The key to the rich chemistry of the Fischer carbene complexes is that they can undergo reactions at several sites; they include modification of the carbene fragment, reaction on the side chain (R-group) or the metal (ligand exchange or change in the oxidation state), insertion of unsaturated organic molecules into the metal–carbon bond, etc. After such elaborations of the carbene complex, removal of the metal to complete the synthesis of the target organic molecule is usually a simple task.

The present chapter cannot possibly do justice to this rich chemistry and no such attempt will be made. Instead the reader is being referred to some of the main reviews that have been published on this subject over the last 20 years. References 8, 11-15, 18, 22 and 23 are reviews of broad scope, while Refs. 9, 10, 16, 17, 19–21,

24 and 25 cover many specialized topics such as carbene and carbyne complexes of ruthenium, osmium, and iridium,⁹ cyclopropanation reactions with olefins,¹⁰ monoand dinuclear (pentamethylcyclopentadienyl)iron carbene complexes,¹⁶ applications of carbene complexes to amino acid synthesis,¹⁷ carbene–alkyne–alkene cyclizations,¹⁹ β -amino substituted α , β -unsaturated Fischer carbene complexes,²⁰ 1-(alkynyl)carbene complexes,²¹ 1-metala-1,3,5-hexatrienes and related compounds²⁴ and the great range of products that can be obtained from α , β -unsaturated carbenepentacarbonylchromium complexes.²⁵

The most prominent systems studied thus far are the Fischer carbene complexes of the Group 6 metals, i.e., Cr, Mo and W, e.g., **4**. One important process that will be discussed at some length in this chapter is nucleophilic substitution, e.g., the replacement of the MeO group by a group with a different heteroatom such as an amino or thioalkyl group.¹ This reaction proceeds via a tetrahedral intermediate (equation 1) and is similar to nucleophilic substitutions on carboxylic esters.



$$(CO)_{5}W = C \bigvee_{Ph}^{OCH_{3}} + Nu^{-} \longleftrightarrow (CO)_{5}W - \bigcup_{Nu}^{OCH_{3}} + CH_{3}O^{-}$$

$$(CO)_{5}W = C \bigvee_{Ph}^{Nu} + CH_{3}O^{-}$$

$$(1)$$

Another common reaction is based on the acidity of the α -carbon proton, i.e., the carbon attached to the carbone carbon, a topic that will be discussed in detail in this chapter. The carbanion that results from the deprotonation of the α -carbon can act as a nucleophile²⁶ and react with a variety of electrophiles, especially in the presence of Lewis acid catalysis, to form many different products (see, e.g., equations 2 or 3^{27,28}). The basicity and nucleophilicity of the carbanion can be enhanced by substituting the MeO group by a Me₂N group²⁹ or replacing a CO ligand with an electron donating one, e.g., R₃P.³⁰



$$(CO)_5Cr = C \xrightarrow{OMe}_{CH_3} + ArCH = O \xrightarrow{Et_3N}_{TMSCl} (CO)_5Cr = C \xrightarrow{OMe}_{6} Ar$$
(3)

In the presence of weak bases such as pyridine and others the reaction of the carbanion with BH⁺ may lead to enol ether formation.^{23,31–33} The mechanism of this reaction is usually assumed to involve protonation of the carbanion on the metal followed by a reductive-elimination (equation 4)²³ but evidence discussed at the end of this chapter suggests a different mechanism.



SCOPE OF THIS CHAPTER

The thrust of this chapter deals with reactions of Fischer carbene complexes that have been the subject of kinetic and/or thermodynamic studies. The number of these is relatively limited. They include the reactions of equations (1) and (4), proton transfers that generate carbanions such as 5 (see equation 2), reactions at the metal center such as the loss or exchange of ligands as well as rearrangement reactions.

On the other hand, there have been hardly any kinetic investigations of the many reactions that have proven to be among the most useful or unique applications of Fischer carbene complex chemistry. Hence they are not part of this chapter although a few of the most prominent ones need to be mentioned. They include cyclopropanation of electrophilic olefins^{8,10,23,34} (e.g., equation 5), metathesis (e.g., equation 6)^{1,23,35–37} and reactions of α , β -unsaturated carbene complexes such as **6** (see equation 3) that lead to what de Meijere *et al.*²⁵ have called an





"incredible range of products," including [4 + 2], [3 + 2] and [2 + 2] cycloadditions, Michael additions, aldol reactions, intramolecular cyclizations and, most prominently, the Dötz reaction.^{8,18,23,25,38} This latter reaction, whose mechanism is quite complex, involves the combination of unsaturated alkoxy carbene complexes with alkynes to yield hydroquinone derivatives (e.g., equation 7).

$$(CO)_5Cr = C \xrightarrow{OMe}_{C_6H_5} + R_1 - C \equiv C - R_2 \longrightarrow OH \xrightarrow{OH}_{C_1(CO)_3} Cr(CO)_3 \qquad (7)$$

It should be noted that a key element in most of these reactions is that they are initiated by the coordination of the substrate to the metal.³⁹ Since Fischer carbene complexes are coordinatively saturated, the coordination of the substrate needs to be preceded by the loss of a ligand (see, e.g., equation 8, S = substrate). This ligand loss can be initiated thermally as well as photochemically. This loss of CO is usually the rate-limiting step which is a major reason why kinetic experiments give little information about the complex steps that follow the initial ligand loss.

$$(CO)_5 Cr = C \begin{pmatrix} OMe \\ R \end{pmatrix} \xrightarrow{-CO} (CO)_4 Cr = C \begin{pmatrix} OMe \\ R \end{pmatrix} \xrightarrow{S} (CO)_4 Cr = C \begin{pmatrix} OMe \\ R \end{pmatrix}$$
(8)

Last, but not least, the rich photochemistry^{17,40–43} of Fischer carbene complexes that has found important synthetic applications needs to be mentioned.

2 Reactions at the metal

The generation of reactive, coordinatively unsaturated intermediates by loss of a CO ligand from carbene complexes such as **7** is a crucial step in numerous reactions of such complexes, the most prominent one being the Dötz reaction.^{8,18,23,25,38} Other

examples include thermal decomposition, substitution of CO by phosphines, reactions with alkenes and others.



THERMOLYSIS

Reversible loss of CO was demonstrated in the thermolysis of the cyclic complex **8** which leads to the dimeric product **9** by the mechanism of equations (9).⁴⁴



Reversibility of the first step was indicated by a large reduction of the rate of decomposition of **8** in the presence of added CO. For example, at 139.4°C in decalin, the second-order rate constant for the decomposition of **8** is reduced 27-fold under 1480 mm Hg CO pressure. Furthermore, exchange of ¹³CO with **8** was found to be first order in the concentration of **8** and approximately 40-fold faster than thermal decomposition.

Additional evidence for reversible CO loss from several carbene complexes such as 11-15 was obtained from kinetic measurements of ¹³CO exchange rates determined by ¹³C NMR.⁴⁵ The results are summarized in Table 1. For the phenylmethoxycarbene complexes (11-13), CO dissociation from the tungsten complex is seen to be much slower than for the chromium and molybdenum derivatives. Changing the methoxy to a phenyl group (14 versus 13) increases the exchange rate which can be understood in terms of the reduced π -donor ability of

| Carbene complex | <i>T</i> (°C) | $k (\times 10^5 \mathrm{s}^{-1})$ | ΔG^{\ddagger} (kcal mol ⁻¹) |
|---|-----------------|-----------------------------------|---|
| $(CO)_5Cr=C(OMe)Ph (11)$ | 44^a | 18.0 | 24.0 |
| $(CO)_5Mo=C(OMe)Ph(12)$ | 27^a | 4.9 | 23.4 |
| $(CO)_5W=C(OMe)Ph(13)$ | 77^b | 2.3 | 28.0 |
| $(CO)_5W=C(Ph)_2$ (14) | 40^a | 4.9 | 24.5 |
| $(CO)_5 Cr = C(O) CH_2 CH_2 CMe_2$ (15) | 77 ^b | 1.1 | 28.5 |

Table 1 Rate constants and ΔG^{\ddagger} values for CO dissociation from carbene complexes

Reference 45. ^{*a*}In hexane. ^{*b*}In toluene.

the phenyl group relative to the methoxy group.⁴⁵ This reduced π -donor effect is partially offset by stronger π -donation to the carbene carbon from the metal, which reduces the π -donation from the metal to the CO ligands and leads to weaker bonding to CO. The faster CO dissociation from **11** compared to **15** has a similar origin, i.e., the alkyl group in **15** is a better electron donor than the phenyl groups in **11**.



An important conclusion from this work is that the measured CO dissociation rates are fast compared to the overall rates of those reactions that are believed to be initiated by CO loss from the corresponding carbene complex, lending strong support to the notion that CO loss *is* the crucial first step. Those processes include the above-mentioned hydrogenolysis, reaction with alkenes and vinyl esters, substitution of CO by phosphines, the Dötz reactions, and others.

SUBSTITUTION OF CO BY PHOSPHINES

With respect to CO substitution by phosphines, it is noteworthy that there is very good agreement between the kinetic results obtained by Werner *et al.*⁴⁶ and the rate constant of 1.8×10^{-4} s⁻¹ listed in Table 1 for CO dissociation from **11**. Werner *et al.*⁴⁶ investigated the reaction of **16** with a series of tertiary phosphines in *n*-decane and some other solvents.



They found that the reaction followed mixed first- and second-order kinetics according to equation (10). With

$$-\frac{d[16]}{dt} = k_{I}[16] + k_{II}[16][R_{3}P]$$
(10)

 $(C_6H_{11})_3P$, $(p-CH_3C_6H_4)_3P$, $(C_6H_5)_3P$, $(C_6H_5)_2(C_2H_5)P$ and $(C_6H_5)(C_2H_5)_2P$ the k_I term is dominant while with the more basic $(C_2H_5)_3P$ and $(n-C_4H_9)_3P$ the k_{II} term becomes significant at high phosphine concentrations.

The $k_{\rm I}$ term was attributed to the S_N1-type mechanism of equations (11a) and (11b)





with $k_2[R_3P] > (\gg)k_{-1}[CO]$ which renders the dissociation step $(k_1 = k_1)$ rate limiting. Based on the structure of the reaction product (17), the CO being lost is likely to come from a *cis* position.

Some representative k_1 values are $2.8 \times 10^{-4} \text{ s}^{-1}$ for **16a** at 68.2°C ($\Delta H^{\ddagger} = 27.5 \text{ kcal mol}^{-1}$, $\Delta S^{\ddagger} = 3.6 \text{ cal mol}^{-1} \text{ deg}^{-1}$) and $2.5 \times 10^{-4} \text{ s}^{-1}$ for **16b** at 44.5°C ($\Delta H^{\ddagger} = 27.0 \text{ kcal mol}^{-1}$, $\Delta S^{\ddagger} = 10.8 \text{ cal mol}^{-1} \text{ deg}^{-1}$).

The mechanism of the second-order term could not be established with any degree of certainty. One possibility discussed by the authors is an S_N2 -type mechanism although this would require an unusual seven coordinate metal complex at the transition state. Alternatively, the phosphine could first attack the carbene carbon to form **19** followed by loss of CO and migration of R_3P to the metal as shown in equations (12). This latter mechanism found support in a subsequent study by

Fischer *et al.*⁴⁷ in which rapid reversible R_3P addition to **16** (equation 12a) as well as to the tungsten analog **20** could be demonstrated.



20a: R' = CH₃ **20b**: R' = Ph

Equilibrium constants, $K'_1 = k'_1/k'_{-1}$, for equation (12a) were determined in toluene for the reactions of **16a** with C₆H₅(C₂H₅)₂P and (C₄H₉)₃P as well as for the reaction of **20a** with (C₄H₉)₃P. Some of the results are summarized in Table 2. They show a strong temperature dependence. For the reaction of **16a**, K'_1 is quite small at the temperatures used in the kinetic study (58.8–68.8°C for the reaction of **16a** with (n-(C₄H₉)₃P).⁴⁶ This means $K'_1[R_3P] < (\ll)1$ and hence the rate law for the mechanism of equations (12a) and (12b) is given by equation (13), implying that k_{II} in

$$-\frac{d[\mathbf{16a}]}{dt} = K_1' k_2' [\mathbf{16a}] [\mathbf{R}_3 \mathbf{P}]$$
(13)

Equation (10) is equal to $K'_1 k'_2$. The fact that $K'_1[R_3P] < (\ll)1$ is also consistent with the first term in equation (10) being independent of $[R_3P]$; if $K'_1[R_3P] \approx 1$, the first term would be given by $k_I[\mathbf{16}]/(1 + K'_1[R_3P])$ and show a decrease with increasing $[R_3P]$.

As to the details of reaction (12b), the most likely mechanisms involve either loss of CO from **19** (equation 14) and subsequent migration of Ph_3P from the carbene

| Carbene complex | <i>T</i> (°C) | $K'_1 (M^{-1})$ | ΔH° (kcal mol ⁻¹) | $\Delta S^{\circ} (\text{cal mol}^{-1} \text{deg}^{-1})$ |
|--|---------------|----------------------|--|--|
| $(CO)_{5}Cr = C(OMe)CH_{2}; (20a; R' = CH_{2})$ | 10 | 30.2 | | |
| (| 17 | 19.4 | | |
| | 25 | 6.22 | | |
| | 30 | 3.69 | | |
| | 38 | 1.98 | | |
| | 45 | 0.68 | | |
| | 52 | 0.46 | | |
| | 60 | 0.15 | -20.0 | -63.8 |
| $(CO)_5W = C(OMe)CH_2$; (20b : R' = CH_2) | 4 | 8.93×10^{4} | | |
| (| 14 | 5.25×10^{3} | | |
| | 30 | 1.35×10^{3} | | |
| | 42 | 5.00×10^{2} | | |
| | 45 | 4.51×10^{3} | -15.1 | - 35.6 |

Table 2 Equilibrium constants for $(C_4H_9)_3P$ addition to $(CO)_5M=C(OMe)CH_3$ in toluene

Ref. 47.

carbon to the metal, or addition of an external Ph_3P to the metal of **21** with loss of Ph_3P from the carbon.⁴⁸



THE DÖTZ REACTION

A prototypical example of the Dötz benzannulation reaction^{8,18,23,25,38} is shown in equation (15).



Above 60°C, the naphthol products typically undergo migration of the $(CO)_3Cr$ moiety, e.g., equation (16).



Reaction (15) (with R' = H and $R'' = C_6H_5$, 4-MeC₆H₄, 4-CF₃C₆H₄ and Me; and with R' = Me and R'' = 4-CF₃C₆H₄) was investigated kinetically in dibutyl ether⁴⁹ under pseudo-first-order conditions ([**22**]:[**21**] \ge 10). The reversible loss of CO led to complex kinetics that was difficult to evaluate. However, when the reaction was conducted either in the presence of a saturated solution of CO or under conditions that led to the continuous removal of CO so that [CO] \approx 0, first-order kinetics was observed. The concentration dependence of the rate was shown to be consistent with the mechanism of Scheme 1 where **26** is a steady-state intermediate and hence the



Scheme 1

rate law is given by equation (17). The kinetic parameters

$$-\frac{\mathrm{d}[\mathbf{13}]}{\mathrm{d}t} = \frac{k_1 k_2 [\mathbf{22}]}{k_{-1} [\mathrm{CO}] + k_2 [\mathbf{22}]} [\mathbf{13}]$$
(17)

are summarized in Table 3. The positive ΔS^{\ddagger} value for the k_1 step is consistent with rate-limiting CO dissociation; it contrasts with the strongly negative ΔS^{\ddagger} values obtained for reactions where nucleophilic attack of the carbene complex is rate limiting, as discussed in a section titled "Insertion reactions initiated by nucleophilic attack." The trend in the k_2/k_{-1} ratios is also consistent with the proposed mechanism; since k_{-1} is independent of the alkyne, these ratios mirror changes in k_2 whose increase with the increasing electron donating effect of R["] reflect enhanced nucleophilicity of the alkyne.

Based on a density functional theory (DFT) study of the model reaction of $(CO)_5Cr=C(OH)CH=CH_2$ with HC=CH, Torrent *et al.*⁵⁰ recently proposed that an association type mechanism according to which insertion of the alkyne into the Cr=C bond followed by CO dissociation may be energetically more favorable than the mechanism of Scheme 1. However, their claims have been rejected because they are contradicted by the above described kinetic results.⁵¹

Table 3 Kinetic parameters for the benzannulation reaction (15) of $(CO)_5Cr=C(OMe)Ph$ (11) with 22 (R' = H) in dibutyl ether at 56.8°C

| R″ | $k_1^a (\times 10^{-3} \mathrm{s}^{-1})$ | k_2/k_{-1} |
|-----------------------------------|--|--------------|
| $4-CF_3C_6H_4$ | 1.07 | 0.0043 |
| C ₆ H ₅ | 1.07 | 0.0050 |
| 4-MeC ₆ H ₄ | 1.07 | 0.0055 |
| CH ₃ | 1.07 | 0.0260 |

Reference 49.

 ${}^{a}\Delta H^{\ddagger} = 25.8 \text{ kcal mol}^{-1}, \Delta S^{\ddagger} = 6.22 \text{ cal mol}^{-1} \text{ deg}^{-1}.$

| X | NR_2 | $k (\times 10^3 \mathrm{s}^{-1})$ | ΔH^{\ddagger} (kcal mol ⁻¹) | ΔS^{\ddagger} (cal mol ⁻¹ deg ⁻¹) |
|-----------------------------------|--|------------------------------------|---|--|
| Cl | NMe ₂ | 0.109 | 26.2 | 7.0 |
| Cl | NC_5H_{10} | 0.394 | 23.9 | 2.0 |
| Cl | NEt ₂ | 1.80 | 23.7 | 4.4 |
| Cl | NPh ₂ | 11.3 | 19.3 | -6.1 |
| Br^{a} | NEt ₂ | 905 ^b | 18.0 | - 1.0 |
| Cl Cl Cl Br ^a | $\begin{array}{c} NC_5H_{10} \\ NEt_2 \\ NPh_2 \\ NEt_2 \end{array}$ | $0.394 \\ 1.80 \\ 11.3 \\ 905b$ | 23.9 23.7 19.3 18.0 | 2.0 4.4 -6.1 -1.0 |

Table 4 Rate constants and activation parameters for the reaction of $(CO)_5M=C(X)NR_2$ in 1,1,2-trichloroethane (equation 18)

Reference 53. At 40°C.

^{*a*}Reference 52 in CH_2Cl_2 .

^{*b*}Calculated based on ΔH^{\ddagger} and ΔS^{\ddagger} .

REARRANGEMENT REACTIONS WITH LOSS OF CO

Carbene complexes of the type **28** with X being a halogen spontaneously rearrange to the dialkylaminocarbyne complex **29** under elimination of CO.^{52,53}



In an initial kinetic study (R = Et; X = Cl and Br)⁵² the reaction was found to follow a first-order rate law and yield rate constants that are much higher for X = Br than for X = Cl. An extension of the work to include different R₂N groups (Me₂N, C₅H₁₀N and Ph₂N) was reported in 1985.⁵³ Some of the results are summarized in Table 4. They show that increasing the bulk of the dialkylamino group enhances the rate; for example, the change from NMe₂ to NPh₂ increases the *k* value 100-fold and, based on low temperature experiments, it was estimated that at 40°C the N(*i*-Pr)₂ derivative may be *ca*. 10⁷-fold more reactive than the NMe₂ derivative. There is also a dramatic increase in the rate upon changing X = Cl to X = Br. Furthermore, the rate increases with increasing solvent polarity although the effect is quite modest.

Other mechanistically relevant observations are the absence of any rate effect by added chloride ions, by running the reactions under CO pressures up to 78 atm, or by added radical initiators such as azobis(isobutyronitrile) or radical inhibitors such as hydroquinone. Furthermore, addition of high concentrations of PPh₃ in the reaction of **28** ($R_2N = Et_2N$, X = Cl) does not effect the rates either but it leads to a different product **30**.



30

However, CO/PPh₃ exchange in **29** had been shown to be much faster than formation of **30** from **28**,⁵⁴ hence one concludes that **30** must have been formed from **29** rather than from **28**. Similar results have been reported for the rearrangement of other aminocarbene complexes such as **28** ($R_2N = Et_2N$) with X = SeR,⁵⁵ TePh,⁵⁶ SnPh₃⁵⁷ and PbPh₃.⁵⁸

In proposing a mechanism that would be consistent with all experimental observations, Fischer *et al.*⁵³ discussed the following possibilities.

(1) Rate-limiting loss of CO followed by rapid migration of X to the metal, equation (19).



(2) Heterolytic C–X bond cleavage followed by loss of CO and addition of X^- to the metal, equation (20).

$$(CO)_5Cr = C \xrightarrow{X} (CO)_5Cr = C = \overset{+}{NR_2} + X^- \xrightarrow{-CO} X(CO)_4Cr \equiv C - NR_2$$
28
29
(20)

(3) Homolytic C–X bond cleavage followed by loss of CO and addition of X to the metal, equation (21).



(4) Intramolecular nucleophilic attack of one of the *cis*-CO ligands by X, followed by loss of CO and rearrangement, equation (22).



(5) Intramolecular attack by X on the metal that is concerted with the loss of CO, equation (23).

$$(CO)_{5}Cr = C \xrightarrow{X}_{NR_{2}} \left(\begin{array}{c} OC & X & CO \\ OC & Cr \equiv C - NR_{2} \\ OC & CO \end{array} \right)^{\ddagger} \xrightarrow{-CO} X(CO)_{4}Cr \equiv N - Et$$

$$29$$

$$(23)$$

Mechanism (1) was excluded because the rate remains unaffected when the reaction is conducted under high CO pressure and the ΔS^{\ddagger} values are much lower than is typical for reactions that are initiated by CO loss (see the section on Substitution of CO by phosphines). Mechanism (2) was excluded based on the rather small solvent dependence. Mechanism (3) was rejected because neither radical initiators nor radical scavengers have any influence on the reaction, and there is no correlation between ΔH^{\ddagger} and the homolytic C–X bond energies. Intramolecular attack by X on one of the CO ligands (mechanism 4) is conceivable when X contains lone pairs (Cl, Br, SePh and TePh) but not with X = SnPh₃ or PbPh₃ and was also excluded.

In contrast, all observations are consistent with the concerted mechanism (5), including the small ΔS^{\ddagger} values typical for intramolecular processes.

PHOTOCHEMICAL REACTIONS

The early work of Fischer *et al.*⁵⁹ showed that irradiation of several complexes of the type 7 (M = Cr or W) in the presence of phosphines led to tetracarbonyl phosphine

complexes, presumably the result of photoinduced loss of CO (equation 24) and subsequent trapping of $(CO)_4M=C(OR)R'$ by the phosphine.

$$(CO)_{5}M = C \xrightarrow{OR} \frac{hv}{R_{3}P} \quad cis \text{ and } trans - (CO)_{4}(R_{3}P)M = C \xrightarrow{OR} + CO \qquad (24)$$
7
32

Even though work by Casey *et al.*^{60–62} appeared to be consistent with Fischer *et al.*'s results, other reports did not.^{63,64} A thorough study of the photolysis of **13** confirmed that UV radiation of this carbene complex induces CO loss (equation 25) to form **33**.⁶⁵



When the reaction is carried out in CH_3CN solution, **33** gets trapped immediately to form **34**, which is the observed product and can be isolated. The reaction is very clean as indicated by sharp isosbestic points in the UV and IR. Furthermore, **13** can be regenerated by reacting **34** with CO.



In THF solution, the photoproduct is the corresponding THF complex 35 while photolysis of 13 in the presence of PhC=CPh in hexane solution leads to an insoluble complex whose structure could either be that of 36 or 37.



Additional detail about the photodissociation of CO from **13** has come from timeresolved Raman spectroscopy.^{66,67} It was shown that the loss of CO occurs within the duration of a nanosecond laser pulse into the LF band and leads to a transient species that decays on a microsecond time scale to regenerate **13**; in solvents such as CH_2Cl_2 , benzene or *n*-hexane formation and decay of this transient species are the only observable processes while in acetonitrile there is also rapid formation of **34**. The suggested sequence of reactions is shown in Scheme 2.



Scheme 2

The proposed structure of the transient species **38** implies "self-solvation" by an agostic interaction of the hydrogen of the methoxy group with the metal which competes with the coordination of CH_3CN to the metal; formation of both **34** and **38** occur on the same time scales. An additional species of unknown structure (**39**) has been postulated in order to explain why the rate of regeneration of **13** is independent of [CO].

The photochemistry of **11** and **13** in inert gas matrices at 10 K has also been reported.⁶⁸ Irradiation with visible light into the low-energy metal-to-ligand charge transfer (MLCT) band (\sim 370 nm) was shown to induce complete conversion of the *anti* to the *syn* configurations (equation 27).



Irradiation into the LF band at ~ 350 nm leads to the same *anti* \rightarrow *syn* conversion but there is subsequent loss of CO which, for the tungsten complex, is followed by a third process. It was suggested that this third process leads to **38**, the transient species that had been reported by McGarvey's group.^{66,67}

Applying time-resolved Raman spectroscopy coupled with matrix isolation techniques to additional carbene complexes such as **40–42**, McGarvey and coworkers⁶⁹ were able to confirm the findings (equation 27) reported by Stufkens and co-workers⁶⁸ They showed that the *anti* \rightarrow *syn* conversion occurs in less than 10 ns. The lifetime of the *syn* isomer which spontaneously reverts back to the *anti* isomer is strongly dependent on the structure of the carbene complex and solvent polarity. The reported lifetimes are summarized in Table 5. The lifetimes are substantially longer for the methyl compared to the aryl carbene complexes and also increase with increasing solvent polarity. One factor that may contribute to the shorter lifetimes of the aryl derivatives is steric crowding in the *syn* isomers although "conventional" steric crowding cannot be the whole story⁶⁹ since the lifetime of the even more crowded *syn* isomer of (CO)₅W=C(OEt)(SiPh₃) is similar to that for **41** and **42**.⁷⁰ However, for the aryl carbene complexes the crowding may force the aryl group to rotate out of the plane of the carbene which reduces the π -overlap with the carbene carbon and hence leads to destabilization.⁶⁹



The solvent dependence suggests stabilization of the *syn* isomers and/or a destabilization of the transition state by more polar solvents but the origin of these stabilizing/destabilizing effects was not discussed.⁶⁹

A recent laser flash photolysis study of **13** using time-resolved IR and UV–Vis spectroscopy⁷¹ has confirmed basic conclusions derived from the previous

| Solvent | (CO) ₅ W=C(OMe)CH ₃ | $(CO)_5W=C(OMe)C_6H_4-4-Me$ | (CO) ₅ W=C(OEt)Ph | $(CO)_5W = C(OMe)Ph^a$ |
|---------------------------------|---|-----------------------------|------------------------------|------------------------|
| <i>n</i> -Hexane | 126 | 1.10 | 4.65 | 2.9 |
| Cyclohexane | 294 | 1.62 | 5.0 | 4.6 |
| CH ₂ Cl ₂ | 1200 | 3.70 | 14.3 | 11.1 |
| Methanol | 2642 | 6.6 | 20.7 | |
| Acetonitrile | 3356 | 7.1 | 23 | |

Table 5 Lifetimes (in μ s) of *syn* isomers of tungsten carbene complexes at 20°C

Reference 69.

^{*a*}Reference 66.



investigations mentioned above and also provided additional quantitative kinetic information. Scheme 3^{71} provides an overview of the various processes that occur upon excitation of the *anti* isomer of **13** with a laser flash at 308 nm. The initially formed upper ligand field excited state, $[13^*]_{anti}$, can undergo three processes: a radiationless decay back to the groundstate, isomerization to the *syn* isomer (quantum yield $\approx 30\%$), and loss of CO to form the penta-coordinated tetracarbonyl carbene complex $[43]_{anti}$ (quantum yield $\approx 1.7\%$). This latter intermediate may revert back to $[13]_{anti}$ upon irreversible addition of CO or reversibly add a ligand (L) such as the solvent (L = S) or other species such as N₂, H₂O, CH₃CN or an olefin.

On the microsecond time scale of the reported experiments, **43** cannot be observed; hence the first detectable tetracarbonyl species was attributed to **44** with (L = S). A conversion of **44**-S into **44**-N₂ could be observed during the first 10 μ s in nitrogen-purged solutions, or into **44**-H₂O in solutions purged with wet argon; these conversions are assumed to occur via **43** as the intermediate. The data also provided a rate constant of $3.5 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$ for the exchange of the solvent or N₂ ligands by CO.

3 Reactions at the carbene carbon

GENERAL FEATURES

Nucleophilic substitution of an alkoxy group of carbene complexes such as 7 is one of the prototypical reactions of Fischer carbene complexes. It has generally been

assumed to proceed via a tetrahedral intermediate (45) in analogy to the reaction of carboxylic esters with nucleophiles.

$$(CO)_{5}M = C \begin{pmatrix} OR \\ R' \end{pmatrix} + Nu^{-} \qquad \underbrace{k_{1}}_{k_{-1}} \qquad \underbrace{(CO)_{5}M}_{R'} - \underbrace{l}_{R'} OR \\ \downarrow \\ R' \end{pmatrix} = (CO)_{5}M = C \begin{pmatrix} Nu \\ R' \end{pmatrix} + RO^{-} \qquad (28)$$

$$7 \qquad 45 \qquad 46$$

With neutral nucleophiles such as phosphines and amines, the nucleophilic attack on the carbene complex leads to zwitterionic intermediates such as 47-49; if there is an acidic proton on the intermediate as in the case of 49, the reaction can proceed to products by elimination of ROH, otherwise the reaction stops at the adduct stage.



Most synthetic applications have involved common nucleophiles such as amines, $^{1,72-76}_{1,83-87}$ hydrazine, $^{1,77}_{1,83-87}$ oximes, $^{1,78}_{1,83}$ alkoxide ions, $^{1,79}_{1,83-87}$ thiolate ions, $^{1,80-82}_{1,83-87}$ carbanions $^{1,83-87}_{1,83-87}$ as well as others.

AMINE NUCLEOPHILES

The first kinetic investigation of a nucleophilic substitution reaction was that of Werner *et al.*^{91,92} who studied the reaction of **11** with several primary aliphatic amines (*n*-BuNH₂, C₆H₁₁NH₂, C₆H₅CH₂NH₂) in *n*-decane, dioxane, methanol and dioxane–methanol (1/1) mixtures.



In *n*-decane the rate was found to be third order with respect to amine concentration (equation 30), in dioxane second order (equation 31),

$$-\frac{d[11]}{dt} = \frac{d[50]}{dt} = k''[11][RNH_2]^3$$
(30)

$$-\frac{d[\mathbf{11}]}{dt} = \frac{d[\mathbf{50}]}{dt} = k''[\mathbf{11}][\mathrm{RNH}_2]^2$$
(31)



Scheme 4

while in dioxane-methanol mixtures or pure methanol a mixed first- and secondorder dependence on amine was reported (equation 32).

$$-\frac{d[\mathbf{11}]}{dt} = \frac{d[\mathbf{50}]}{dt} = k'[\mathbf{11}][\mathrm{RNH}_2] + k''_1[\mathbf{11}][\mathrm{RNH}_2]^2$$
(32)

These rate laws indicate that up to three amine molecules may be present in the ratelimiting transition state but an unambiguous mechanistic interpretation is difficult because the sequence by which these amine molecules assemble cannot be ascertained. For the reactions in *n*-decane the authors proposed the mechanism shown in Scheme 4.

The function of the first amine molecule is to activate the substrate by hydrogen bonding to the methoxy group and possibly also to provide acid catalysis to MeO⁻ departure in the product forming step. The nucleophilic attack occurs with an amine molecule that is hydrogen bonded to a third amine which presumably helps stabilize the positive charge that develops on the nitrogen. In dioxane and methanol the R N H \cdots N^H_H R complex may be replaced by an amine–solvent complex, e.g., R N H \cdots O CH₃. Furthermore, in methanol the solvent presumably takes on the role of the amine that hydrogen bonds to the methoxy group.

Support for this mechanism whose last step is probably rate limiting comes from ¹H NMR studies that indicate the presence of association equilibria such as between

the carbene complex and the amine (first step in Scheme 4) and between two amine molecules. The negative ΔH^{\ddagger} and strongly negative ΔS^{\ddagger} values are also consistent with the proposed scheme.

Another aminolysis reaction of a Fischer carbene complex that has been studied kinetically is shown in equation (33).⁹³

$$(CO)_5 W = C \xrightarrow{SMe}_{SMe} \xrightarrow{RNH_2} (CO)_5 W - C \equiv \overset{-}{N} - R$$

$$51 \qquad 52 \qquad (33)$$

Rates were measured with monoamines such as *n*-PrNH₂, *n*-BuNH₂, $C_{6}H_{11}NH_{2}$ and PhCH₂NH₂ as well as diamines such as $H_2N(CH_2)_2NH_2$, $H_2N(CH_2)_3NH_2$, $H_2N(CH_2)_4NH_2$, $H_2N(CH_2)_2NMe_2$ and $H_2N(CH_2)_3NMe_2$; the majority of experiments was performed in THF but some data were also obtained in cyclohexane, pyridine, methanol, cyclohexane–THF and cyclohexane–pyridine mixtures. For the reactions of monoamines in THF the rate law was second-order in amine (equation 34), for the short chain diamines the rate law was first order in amine (equation 35) while for the longer chain diamines the rate law was that of equation (36). For the reactions with the primary amines the reaction was catalyzed by relatively high concentrations of DABCO and 4-picoline.

$$-\frac{d[51]}{dt} = k_2[51][RNH_2]^2$$
(34)

$$-\frac{d[51]}{dt} = k_1[51][RNH_2]$$
(35)

$$-\frac{d[51]}{dt} = k_1[51][RNH_2] + k_2[51][RNH_2]^2$$
(36)

The results with the monoamines were interpreted in terms of the mechanism outlined in Scheme 5 with the conversion of 54 to 55 being the rate-limiting step; the



Scheme 5

intermediacy of **55** which does not accumulate to detectable levels is supported by the fact that reaction of **51** with secondary amines leads to $(CO)_5W=C(SMe)NR_2^{94}$ which is stable. It was noted that the intermediate **54** may have the cyclic structure **56** which would allow direct proton transfer to the leaving group.



The mechanism of Scheme 5 is quite similar to that of Scheme 4 except that there is no association of an amine molecule with the leaving group. Not even methanol appears to hydrogen bond to the sulfur group which is consistent with the much weaker hydrogen bond acceptor ability of sulfur compared to oxygen.

The reduced reaction order and the higher rate constants with diamines was explained in terms of intramolecular catalysis involving an intermediate like **57**.

The postulate of tetrahedral intermediates generated by nucleophilic attack on the carbene carbon (see T^{\pm} in Schemes 4 and 5) is reasonable even though the studies discussed above provide no direct evidence for them. However, support for such intermediates, at least in the reaction of **11** with amines, comes from reports that **11** forms isolable tetrahedral adducts such as **58** by reaction with DABCO or quinuclidine in ether.^{95,96}



58 (X = N or CH)

A more detailed study has provided additional strong evidence for the presence of tetrahedral intermediates in the reaction of **11** with primary amines (*n*-BuNH₂, MeOCH₂CH₂NH₂, ClCH₂CH₂NH₂, H₂NCOCH₂NH₂ and EtO₂CCH₂NH₂) in 20% MeCN-80% water (v/v).^{97,98} Furthermore, because in this highly polar solvent

there are no hydrogen bonding associations of the types observed in non-polar solvents which unduly complicate the situation, more clear cut conclusions could be drawn about mechanistic details. The proposed mechanism is outlined in equation (37).

$$(CO)_{5}Cr = C \xrightarrow{OMe}_{Ph} \underbrace{k_{1}[RNH_{2}]}_{k_{-1}} \underbrace{(CO)_{5}Cr}_{K} \xrightarrow{-} C \xrightarrow{OMe}_{Ph} \underbrace{k_{3}^{A}[RNH_{2}]}_{RNH_{2}} (CO)_{5}Cr = C \xrightarrow{H}_{NR} + MeOH$$

$$11 \xrightarrow{T^{\pm}} \underbrace{k_{3}^{OH}[OH^{-}]}_{S0} 50$$

$$(37)$$

The k_3^{A} and k_3^{OH} steps refer to base catalysis by the amine and by OH⁻, respectively; the k_2 step represents spontaneous, possibly intramolecularly catalyzed conversion of T[±] to the product although its contribution to the overall reaction is negligible and is not included in the following kinetic analysis.

Applying the steady-state approximation to T^{\pm} leads to the rate law of equation (38)

$$-\frac{d[11]}{dt} = \frac{d[50]}{dt} = k_{obsd}[11]$$
(38)

with the pseudo-first-order rate constant k_{obsd} given by equation (39), or the second-order rate constant,

$$k_{\text{obsd}} = \frac{k_1 (k_3^{\text{A}} [\text{RNH}_2] + k_3^{\text{OH}} [\text{OH}^-])}{k_{-1} + k_3^{\text{A}} [\text{RNH}_2] + k_3^{\text{OH}} [\text{OH}^-]} [\text{R}_2 \text{NH}]$$
(39)

 $k_{\rm A}$, given by equation (40). The results are completely consistent with equation (40).

$$k_{\rm A} = \frac{k_{\rm obsd}}{[\rm RNH_2]} = \frac{k_1 (k_3^{\rm A} [\rm RNH_2] + k_3^{\rm OH} [\rm OH^-])}{k_{-1} + k_3^{\rm A} [\rm RNH_2] + k_3^{\rm OH} [\rm OH^-]}$$
$$= \frac{k_1 ((k_3^{\rm A}/k_{-1})[\rm RNH_2] + (k_3^{\rm OH}/k_{-1})[\rm OH^-])}{1 + (k_3^{\rm A}/k_{-1})[\rm RNH_2] + (k_3^{\rm OH}/k_{-1})[\rm OH^-]}$$
(40)

Representative plots of k_A versus [RNH₂] and [OH⁻] are shown in Figs. 2 and 3, respectively. At low [RNH₂] and low [OH⁻] equation (40) simplifies to equation (41), because (k_3^A/k_{-1}) [RNH₂] + (k_3^{OH}/k_{-1}) [OH⁻] \ll 1,

$$k_{\rm A} = k_1 \frac{k_3^{\rm A}}{k_{-1}} [{\rm RNH}_2] + k_1 \frac{k_3^{\rm OH}}{k_{-1}} [{\rm OH}^-]$$
(41)

i.e., k_A increases linearly with increasing [RNH₂] or [OH⁻].

At high $[RNH_2]$ and/or high $[OH^-]$ equation (40) is reduced to equation (42).

$$k_{\rm A} = k_1 \tag{42}$$



Fig. 2 Reaction of $(CO)_5Cr=C(OMe)Ph$ (11) with glycine ethyl ester in 20% MeCN-80% water at 25°C. Second-order rate constants as a function of amine concentration. (Figure taken from Ref. 97, by permission from the American Chemical Society.)



Fig. 3 Reaction of $(CO)_5Cr=C(OMe)Ph$ (11) with glycine ethyl ester in 20% MeCN-80% water at 25°C. Second-order rate constants as function of $[OH^-]$. (Figure taken from Ref. 97, by permission from the American Chemical Society.)

| RNH ₂ | pK_a^{AH} | $k_1 (M^{-1} s^{-1})$ | $k_3^{\rm A}/k_{-1} \ ({ m M}^{-1})$ | $k_3^{\text{OH}}/k_{-1} \ (\text{M}^{-1})$ | $k_3^{\mathrm{OH}}/k_3^{\mathrm{A}}$ |
|--|-------------|-----------------------|--------------------------------------|--|--------------------------------------|
| CH ₃ (CH ₂) ₃ NH ₂ | 10.67 | 2900 | 33.7 | 8.1×10^{3} | 2.40×10^{2} |
| CH ₃ OCH ₂ CH ₂ NH ₂ | 9.52 | 400 | 21.1 | 8.4×10^{4} | 3.98×10^{3} |
| CICH ₂ CH ₂ NH ₂ | 8.61 | 91 | 19.5 | 2.8×10^{5} | 1.44×10^{4} |
| H ₂ NCHOCH ₂ NH ₂ | 8.03 | 100 | 17.3 | 1.25×10^{6} | 7.22×10^{4} |
| EtOCOCH ₂ NH ₂ | 7.70 | 36 | 9.7 | 1.49×10^{6} | 1.54×10^{5} |

Table 6 Kinetic parameters for the reaction of $(CO)_5Cr=C(OMe)Ph$ (11) with primary amines in 20% MeCN-80% water (v/v) at 25°C (equation 37)

Reference 97.

As can be seen in Figs. 2 and 3, the plateau values for the amine and OH^- concentration dependence are the same (k_1) , as they should be.

Detailed analysis of the data allowed an evaluation of the k_1 , k_3^A/k_{-1} , k_3^{OH}/k_{-1} and k_3^{OH}/k_3^A values summarized in Table 6. The ratios, in particular the k_3^{OH}/k_3^A ratios, provided crucial information about the mechanism of the general base catalyzed conversion of T^{\pm} to products. There are two possible mechanisms that are indistinguishable on the basis of the rate law alone. One mechanism involves ratelimiting deprotonation of T^{\pm} by RNH₂ or OH⁻, respectively, followed by rapid methoxide ion expulsion from T^- as shown in equation (43).



For this case the meaning of k_3^A and k_3^{OH} in equations (37) and (40) is that defined by equations (44) and (45), respectively.

$$k_3^{\mathrm{A}} = k_{3\mathrm{p}}^{\mathrm{A}} \tag{44}$$

$$k_3^{\rm OH} = k_{3p}^{\rm OH} \tag{45}$$

The aminolysis of carboxylic esters^{99–101} to which the reaction of Fischer carbene complexes with amines has frequently been compared,^{1,92,93} typically proceeds by a similar mechanism with rate-limiting deprotonation of the corresponding zwitterionic tetrahedral intermediate (**59**) and so do many S_NAr reactions where the deprotonation of the zwitterionic Meisenheimer complex (e.g., **60**) is rate

limiting.^{102–104}



The second mechanism is shown in equation (46).



Here the proton transfer is a fast equilibrium and general acid-catalyzed loss of the methoxide ion by RNH_3^+ and water, respectively, is rate limiting. For this mechanism k_3^{A} and k_3^{OH} are given by equations (47) and (48), respectively,

$$k_3^{\rm A} = k_3^{\rm AH} K_a^{\pm} / K_a^{\rm AH} \tag{47}$$

$$k_3^{\rm OH} = k_3^{\rm H_2O} K_{\rm a}^{\pm} / K_{\rm w} \tag{48}$$

with K_a^{AH} being the acidity constant of RNH₃⁺ and K_w the ionic product of the solvent. An example where this second mechanism prevails is the aminolysis of strongly activated vinylic compounds such as β -methoxy- α -nitrostilbene (**61**), i.e., deprotonation of the corresponding zwitterionic intermediate (**62**) is fast and methoxide ion departure is rate limiting.^{105,106}

For the aminolysis of **11**, rate-limiting deprotonation of T^{\pm} can be excluded on the basis of the k_3^{OH}/k_3^A ratios. These ratios increase from 240 for the *n*-BuNH₂ reaction to 1.54×10^5 for the glycine ethyl ester reaction (Table 6). If proton transfer were rate limiting, these ratios should, according to equations (44) and (45), be given by equation (49).

$$k_3^{\rm OH}/k_3^{\rm A} = k_{3p}^{\rm OH}/k_{3p}^{\rm A} \tag{49}$$

The rate constant k_{3p}^{OH} refers to an essentially diffusion controlled proton transfer; it should be independent of the amine and have a value around $10^{10} \text{ M}^{-1} \text{ s}^{-1}$.¹⁰⁷ The rate constant k_{3p}^{A} should also be independent of the amine since an increase in the acidity of RNH₃⁺ should have a similar effect on the pK_a of T_A^{\pm} ; hence the pK_a *difference* between T_A^{\pm} and the respective RNH₃⁺ is expected to be constant. Furthermore, the pK_a^{\pm} of T^{\pm} is likely to be somewhat lower than the pK_a of the respective RNH₃⁺,⁹⁷ which would make the proton transfer from T_A^{\pm} to RNH₂ thermodynamically favorable, with a rate constant close to diffusion control,¹⁰⁷ probably of the order of $4 \times 10^8 - 2 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$.¹⁰⁸ Hence the $k_3^{\text{OH}}/k_3^{\text{A}}$ ratios should be independent of the amine and have values of the order of 5–25. This contrasts with the experimental observation of a 640-fold change between *n*-butylamine and glycine ethyl ester.

On the other hand, the strong increase in the $k_3^{\text{OH}}/k_3^{\text{A}}$ ratio with decreasing basicity of the amine is easily accounted for in terms of the mechanism of equation (46). According to equations (47) and (48), the $k_3^{\text{OH}}/k_3^{\text{A}}$ ratio is given by equation (50). The K_a^{AH}/K_w ratio is proportional to K_a^{AH} while the $k_3^{\text{H}_2O}/k_3^{\text{AH}}$ decreases somewhat with decreasing amine basicity because RNH₃⁺ becomes a better acid catalyst but overall $k_3^{\text{OH}}/k_3^{\text{A}}$ increases with decreasing amine basicity.

$$\frac{k_3^{\rm OH}}{k_3^{\rm A}} = \frac{k_3^{\rm H_2O}K_a^{\rm AH}}{k_3^{\rm AH}K_{\rm w}}$$
(50)

PHOSPHINE AND PHOSPHITE NUCLEOPHILES

With respect to nucleophilic attack on the carbene carbon, there are strong similarities between the reactions of phosphines or phosphites and those of amines. The reactions are exemplified by equation (51).^{1,47,109–111} However, as discussed previously, **19** tends to rearrange to **17** (equation 12b), a process not observed with amine nucleophiles.

$$(CO)_{5}M = C \begin{pmatrix} OMe \\ R' \end{pmatrix} + R_{3}P \implies (CO)_{5}M - \begin{matrix} - & OMe \\ C \\ R' \end{pmatrix} + \begin{matrix} - & P \\ R_{3} \end{pmatrix}$$
(51)
16 19

As discussed earlier, equilibrium constants for the addition of several tertiary phosphines $((n-Bu)_3P, Ph(C_2H_3)_2P \text{ and } (i-Pr)_3P \text{ to } (CO)_5Cr=C(OMe)CH_3 \text{ and } (CO)_5W=C(OMe)CH_3 \text{ in toluene-}d_8 \text{ have been reported by Fischer et al.}^{47}$ (Table 2). In a follow-up study, Fischer¹¹² investigated the temperature and substituent effect on the equilibrium constant for the reaction (52) in toluene (Z = MeO, Me, H, Br and CF₃).

$$(CO)_5 W = C \xrightarrow{OMe} + (n-Bu)_3 P \xrightarrow{K_1} (CO)_5 W - C \xrightarrow{P(n-Bu)_3} Z$$
(52)
63 Z 64

010

| Z | ΔG° (kcal mol ⁻¹) | ΔH° (kcal mol ⁻¹) | ΔS° (cal mol ⁻¹ deg ⁻¹) | $\log K_1$ |
|-----------------|--|--|---|------------|
| MeO | - 1.20 | -12.1 | -43.5 | 1.04 |
| Me | -2.44 | -12.3 | - 39.5 | 2.13 |
| Н | -2.99 | -12.7 | -38.7 | 2.61 |
| Br | -3.44 | -13.4 | - 39.7 | 3.02 |
| CF ₃ | -4.26 | -14.1 | - 39.2 | 3.72 |
| | | | | |

Table 7 Thermodynamic parameters for the reaction of $(CO)_5W=C(OMe)C_6H_4Z$ (63) with $(n-Bu)_3P$ (equation 52) in toluene at $-48^{\circ}C$

Reference 112.

 ΔG° , ΔH° , ΔS° and log K_1 values are summarized in Table 7. The K_1 values show a strong dependence on Z. The log K_1 values for Z = Me, H, Br and CF₃ correlate well with the Hammett σ constants from which one calculates $\rho = 2.2$; the point for Z = MeO deviates negatively by about 0.9 log units from the Hammett line defined by the other substituents.

The negative deviation for the methoxy derivative is also seen in a correlation of log K_1 with the vibrational CO force constants obtained from IR measurements; the more negative ΔS° values are further testimony to the deviant behavior of the methoxy derivative. Even though not stated in these terms by Fischer,¹¹² the most plausible source of the deviations with Z = MeO is the strong π -donor effect of the methoxy group (**63a**) which leads to a substantial stabilization of the carbene complex. The reduced CO vibrational force constants for the methoxy derivative are consistent with this explanation.



Also not discussed by the author are the possible reasons for the rather strong substituent dependence of K_1 as reflected in the ρ value of 2.2. We offer the following interpretation. The positive ρ value implies that electron withdrawing substituents stabilize the adduct (64) and/or destabilize the carbene complex. Since in the adduct there is both a positive and a negative charge, the net substituent effect on this species cannot be very large; in view of the fact that the negative charge is more dispersed than the positive charge, an argument could be made that there should actually be a slight net *destabilization* of 64. Hence the main factor that can account for the positive ρ value is a destabilization of the carbene complex, which is best understood in terms of an increased electron deficiency of the carbene carbon. A

| М | R′ | PR ₃ | $k_1 (M^{-1} s^{-1})$ | $k_{-1} (s^{-1})$ | $K_1^a (\mathrm{M}^{-1})$ |
|----|----|-----------------|-----------------------|-------------------|---------------------------|
| Cr | Me | $P(n-Bu)_3$ | 34.7 | 0.70 | 50 |
| Mo | Me | $P(n-Bu)_3$ | 263 | b | >250 |
| W | Me | $P(n-Bu)_3$ | 225 | 1.2 | 200 |
| Cr | Me | $P(n-OBu)_3$ | 1.15 | 0.24 | 4.8 |
| Mo | Me | $P(n-OBu)_3$ | 10.4 | 0.26 | 40 |
| W | Me | $P(n-OBu)_3$ | 7.27 | 0.35 | 21 |
| Cr | Ph | $P(n-Bu)_3$ | 111 | 20.9 | 5.3 |
| Mo | Ph | $P(n-Bu)_3$ | 830 | 8.0 | 100 |
| W | Ph | $P(n-Bu)_3$ | 560 | 10.5 | 53 |
| Cr | Ph | $P(n-OBu)_3$ | 3.00 | 0.095 | 32 |
| Мо | Ph | $P(n-OBu)_3$ | 28.5 | b | > 140 |
| W | Ph | $P(n-OBu)_3$ | 13.4 | 0.32 | 42 |

Table 8 Rate and equilibrium constants for the reaction of PR_3 with $(CO)_5M=C(OMe)R'$ in acetone at 25°C (equation 51)

Reference 113.

^{*a*}Calculated as k_1/k_{-1} .

^bToo small for an accurate determination.

similar conclusion has recently been reached regarding the substituent effect on the reaction of **63** with *n*-BuNH₂ in 50% MeCN-50% water.⁹⁸

Reactions of the type of equation (51) have also been the subject of a kinetic investigation.¹¹³ Rate constants in the forward (k_1) and reverse direction (k_{-1}) were determined for a number of combinations of R, R', M in acetone at 25°C. The results are summarized in Table 8. A major conclusion drawn from these results is that steric effects are important. In the absence of steric effects one would expect K_1 to show the following patterns: (i) It should be larger for $P(n-Bu)_3$ than for $P(n-OBu)_3$ because the former is more basic. (2) It should be larger for R' = Ph than R' = Me because the former is more electron withdrawing. For the methyl carbene complex (R' = Me) K_1 is indeed larger with $P(n-Bu)_3$ than with $P(n-OBu)_3$ but for the bulkier phenylcarbene complex (R' = Ph) the K_1 values for the two nucleophiles are comparable or even smaller with $P(n-Bu)_3$ in the case of the chromium complex. This reversal reflects the larger size of $P(n-Bu)_3$ relative to $P(n-OBu)_3$.¹¹³

Interestingly, most of the steric effect that affects the equilibrium constants is expressed in k_{-1} while k_1 is relatively unaffected. This is clearly seen in the large increase in k_{-1} for the change from R' = Me to R' = Ph with $P(n-Bu)_3$ as the nucleophile. These results suggest a transition state with little P-C bond formation in the k_1 direction.

Phosphine and phosphite addition to dithiocarbene complexes such as **51** has also been reported.^{113,114} Different products are obtained depending on whether the reaction is with a tertiary or a secondary phosphine (Scheme 6). The rate law for the



reactions of **51** with PEt₃, HPMe₂, P(Ph)₂Me, HPPh₂ and P(OMe)₃ is given by equation (53).¹¹⁴

$$-\frac{\mathrm{d}[\mathbf{51}]}{\mathrm{d}t} = k[\mathbf{51}][\mathrm{phosphine}]$$
(53)

The mechanism proposed for the reaction with tertiary phosphines is shown in equation (54).

~ · ·

Two possibilities were discussed: rate-limiting nucleophilic attack $(k_2 \gg k_{-1}; k_2 \gg k_1)$ and rate-limiting rearrangement of the tetrahedral intermediate to the product $(k_{-1} \gg k_2; k_{-1} \gg k_1)$. The authors favored the first based on the trend in the experimental k values that follow the basicities of the phosphines, i.e., PEt₃ > HPMe₂ \approx PPh₂(Me) > HPPh₂ > P(OMe)₃. This interpretation is supported by the fact that HPPh₂ fits into this order even though the reaction leads to a different product but apparently also involves rate-limiting nucleophilic attack on the carbene carbon. The proposed mechanism for this latter reaction is shown in Scheme 7. The rate-limiting nucleophilic addition of the phosphine is followed either by a direct rapid rearrangement involving H atom migration to



Scheme 7

the carbon and P migration to the metal (path a) or by step-wise rearrangement according to path b.

ALCOHOLS AND ALKOXIDE ION NUCLEOPHILES

In 1968, Kreiter¹¹⁵ reported that the ethoxymethylcarbene complex **66** reacts with excess of methanol in the presence of catalytic amounts of NaOMe at room temperature to yield the corresponding methoxymethyl carbene complex. This reaction implies a process such as shown in equation (55).



Similar conclusions were reached by Schubert and Fischer¹¹⁶ who showed that **68** in CD_3OD in the presence of catalytic amounts of $NaOCD_3$ is rapidly converted to **69**, and **11** is converted to **70**. No direct evidence for the tetrahedral intermediate was obtained, though, because it reacts further to form products such as **71**.^{116,117}



| Parameter | 11 | 13 |
|---|-------|-------|
| $\frac{1}{k_1 (M^{-1} s^{-1})}$ | 77.1 | 186 |
| $k_{-1} (s^{-1})$ | 1.10 | 1.68 |
| $K_1^{a} (M^{-1})$ | 70.1 | 111 |
| $k_{-1}^{\rm BH} ({\rm M}^{-1} {\rm s}^{-1}) ({\rm C}_{6}{\rm H}_{5}{\rm OH})^{b}$ | 9.43 | 16.8 |
| $k_{-1}^{\rm BH} ({\rm M}^{-1} {\rm s}^{-1}) (3.5 - {\rm Cl}_2 {\rm C}_6 {\rm H}_3 {\rm OH})^b$ | 772 | 1500 |
| $\log k_0^c$ | ~0.96 | ~1.25 |

Table 9 Rate and equilibrium constants for the reaction of $(CO)_5Cr=C(OMe)Ph$ (11) and $(CO)_5W=C(OMe)Ph$ (13) with MeO⁻ in methanol at 25°C (equation 56)

Reference 118.

^{*a*} K_1 obtained as k_1/k_{-1} .

^bGeneral acid-catalyzed conversion of **72** to **11** and **13** by C₆H₅OH and 3,5-Cl₂C₆H₃OH, respectively. ${}^{c}k_{0}$ is the intrinsic rate constant, see text.

Direct evidence for the symmetrical dimethoxy adducts **72** (M = Cr and W) was obtained by recording time-resolved UV–Vis spectra of its formation during a period of about 1 s starting 10 ms after mixing **11** or **13** with a *ca.* 0.2 M NaOMe solution in methanol.¹¹⁸

$$(CO)_{5}M = C \xrightarrow{OMe}_{Ph} + MeO^{-} \xrightarrow{k_{1}} (CO)_{5}M - C \xrightarrow{Ph}_{OMe}$$
(56)

$$11 M = Cr \qquad 72$$

Acidification of the solution within 1-2 s after formation of **72** leads to almost quantitative recovery of **11** or **13**, respectively, but when acidification was delayed significantly little or no (CO)₅M=C(OMe)Ph could be recovered. NMR and low temperature IR experiments provided additional evidence for formation of **72**.

A stopped-flow kinetic study of this reaction¹¹⁸ yielded the results summarized in Table 9. The k_{-1}^{BH} values refer to general acid catalyzed MeO⁻ expulsion, most probably by a concerted process with a transition state such as **73**; in the forward direction this would correspond to general base catalyzed addition of methanol.

$$\begin{array}{c} & & \text{OMe} \\ & & & \downarrow \\ (\text{CO})_5\text{M} \stackrel{\bullet}{\longrightarrow} \stackrel{\bullet}{\text{C}} \stackrel{\bullet}{\longrightarrow} \text{Ph} \\ & & & \text{OMe} \\ & & & & \\ & & & \\ & & & & \\ & & & \\ & & & & \\ & & & \\ & & & \\ & & & & \\ &$$
Table 9 includes an entry for log k_0 , with k_0 being the *intrinsic* rate constant of the reaction which refers to the hypothetical situation where $k_1 = k_{-1}$ and $K_1 = 1$ or $\Delta G^\circ = 0$. The significance of k_0 will be discussed below.

We note that the rate and equilibrium constants depend very little on the metal. This finding is reminiscent of the situation with phosphine addition (equation 51) where the kinetic and thermodynamic parameters are very similar for the Cr and W carbene complexes (Table 8).

As pointed out earlier, nucleophilic addition to Fischer carbene complexes is closely related to addition to carboxylic acid esters. The ester analog of reaction (56) is equation (57).

$$O = C \begin{pmatrix} OMe \\ Ph \end{pmatrix} + MeO^{-} \qquad \underbrace{k_{1}}_{k_{-1}} \quad O - \bigcup_{i=1}^{OMe} Ph \qquad (57)$$

However, the tetrahedral adduct (74) cannot be observed in methanol solution because the equilibrium of equation (57) is very unfavorable. An approximate K_1 value between 10^{-7} and 5×10^{-7} M⁻¹ may be estimated for equation (57), based on a combination of sources discussed in Ref. 118. This estimate suggests that formation of 72 is $(2 \times 10^8) - 10^9$ -fold more favorable than formation of 74, i.e., the (CO)₅M moiety is much more effective in stabilizing negative charge than oxygen. This enhanced stabilization is in large measure attributable to a resonance effect brought about by delocalization of the negative charge into the CO ligands of 72. Independent evidence for such charge delocalization comes from IR data. The CO stretching frequency shifts of the order of 80–90 cm⁻¹ observed upon conversion of 11 to 72¹¹⁸ indicate a reduction in the C=O double bond character of the CO ligands.

Additional insights into the resonance effect on the stabilization of **72** comes from a consideration of the rate constants k_1 and k_{-1} , or, more precisely, the *intrinsic* rate constants. The general definition of the intrinsic rate constant, k_0 , of a reaction with forward and reverse rate constants k_1 and k_{-1} , respectively, is $k_0 = k_1 = k_{-1}$ when $K_1 = 1$; if dealing with free energies, one can define an intrinsic barrier, ΔG_0^{\ddagger} , as $\Delta G_0^{\ddagger} = \Delta G_1^{\ddagger} = \Delta G_{-1}^{\ddagger}$ when $\Delta G^\circ = 0$.^{119,120} The significance of k_0 or ΔG_0^{\ddagger} is that they are purely kinetic measures of chemical reactivity because they separate out thermodynamic effects from kinetic effects and hence they allow meaningful comparisons of reactivity between different reactions.

An extensive body of research has shown that reactions that lead to resonance stabilized products or involve resonance stabilized reactants that lose their resonance during the reaction invariably have higher intrinsic barriers or lower intrinsic rate constants than comparable reactions that do not involve resonance effects.^{121–123} The underlying reason for the higher ΔG_0^{\ddagger} or lower k_0 values is that

the transition state is imbalanced in the sense that resonance development in the product lags behind, or the loss of resonance in the reactant is more advanced than bond changes. Such transition state imbalances have typically been inferred from Brønsted type structure reactivity coefficients. Most documented examples are from the field of proton transfers from carbon acids to a buffer base not subject to resonance effects (more on this in Section 4). In these reactions the imbalance manifests itself in the Brønsted α -value (change in the rate constant for deprotonation of the carbon acid arising from the change in a remote substituent of the acid) being substantially greater than the Brønsted β -value (change in the rate constant for deprotonation of the carbon acid arising from a change in the buffer base basicity).^{121–123} The best-known example is the deprotonation of substituted phenylnitromethanes by amines (equation 58) where $\alpha = 1.29$ and $\beta = 0.53$.¹²⁴



The reason why α exceeds β (β is commonly assumed to be an approximate measure of the degree of proton transfer at the transition state^{125–127}) is that at the transition state (**75**) the partial negative charge is mainly localized on the carbon and hence closer to the Z substituent than in the product ion where the charge is delocalized into the nitro group.

In nucleophilic addition reactions, the imbalance results in α_{nuc}^n exceeding β_{nuc}^n ; α_{nuc}^n and β_{nuc}^n are normalized structure–reactivity coefficients that are the analogs of α and β , respectively, in proton transfers.¹²⁸ Note that since α_{nuc}^n is usually obtained from the variation of a phenyl substituent, it is equal to the normalized Hammett ρ value. A representative example is shown in equation (59) where $\alpha_{nuc} = 0.67$ and $\beta_{nuc}^n = 0.37$.¹²⁹ Again, the exalted α_{nuc}^n coefficient is the result of the negative charge at the transition state **76** being closer to the Z substituent than it is in the adduct.



The connection between enhanced intrinsic barriers and the lack of synchrony between resonance effects and bond changes is a manifestation of a more general phenomenon called the principle of non-perfect synchronization (PNS). This principle applies to other types of transition state imbalances as well and states that any product stabilizing factor whose development at the transition state lags behind bond changes, or any reactant stabilizing factor that is lost ahead of bond changes, increases the intrinsic barrier or lowers the intrinsic rate constant.^{121–123} Besides resonance, the kinds of stabilizing factors referred to here include solvation, hydrogen bonding effects and electrostatic effects. For factors that are *destabilizing*, e.g., steric effects, the changes in ΔG_0^{\ddagger} or k_0 are in the opposite direction and the same is true for stabilizing factors that develop ahead of bond formation, e.g., soft–soft, hard–hard acid–base interactions.

Intrinsic rate constants may be calculated based on the Marcus equation¹²⁰ although we prefer, when sufficient data are available, to obtain k_0 from interpolated and extrapolated Brønsted plots.^{121–123} For reactions (56) and (57) there were not enough data available for the construction of Brønsted plots and hence k_0 was estimated based on the simplest version of the Marcus formalism (equation 60). The log k_0 values are approximately 0.96 and 1.25 for the reaction of **11** and **13** (equation 56), respectively (Table 9).

$$\log k_0 = \log k_1 - 0.5 \log K_1 \tag{60}$$

In comparison, the estimated log k_0 value for reaction (57) is 3.1,¹¹⁸ i.e., k_0 for the Fischer carbenes is two orders of magnitude lower than for the ester. This is consistent with the fact that **72** derives a significant part of its stabilization from resonance while **74** does not.

The lowering of $\log k_0$ for the reactions of **11** relative to the reaction of methylbenzoate, although substantial, is not as dramatic as that for the reaction of MeO⁻ or OH⁻ with nitro activated electrophiles such as **77**¹³⁰ (log $k_0 \approx -0.76$) or **61** (log $k_0 \approx -3.9$).¹³¹ These latter log k_0 values show that the π -acceptor effect of nitro groups is significantly stronger than that of the (CO)₅M moiety, a point to which we shall return in Section 4.



In an effort to examine whether reaction (56) or similar reactions in fact have an imbalanced transition state, substituent effects on the rate and equilibrium constants of the reaction of 78 with alkoxide ions (equation 61) were determined.¹³²



For **78a** the investigation included Z = 4-Me₂N, 4-MeO, 4-Me, H, 4-F, 3-Cl and 4- CF_3 , for 78b Z = 4-MeO, 4-Me, H, 4-F and 4-Cl; the nucleophile/solvent combinations used were MeO- in methanol and in 90% MeCN-10% methanol (v/v), and OH⁻, HC \equiv CCH₂O⁻ and CF₃CH₂O⁻ in 50% MeCN-50% water (v/v). All reactions were at 25°C.

Some of the Hammett ρ values obtained in this study are summarized in Table 10 while β_{nuc} , β_{nuc}^{n} , etc. for the reactions with HC=CCH₂O⁻ and CF₃CH₂O⁻ are reported in Table 11. With respect to the question of imbalance, the α_{nuc}^{n} values average around 0.56 \pm 0.05 while for β_{nuc}^n the average is ≤ 0.44 , i.e., $\alpha_{nuc}^n > \beta_{nuc}^n$, which suggests a transition state (80) with the partial negative charge residing mainly on the metal rather than dispersed into the CO ligands. The size of the imbalance is relatively modest, which is consistent with the fact that the intrinsic rate constants are only reduced by about two orders of magnitude relative to k_0 for the ester reaction (equation 57).



80

Another type of transition state imbalance was noted by determining substituent effects on the rate and equilibrium constants for buffer-catalyzed MeOH addition to **78a** (equation 62).



78a

Table 10 Hammett ρ values for the reactions of alkoxide ions with (CO)₅Cr=C(OMe)C₆H₄Z (**78a**) and (CO)₅W=C(OMe)C₆H₄Z (**78b**) at 25°C (equation 61)

| Reaction | $\rho(k_1)$ | $\rho(k_{-1})$ | $\rho(K_1)$ | $\rho_{\rm n}(k_1) = \alpha_{\rm nuc}^{\rm n}$ | $\rho_{\rm n}(k_{-1})$ |
|---|---|---|---|---|--|
| $78a + MeO^{-a} 78b + MeO^{-a} 78a + HC \equiv CCH_2O^{-b} 78b + HC \equiv CCH_2O^{-b}$ | $\begin{array}{c} 2.67 \pm 0.10 \\ 2.37 \pm 0.22 \\ 1.39 \pm 0.14 \\ 1.68 \pm 0.17 \end{array}$ | $\begin{array}{c} -2.24 \pm 0.15 \\ -1.67 \pm 0.17 \\ -1.06 \pm 0.11 \\ -1.39 \pm 0.16 \end{array}$ | $\begin{array}{c} 4.92 \pm 0.12 \\ 4.04 \pm 0.08 \\ 2.45 \pm 0.24 \\ 3.07 \pm 0.01 \end{array}$ | $\begin{array}{c} 0.54 \pm 0.02 \\ 0.59 \pm 0.05 \\ 0.57 \pm 0.01 \\ 0.55 \pm 0.05 \end{array}$ | $\begin{array}{c} -0.46 \pm 0.2 \\ -0.41 \pm 0.05 \\ -0.43 \pm 0.01 \\ -0.45 \pm 0.05 \end{array}$ |

Reference 132.

^{*a*}In MeOH. ^{*b*}In 50% MeCN–50% water (v/v).

| | 78a (Z = 4-Cl) | 78b (Z = 4-Cl) |
|--|---------------------------|---------------------------|
| $\beta_{\text{nuc}} = d \log k_1 / d p K_{\text{a}}^{\text{ROH}}$ $\beta_{\text{lg}} = d \log k_{-1} / d p K_{\text{a}}^{\text{ROH}}$ | $0.42 \le -0.49$ | $0.54 \le -0.57$ |
| $\beta_{\rm eq} = {\rm d} \log k_1 / {\rm d} {\rm p} K_{\rm a}^{\rm ROH}$ | ≥0.91 | ≥1.11 |
| $\beta_{\text{nuc}}^{\text{n}} = d \log k_1 / d \log K_1$ $\beta_{\text{lg}}^{\text{n}} = d \log k_{-1} / d \log K_1$ | $\leq 0.46 \\ \leq -0.54$ | $\leq 0.42 \\ \leq -0.51$ |

Table 11 Brønsted coefficients for the reactions of $HC \equiv CCH_2O^-$ and $CF_3CH_2O^-$ with **78a** (Z = 4-Cl) and **78b** (Z = 4-Cl) in 50% MeCN-50% water at 25°C (equation 61)

Reference 132.

Normalized ρ values for this reaction catalyzed by three different aryloxide ions are reported in Table 12 while Brønsted coefficients for this type of catalysis are summarized in Table 13. The rather low $\beta(k_1^B)$ values (high $\alpha(k_{-1}^{BH})$) suggest that the proton transfer from the attacking methanol nucleophile to the buffer base has made much less progress than the C–O bond formation at the transition state (or that in the reverse direction protonation of the departing MeO⁻ by the buffer acid is ahead of C–O bond cleavage), as shown in **82** (δ^+ on MeO group).



Finally, although not relevant with respect to the question of imbalance, some comments regarding the actual (non-normalized) ρ values for reaction (61) summarized in Table 10 are in order. The $\rho(K_1)$ and $\rho(k_1)$ values are seen to be significantly smaller and the $\rho(k_{-1})$ significantly less negative for the reactions of HC=CCH₂O⁻ in 50% MeCN-50% water than for the reactions of MeO⁻ in methanol. A combination of two factors appears to be responsible for this result. One

| Table 12 | Normalized | Hammett | ρ values | for | buffer-catalyzed | MeOH | addition | to |
|--------------|--------------------------------------|-------------|---------------|------|------------------|------|----------|----|
| $(CO)_5Cr=C$ | (OMe)C ₆ H ₄ Z | (78a) in Me | eOH (equa | tion | 62) | | | |

| B ⁻ | $\rho_{\rm n}(k_1^{\rm B})$ | $ ho_{ m n}(k_{-1}^{ m BH})$ |
|--|-----------------------------|------------------------------|
| $C_6H_5O^-$ | 0.62 ± 0.07 | -0.38 ± 0.07 |
| $4\text{-BrC}_6\text{H}_4\text{O}^-$ | 0.58 ± 0.07 | -0.42 ± 0.07 |
| 3,5-Cl ₂ C ₆ H ₃ O ⁻ | 0.59 ± 0.06 | -0.41 ± 0.06 |

Reference 132.

| Z | $oldsymbol{eta}(k_1^{ m B})$ | $\alpha(k_{-1}^{ m BH})$ |
|-----------|--------------------------------|------------------------------------|
| 4-MeO | 0.17 ± 0.02 | 0.83 ± 0.02 |
| 4-Me H | 0.16 ± 0.02 0.17 + 0.04 | 0.84 ± 0.02 0.83 ± 0.04 |
| 4-F | 0.12 ± 0.02 | 0.88 ± 0.02 |
| 4-Cl | 0.10 ± 0.04 | 0.90 ± 0.04 |

Table 13 Brønsted coefficients for buffer catalysis of MeOH addition to $(CO)_{5-}$ Cr=C(OMe)C₆H₄Z (**78a**) in MeOH (equation 62)

Reference 132.

is the higher polarity of the aqueous acetonitrile ($\varepsilon = 58.8$) compared to that of methanol ($\varepsilon = 32.6$). The other is the change in the nucleophile; since the HC=CCH₂O group in **79** or **80** is more electron withdrawing than the MeO group, it contributes more to the stabilization of the negative charge than the MeO group. This, just as is the case with a more polar solvent, reduces the requirement for stabilization of the charge by the Z substituent.

THIOL AND THIOLATE ION NUCLEOPHILES

In 1972, Fischer *et al.*⁸⁰ reported the successful conversion of methoxy carbene complexes of the type 16 (M = Cr and W; R' = Me and Ph) into the thioalkyl carbene

$$(CO)_5 M = C \xrightarrow{OMe}_{R'} \xrightarrow{RSH}_{(CO)_5 M} = C \xrightarrow{SR}_{R'} + MeOH$$
(63)
16 83

complexes **83** by treatment of the solid **16** with RSH (R' = Me, Et and Ph) in the absence of any solvent. In fact no reaction could be observed either in polar or non-polar solvents.⁸⁰ However, **83** (R = Ph) was obtained in benzene-methanol mixtures when NaSPh was used as the nucleophile⁸¹ in the reaction with **16** (M = Cr, Mo and W; R' = Me). In this case there is an initial rapid formation of a stable intermediate that was assumed to be the tetrahedral adduct **84**. Subsequent addition of HCl to **84** led to conversion to products.

$$\overbrace{(CO)_5M}^{-} \overbrace{C-R'}^{OMe}$$

84

Using Na₂CO₃ as a base catalyst in methanol, Aumann and Schröder⁸² reacted **85** (R' = Me and Ph) with several thiols (R = C₆H₁₁, Et, Ph and alkyl) to obtain the corresponding alkylthic carbene complexes, again assuming that the reaction involved a tetrahedral intermediate.

$$(CO)_{5}Cr = C \xrightarrow{OEt}_{R'} + RSH \xrightarrow{Na_{2}CO_{3}}_{MeOH} (CO)_{5}Cr = C \xrightarrow{SR}_{R'} + MeOH$$

$$85 \qquad 86 \qquad (64)$$

Direct evidence for the tetrahedral intermediate and its spectroscopic characterization along with a detailed kinetic study of the reactions of **11**, **13**, **42** and **87** with a series of alkane thiolate ions (*n*-PrS⁻, HOCH₂CH₂S⁻, MeO₂CCH₂CH₂S⁻ and MeO₂CCH₂S⁻) in 50% MeCN-50% water (v/v) were reported by Bernasconi *et al.*^{133,134} Some of the salient features of these reactions can be summarized as follows.

$$(CO)_5 M = C \bigvee_{Ph}^{OMe} (CO)_5 M = C \bigvee_{Ph}^{OEt} (CO)_5 M = C \bigvee_{Ph}^{SR}$$

$$(CO)_5 M = C \bigvee_{Ph}^{SR} (CO)_5 M = C \bigvee_{Ph}^{SR}$$

$$(CO)_5 M = C \bigvee_{Ph}^{SR} (CO)_5 M = C \bigvee_{Ph}^{SR}$$

$$(CO)_5 M = C \bigvee_{Ph}^{SR} (CO)_5 M = C \bigvee_{Ph}^{SR}$$

$$(CO)_5 M = C \bigvee_{Ph}^{SR} (CO)_5 M = C \bigvee_{Ph}^{SR}$$

$$(CO)_5 M = C \bigvee_{Ph}^{SR} (CO)_5 M = C \bigvee_{Ph}^{SR}$$

$$(CO)_5 M = C \bigvee_{Ph}^{SR} (CO)_5 M = C \bigvee_{Ph}^{SR} (CO)_5 M = C \bigvee_{Ph}^{SR}$$

In the presence of low thiolate ion concentrations at a pH below the pK_a^{RSH} of the thiol, a clean conversion of **11**, **13**, **42** or **87** to **88** was observed. This is shown in Fig. 4 for a representative example. The figure shows a sharp isosbestic point at 422 nm, indicating that there is no accumulation of an intermediate to detectable levels. A different situation arises at relatively high thiolate ion concentrations at a pH close to or higher than pK_a^{RS} . Here the carbene complex is rapidly converted into a new species whose UV spectrum (dashed line in Fig. 4) resembles neither that of the starting material nor that of the product. This species is stable for several minutes, but upon acidification of the solution yields the substitution product. This species is the tetrahedral intermediate, a conclusion that is not only consistent with the UV spectrum but also with IR, ¹H NMR and ¹³C NMR spectra.¹³⁴

The above observations were interpreted in terms of equation (65). At high pH and high [RS⁻] the first step is fast and the equilibrium favors the intermediate while



Fig. 4 Conversion of 5×10^{-5} M (CO)₅W=C(OMe)Ph (**13**) (spectrum A) to (CO)₅. W=C(SR)Ph (**90b**) (R = MeOCOCH₂CH₂, spectrum B) in the presence of 7.94×10^{-5} M MeOCOCH₂CH₂S⁻ and 10^{-3} M MeOCOCH₂CH₂SH in an *N*-methylmorpholine buffer at pH 7.56 in 50% MeCN-50% water (v/v) at 20°C. Spectra taken every 10 s for 150 s. Spectrum C corresponds to intermediate (**89**) (equation 65) generated in the presence of 10^{-3} M MeOCOCH₂CH₂S⁻ at pH 10.69. (Figure taken from Ref. 134, by permission from the American Chemical Society.)

product formation is slow.



In fact spontaneous conversion of **89** to products (k_2 step) is so slow that product formation only occurs after addition of acid, indicating H⁺ and/or buffer and catalyzed ($k_2^H a_{H^+}$ and k_2^{BH} [BH], respectively) alkoxide ion departure; this is reminiscent of the observations with NaSPh in benzene–methanol mixtures where product formation only occurs after addition of HCl.⁸¹ At low pH, **89** turns into a steady-state intermediate because the equilibrium of the first step is no longer favorable and a_{H^+} and/or [BH] are high enough to ensure rapid product formation via the $k_2^H a_{H^+}$ and/or k_2^{BH} [BH] pathways; this essentially corresponds to the situation in methanol in the presence of Na₂CO₃ as reported by Aumann and Schröder.⁸²

Kinetic analysis confirmed the above picture and allowed a determination of k_1 , k_{-1} , $k_2^{\rm H}$ and $k_2^{\rm BH}$ (B = N-methylmorpholinium ion). A summary of the various rate and equilibrium constants is provided in Table 14. Several conclusions of general interest emerge from these data.

(1) As pointed out earlier, the reactivity of pentacarbonyl-type Fischer carbene complexes towards nucleophiles is much higher than that of carboxylic esters. For example, K_1 for MeO⁻ addition to **11** (equation 56) is approximately $2 \times 10^8 - 10^9$ higher than for MeO⁻ addition to methylbenzoate. A comparison of K_1 for *n*-PrS⁻ addition to **11** (1.06 × 10⁴ M⁻¹) with Guthrie's¹³⁵ estimate of $K_1 \approx 7.9 \times 10^{-14} \text{ M}^{-1}$ for EtS⁻ addition to acetic acid in water (equation 66) suggests an even greater reactivity difference ($\approx 1.3 \times 10^{17}$ -fold) between the Fischer carbene complex and the acryl analog.

$$CH_{3}COH + EtS^{-} = O - OH_{1} - CH_{3}$$

$$CH_{3}COH + EtS^{-} = O - OH_{1} - CH_{3}$$

$$SEt$$
(66)

This enhanced reactivity difference for thiolate addition has been attributed to the combination of two factors.¹³⁴ The first is the favorable soft acid–soft base^{136,137} interaction for the reaction of the soft carbene complex with the soft thiolate ion which contrasts with the unfavorable hard acid (CH₃COH)–soft base (EtS⁻) interaction in reaction (66). The second is the favorable hard acid–hard base interaction between the hard MeO⁻ with the hard methylbenzoate which contrasts with the unfavorable soft acid–hard base interaction of the carbene complex with MeO⁻.

(2) Relative to their proton basicities, the thiolate ions are much more reactive towards Fischer carbene complexes than alkoxide ions. For example, for the reaction of n-PrS⁻ (p $K_a^{\text{RSH}} = 11.94$) with **11**, $K_1 = 1.06 \times 10^4 \text{ M}^{-1}$ and $k_1 = 1.34 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$ (Table 14), while for the reaction of HC=CCH₂O⁻ (p $K_a^{\text{ROH}} = 13.68$) with **11**, $K_1 = 1.67 \times 10^2 \text{ M}^{-1}$ and $k_1 = 47 \text{ M}^{-1} \text{ s}^{-1}$.¹³² These comparisons demonstrate the well-known high carbon basicity and high nucleophilicity of sulfur bases which have been mainly attributed to their strong polarizability^{138,139} or softness.^{136,137} As pointed out above, the softness of the carbene complex enhances the effect of the soft nucleophile. An additional factor,

| | $CH_{3}CH_{2}CH_{2}S^{-}$ $(pK_{a}^{RSH} = 11.94)$ | $HOCH_2CH_2S^-$ $(pK_a^{RSH} = 10.79)$ | $\frac{\text{MeO}_2\text{CCH}_2\text{CH}_2\text{S}^-}{(pK_a^{\text{RSH}} = 10.69)}$ | $MeO_2CCH_2S^-$ $(pK_a^{RSH} = 9.45)$ |
|--|---|---|---|---------------------------------------|
| $ \frac{(\text{CO})_{5}\text{Cr}=\text{C(OMe)Ph (11)}}{k_{1} (\text{M}^{-1} \text{s}^{-1})} \\ \frac{k_{2}^{\text{H}} (\text{M}^{-1} \text{s}^{-1})}{k_{-1} (\text{s}^{-1})} \\ \frac{k_{-1} (\text{s}^{-1})}{K_{1} = k_{1}/k_{-1} (\text{M}^{-1})} \\ \frac{k_{2}^{\text{BH}} (\text{M}^{-1} \text{s}^{-1})^{a}}{k_{2}^{\text{BH}} (\text{M}^{-1} \text{s}^{-1})^{a}} $ | 1.34×10^{4} 5.39×10^{8} 1.26 1.06×10^{4} | $2.25 \times 10^{4} \\ 3.50 \times 10^{8} \\ 5.01 \\ 4.49 \times 10^{3} \\ 1.39 \times 10^{2} \\ \end{cases}$ | $2.30 \times 10^{4} \\ 1.99 \times 10^{8} \\ 6.55 \\ 3.51 \times 10^{3} \\ 6.88 \times 10^{1}$ | 6.76×10^4 |
| $(CO)_{5}W=C(OMe)Ph (13)$ $k_{1} (M^{-1} s^{-1})$ $k_{2}^{H} (M^{-1} s^{-1})$ $k_{-1} (s^{-1})$ $K_{1} = k_{1}/k_{-1} (M^{-1})$ $k_{2}^{BH} (M^{-1} s^{-1})^{a}$ | 4.21×10^{4} 5.69×10^{8} 3.11 1.35×10^{4} | 6.52×10^{4} 3.76×10^{8} 14.0 4.66×10^{3} 2.17×10^{2} | 7.26×10^4 2.05×10^8 16.3 4.45×10^3 1.46×10^2 | 1.73×10^{5} |
| $(CO)_{5}Cr=C(OEt)Ph (42)$ $k_{1} (M^{-1} s^{-1})$ $k_{2}^{H} (M^{-1} s^{-1})$ $k_{-1} (s^{-1})$ $K_{1} = k_{1}/k_{-1} (M^{-1})$ $k_{2}^{BH} (M^{-1} s^{-1})^{a}$ | 7.05×10^{3} 4.23×10^{8} 1.38 5.11×10^{3} | 1.20×10^{4} 2.67×10^{8} 9.08 1.32×10^{3} 8.40×10^{1} | 1.20×10^{4} 1.68×10^{8} 9.88 1.21×10^{3} 4.33×10^{1} | 3.43×10^{4} |
| $(CO)_{5}W=C(OEt)Ph (87)$ $k_{1} (M^{-1} s^{-1})^{a}$ $k_{2}^{H} (M^{-1} s^{-1})$ $k_{-1} (s^{-1})$ $K_{1} = k_{1}/k_{-1} (M^{-1})$ $k_{2}^{BH} (M^{-1} s^{-1})^{a}$ | 2.33×10^{4} 5.12×10^{8} 2.98 ± 0.33 7.82×10^{3} | 4.04×10^{4} 2.65×10^{8} 20.7 1.95×10^{3} 9.23×10^{1} | $\begin{array}{c} 4.25 \times 10^{4} \\ 1.84 \times 10^{8} \\ 26.4 \\ 1.61 \times 10^{3} \\ 6.47 \times 10^{1} \end{array}$ | 1.05×10^5 |

 Table 14
 Rate and equilibrium constants for the reactions of 11, 13, 47 and 87 with various thiolate ions in 50% MeCN-50% water, v/v) at 25°C (equation 65)

Reference 134.

 $^{a}BH = N$ -methylmorpholinium ion.

particularly important in hydroxylic solvents, is the weaker solvation of thiolate ions compared to that of alkoxide ions.

(3) Using equation (60), one can calculate approximate intrinsic rate constants for thiolate ion addition. For the reaction of n-PrS⁻ with 11 and 13 one obtains $\log k_0 \approx 2.11$ and ≈ 2.56 , respectively. This compares with $\log k_0 \approx 0.96$ and ≈ 1.25 for the reaction of MeO⁻ in methanol with **11** and **13**, respectively, or log $k_0 \approx 0.74$ and ≈ 0.94 for the reactions of HC=CCH₂O⁻ in 50% MeCN-50% water with 11 and 13, respectively.¹³² The differences in the intrinsic rate constants between the thiolate and alkoxide ion reactions are believed to be the result of transition states that are imbalanced with respect to the factors that are responsible for the differences in the equilibrium constants, i.e., the strong solvation of the alkoxide ions¹³⁴ and the softsoft interactions in the thiolate ion reactions. Specifically, following generally observed behavior,^{140,141} the partial desolvation of the nucleophile that occurs as it enters the transition state should be more advanced than bond formation according to the rules of the PNS.^{121,122} This has the effect of reducing k_0 and more so for the reactions with the more strongly solvated alkoxide ions. Regarding the soft-soft interactions, it is not unreasonable to assume that they are more advanced than bond formation at the transition state,¹⁴² which has the effect of enhancing k_0 for the thiolate ion reactions. Hence both factors reinforce each other in increasing the difference in the intrinsic rate constants of these reactions.

(4) Nucleophilic addition to the tungsten carbene complexes is slightly favored over addition to chromium complexes. The effect on K_1 is minimal but somewhat larger on k_1 . The $K_1(W)/K_1(Cr)$ ratios vary between 1.27 and 1.53, the $k_1(W)/k_1(Cr)$ ratios between 2.56 and 3.54 (Table 14). Similar results were found for MeO⁻ addition to **11** and **13**: $K_1(W)/K_1(Cr) = 1.58$, $k_1(W)/k_1(Cr) = 2.41$ (Table 9). The rather small dependence of K_1 on the metal may be the result of the extensive dispersion of the negative charge into the CO ligands of the adducts. Since this leaves little charge on the metal, the identity of the metal has only a small influence on the stability of the adduct. An additional factor is related to the strong contribution of **91** to the structure of **11** or **13**. Because **91** leads to substantial negative charge on the negative charge by the (CO)₅W moiety in the adduct is partially offset by a similar effect on the carbene complex, a point to which we shall return in Section 4.



The fact that the k_1 values show a somewhat greater dependence on the metal than K_1 and that not only k_1 but also k_{-1} is larger for the tungsten complexes is noteworthy; it means that the *intrinsic* rate constant is somewhat higher for the tungsten complexes than for the chromium complexes. This can be understood with

reference to equation (67). If, due to the typical imbalance, the fractional charge on the *metal* is greater at the transition state **92** than in the adduct **93** and tungsten is better able to support negative charge than chromium, the greater stabilization resulting from the change from chromium to tungsten will affect the transition state more than the adduct and increase k_1 more than K_1 .



(5) The mechanism of the acid-catalyzed leaving group departure is probably concerted, with transition states **94** for the $k_2^{\rm H}$ and **95** for the $k_2^{\rm BH}$ step, respectively. There is an increase in $k_2^{\rm H}$ and $k_2^{\rm BH}$ with increasing basicity of the thiolate ion. This indicates the operation of an electronic push by the RS group, presumably because of developing resonance (**96**) in the transition state. From the slopes of plots of log $k_2^{\rm H}$ versus $pK_a^{\rm RSH}$, $\beta_{\rm push}$ values in the order of 0.3 were obtained.



With respect to the dependence on the leaving group, the $k_2^{\rm H}$ and $k_2^{\rm BH}$ values are slightly larger for the methoxy compared to the ethoxy derivative. In acid-catalyzed alkoxide ion departure the relative progress of C–O bond cleavage versus oxygen protonation determines which leaving group leaves faster. If C–O bond cleavage is ahead of proton transfer, the energetics of the reaction should be more reflective of

Table 15 Brønsted-type coefficients for the dependence of K_1 , k_1 and k_{-1} on pK_a^{RSH} in the reactions of (equation 65)

| | 11 | 13 | 42 | 87 |
|---|----------------|----------------|--------------|----------------|
| $\beta_{\text{nuc}} = d \log k_1 / d p K_a^{\text{RS}}$ $\beta_{\text{lg}} = d \log k_{-1} / d p K_a^{\text{RSH}}$ | -0.28 -0.55 | -0.25 -0.57 | -0.28 - 0.70 | -0.26 -0.75 |
| $\beta_{\rm eq} = d \log K_1 / d p K_{\rm a}^{\rm RSH}$ | 0.36 | 0.39 | 0.50 | 0.54 |

Reference 134.

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C-O bond cleavage than of the proton transfer and hence the less basic leaving group MeO^- should leave faster. On the other hand, if proton transfer is ahead of C-O bond cleavage the opposite holds and the ethoxy derivative should react faster. The results suggest that the former situation applies.

The dependence of K_1 , k_1 and k_{-1} on pK_a^{RSH} yielded the Brønsted type coefficients summarized in Table 15. The most interesting feature is that β_{nuc} is negative, i.e., nucleophilic reactivity of the thiolate ions *decreases* with increasing basicity. Negative β_{nuc} values are unusual but not without precedent.^{143–146} According to Jencks et al.,¹⁴³ they may result from the requirement of partial desolvation of the nucleophile prior to reaction, as shown in Scheme 8. In terms of Scheme 8, the experimental k_1 values correspond to $K_d k_1'$ with K_d being the equilibrium constant for partial desolvation of the nucleophile. In the simplest approach, β_{nuc} may be approximated by equation (68). Since desolvation becomes more difficult as the basicity of RS⁻ increases, $\beta_d < 0$. If β'_{nuc} is small because of an early transition state, β_{nuc} may be dominated by β_d and become negative. A more elaborate discussion which also takes into account the desolvation effect on pK_a^{RSH} is presented in Ref. 134.

$$\beta_{\text{nuc}} = \frac{d \log k_1}{d p K_a^{\text{RSH}}} = \frac{d \log K_d k_1'}{d p K_a^{\text{RSH}}} = \frac{d \log K_d}{d p K_a^{\text{RSH}}} + \frac{d \log k_1'}{d p K_a^{\text{RSH}}} = \beta_d + \beta_{\text{nuc}}$$
(68)

A recent study of thiol/thiolate ion addition to alkylthio Fischer carbene complexes has led to additional insights into the structure–reactivity relationships of these nucleophilic addition reactions and also provides evidence for metal protonation of tetrahedral adducts.¹⁴⁷ Two reaction series were investigated in 50% MeCN–50% water (v/v) at 25°C. One is the reaction of **97** with a series of RS⁻/RSH (R = *n*-Pr, Me, HOCH₂CH₂, MeO₂CCH₂CH₂ and MeO₂CCH₂), the other is the reaction of **98** with HOCH₂CH₂S⁻/HOCH₂CH₂SH; in this case the phenyl substituent in the tungsten complexes was varied (Z = CF₃, Cl, F, H, Me, MeO and Me₂N).





Fig. 5 Plots of k_{obsd} versus [H⁺] according to equation (70) for the reaction of **99** (M = W) with HCl in 50% MeCN-50% water (v/v) at 25°C. \Box , Z = MeO; \blacksquare , Z = Me; \bigcirc , Z = H; \bullet , Z = F; \triangle , Z = Cl. (Figure taken from Ref. 147, by permission from the American Chemical Society.)

The reactions can be described by equation (69) which includes a rapid protonation equilibrium that becomes significant in highly acidic solutions.



The metal protonated adducts (100) could not be detected directly because they are very acidic ($pK_a^{MH} < 1$). This means they cannot be generated at the required pH because the thiolate ions are protonated under these conditions and the corresponding thiols are not reactive enough to add to the carbene complex. However there is kinetic evidence for 100. When 99 that has been generated at high pH is reacted with HCl, the pseudo-first-order rate constant for the conversion of 99 back to 98 shows a non-linear dependence on a_{H^+} , as shown in Fig. 5 for some representative examples. This dependence is consistent with H⁺-catalyzed

| Carbene complex | pK_a^{MH} | $k_{-1}^{\rm H} ({\rm M}^{-1} {\rm s}^{-1})$ | Carbene complex | $pK_a^{\rm MH}$ | $k_{-1}^{\rm H} ({\rm M}^{-1}{\rm s}^{-1})$ |
|----------------------|-------------|--|---------------------|-------------------------|---|
| 98a (Z = H) | 0.74 | 15.2 | 98b (Z = H) | $0.37 \\ 0.06 \\ -0.47$ | 11.7 |
| 98b (Z = MeO) | 0.62 | 40.4 | 98b (Z = F) | | 5.31 |
| 98b (Z = Me) | 0.54 | 30.0 | 98b (Z = Cl) | | 1.91 |

Table 16 pK_a^{MH} and k_{-1}^{H} values for the reactions of various alkylthic carbene complexes with HOCH₂CH₂S⁻ in 50% MeCN-50% water (v/v) at 25°C (equation 69)

Reference 147.

expulsion of RS^- from **99** preceded by fast equilibrium protonation on the metal and can be described by equation (70).

$$k_{\rm obsd} = \frac{K_{\rm a}^{\rm MH} k_{-1}^{\rm H} a_{\rm H^+}}{K_{\rm a}^{\rm MH} + a_{\rm H^+}}$$
(70)

The pK_a^{MH} values obtained from this analysis are summarized in Table 16. This study was the first to provide such pK_a^{MH} values. The protonated adducts (100) bear some resemblance to hydrido transition metal complexes such as 101 whose pK_a^{MH} values have been determined in water, methanol and acetonitrile;¹⁴⁸ in water $pK_a^{MH} = 5.4$ (Cr) and 8.0 (W); in methanol $pK_a^{MH} = 6.4$ (Cr) and 9.0 (W), respectively. In 50% MeCN-50% water, the pK_a^{MH} values would be expected to be somewhat higher than in water, perhaps similar to the values in methanol. The fact that the pK_a^{MH} values for 100 are several units lower than those for 101 is consistent with the effect of replacing the electron donating cyclopentadienyl ligand with two electron withdrawing CO ligands and the C(SR)₂C₆H₄Z moiety. What is more difficult to understand is why there is such a small metal dependence of the pK_a^{MH} values of 100 but a strong dependence for 101.

$$\overset{H}{\underset{J}{\eta-C_5H_5M(CO)_3}}$$

101

Structure–reactivity comparisons of the reactions of thioalkyl carbene complexes with those of the alkoxy analogs reveal interesting differences as well as similarities. Table 17 provides a comparison of rate and equilibrium constants between the reactions of n-PrS⁻ with 97 and 11 or 13. The following points are noteworthy.

(1) The equilibrium constants for the MeS carbene complexes are over a 100-fold larger than for the MeO carbene complexes. This must be the result of the weaker π -donor effect of the MeS (96) group compared to that of the MeO (91) group. ($\sigma_{\rm R} = 0.43$ and 0.15 for MeO and MeS, respectively),¹⁴⁹ i.e., the MeS carbene complex is not as strongly stabilized as the MeO analog. The stronger steric hindrance in the adduct of the MeS derivative ($E_{\rm S} = -0.55$ and -1.07 for MeO

Rate and equilibrium constants for the reaction of n-PrS⁻ with 97a, 97b, 11 and 13 in 50% MeCN-50% water (v/v) at 25°C (equation Table 17 65)

| | (CO) ₅ Cr=C(SMe)Ph 97a | (CO) ₅ Cr=C(OMe)Ph 11 | (CO) ₅ Cr=C(SMe)Ph 97b | (CO) ₅ W=C(OMe)Ph 13 |
|---|--|----------------------------------|---|---------------------------------|
| k_{1}^{RS} (M ⁻¹ s ⁻¹) | 5.33×10^{2} | 1.34×10^{4} | 1.58×10^{3} | 4.21×10^{4} |
| $k_{-1} (s^{-1})$ | 4.85×10^{-4} | 1.26 | 3.33×10^{-4} | 3.11 1.25 × 10 ⁴ |
| K_1^{KS} (M ⁻¹) $k^{\text{H}} a^{a}$ (M ⁻¹ s ⁻¹) | 1.10×10^{3} 5.09×10^{3} | 1.06 X 10 | 4.75×10^{-3} 3.86×10^{3} | 1.35 × 10 ⁻ |
| $k_2^{\text{Hb}} (\text{M}^{-1} \text{ s}^{-1})$ | | 5.39×10^{8} | | 5.69×10^{8} |

Reference 147.

^{*a*}Refers to H⁺-catalyzed loss of MeS⁻ from the respective adduct. ^{*b*}Refers to H⁺-catalyzed loss of MeO⁻ from the respective adduct.

and MeS, respectively)¹⁵⁰ and the weaker inductive effect of MeS ($\sigma_{\rm F} = 0.30$ and 0.20 for MeO and MeS, respectively),¹⁴⁹ both of which would make adduct formation less favorable for the MeS compared to the MeO carbene complexes are not nearly important enough to offset the influence of π -donor effects.

- (2) In contrast to the equilibrium constants, the k_1 values for the MeS carbene complexes are about 25-fold lower than those for the MeO analogs. This reversal indicates that the *intrinsic* rate constants, k_0 , for the MeO complexes are substantially larger than for the MeS complexes. These differences in the intrinsic rate constants are, once again, indicative of transition state imbalances where one or several of the factors – inductive, steric and π -donor effects – either lag behind or are ahead of bond formation. As a result of the imbalances the relative importance of these factors in how they affect the rate constants is different from how they affect the equilibrium constants and this is the reason why the intrinsic rate constants differ. A detailed analysis led to the conclusion that the steric effect whose development runs ahead of C-S bond formation and the lag in the charge delocalization into the (CO)₅M moiety which leads to a stronger transition state stabilization by the inductive effects of the MeX group when X = O than when X = S are mainly responsible for the differences in the intrinsic rate constants.¹⁴⁷ A potential third contribution is an imbalance involving the π -donor effects of the MeO and MeS groups.
- (3) The determination of the substituent effect on the reaction of **98b** with HOCH₂CH₂S⁻/HOCH₂CH₂SH yielded the Hammett ρ values summarized in Table 18. The ρ values for the $k_1^{\text{RS}} k_{-1}$ pathway are strikingly smaller than the corresponding ρ values for alkoxide ion addition to **78c** or **78b** (equation 61). For example, for the reaction of HC=CCH₂O⁻ with **78b**, $\rho(k_1) = 1.68$, $\rho(k_{-1}) = -1.39$ and $\rho(K_1) = 3.07$ (Table 10) while the corresponding values for the reaction of HOCH₂CH₂S⁻ with **98b** are 0.78, -0.67 and 1.45, respectively. Since one reaction involves an alkoxy carbene complex with an oxyanion nucleophile while the other is between an alkylthio carbene complex and a thiolate ion, it is not clear which of these two changes is responsible for large difference in the ρ values. The results of a substituent effect study on the intramolecular reaction (71)¹⁵¹ (note that other aspects of reaction (71) will be discussed below) shed some light on this question.



The reported ρ values ($\rho(k_1) \approx 0.96$, $\rho(k_{-1}) = -1.65$ and $\rho(K_1) \approx 2.67$) are much closer to those for the reaction of HC=CCH₂O⁻ with the alkylthio

| Parameter | ρ | Parameter | ρ |
|-------------------------------------|-------|--------------------|--------|
| k_1^{RS} | 0.78 | k_1^{RSH} | - 1.04 |
| k_{-1} | -0.67 | k_{-1}^{H} | -2.65 |
| $K_1^{\rm RS} = k_1^{\rm RS} / k_1$ | 1.45 | $K_1^{ m MH}$ | 2.08 |

Table 18 Hammett ρ values for the reaction of HOCH₂CH₂S⁻/HOCH₂CH₂SH with **98b** (Z = CF₃, Cl, F, H. Me, MeO and Me₂N) in 50% MeCN-50% water (v/v) at 25°C (equation 65)

Reference 147.

complex (Table 18). This implies that it is the nature of the nucleophile rather than the identity of the group attached to the carbone carbon that dramatically changes the response to substituent effects. In other words, $\rho(K_1)$ is only modestly affected by the change from a methoxy to an alkylthio group in the reactions with oxyanions, but $\rho(K_1)$ is much smaller in the reaction with a thiolate ion than in the reaction of an oxyanion with the same carbone complex.

These results may be understood in the context of soft-hard acid-base theory.^{136,137} As mentioned earlier, the Fischer carbene complexes can be regarded as soft electrophiles, especially the alkylthic complexes. Hence, the adducts **99** formed by the reaction of **98b** with a thiolate ion nucleophile enjoy enhanced stability due to the symbiotic effect¹³⁶ of adding a soft nucleophile. This stabilization apparently reduces the need for additional stabilization by the phenyl substituent, which translates to a reduced $\rho(K_1)$ value.

INTRAMOLECULAR NUCLEOPHILIC REACTIONS

Fischer carbene complexes of the type **104** and **105** undergo a cyclization reaction according to the general Scheme 9 (X = O or S);^{151,152} some aspects of this process (M = W, X = S, equation 71) have already been discussed above. The rate and equilibrium constants for the cyclization step via the k_1-k_{-1} pathway are summarized in Table 19. As had been observed when comparing the relative reactivities of the MeS and MeO complexes **97** versus **11** and **13**, respectively, towards *n*-PrS⁻ (Table 17), we note again that the equilibrium constants are higher

$$(CO)_{5}M = C \begin{pmatrix} XCH_{2}CH_{2}OH \\ Ar \end{pmatrix} \begin{pmatrix} K_{OH}[OH^{-}] \\ Ar \end{pmatrix} (CO)_{5}M = C \begin{pmatrix} XCH_{2}CH_{2}O^{-} \\ Ar \end{pmatrix} \begin{pmatrix} k_{1} \\ k_{-1} \end{pmatrix} (CO)_{5}M - C - O \\ Ar \end{pmatrix} \begin{pmatrix} k_{1} \\ k_{-1} \end{pmatrix} \begin{pmatrix} K_{OH}[OH^{-}] \\ K_{-1} \end{pmatrix} \begin{pmatrix} k_{1} \\ k_{-1} \end{pmatrix} \begin{pmatrix} K_{OH}[OH^{-}] \\ K_{-1} \end{pmatrix} \begin{pmatrix} K_{OH}[$$

Scheme 9

Table 19 Rate and equilibrium constants for the cyclization step through the $k_1 - k_{-1}$ pathway of Scheme 9, intrinsic rate constants and effective molarities

| Reactants | <i>K</i> ₁ | $k_1 (s^{-1})$ | $k_{-1} (s^{-1})$ | $\log k_0^a$ | $\mathrm{EM}(K_1)$ (M) | $\mathrm{EM}(k_1)$ (M) |
|--|--|---|---|--|--|--|
| $\begin{array}{c} ({\rm CO})_5{\rm Cr}{=}{\rm C}({\rm OCH}_2{\rm CH}_2{\rm OH}){\rm Ph}\ ({\bf 104a}) \\ ({\rm CO})_5{\rm W}{=}{\rm C}({\rm OCH}_2{\rm CH}_2{\rm OH}){\rm Ph}\ ({\bf 104b}) \\ ({\rm CO})_5{\rm Cr}{=}{\rm C}({\rm SCH}_2{\rm CH}_2{\rm OH}){\rm Ph}\ ({\bf 105a}) \\ ({\rm CO})_5{\rm W}{=}{\rm C}({\rm SCH}_2{\rm CH}_2{\rm OH}){\rm Ph}\ ({\bf 105b}) \\ ({\rm CO})_5{\rm Cr}{=}{\rm C}({\rm OMe}){\rm Ph}\ + {\rm HC}{\equiv}{\rm CCH}_2{\rm O}^b \\ ({\rm CO})_5{\rm W}{=}{\rm C}({\rm OMe}){\rm Ph}\ + {\rm HC}{\equiv}{\rm CCH}_2{\rm O}^b \end{array}$ | $\approx 2.12 \times 10^{5} 9.03 \times 10^{4} \approx 1.06 \times 10^{6} \approx 3.01 \times 10^{5} 167^{c} 167^{c} $ | $\approx 7.21 \times 10^{3}$ 1.81×10^{3} ≈ 30.5 ≈ 55.3 47.0^{d} 84.0^{d} | $3.40 \times 10^{-2} 2.00 \times 10^{-2} 2.88 \times 10^{-5} 1.83 \times 10^{-5} 0.281 0.504$ | ≈ 1.19 ≈ 0.78 ≈ -1.53 ≈ -1.50 0.56 0.81 | 2.7×10^4 1.1×10^4 | 7.0×10^2 9.9 × 10 ¹ |

In 50%MeCN-50% water (v/v) at 25°C, Ref. 152.

^{*a*}log k_0 obtained as log $k_1 - 0.5 \log K_1$.

^bReference 132.

^cIn units of M^{-1} . ^dIn units of $M^{-1} s^{-1}$.

| Carbene complex | <i>K</i> ₂ (M) | $k_2 (s^{-1})$ | $k_{-2}^{\rm H} ({\rm M}^{-1}{\rm s}^{-1})$ | $\log k_0^a$ |
|--|---|--|---|--|
| $\begin{array}{l} ({\rm CO})_5{\rm Cr}{=}{\rm C}({\rm OCH_2CH_2OH}){\rm Ph}\ ({\bf 104a})\\ ({\rm CO})_5{\rm W}{=}{\rm C}({\rm OCH_2CH_2OH}){\rm Ph}\ ({\bf 104b})\\ ({\rm CO})_5{\rm Cr}{=}{\rm C}({\rm SCH_2CH_2OH}){\rm Ph}\ ({\bf 105a})\\ ({\rm CO})_5{\rm W}{=}{\rm C}({\rm SCH_2CH_2OH}){\rm Ph}\ ({\bf 105b}) \end{array}$ | $\begin{array}{c} 1.36 \times 10^{-8} \\ 4.59 \times 10^{-9} \\ 5.26 \times 10^{-9} \\ 1.50 \times 10^{-8} \end{array}$ | $1.505.032.56 \times 10^{-2}7.11 \times 10^{-2}$ | $\begin{array}{c} 1.10 \times 10^5 \\ 1.12 \times 10^9 \\ 5.04 \times 10^6 \\ 4.74 \times 10^6 \end{array}$ | ≈ 5.11 ≈ 4.88 ≈ 2.56 ≈ 2.76 |

Table 20 Rate and equilibrium constants for the cyclization steps through the k_2-k_{-2} pathway of Scheme 9; intrinsic rate constants

In 50%MeCN-50% water (v/v) at 25°C, Ref. 152.

^{*a*}log k_0 obtained as log $k_2 - 0.5 \log K_2$.

for the thia carbene compared with oxa complexes but the opposite is true when the k_1 values are being compared. The implication is again that the intrinsic rate constants (included in Table 19) are lower for the alkylthio derivatives. This state of affairs can be explained as before, i.e., the higher K_1 values for the alkylthio complexes result from the weaker π -donor effect of the sulfur group while the lower intrinsic rate constants arise from several transition state imbalances. One is early development of the steric effect (lowers k_0 for the thia complexes more than for the oxa complexes); another is the delayed delocalization of the negative charge into the (CO)₅M moieties that allows enhanced transition state stabilization by an inductive effect which is stronger for the oxa than the thia complexes.



Table 19 includes the results for the *intermolecular* reactions of $HC\equiv CCH_2O^$ with $(CO)_5M=C(OMe)Ph$ discussed earlier. Comparison with the intramolecular reactions of **102** yields the effective molarities $(EMs)^{153,154}$ given in the table. The EM values in Table 19 have been corrected for the difference in the pK_a values of the oxyanion nucleophiles.¹⁵² These EM values are at the low end of the range which is common for the formation of 5-membered rings in conformationally flexible systems $(10^4 - 10^8 \text{ M}).^{153,154}$

The results for the $k_2 - k_{-2}^{\rm H}$ pathway are reported in Table 20. One possibility is that this pathway proceeds in two steps as shown in equation (72),¹⁵² with a transition state for the rate-limiting step such as **107**. According to equation (72), $k_{-2}^{\rm H}$ in Scheme 9 takes on the meaning of $k'_{-2}/K_{\rm a}^{\pm}$; another possibility is that the reaction is concerted with a transition state such as **108**. Since no buffer catalysis could be

observed, the two-step mechanism seems more likely.

$$(CO)_{5}M = C \xrightarrow{XCH_{2}CH_{2}OH}_{Ar} \xrightarrow{k_{2}} (CO)_{5}M - \overset{-}{C} - \overset{-}{OH}_{Ar} \xrightarrow{k_{a}^{+}} (CO)_{5}M - \overset{-}{\overset{-}{C} - O}_{Ar} \xrightarrow{k_{a}^{+}}_{H^{+}} (CO)_{5}M - \overset{-}{\overset{-}{C} - O}_{Ar} \xrightarrow{k_{a}^{+}}_{Ar} (72)$$
106

$$(CO)_{5}M \xrightarrow{\delta-} C \cdots OH \qquad (CO)_{5}M \xrightarrow{\delta-} C \cdots O \cdots H \cdots OH_{2} \qquad (CO)_{5}M \xrightarrow{\delta-} C \cdots O$$

The most noteworthy feature of the results for the $k_2 - k_{-2}^{H}$ pathway is that, while the *relative* log k_0 values for the oxa versus the thia complexes are close to those for the k_1-k_{-1} pathway, the *absolute* values are about 2.5 log units higher for the $k_2-k_{-2}^{H}$ pathway.

There are at least two factors that are likely to contribute to this difference in the intrinsic rate constants for the two pathways. One is that in the k_1-k_{-1} pathway the nucleophile is a strongly basic alkoxide ion. The strong solvation of such oxyanions is known to reduce their kinetic activity.^{140,141} This reduction is a classic PNS¹²¹⁻¹²³ effect and the result of a transition state where the loss of solvation is ahead of bond formation. As discussed earlier, the lower intrinsic reactivity of alkoxide ions relative to thiolate ions was interpreted along similar lines.

The second factor is an electrostatic effect at the transition state. The transition state for the k_1-k_{-1} pathway is shown as **109**; the most likely transition state for the $k_2-k_{-2}^{\rm H}$ pathway is **107**. In **109** there is a destabilizing effect resulting from the two partial negative charges, while in **107** there is a stabilizing interaction between the positive and negative charge.

HYDROXIDE ION AND WATER AS NUCLEOPHILES

The hydrolysis of Fischer carbene complexes has received little attention in the early literature, probably because the major interest in these compounds has been their potential utility in synthesis. The first hydrolysis study was reported in 1993 by Aumann *et al.*¹⁵⁵ who investigated the reaction of several carbene complexes of the type **110** (R = Ph, CH=CHPh, C₄H₃S, CH=CHC₆H₃S and C=CPh) in THF



containing small quantities of water (equation 73).

$$(CO)_5Cr = C \bigvee_{R}^{OEt} + H_2O + C_6H_{12}N_4 \longrightarrow RCH = O + (CO)_5CrC_6H_{12}N_4 + EtOH$$
110

In the presence of the tricyclic amine urotropin (hexamethylenetetramine, $C_6H_{12}N_4$) the aldehyde RCH=O was formed in \geq 90% yield with all R groups except when R is C=CPh; in this latter case the triple bond undergoes nucleophilic attack by the amine.

Based on their product analysis Aumann *et al.*¹⁵⁵ proposed the mechanism shown in Scheme 10. It involves two principal stages. The first is a nucleophilic substitution of the ethoxy group of **110** which presumably proceeds through a tetrahedral intermediate (T_{OH}^-) . The nucleophile, OH⁻, is generated in the acid–base equilibrium between water and urotropin. The second stage is the conversion of **112** into the corresponding aldehyde, a known reaction reported by Fischer and Maasböl;⁵ since under basic conditions the product of the first stage is undoubtedly present in its anionic form **111**, fast protonation of **111** precedes the product forming step. Note that **112** with R = Ph or Me as well as the tungsten analogs are isolable compounds that have been fully characterized.¹⁵⁶

A kinetic study of the hydrolysis of **11**, **13**, **42** and **87** in 50% MeCN–50% water (v/v) was reported in 1997.¹⁵⁷ Rates were determined in KOH and HCl solutions as well as in triethylamine, *N*-morpholine and acetate buffers, covering a pH range 2–14. Two kinetic processes were observed. The first one was relatively fast and corresponds to the substitution of the alkoxy groups by OH. This substitution undoubtedly proceeds via a tetrahedral intermediate but, in contrast to the reactions of similar carbene complexes with alkoxide or thiolate ions, this intermediate does not accumulate to detectable levels. The second, slower process refers to the

(73)



Fig. 6 Rate–pH profile of the hydrolysis of $(CO)_5Cr=C(OMe)CH=CHPh$ (113) in 50% MeCN–50% water (v/v) at 25°C. (Figure taken from Ref. 157, by permission from the American Chemical Society.)

conversion of $(CO)_5M=C(OH)R$ to the corresponding aldehyde (RCH=O) and $(CO)_5MOH$.



The kinetic study focused on the first process. A representative rate-pH profile is shown in Fig. 6 from which a k_{H_2O} for the water reaction (plateau) and a k_{OH} for the OH⁻ reaction were extracted. The k_{H_2O} and k_{OH} values are reported in Table 21. Not included in the table are k_B values that represent general base catalysis of the water reaction.

Table 21 Summary of $k_{H_{2O}}$ and k_{OH} values for the hydrolysis of several carbene complexes in 50% MeCN-50% water (v/v) at 25°C (Scheme 11)

| Carbene complex | $k_{\rm H_2O}~({\rm s}^{-1})$ | $k_{\rm OH} ({\rm M}^{-1}{\rm s}^{-1})$ |
|--|-------------------------------|---|
| $(CO)_5Cr=C(OMe)Ph(11)$ | 2.9×10^{-3} | 26.6 |
| $(CO)_5W=C(OMe)Ph(13)$ | 2.8×10^{-3} | 26.3 |
| $(CO)_5Cr=C(OEt)Ph$ (42) | 4.5×10^{-4} | 10.5 |
| (CO) ₅ Cr=C(OEt)Ph (112) | 2.8×10^{-3} | 17.6 |
| (CO) ₅ Cr=C(OMe)CH=CHPh (113) | 1.8×10^{-4} | 14.6 |
| | | |

Reference 157.



A central question that was addressed in this study is why no tetrahedral intermediates could be detected; based on the reactions of alkoxide ions with carbene complexes such as **11** or **13** (equation 61) one might have expected that, at high pH, these intermediates would accumulate to detectable levels. Two requirements must be met for an intermediate to become detectable in any two-step reaction. The first is that the equilibrium of the first step favors the intermediate, the second is that the rate of intermediate formation is faster than its conversion to products. Applied to equation (74), this would translate into $K_1^{OH}[OH^-] > 1$ and $k_1^{OH}[OH^-]/k_2^{OH} > 1$.

$$(CO)_{5}M = C \xrightarrow{OMe}_{Ph} \underbrace{k_{1}^{OH}[OH^{-}]}_{k_{-1}^{H_{5}O}} \xrightarrow{(CO)_{5}M} \underbrace{-C}_{OH} \xrightarrow{k_{2}^{OH}}_{OH} (CO)_{5}M = C \xrightarrow{OH}_{Ph} + MeO^{-}$$

$$11M = Cr$$

$$13M = W$$

$$(74)$$

Since K_1^{OH} can reasonably be expected to be at least as large and probably larger than the equilibrium constant for addition of the much less basic HC=CCH₂O⁻ to (CO)₅Cr=C(OMe)Ph ($K_1^{RO} = 167 \text{ M}^{-1}$),¹³² the condition that $K_1^{OH}[OH^-] > 1$ should easily be met at high pH. Furthermore, since the k_1^{OH} values (26 M⁻¹ s⁻¹) are comparable to $k_1^{RO} = 47 \text{ M}^{-1} \text{ s}^{-1}$ for HC=CCH₂O⁻¹³² and expulsion of MeO⁻ from **113** should proceed at a similar rate as expulsion of MeO⁻ from the corresponding HC=CCH₂O⁻ adduct, one expects $k_1^{OH}[OH^-]/k_2^{OH} > 1$ at high pH. Thus, even though both requirements for detectability of **114** appear to be met, the intermediate is not observable.

The reason for this apparent contradiction is that for **114** there are two additional pathways available for its conversion to products which result from the acidic nature of the OH group. These pathways are shown in the expanded Scheme 11 which also includes the various pathways in acidic solution, i.e., nucleophilic attack by water $(k_1^{H_2O})$ and H⁺-catalyzed expulsion of OH⁻ (k_{-1}^H) and RO⁻ (k_2^H) from T⁻_{OH} as well as H⁺-catalyzed expulsion of RO⁻ from T²⁻_{OH} (k_3^H) ; k_2^i represents RO⁻ departure that is



intramolecularly acid catalyzed by the OH group. It is this k_2^i and/or the pathways through T_{OH}^{2-} that lead to a more rapid collapse of \overline{T}_{OH} to products than via $k_2^{H_2O}$ and prevent \overline{T}_{OH} from becoming detectable. As a result, nucleophilic attack is rate limiting, i.e., $k_{OH} = k_1^{OH}$ and $k_{H_2O} = k_1^{H_2O}$. The driving force that makes the k_2^i step a potentially more favorable pathway

The driving force that makes the k_2^1 step a potentially more favorable pathway than the $k_2^{\text{H}_2\text{O}}$ step is that the acidity of the OH group increases strongly along the reaction coordinate which renders proton transfer to the departing RO⁻ thermodynamically favorable at the transition state, a pre-condition for effective acid catalysis.¹⁵⁸ In the pathway through T_{OH}^{2-} , the negative charge on the oxygen provides an extra electronic "push" in the form of transition state stabilization ($k_3^{\text{H}_2\text{O}}$ step) by the developing resonance of **116**. For this reason the collapse of T_{OH}^{2-} should be much faster than the collapse of T_{OH}^{-} .

Another interesting issue in the hydrolysis of alkoxy carbene complexes is the finding that replacement of the phenyl group with an alkyl group changes the mechanism even though the final products are the same (e.g., equation 75). The mechanism involves deprotonation of the alkyl group and hydrolytic breakdown of the anion, as discussed in Section 4.

$$(CO)_5 M = C \xrightarrow{OR} \xrightarrow{OH^-/H_2O} CH_3 CH = O + (CO)_5 MOH^- + ROH$$
(75)

0.0

Replacing the alkoxy group by an alkylthio group does not change the mechanism (Scheme 12) but it leads to a much more complex situation regarding the nature of the rate-limiting steps of the hydrolysis process.¹⁵⁹ This can be seen by inspection of the rate-pH profiles in Fig. 7 which suggest multiple changes in the rate-limiting step as the pH changes. The following interpretation of the rate-pH profiles was proposed.¹⁵⁹

Region I (pH ≥ 12.5 for (CO)₅Cr=C(SMe)Ph and ≥ 11.5 for (CO)₅-W=C(SMe)Ph). Nucleophilic attack by OH⁻ is rate limiting, i.e., the observed pseudo-first-order rate constant is given by equation (76). This means that the



Fig. 7 Rate-pH profiles of hydrolysis of $(CO)_5M=C(SMe)Ph$ (**90a**), \bigcirc ; **90b**, •; in 50% MeCN-50% water (v/v) at 25°C. (Figure taken from Ref. 159, by permission from the American Chemical Society.)

relationships of equations (77) and (78) apply. Note that in equation (78) the $k_2^H a_{H^+}$ term has been omitted because it is negligible down to pH 6 (see below).

$$k_{\rm obsd} = k_1^{\rm OH} [\rm OH^-] \tag{76}$$

$$k_1^{\text{OH}}[\text{OH}^-] \gg k_1^{\text{H}_2\text{O}} \quad \text{and} \quad k_{-1}^{\text{H}_2\text{O}} \gg k_{-1}^{\text{H}}a_{\text{H}^+}$$
(77)

$$k_{2}^{\rm H_{2}O} + k_{2}^{\rm i} + k_{a}^{\rm T} k_{3}^{\rm H_{2}O} / a_{\rm H^{+}} + k_{a}^{\rm T} k_{3}^{\rm H} \gg k_{-1}^{\rm H_{2}O}$$

$$\tag{78}$$

Region II (pH ~10 to ~12.5 for (CO)₅Cr=C(SMe)Ph, pH ~10 to ~11.5 for (CO)₅W=C(SMe)Ph)). Nucleophilic attack by water is rate limiting. Just as is the case for the hydrolysis of alkoxy carbene complexes, there is also general base catalysis of water attack, hence k_{obsd} is given by equation (79) (k_1^B [B] term not shown in Schemes 11 and 12). This rate law implies that the relationships of equations (80) and (81) hold; the $k_2^H a_{H^+}$ term is again negligible and omitted from equation (81).

$$k_{\rm obsd} = k_1^{\rm H_2O} + k_1^{\rm B}[{\rm B}] \tag{79}$$

$$k_2^{\rm H_2O} \gg k_1^{\rm OH} [\rm OH^-] \text{ and } k_{-1}^{\rm H} a_{\rm H^+} \gg k_{-1}^{\rm H_2O}$$
 (80)

$$k_2^{\rm H_2O} + k_2^{\rm i} + K_a^{\rm T} k_3^{\rm H_2O} / a_{\rm H^+} + K_a^{\rm T} k_3^{\rm H} \gg k_{-1}^{\rm H} a_{\rm H^+}$$
(81)

Region III (pH \sim 6 to \sim 10). In this pH range, equation (80) remains valid but equation (81) changes to equation (82) which means that the nucleophilic attack step is now a rapid

$$k_{-1}^{\rm H}a_{\rm H^+} \gg k_2^{\rm H_2O} + k_2^{\rm i} + K_{\rm a}^{\rm T}k_3^{\rm H_2O} / a_{\rm H^+} + K_{\rm a}^{\rm T}k_3^{\rm H}$$
(82)

pre-equilibrium and hence k_{obsd} is given by equation (83), with $K_1^{H_2O} = k_1^{H_2O}/k_{-1}^{H_1}$ being

$$k_{\text{obsd}} = \frac{k_1^{\text{H}_2\text{O}}}{a_{\text{H}^+}} (k_2^{\text{H}_2\text{O}} + k_2^{\text{i}} + K_a^{\text{T}} k_3)$$
(83)

the equilibrium constant for nucleophilic attack by water. The fact that the slope of the rate – pH profile is approximately unity indicates that neither the $k_2^{\rm H}a_{\rm H^+}$ term nor the $K_{\rm a}^{\rm T}k_3^{\rm H_2O}/a_{\rm H^+}$ term contribute significantly to $k_{\rm obsd}$ in this region and hence these two terms have been omitted from equations (82) and (83).

Region IV (pH \leq 6). Below pH 6, the $k_2^{\rm H}a_{\rm H^+}$ term finally becomes important; it now represents the dominant pathway for the rate-limiting collapse of $T_{\rm OH}^-$ (equation 84), equation (82) becomes equation (85), and equation (83) becomes equation (86).

$$k_2^{\rm H}a_{\rm H^+} \gg k_2^{\rm H_2O} + k_2^{\rm i} + K_a^{\rm T}k_3^{\rm H}$$
(84)

$$k_{-1}^{\rm H}a_{\rm H^+} \gg k_2^{\rm H}a_{\rm H^+} \tag{85}$$

$$k_{\text{obsd}} = K_1^{\text{H}_2\text{O}} k_2^{\text{H}} \tag{86}$$

Why are the rate-pH profiles so different for complexes with thiomethyl compared to those with alkoxy leaving groups? For the hydrolysis of the alkoxy carbene complexes there are no changes from rate-limiting nucleophilic attack at high pH to rate-limiting collapse of T_{OH} at low pH, i.e., nucleophilic attack is rate limiting over the entire rate-pH profile. The key feature responsible for the difference between the two types of carbene complexes is that oxyanion (OH⁻ as well as RO^-) departure from $\mathrm{T}^-_{\mathrm{OH}}$ is very sensitive to H^+ catalysis while for thiolate ion departure the sensitivity to H⁺ catalysis is much weaker. Hence, for the hydrolysis of the alkoxy carbene complexes, H^+ catalysis of conversion of T_{OH}^- to products and of conversion back to reactants becomes important in similar pH ranges. This means that lowering the pH affects the rates of conversion of T_{OH} to reactants and products in a comparable way so that no change in rate-limiting steps occurs. In contrast, for the hydrolysis of thioalkyl carbene complexes, $k_2^{\rm H}$ is so small that at pH values where the $k_{-1}^{\rm H}a_{\rm H^+}$ term starts to become important, the $k_{2}^{\rm H}a_{\rm H^+}$ terms is still negligible. Hence in the pH range 6-10 where H⁺ catalysis of OH⁻ departure $(k_{-1}^{\rm H}a_{\rm H^+})$ is already strong, H^+ catalysis of RS⁻ departure $(k_2^{\rm H}a_{\rm H^+})$ is insignificant which renders the $(k_{-1}^{H}a_{H^{+}})$ process to be faster than the conversion of T_{OH}^{-} to products and leads to the observed change in rate-limiting step. As the pH is lowered further, the $k_2^{\rm H}a_{\rm H^+}$ term eventually becomes significant but the $k_{-1}^{\rm H}a_{\rm H^+}$ has also increased so that the collapse of T_{OH}^- to products remains rate limiting.

The lower sensitivity to H^+ catalysis of thiolate compared to alkoxide ion departure has been observed in other reactions and is a general phenomenon; a detailed discussion has been presented in connection with the study of the acid-catalyzed breakdown of alkoxide and thiolate ion adducts of benzylidene Meldrum's acid type adducts.¹⁶⁰ The main reason for this reduced sensitivity appears to be the lower basicity of RS⁻ compared to RO⁻ or OH⁻ ions. However, the mechanism for H⁺ catalysis is probably the same for all leaving groups and most likely represents a concerted reaction with a transition state such as **117** or **118** (B = H₂O).



INSERTION REACTIONS INITIATED BY NUCLEOPHILIC ATTACK

There are a number of reactions that involve insertion of an alkyne or nitrile between the metal and the carbene carbon that are initiated by a nucleophilic addition to the carbene carbon. Many of these reactions have been reviewed;¹ examples that have been subjected to kinetic investigations are shown in equations $(87)^{161}$ and (88).^{162–164}



Rates of reaction (87) were measured for **78a** ($Z = CF_3$ and H) and **78b** ($Z = CF_3$, Br, H, Me and MeO) at various temperatures in *n*-octane.¹⁶¹ The reaction was found

| Carbene complex | $k = k_1^a (M^{-1} s^{-1})$ | ΔH^{\ddagger} (kcal mol ⁻¹) | ΔS^{\ddagger} (cal mol ⁻¹ deg ⁻¹) |
|-----------------------|-----------------------------|---|--|
| 78a $Z = CF_3$ | 1.71 | 6.70 | - 34.7 |
| 78a Z = H | 0.26 | 8.01 | -34.0 |
| 78b $Z = CF_3$ | 5.36 | 6.00 | -34.7 |
| 78b $Z = Br$ | 1.76 | 7.56 | -31.6 |
| 78b Z = H | 0.873 | 7.82 | - 32.1 |
| 78b $Z = Me$ | 0.334 | 8.73 | -31.1 |
| 78b Z = MeO | 0.105 | 9.43 | - 30.9 |
| | | | |

Table 22 Second-order rate constants and activation parameters for the reactions of $(CO)_5Cr=C(OMe)C_6H_4Z$ (**78a**) and $(CO)_5W=C(OMe)C_6H_4Z$ (**78b**) with $CH_3C=CNEt_2$ (equation 87) in *n*-octane at 20°C

Reference 161.

^aSee Scheme 13.

to be overall second order, i.e., first order in each of the reactants. Second-order rate constants at 20°C as well as ΔH^{\ddagger} and ΔS^{\ddagger} values are summarized in Table 22.

On the basis of these results the mechanism shown in Scheme 13 was proposed in which the k_1 step is rate limiting. In support of rate-limiting nucleophilic attack, Fischer and Dötz¹⁶¹ cite the fact that the second-order rate constants (Table 22) increase strongly as the Z substituent becomes more electron withdrawing (the reasons why electron withdrawing substituents enhance nucleophilic additions even with neutral nucleophiles have been discussed under "Phosphine and phosphite nucleophiles"), and that they correlate well with the equilibrium constants of (*n*-



Scheme 13

Bu)₃P addition to **63** (equation 52). The strongly negative ΔS^{\ddagger} values are also consistent with a bimolecular reaction that leads to a zwitterionic intermediate such as **122**.

Three kinetic investigations have been reported on reactions of **120** with nitriles (equation 88). The first two involved dialkylcyanamides as the nitriles, equation (89), with $RR' = Me_2$, MeEt, $(n-Pr)_2$ and $(i-Pr)_2$.^{162,163} With $Me_2N-C=N$ rates were determined for five derivatives of **120** ($Z = CF_3$, Br, H, Me and MeO) at different temperatures and also in a variety of solvents.



With the other nitriles the investigation was limited to **120** with Z = H. The reactions were again essentially second order, although in some cases a small first-order term was detected at high temperature. Using the same line of reasoning as for reaction (87) – strong acceleration by electron withdrawing substituents, large negative ΔS^{\ddagger} values and correlation of *k* with the equilibrium constants of equation (52) – it was concluded that reaction (89) must involve a rate-limiting nucleophilic attack by the nitrile on the carbene carbon of **120** (*k*₁) followed by rapid rearrangements as outlined in Scheme 14. The slight increase in the rate constants upon changing the methyl groups on the cyanamides to the more electron donating ethyl and propyl groups is also consistent with their mechanistic conclusion. As to the rearrangement of **125** into **124** via **126**, it is strongly analogous to that of **122** rearranging to **119** via **123** in Scheme 13.

In comparing the nucleophilic attack of $RR'N-C\equiv N$ on **120** with that of $CH_3C\equiv CNEt_2$ on **78b** one would expect the former reaction to be significantly faster because $RR'N-C\equiv N$ should be a stronger nucleophile and **120** should be more reactive than **78b** since it lacks the strong π -donor stabilization by the methoxy group. However, comparison of the *k* values in Tables 22 and 23 reveals the opposite behavior. Even though Fischer^{162,163} did not comment on this point, the most likely explanation of the observed reactivity patterns is that the two phenyl groups in **120** lead to strong steric crowding at the transition state of the k_1 step. The importance of steric effects in reactions of carbene complexes with nucleophiles has been pointed out earlier (see the section on "Phosphine addition").

The third kinetic study involved the reaction of **120** (Z = H) with arylnitriles and acetonitrile conducted in 1,1,2-trichloromethane at 50.6°C.¹⁶⁴ Even though **121** is the main product (equation 88), byproducts such as **127**, **128** and even traces of diphenylketene were detected.



Scheme 14

$$(CO)_5 W - N = C - R$$
 $(CO)_4 W (N = C - R)_2$
127 128

These byproducts are formed by a dissociative mechanism which is reflected in a rate law (equation 90) that includes a first-order term.

$$-\frac{d[120]}{dt} = k_{\rm II}[120][{\rm RCN}] + k_{\rm I}[120]$$
(90)

The following rate constants were obtained in 1,1,2-trichloroethane at 50.6°C: $k_{\rm I} = 4.5 \times 10^{-4} \, {\rm s}^{-1}$; $k_{\rm II} = 0.6 \times 10^{-4} \, {\rm M}^{-1} \, {\rm s}^{-1}$ for MeCN, $4.4 \times 10^{-4} \, {\rm M}^{-1} \, {\rm s}^{-1}$ for PhC=N, $7.9 \times 10^{-4} \, {\rm M}^{-1} \, {\rm s}^{-1}$ for *p*-MeOC₆H₄C=N and $1.97 \times 10^{-3} \, {\rm M}^{-1} \, {\rm s}^{-1}$ for *p*-Me₂NC₆H₄C=N. The trend in the $k_{\rm II}$ values for the arylnitriles is consistent with increased nucleophilicity due to the electron donating substituents and with the increasing importance of the $k_{\rm II}$ pathway relative to the $k_{\rm I}$ pathway. The mechanism of the $k_{\rm II}$ pathway was assumed to be analogous to that in Scheme 14, again with nucleophilic attack being the rate-limiting step. The fact that $k_{\rm II}$ for the arylnitriles is

| Z | $k = k_1^{a} (\mathbf{M}^{-1} \mathbf{s}^{-1})$ | ΔH^{\ddagger} (kcal mol ⁻¹) | ΔS^{\ddagger} (cal mol ⁻¹ deg ⁻¹) |
|--------------------------------------|---|---|--|
| Me ₂ N-C | $E \equiv N^b$ | | |
| MeO | 0.0182 | 11.0 | -28.9 |
| Me | 0.0459 | 10.6 | -28.5 |
| Н | 0.0836 | 9.3 | -31.8 |
| Br | 0.130 | 9.1 | -31.6 |
| CF ₃ | 0.337 | 8.9 | - 30.4 |
| Et N-0 Me | $C = N^{c}$ | 9 7 | - 29.6 |
| | 0.120 | 2.1 | 29.0 |
| (<i>i</i> -Pr) ₂ N- H | $-C \equiv N^c \\ 0.192$ | 9.5 | - 29.4 |
| $(Pr)_2N-C$ | $C \equiv N^c$ | | |
| H | 0.214 | 9.0 | -31.1 |
| ^a See Sche | me 14. | | |

Table 23 Second-order rate constants and activation parameters for the reactions of RR'N– C=N with $(CO)_5W=C(OMe)C_6H_4Z$ (**120**) in methylcyclohexane at 20°C

^bReference 162.

^cReference 163.

higher than for MeC=N, despite their lower basicity, was attributed to the capability of the phenyl ring to delocalize the positive charge that forms on the nitrile fragment in the first intermediate $(129 \leftrightarrow 129' \leftrightarrow 129'')$.



For the $k_{\rm I}$ pathway, dissociation of CO from the carbene complex was assumed to be rate limiting, in agreement with the fact that the measured $k_{\rm I}$ value is the same as the rate constant for thermolysis of **120** measured independently. The proposed mechanism for the dissociative pathway is shown in Scheme 15 (shown for Z = H).



Scheme 15

Another type of insertion reaction that is initiated by nucleophilic attack on the carbon carbon of 120 is shown in equation (91).



This reaction has been the subject of a kinetic study in dibutyl ether (R = Et with Z = MeO, Me, H, Br and CF₃; R = Me with Z = H; R = Ph with Z = H) and *n*-octane (R = Et with Z = H).¹⁶⁵ In dibutyl ether the reaction is strictly second order, i.e., first order in **120** and first order in isothiocyanate, while in *n*-octane there is both a second- and a third-order term (equation 92).

$$-\frac{d[120]}{dt} = k_{\rm II}[120][132] + k_{\rm III}[120][130]^2$$
(92)

The dependence of $k_{\rm II}$ shows the same characteristics as for the nitrile insertions discussed above – acceleration by electron withdrawing Z substituents in the carbene complex, increased rates when the R group becomes more electron donating, correlation of $k_{\rm II}$ with the equilibrium constants of equation (52), and strongly negative ΔS^{\ddagger} values – and suggests rate-limiting nucleophilic attack by the isothiocyanate on the carbene complexes. The proposed mechanism is shown in Scheme 16. Note that even though there is no direct evidence that the initial nucleophilic attack is through sulfur rather than nitrogen, the product strongly suggests that the former is the case.



Scheme 16

4 Acid-base reactions at the α -carbon

GENERAL FEATURES AND METHODS

The first indication that Fischer carbene complexes of the type **40** or **68** have pronounced acidic properties was Kreiter's¹⁶⁶ observation of the rapid conversion of **68** to **134a** in dilute NaOMe/MeOD solutions. Casey and Anderson¹⁶⁷ subsequently showed that in THF the acidity of **68** is approximately the same as that of *p*-cyanophenol whose pK_a is about 8.¹⁶⁸ In quoting this work, a number of authors, as recently as in 2000,²⁵ have rather misleadingly referred to **68** as an acid with a pK_a of 8. This inference, of course, cannot be correct since *p*cyanophenoxide ion and the conjugate base of **68** should respond quite differently to a solvent change from water to THF; indeed the pK_a^{CH} (the superscript CH will be used throughout this section to distinguish pK_a^{CH} from the pK_a of other acids) of **68** in water is 12.3,¹⁶⁹ not 8, and, in 50% MeCN–50% water (the standard solvent used for most pK_a^{CH} and proton transfer determinations of Fischer carbene complexes¹⁷⁰), 12.5.¹⁷¹ The problem in defining and determining pK_a^{CH} values in organic solvents such as THF will be taken up in more detail in a section titled "Thermodynamic acidities in organic solvents."

$$(CO)_5 M = C \bigvee_{CH_3}^{OMe} (CO)_5 M = C \bigvee_{CD_3}^{OMe} CD_3$$

$$68 M = Cr \qquad 134a M = Cr \\ 40 M = W \qquad 134b M = W$$

It is perhaps surprising that no pK_a^{CH} measurements in water for even the most prototypical of all Fischer carbene complexes such as **68** had been reported before 1989.¹⁶⁹ The most likely explanation for this state of affairs is that in aqueous solution the conjugate anions of carbene complexes hydrolyze rapidly which precludes equilibrium determinations by classical photometric methods. In fact, most pK_a^{CH} values of Fischer complexes in aqueous solution were obtained by kinetic rather than equilibrium methods. For **68** the pK_a^{CH} was determined in KOH solution by applying the relationship $K_a^{CH} = (k_1^{OH}/k_{-1}^{H_2O})K_w$ where k_1^{OH} is the rate constant for deprotonation of the carbene complex by OH^- , $k_{-1}^{H_2O}$ is the rate constant for protonation of the anion by water (equation 93) and K_w is the ionic product of the solvent;^{169,171} k_1^{OH} and $k_{-1}^{H_2O}$ were determined as the slope and intercept, respectively, of a plot of the observed pseudo-first-order rate constant for equilibrium approach versus [OH⁻] (equation 94).

$$(CO)_{5}M = C \begin{pmatrix} OMe \\ CH_{3} \end{pmatrix} \begin{pmatrix} k_{1}^{OH}[OH^{-}] + k_{1}^{B}[B] \\ k_{-1}^{H_{2}O} + k_{-1}^{BH}[BH^{+}] \end{pmatrix} (CO)_{5}M - C \begin{pmatrix} OMe \\ CH_{2} \end{pmatrix} (93)$$
68
135

$$k_{\rm obsd} = k_1^{\rm OH} [\rm OH^-] + k_{-1}^{\rm H_2 \rm O}$$
(94)

This methodology was successfully applied to other carbene complexes with relatively high pK_a^{CH} values¹⁷²⁻¹⁷⁵ but for the more acidic complexes it fails because $k_{-1}^{H_2O}$ becomes very small relative to the $k_1^{OH}[OH^-]$ term and hence cannot be evaluated accurately.^{171,176} In these cases the pK_a^{CH} values were obtained by measuring proton transfer rates in buffer solutions ($k_1^{B}[B]$ and $k_{-1}^{BH}[BH]$ terms in equation 93) of buffers that have pK_a^{BH} values close to pK_a^{CH} . In such solutions k_{obsd} is given by equation (95) (K_a^{BH} is the acidity constant of BH) and the slope of a plot of k_{obsd} versus [B] is given by equation (95). From a plot of these slopes versus a_{H^+} according to equation (96) one then obtains k_1^B and k_{-1}^{BH}/K_a^{BH} :

$$k_{\text{obsd}} = k_1^{\text{OH}}[\text{OH}^-] + k_{-1}^{\text{H}_2\text{O}} + \left(k_1^{\text{B}} + k_{-1}^{\text{BH}} \frac{a_{\text{H}^+}}{K_a^{\text{BH}}}\right)$$
(95)

slope =
$$k_1^{\rm B} + k_{-1}^{\rm BH} \frac{a_{\rm H^+}}{K_{\rm a}^{\rm BH}}$$
 (96)

The kinetic experiments described above were typically extended to include numerous other buffers in order to provide the respective k_1^{B} and k_{-1}^{BH} values. For
buffers whose pK_a^{BH} values were significantly below the pK_a^{CH} of the carbene complex, the kinetic experiments had to be conducted in the direction anion + BH \rightarrow carbene complex + B in order to make the reaction thermodynamically favorable. This was achieved by generating the anion in a KOH solution prior to mixing it with the buffer. However, because of the very short lifetime of the anion which was typically a few seconds due to its rapid hydrolytic decomposition (more on this in the section on the hydrolysis of ionizable carbene complexes), these experiments had to be conducted in a double mixing stopped-flow apparatus. This apparatus allows mixing of the anion solution with the buffer solution within 20– 30 ms after it has been generated in the presence of KOH, i.e., before significant decomposition can occur.

For the most acidic carbene complexes such as **136** $(pK_a^{CH} = 4.18)^{177}$ and **137** $(pK_a^{CH} = -0.03)$,¹⁷⁷ the proton transfer reactions were conducted in HCl and carboxylate buffer solutions.



In this case, the reaction pathways are the ones of equation (97)

$$C_{5}H_{5}(NO)(PPh_{3})Re \xrightarrow{k_{1}}^{H} k_{-1}^{H}a_{H'} + k_{-1}^{B}[B] C_{5}H_{5}(NO)(PPh_{3})Re \xrightarrow{Se} (97)$$
136 138

with $k_1^{\text{H}_2\text{O}}$ and k_{-1}^{H} referring to the deprotonation of the carbene complex by water and protonation of the anion by the hydronium ion, respectively, while k_1^{B} and k_{-1}^{BH} have the same meaning as in equation (93). For **137** the pK_a^{CH} was obtained in HCl solution where k_{obsd} is given by equation (98). Hence $K_a^{\text{CH}} = k_1^{\text{H}_2\text{O}}/k_{-1}^{\text{H}}$ with $k_1^{\text{H}_2\text{O}}$ and k_{-1}^{H} representing intercept and slope, respectively, of a plot of k_{obsd} versus a_{H^+} . For **136** the pK_a^{CH} was determined in chloroacetate buffers as described above for the amine buffers (equation 96).

$$k_{\rm obsd} = k_1^{\rm H_2O} + k_{-1}^{\rm H} a_{\rm H^+} \tag{98}$$

KINETIC AND THERMODYNAMIC ACIDITIES

As pointed out earlier, most data on pK_a^{CH} values and rate constants for proton transfer were obtained in 50% MeCN-50% water (v/v), a solvent that is essentially aqueous but which provides better solubility for some sparingly soluble Fischer carbene complexes. Some acidity measurements were reported in other solvents as

well, in particular in MeCN, THF, DMSO and acetone as will be elaborated upon in a later section.

Table 24 provides a summary of pK_a^{CH} values, the rate constants k_1^{OH} and $k_{-1}^{H_2O}$ (e.g., equation 93), the rate constants $k_1^{H_2O}$ and $k_{-1}^{H_1}$ (e.g., equation 97), and the corresponding *intrinsic* rate constants k_0^{OH} (for the $k_1^{OH} - k_{-1}^{H_2O}$ pathway) and $k_0^{H_2O}$ (for the $k_1^{H_2O} - k_{-1}^{H}$ pathway). These latter were estimated according to equations (99) and (100), respectively, where K_1^{OH} in equation (99) is defined as $k_1^{OH} / k_{-1}^{H_2O} = K_a^{CH} / K_w$.

$$\log k_0^{\rm OH} = \log k_1^{\rm OH} - 0.5 \log K_1^{\rm OH}$$
(99)

$$\log k_0^{\rm H_2O} = \log k_1^{\rm H_2O} - 0.5 \log K_{\rm a}^{\rm CH}$$
(100)

It needs to be emphasized that the intrinsic rate constants obtained via equations (99) and (100) are subject to significant uncertainty, especially those for which $\log K_1^{OH}$ or $\log K_a^{CH}$ differ substantially from zero. This is because equations (99) and (100) are based on the simplest form of the Marcus^{119,120} formalism (first two terms in equation 101) which may be a good approximation when ΔG° is close to zero but less so when $|\Delta G^{\circ}|$ becomes large (third term in equation 101). The $\log k_0$ values for these latter cases are identified by italics in the table.

$$\Delta G^{\ddagger} = \Delta G_0^{\ddagger} + 0.5 \Delta G_0^{\ddagger} + \frac{(\Delta G^{\circ})^2}{16 \Delta G_0^{\ddagger}}$$
(101)

The kinetic data from the reactions conducted in buffers were also used to determine the corresponding intrinsic rate constants. They were calculated from statistically corrected Brønsted plots of $\log(k_1^{\rm B}/q)$ or $\log(k_{-1}^{\rm BH}/p)$ versus $pK_a^{\rm BH} - pK_a^{\rm CH} + \log(p/q)$ where *p* is the number of equivalent protons on BH and *q* is the number of equivalent basic sites on B. Interpolation or extrapolation of such plots to $pK_a^{\rm BH} - pK_a^{\rm CH} + \log(p/q) = 0$ yields $\log k_0$. Such $\log k_0$ values for the reactions with primary aliphatic and secondary alicyclic amines as well as carboxylate ions are summarized in Table 25. It those cases where $\log k_0$ could be obtained from interpolation or short extrapolations the $\log k_0$ values can be assumed to be quite reliable but those obtained from long extrapolations are less accurate; the latter ones are again identified by italics.

The significance of intrinsic rate constants in the context of our understanding chemical reactivity has already been discussed in a section titled "Alcohols and alkoxide ion nucleophiles" and will be further elaborated upon below.

EFFECT OF STRUCTURE ON pKA VALUES

(CO)₅M type complexes

The pK_a^{CH} values of the neutral (CO)₅M type complexes that are summarized in Table 24 and span a range 8.4–14.5; the two cationic rhenium carbene complexes which are substantially more acidic ($pK_a^{CH} = 4.18$ and -0.03, respectively) will be dealt with below.

| Entry | | | pK_a^{CH} | $k_1^{\text{OH}} (\text{M}^{-1} \text{ s}^{-1})$ | $k_{-1}^{\rm H_2O}~({\rm s}^{-1})$ | $k_1^{\rm H_2O}~({\rm s}^{-1})$ | $k_{-1}^{\rm H} ({\rm M}^{-1} {\rm s}^{-1})$ | $\log k_0^{\rm OH}$ | $\log k_0^{\mathrm{H_2O}}$ |
|-------|------------------------------|------------------------------------|-------------|--|------------------------------------|---------------------------------|--|---------------------|----------------------------|
| 1 | $(CO)_5Cr$ | (8) ^{<i>a</i>} | 14.47 | 74.0 | 14.0 | | | 1.51 | |
| 2 | (CO) ₅ Cr | (141) ^{<i>a</i>} | 13.41 | 39.5 | 0.66 | | | 0.70 | |
| 3 | $(CO)_5Cr = C'_{CH_3}^{OEt}$ | (66) ^{<i>a</i>} | 12.98 | 241 | 1.48 | | | 1.38 | |
| 4 | $(CO)_5 Mo = C CH_3$ | (12) ^{<i>a</i>} | 12.81 | 181 | 0.75 | | | 1.07 | |
| 5 | $(CO)_5Cr = C CH_3$ | (68) ^{<i>a</i>} | 12.50 | 456 | 0.91 | | | 1.31 | |
| 6 | $(CO)_5W = C CH_3$ | (40) ^{<i>a</i>} | 12.36 | 284 | 0.42 | | | 1.04 | |

Table 24 Summary of pK_a^{CH} values and rate constants for proton transfer from carbene complexes to OH⁻ in 50% MeCN-50% water (v/v) at 25°C

(continued on next page)

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| Entry | | | | p <i>K</i> _a ^{CH} | $k_1^{\text{OH}} (\text{M}^{-1} \text{ s}^{-1})$ | $k_{-1}^{\rm H_2O}~({\rm s}^{-1})$ | $k_1^{\rm H_2O}~({\rm s}^{-1})$ | $k_{-1}^{\rm H} ({\rm M}^{-1} {\rm s}^{-1})$ | $\log k_0^{\mathrm{OH}}$ | $\log k_0^{\mathrm{H_2O}}$ |
|-------|--|--|-------------------------------------|---------------------------------------|--|------------------------------------|---------------------------------|--|--------------------------|----------------------------|
| 7 | $(CO)_5Cr = C$ | ,OMe | (145) ^{<i>a</i>} | 12.32 | 46.8 | 6.40×10^{-2} | | | -0.24 | |
| 8 | (CO) Cr = C' | CH ₂ CH ₃ OMe | (146) ^{<i>a</i>} | 12.27 | 2.97 | 3.57×10^{-3} | | | -0.99 | |
| 9 | | CH(CH ₃) ₂ OMe | $(144a)^{a}$ | 10.40 | 115 | 1.90×10^{-3} | 4.10×10^{-4} | 1.02×10^{7} | -0.34 | |
| 10 | (CO)5Cr=C | CH ₂ Ph OMe | (144b) ^{<i>a</i>} | 10.18 | 140 | 1.35×10^{-2} | | | -0.35 | ≈ <i>1.67</i> |
| 11 | $(CO)_5 W = C$ | CH ₂ Ph SMe | (139a) ^b | 9.05 | 1.20×10^{-3} | 8.30×10^{-4} | 1.61×10^{-2} | 1.73×10^{7} | 0.01 | ≈2.72 |
| 12 | $(CO)_5Cr = C$ | CH ₃ SMe | $(139b)^{b}$ | 8.37 | 8.37×10^{2} | 1.21×10^{-4} | 5.23×10^{-2} | 1.17×10^{7} | -0.49 | ≈2.90 |
| 12 | $(CO)_5W = C$ | CH ₃ | (13 6) ^c | 4 19 | 1.61×10^4 | 2.07×10^{-8} | 1.85×10^{-2} | 2.80×10^{2} | ~ - 1.29 | 0.36 |
| 15 | C ₅ H ₅ (NO)(PPh | A ₃)Re | (130) | 4.10 | 1.01 × 10 | 5.07 × 10 | 1.05 × 10 | 2.00 × 10 | ~ -1.28 | 0.50 |

| Table 24 (| 'able 24 (continued) | | | | | | | |
|------------------------|---|-------------|--|------------------------------------|---------------------------------|--|---------------------|----------------------------|
| Entry | | pK_a^{CH} | $k_1^{\text{OH}} (\text{M}^{-1} \text{ s}^{-1})$ | $k_{-1}^{\rm H_2O}~({\rm s}^{-1})$ | $k_1^{\rm H_2O}~({\rm s}^{-1})$ | $k_{-1}^{\rm H} ({ m M}^{-1} { m s}^{-1})$ | $\log k_0^{\rm OH}$ | $\log k_0^{\mathrm{H_2O}}$ |
| 14 C | $_{5}H_{5}(NO)(PPh_{3})Re$ (137) ^c | -0.03 | $\approx 10^5$ | $\approx 6 \times 10^{-4}$ | 92.5 | 78.3 | ≈ - 2.61 | ≈1.95 |
| ^a Reference | e 170. | | | | | | | |

^bReference 176. ^cReference 177.

| Entry | Carbene complex | | pK ^{CH} _a | $\log k_0^{\mathrm{B}}(\boldsymbol{\beta})$ | | |
|-------|--------------------------------|------------------------------------|-------------------------------|---|-------------------|-------------------|
| | | | | RNH ₂ | R ₂ NH | RCOO ⁻ |
| 1 | $(CO)_5Cr = \bigcirc \bigcirc$ | (8) ^{<i>a</i>} | 14.47 | | 3.51 (0.62) | |
| 2 | $(CO)_5 M = C$ | (12) ^{<i>a</i>} | 12.81 | 2.95 (0.59) | 3.73 (0.68) | |
| 3 | $(CO)_5Cr = C'_{CU}$ | (68a) ^{<i>a</i>} | 12.50 | 3.04 (0.61) | 3.70 (0.62) | |
| 4 | $(CO)_5W = C$ | (40) ^{<i>a</i>} | 12.36 | 2.73 (0.64) | 3.18 (0.49) | |
| 5 | $(CO)_5 Cr = C $ | (145) ^{<i>a</i>} | 12.32 | | 2.34 (0.62) | |
| 6 | $(CO)_5Cr = C$ | (146) ^{<i>a</i>} | 12.27 | | -0.11 (0.62) | |
| | $CH(CH_3)_2$ | | | | | |

Table 25 Summary of intrinsic rate constants and Brønsted β values for the proton transfer from carbene complexes to primary aliphatic amines (RNH₂), secondary alicyclic amines (R₂NH) and carboxylate ions in 50% MeCN-50% water (v/v) at 25°C

| Table 25 (c | Fable 25 (continued) | | | | | | | | |
|-------------|----------------------------------|------------------------------------|-------------|---|-------------------|-------------|--|--|--|
| Entry | Carbene complex | | pK_a^{CH} | $\log k_0^{\mathrm{B}}(\boldsymbol{\beta})$ | | | | | |
| | | | | RNH ₂ | R ₂ NH | $RCOO^{-}$ | | | |
| 7 | $(CO)_5Cr = C$ | $(144a)^{a}$ | 10.40 | 1.51 (0.54) | 1.86 (0.48) | 0.79 (0.33) | | | |
| 8 | $(CO)_5 W = C'$ | $(\mathbf{144b})^a$ | 10.18 | 1.55 (0.51) | 1.93 (0.47) | | | | |
| 9 | $(CO)_5Cr = C$ | (139a) ^b | 9.05 | 2.09 (0.48) | 2.61 (0.45) | 2.17 (0.44) | | | |
| 10 | $(CO)_5W = C$ | (139b) ^b | 8.37 | 2.17 (0.47) | 2.50 (0.49) | 2.27 (0.43) | | | |
| 11 | $C_{5H_{5}}^{+}(NO)(PPh_{3})Re $ | (136) ^{<i>c</i>} | 4.18 | 0.14 (0.53) | 0.93 (0.49) | 0.72 (0.28) | | | |
| 12 | $C_{5}H_{5}(NO)(PPh_{3})Re$ | (137) ^c | -0.03 | 0.86 (0.33) | | 2.85 (0.12) | | | |

^{*a*}Reference 170. ^{*b*}Reference 176. ^{*c*}Reference 177.

The powerful electron withdrawing effect of the (CO)₅M moieties is best appreciated by comparing these pK_a^{CH} values with those of corresponding esters. For example, (CO)₅Cr=C(OEt)CH₃ is 13.5 pK_a units more acidic than CH₃CO₂Et ($pK_a^{CH} = 26.5$)¹⁷⁸ while (CO)₅Cr=C(SMe)CH₃ is 12 pK_a units more acidic than CH₃COSEt ($pK_a^{CH} = 21.0$);¹⁷⁹ the fact that the pK_a^{CH} values for the carbene complexes refer to 50% MeCN-50% water as the solvent is inconsequential since these pK_a^{CH} values are nearly the same as in water.¹⁷¹

The strong electron withdrawing effect of the (CO)₅M groups is mainly the result of the stabilization of the conjugate anion by dispersion of the negative charge into the CO ligands, the same effect that enhances the equilibrium constants for nucleophilic addition to the carbene carbon discussed in this chapter. Regarding the variation of the pK_a^{CH} values with structure the following points are noteworthy.

(1) The factor that affects the pK_a^{CH} values the most is the nature of the π -donor attached to the carbone carbon. For example, replacing the methoxy group in **68** or **40** with a thiomethyl group (**139**) (entry 5 versus 11 and 6 versus 12) lowers the pK_a^{CH} by 3.5–4.0 units.



139a M = Cr**139b** M = W

This increase in acidity is mainly the result of the weaker π -donor effect of the MeS ($\sigma_{\rm R} = -0.15$)¹⁴⁹ compared to that of the MeO group ($\sigma_{\rm R} = -0.43$)¹⁴⁹ which leads to a weaker stabilization of the neutral thia compared to the oxa carbene complexes. The higher $pK_a^{\rm CH}$ values of entries 1–3 compared to that of 5 can be attributed to π -donation by the respective oxygens in entries 1–3 that is stronger than in 5. In the case of entry 3, the ethyl group is a better stabilizer of the oxyanion resonance structure of the carbene complex (140). For the cyclic carbene complexes (entries 1 and 2), the π -donor effect is enhanced further because, by virtue of the cyclic structures of entries 1 and 2, the oxygen is locked into a position for better π -overlap with carbene carbon (142). ⁵³Cr NMR data are in agreement with this assessment.⁶



The notion that the π -donor effect is a dominant factor is also supported by the fact that **143a** and **143b** are not acidic enough to be deprotonated in 50% MeCN–

50% water although its acidity was determined in pure acetonitrile where its pK_a^{CH} is about 10 units higher than that of entry 5¹⁸⁰ (more on this under "Thermodynamic acidities in organic solvents").

(2) Substituting one of the α hydrogens by phenyl groups (144) increases the acidity by more than 2 p K_a -units (entry 9 versus 5 and 10 versus 6). This acidifying effect is mainly the result of additional resonance stabilization of the anion provided by the phenyl group.



(3) Substituting one or two α hydrogens by methyl groups (**145** or **146**) has an acidifying influence (entries 7 and 8 versus 5) that becomes more apparent when statistical corrections for the varying number of acidic protons are applied. These corrected pK_a^{CH} values are 12.98 for entry 5, 12.62 for 7 and 12.27 for 8. The acidity enhancement may be attributed to an increase in stability of the respective anions (entry 8 > entry 7 > entry 5) caused by the methyl groups. Assuming that the dominant resonance form of the anions has the negative charge mainly delocalized into the (CO)₅Cr moiety (**147**) the increased anion stability in the order 8 > 7 > 5 simply reflects the well-known stabilization of alkenes by methyl groups.



A similar situation exists when the pK_a^{CH} values of entry 1 (14.47) and entry 2 (13.41) are compared: here the acidifying effect of the methyl group is even more dramatic than in the comparison of entries 7 and 8 with 5, perhaps because π overlap in the anion is enhanced due to the cyclic nature of the carbene complex, which results in a more fully developed C=C double bond.

(4) The pK_a^{CH} values are not very sensitive to the metal. This is seen by a comparison of entries 4–6, of 9 with 10, and of 11 with 12. These results are reminiscent of the observations made in comparing equilibrium constants for the addition of nucleophiles to the carbene carbon (see, e.g., Tables 8, 9, 14, 17 and 19); they imply that the stabilization of the respective anions by the (CO)₅M moiety depends little on the metal. The simplest explanation for this state of affairs is that it is mainly the CO ligands rather than the metal that are responsible for the stabilization of the anionic charge. It is also possible that we are dealing with a compensating effect between the influence of the metal on the stability of the anion

and that of the carbene complex. Thus, the $(CO)_5W$ moiety may be somewhat more effective in stabilizing negative charge of the anion than the $(CO)_5Cr$ moiety but there will also be greater stabilization of the negative charge on the resonance structure **148** (M = W) of the carbene complex itself.



Because of this compensation effect the difference in the pK_a^{CH} values of entries 5 and 6 remains small. The fact that the pK_a^{CH} difference between entries 11 and 12 $(\Delta pK_a^{CH} = 0.68)$ is larger than that between entries 5 and 6 $(\Delta pK_a^{CH} = 0.14)$ supports this explanation: the resonance structure **149** of entries 11 and 12 is of lesser importance than **148** and hence there is less negative charge on the (CO)₅M moieties of entries 11 and 12 than those of 5 and 6. As a result there is less compensation and less reduction in ΔpK_a^{CH} between the chromium and tungsten carbene complex. The same type of compensating effects may be involved in explaining the small metal dependence of the equilibrium constants for nucleophilic addition to the carbene carbon (Tables 8, 9, 14, 17 and 19).

Rhenium carbene complexes

The two rhenium carbene complexes are much more acidic than the $(CO)_5M$ -type carbene complexes. There are several reasons why they are so acidic. Probably the most important one is that they are cationic which makes the $C_5H_5(NO)(PPh_3)Re$ moiety a stronger π -acceptor than the $(CO)_5M$ moieties, despite the fact that in the latter there are five electron withdrawing CO ligands on the metal while there is only one electron withdrawing NO ligand in the rhenium complexes. Or, in other words, the dominant resonance structures of the conjugate bases of the rhenium complexes, **138** and **150**, respectively, are neutral molecules and hence more stable than the anionic conjugate bases of the $(CO)_5M$ type complexes.



An additional driving force for the deprotonation of the Re complexes is that their conjugate bases are aromatic, i.e., **138** is a selenophene, **150** a benzothiopene derivative. The fact that entry 14 ($pK_a^{CH} = -0.3$) is substantially more acidic than entry 13 ($pK_a^{CH} = 4.18$) is probably in large measure due to the stronger aromaticity of the benzothiophene moiety compared to the selenophene moiety.¹⁸³

| Carbon acid | Solvent | pK _a ^{CH} | $\log k_0^{\mathrm{B}}$ | | |
|--|------------------|-------------------------------|-------------------------|-------------------|-------------------|
| | | | RNH ₂ | R ₂ NH | RCOO ⁻ |
| HCN | H ₂ O | 9.0 | ~8.60 | | ~9.1 |
| $CH_2(CN)_2$ | H ₂ O | 11.2 | | \sim 7.0 | |
| 9-Cyanofluorene | 50% DMSO | 9.53 | 3.76 | 4.58 | |
| Meldrum's acid | 50% DMSO | 4.70 | | 3.90 | |
| 4-NO ₂ C ₆ H ₄ CH ₂ CN | 50% DMSO | 12.62 | | 3.70 | |
| 1,3-Indandione | 50% DMSO | 6.35 | 2.44 | 3.13 | 3.18 |
| $CH_2(COCH_3)_2$ | 50% DMSO | 9.12 | 2.06 | 2.75 | |
| 2,4-(NO ₂) ₂ C ₆ H ₃ CH ₂ CN | 50% DMSO | 8.06 | | 2.75 | |
| CH ₃ NO ₂ | 50% DMSO | 11.32 | | 0.73 | |
| PhCH ₂ NO ₂ | 50% DMSO | 7.93 | | -0.25 | -0.59 |
| CH ₃ NO ₂ | H_2O | 10.28 | | -0.59 | |
| PhCH ₂ NO ₂ | H ₂ O | 6.88 | -1.16 | -0.86 | |

Table 26 Intrinsic rate constants for the deprotonation of representative carbon acids in water at 25° C or 50% DMSO-50% water (v/v) at 20° C

Taken from Ref. 122.

INTRINSIC RATE CONSTANTS FOR PROTON TRANSFER

Comparisons with other carbon acids

As discussed in the section "Alcohols and alkoxide ion nucleophiles," the most useful measure of reactivity of a reaction system is its intrinsic barrier or intrinsic rate constant because, in comparing different systems, these quantities correct for potential differences in the thermodynamics of the reactions. A comparison of intrinsic rate constants for the deprotonation of Fischer carbene complexes by buffer bases with those for the deprotonation of other carbon acids is revealing. A representative list of such $k_0^{\rm B}$ values is given in Table 26. They show the well-known trend towards lower $k_0^{\rm B}$ values with increasing charge delocalization/resonance stabilization in the respective carbanions, a trend that results from the lag in the delocalization behind proton transfer at the transition state.

Most log $k_0^{\rm B}$ values for the carbene complexes are between *ca*. 2.0 and 3.7 (Table 25) which places them into the midrange of the log $k_0^{\rm B}$ values for the organic carbon acids listed in Table 26 but definitely above the intrinsic rate constants for the nitroalkanes. An interesting question is whether these findings allow the inference that the π -acceptor strength of the (CO)₅M groups is comparable to that of 1,3-diones, 4-NO₂C₆H₄CH₂CN or 2,4-(NO₂)₂C₆H₃CH₂CN but not quite at the level of nitro groups.

As already mentioned in the above referred sections, the k_0 -reducing effect of resonance stabilization of the carbanion resulting from the deprotonation of organic carbon acids is the consequence of transition state imbalances. These imbalances manifest themselves in the Brønsted α value being greater than the β value, with the

| Amine type | eta^a | Amine | $lpha^b$ |
|-------------------|----------------------------------|-----------------------------|-----------------|
| RNH ₂ | $0.54 \pm 0.04 \\ 0.48 \pm 0.07$ | <i>n</i> -BuNH ₂ | 0.56 ± 0.03 |
| R ₂ NH | | Piperidine | 0.53 ± 0.03 |

Table 27 Brønsted α and β values for the deprotonation of substituted benzyl carbene complexes (equation 102) in 50% MeCN-50% water at 25°C

Reference 184.

 $^{a}\beta = d \log k_{1}^{B}/d p K_{a}^{BH} (Z = H).$

 ${}^{b}\alpha = d\log k_{1}^{\mathrm{B}}/d\log K_{\mathrm{a}}^{\mathrm{CH}}.$

difference between α and β correlating with the strength of the π -acceptor. Hence, if the same rules apply to Fischer carbene complexes, the transition state for their proton transfer reactions should also be imbalanced. However, a recent study of the reaction of substituted benzyl carbene complexes (equation 102) yielded α and β values that, within their experimental uncertainty, are equal to each other (Table 27).¹⁸⁴

$$(CO)_{5}Cr = C \bigvee_{\substack{CH_{2} \\ CH_{2} \\$$

This result raised the question whether there is a fundamental difference between the $(CO)_5Cr$ moiety and purely organic π -acceptors in the way they affect intrinsic barriers, i.e., is it conceivable that the $(CO)_5Cr$ moiety would raise intrinsic barriers without a transition state imbalance? This seems unlikely.

A more reasonable hypothesis is that the transition state *is* imbalanced, as shown in equation (102), but that there is a structural feature characteristic of carbene complexes, absent from other carbon acids, that masks the imbalance by reducing α . The most likely candidate is the π -donor effect of the methoxy group. Inasmuch as the contribution of **151** leads to resonance stabilization of the carbene complexes, this resonance is expected to add to the intrinsic barrier of proton transfer. This is because, as is true for resonance effects in general, its loss at the transition state should be ahead of the proton transfer.¹²¹⁻¹²³ As Z becomes more electron withdrawing, the greater electron deficiency of the carbene complex induces a stronger π -donor effect by the methoxy group. There are two consequences. One is that the increased contribution of the resonance structure **151** partially compensates for the destabilization of the electron deficient carbon by the electron withdrawing inductive effect of Z. The second is that the increased resonance stabilization of the carbene complex increases the intrinsic barrier and hence the rate $(k_1^{\rm B})$ enhancement caused by the inductive effect of Z will be attenuated. This attenuation is proportional to the electron withdrawing strength of Z, as shown schematically in Fig. 8, and hence the slope of the Brønsted plot (α_{CH}) is reduced.



Fig. 8 Schematic representation of how the Brønsted slope (α_{CH}) is lowered as a result of the π -donor effect of the MeO group. (Figure taken from Ref. 184, by permission from the American Chemical Society.)



The notion that the π -donor effect of the methoxy group is responsible for the reduction in α is supported by data on the deprotonation of **152** by amines which indicate a much smaller difference between α and β than in the reaction of substituted phenylnitromethanes.¹⁸⁵ Again, the increased resonance stabilization by the π -donor effect of the MeO group (**153**) that results when Z is made more electron withdrawing is believed to be responsible for the reduced α value.

Dependence of k_0 on carbene complex structure

For the (CO)₅M-type carbene complexes, the intrinsic rate constants depend very little on the metal (Table 25, see entries 4–6), suggesting that the π -acceptor strength of the (CO)₅M moiety is insensitive to the nature of the metal. This is true for the k_0^{B} (Table 25) as well as the k_0^{OH} values (Table 24); it is consistent with the fact that the pK_a^{CH} values also depend very little on M, as pointed out earlier.

The effect of changing the π -donor strength of alkoxy groups is also minimal, as is apparent by comparing entries 1, 3 and 5. On the other hand, the change from a MeO to a MeS group leads to a substantial lowering of k_0 ($\Delta \log k_0^{OH} = -1.30$, $\Delta \log k_0^{RNH_2} = -0.95$ and $\Delta \log k_0^{R_2NH} = -1.09$ for M = Cr; $\Delta \log k_0^{OH} = -1.53$, $\Delta \log k_0^{RNH_2} = -0.56$ and $\Delta \log k_0^{R_2NH} = -0.68$ for M = W). There are probably several factors that contribute to the decrease in the k_0 values. One such factor is the result of the imbalanced nature of the transition state which renders the intrinsic rate constants sensitive to the difference in the inductive/field effect of the MeO and MeS groups. These groups being closer to the negative charge at the transition state than in the anion (equation 103), their electron withdrawing effect should result in a transition state stabilization that is disproportionately strong relative to their stabilization of the anion. This will increase the intrinsic rate constant in both cases but more so for the MeO complex because of the stronger inductive effect of the MeO group compared to the MeS group.

$$\underbrace{\overset{\delta-}{(CO)_5M}}_{CH_3} \overset{\delta+}{=} B^{\nu} \underbrace{\overset{\delta-}{=}}_{CH_3} \underbrace{\overset{\delta-}{(CO)_5M}}_{H_2C\cdots H\cdots B^{\nu+\delta}} \overset{\delta+}{=} \underbrace{\overset{\bullet-}{(CO)_5M}}_{CH_2} \underbrace{\overset{XMe}{=}}_{CH_2} \underbrace{\overset{XMe}{=}}_{CH_2}$$
(103)

A second factor is the larger size of the MeS group which probably reduces k_0 for the MeS relative to the MeO complexes by a steric effect. As discussed in some detail elsewhere,¹⁷⁶ there is some independent evidence based on the relatively small differences between $\log k_0^{R_2NH}$ and $\log k_0^{RNH_2}$ for the reactions of entries 11 and 12 that steric effects indeed play a role in the reactions of these carbene complexes.

A third factor is electrostatic interaction between the base and the partial positive charge on the MeX group that arises from its π -donor effect. With amine bases ($\nu = 0$) there is destabilization of the transition state by electrostatic repulsion. Since the π -donor effect of MeO is stronger than that of MeS and hence $\delta +$ on MeX is larger with X = O than X = S, the transition state destabilization will be greater for the MeO complexes. This will reduce $|\Delta \log k_0^{\text{RNH}_2}|$ and $|\Delta \log k_0^{\text{R}_2\text{NH}}|$. With OH⁻ as the base ($\nu = -1$) there is stabilization of the transition state that is more effective for the MeO complexes. This will enhance $|\Delta \log k_0^{\text{OH}}|$. The significantly more negative $\Delta \log k_0^{\text{OH}}$ values compared to the $\Delta \log k_0^{\text{RMH}_2}$ and $\Delta \log k_0^{\text{R}_2\text{NH}}$ values are consistent with the above analysis. Interestingly, these electrostatic effects are strong enough to override the large steric effect expected for the amine reactions, especially those with the secondary amines, compared to that on the OH⁻ reactions.

Another factor that has a strong effect on the intrinsic rate constants is the substitution of a hydrogen on the α -carbon by a phenyl or methyl group(s). The decrease in k_0 that results from a phenyl group ($\Delta \log k_0^{OH} = -1.65$, $\Delta \log k_0^{RNH_2} = -1.53$, and $\Delta \log k_0^{R_2NH} = -1.84$ for M = Cr; $\Delta \log k_0^{OH} = -1.39$, $\Delta \log k_0^{RNH_2} = -1.18$, and $\Delta \log k_0^{R_2NH} = -1.25$) can be mainly attributed to the

resonance effect of the phenyl group; this resonance effect is in addition to the resonance effect exerted by the (CO)₅M group and thus further reduces the k_0 values. The fact that the $\Delta \log k_0^{OH}$ and $\Delta \log k_0^{R_2NH}$ values are comparable suggests that steric effects play a minor role if any in the reduction of the intrinsic rate constants.

Methyl groups also lower the intrinsic rate constants significantly $(\log k_0^{OH} = 1.04, -0.24 \text{ and } -0.99 \text{ for entries 5 versus 7 versus 8, respectively;} log <math>k_0^{OH} = 1.51$ and 0.70 for entries 1 and 2, respectively; $\log k_0^{R_2NH} = 3.70, 2.34$ and -0.11 for entries 5, 7, and 8, respectively). These results would be difficult to explain without invoking a transition state imbalance and hence they support our earlier conclusion that the equality between α and β in reaction (102) is not to be taken as evidence against an imbalance. The transition state imbalance contributes in two ways to the lowering of k_0 by the presence of α -methyl groups. One is that C=C double bond formation in the anion lags behind proton transfer and hence the C=C double bond stabilizing effect of methyl groups is barely developed at the transition state. The other leads to a *destabilization* of the transition state by the electron donating effect of the alkyl groups on the partial negative charge of the α -carbon (e.g., **154**).



Another way of looking at the results for the entries 5, 7 and 8 is to compare the effect of adding the methyl groups on the pK_a^{CH} with that on the rate constants for proton transfer. The statistically corrected pK_a^{CH} values are 12.98, 12.62 and 12.27 for entries 5, 7 and 8, respectively, while the respective statistically corrected k_1^{OH} for deprotonation of these carbene complexes by OH⁻ are 152, 23.4 and 10.8 M⁻¹ s⁻¹. The decrease in k_1^{OH} with increasing acidity translates into a Brønsted $\alpha = -2.4$, a highly unusual negative value which is reminiscent of the reactions of CH₃NO₂, CH₃CH₂NO₂ and (CH₃)₂CHNO₂ with OH⁻ where $\alpha = -0.5$,¹⁸⁶ a situation known as the "nitroalkane anomaly."

It should be noted that in the reaction of the isopropyl derivative (entry 8) with the bulky secondary alicyclic amines, steric hindrance contributes to the reduction in $k_0^{\text{R}_2\text{NH}}$, as seen in the large drop in log $k_0^{\text{R}_2\text{NH}}$ from 2.34 for the ethyl to -0.11 for the isopropyl derivative.

THERMODYNAMIC ACIDITIES IN ORGANIC SOLVENTS

A major drawback of water or aqueous acetonitrile as the medium for acidity determinations is that the range of pK_a^{CH} values accessible is relatively narrow and does not allow the study of the acidity of more weakly acidic carbene complexes

than entry 1 (p $K_a^{CH} = 14.47$ in 50% MeCN-50% water). THF, a solvent commonly used in synthetic applications, allows ionization of even the least acidic carbene complexes but ion-pairing effects are so severe in this medium that quantitative measurements are strongly affected or distorted (more on this later). DMSO, a standard solvent for acidity determinations pioneered by Bordwell,¹⁸⁷ tends to oxidize Fischer carbene complexes quite rapidly.^{87,188}

The most suitable solvent appears to be acetonitrile, which has also been used successfully for acidity measurements of hydrido transition metal complexes^{148,189} and transition metal acyl compounds.¹⁹⁰ Fischer carbene complexes are quite stable in this solvent; its polarity is high enough that ion pairing is not a significant problem and pK_a values up to about 36 can be measured.^{191,192}

Acetonitrile

Table 28 summarizes pK_a^{CH} value determined in acetonitrile¹⁹⁴ along with pK_a^{CH} values in 50% MeCN–50% water for the complexes where these are available (values in parentheses). They were obtained spectrophotometrically from measurements of the equilibrium constants of the reaction of the carbene complex with strong amine bases such as TMG (1,1,2,2-tetramethyguanidine), DBU (1,8-diazabicyclo[5.4.0]undec-7-ene), BEMP (2-(*tert*-butylimino)-2-(diethylamino)-1,3-dimethylperhydro-1,3-diazaphosphine and P₂-Et (1-*tert*-butyl-2,2,4,4,4-penta-kis(dimethylamino)-2 λ^5 ,4 λ^5 -catenadi(phosphazene).

The pK_a^{CH} values in pure acetonitrile are, on average, 10.2 units higher than in 50% MeCN-50% water. This difference may be expressed by equation (104)

$${}^{50}\Delta^{A}pK_{a}^{CH} = pK_{a}^{CH}(MeCN) - pK_{a}(50\% MeCN - 50\% water)$$
$$= \log {}^{50}\gamma_{C^{-}}^{A} + \log {}^{50}\gamma_{H^{+}}^{A} - \log {}^{50}\gamma_{CH}^{A}$$
(104)

where ${}^{50}\gamma_{C^-}^{A}$, ${}^{50}\gamma_{H^+}^{A}$ and ${}^{50}\gamma_{CH}^{A}$ are the solvent activity coefficients for the transfer of the anion C⁻, H⁺, and the carbene complex (CH), respectively, from 50% MeCN–50% water ("50") to pure MeCN ("A").¹⁹³ The largest contributor to ${}^{50}\Delta^{A}pK_{a}^{CH}$ must come from the log ${}^{50}\gamma_{H^+}^{A}$ term: for the transfer of H⁺ from pure water ("W") to acetonitrile, log ${}^{W}\gamma_{H^+}^{A} = 8.1$, 195,196 reflecting the much weaker solvation of H⁺ in acetonitrile. The value of log ${}^{50}\gamma_{H^+}^{A}$ is not known but it is probably close to 8.1. This leaves a contribution of perhaps 2.5–3.0 log units of ${}^{50}\Delta^{A}pK_{a}^{CH}$ to be accounted for by log ${}^{50}\gamma_{C^-}^{A} - \log {}^{50}\gamma_{CH}^{A}$ which must result from a combination of weaker solvation of C⁻ (log ${}^{50}\gamma_{C^-}^{A} > 0$) and a stronger solvation of CH (log ${}^{50}\gamma_{CH}^{A} < 0$) in pure acetonitrile.

The Me₂N derivatives (**143a** and **143b**) are seen to be the least acidic carbene complexes with pK_a^{CH} values (31.9 and 31.7, respectively) almost 10 units higher than for the methoxy analog **6** ($pK_a^{CH} = 22.2$). The reduction in acidity reflects the strong π -donor effect at the Me₂N group, which stabilizes the carbene complex; the magnitude of the effect emphasizes the great importance of the resonance form **143'** in determining the electronic structure of Fischer carbene complexes.

| Carbene complex | | pK_a^{CH} | Carbene complex | | pK_a^{CH} |
|--|--------|--------------------------|---|-------|------------------------|
| $(CO)_5Cr = C'^{NMe_2}_{CH_3}$ | (143a) | 31.9 ^{<i>a</i>} | $(CO)_4Cr = C$ Ph_2P CH_3 CH_3 | (158) | 25.4 ^b |
| $(CO)_5W = C_{CH_2}^{NMe_2}$ | (143b) | 31.7 ^{<i>a</i>} | $(CO)_5Cr$ | (8) | $24.4^{b} (14.47)^{c}$ |
| $(CO)_5Cr = C'$ | (155) | 30.8 | $(CO)_5Cr = C$ CH ₃ | (66) | $25.2^{b} (12.98)^{c}$ |
| $(CO)_5Cr = C$ | (156) | 27.6 ^{<i>b</i>} | $(CO)_5Cr = C'OMe$ CH_2CH_3 | (145) | $22.9^{b} (12.36)^{c}$ |
| CH_{3} $(CO)_{4}Cr = C CH_{3}$ $(CO)_{4}Cr = C CH_{3}$ | (157) | 26.5 ^{<i>b</i>} | $(CO)_5Cr = C'OMe$ CH ₃ | (68) | $22.2^{b} (12.50)^{c}$ |

Table 28 Summary of pK_a^{CH} values of Fischer carbene complexes in acetonitrile at 25°C

^{*a*}Reference 180.

^bReference 194.

^cIn 50% MeCN-50% water (Table 24).



Replacing the methyl groups on the nitrogen by a Me₂N and a PhCH₂ group increases the acidity only by about one pK_a unit (**155**), suggesting that the π -donor effect of the N(Me₂N)CH₂Ph group is only slightly weaker than that of the Me₂N group. On the other hand, converting the Me₂N group into an aziridinyl group (**156**) leads to a 4.3 unit drop in pK_a^{CH} because the ring strain counteracts sp² hybridization of the nitrogen and hence reduces its π -donor effect.



Substitution of one CO ligands by a phosphine (**157** and **158**) increases the pK_a^{CH} relative to that of (CO)₅Cr=C(OMe)CH₃ (**68**), reflecting the reduced electron withdrawing effect of the (CO)₄(R₃P)Cr moiety relative to the (CO)₅Cr moiety.



Tetrahydrofuran and dimethyl sulfoxide

Despite the drawbacks mentioned above, a small number of acidity measurements have been reported in THF^{167,197–199} and in DMSO.²⁹ A major problem with THF is that there are no free ions in this solvent which makes it difficult to define a pK_a^{CH} that would be directly comparable to pK_a^{CH} values in polar solvents. Nevertheless, some of these acidity data have recently been summarized in terms of pK_a^{CH} values;²⁹ they are reported as " pK_a^{CH} " in Table 29. The " pK_a^{CH} " values for **68**, **8** and **141** in THF are based on equilibrium measurements of the reaction of the respective carbene complexes with the PPN (see footnote *b* in Table 29) salt of *p*-cyanophenoxide ion;^{167,197,198} " pK_a^{CH} " = 8 was deduced from the fact that the

| Carbene complex | | Solvent | Base | pK_a^{CH} |
|---|--------|---------|---|------------------------|
| $(CO)_5 Cr = C \begin{pmatrix} OMe \\ CH_3 \end{pmatrix}$ | (68) | THF | Li ⁺ MeO ⁻ | 12 ^{<i>a</i>} |
| $(CO)_5Cr$ | (8) | THF | $\frac{\text{PPN}^{+b}\text{4-CNC}_{6}\text{H}_{4}\text{O}^{-}}{\text{PPN}^{+b}\text{4-CNC}_{6}\text{H}_{4}\text{O}^{-}}$ | 8^c 8^d |
| (CO) ₅ Cr | (141) | THF | PPN ^{+b} 4-CNC ₆ H ₄ O ⁻ | 8 ^{<i>d</i>} |
| $(CO)_4Cr = C$ $(CO)_4Cr = C$ (CH_3) | (157) | THF | Li ⁺ t-BuO ⁻ | 18.8 ^e |
| $(CO)_5 Cr = C CH_3^{NMe_2}$ | (143a) | DMSO | K ⁺ 2-naphthylacetonitrile anion | 20.4 ^f |

Table 29 Summary of pK_a^{CH} values of Fischer carbene complexes in THF and DMSO

^aReference 197.

 b PPN⁺ = bis(triphenylphospine)iminium ion.

^cReference 167.

^dReference 198.

^eReference 199.

^fReference 29.

equilibrium constants were about 1.0 for all three complexes while the aqueous pK_a of *p*-cyanophenol is 8.0. The " pK_a^{CH} " value of 12 for **68**¹⁹⁷ was calculated on the basis of the equilibrium constant of the reaction of **68** with lithium methoxide and the pK_a of methanol. The " pK_a^{CH} " value for **157** was derived from measurements with lithium *tert*-butoxide and the known pK_a of *tert*-butanol.¹⁹⁹

The problem with the " pK_a^{CH} " values in THF is that their absolute values have no well defined physical meaning. This is not only because of ion pairing effects but also because they are not based on pK_a values of the proton acceptors in THF but in solvents such as water or methanol. Even the *relative* " pK_a^{CH} " values suffer from gross distortions which render their meaning questionable. For example, the " pK_a^{CH} " value obtained for **68** strongly depends on the base system used for the deprotonation, i.e., PPN⁺/4-CNC₆H₄O⁻ versus Li⁺/MeO⁻. This must be the result of large differences in the tightness or stability of reactant and/or product ion pairs. Or, the " pK_a^{CH} " values of **68**, **8** and **141** are all about the same in THF which

contrasts with the results in acetonitrile (Table 28) where the pK_a^{CH} of **8** is 2.2 units higher than that of **68** and the pK_a^{CH} of **141** is 1.5 units higher than that of **68**.

With respect to the " pK_a^{CH} " of **143a** in DMSO (20.4), it is 11.5 units lower than in acetonitrile. This is a reasonable result. Carbon acids such as 4-NO₂C₆H₄CH₂NO₂ and 3,5-(NO₂)₂C₆H₃CH₂NO₂ have ^D $\Delta^A pK_a^{CH}$ values of 11.7, 12.0 and 11.2, respectively.²⁰⁰ One may express ^D $\Delta^A pK_a^{CH}$ by equation (105) which is analogous to equation (104).

$${}^{\mathrm{D}}\Delta^{\mathrm{A}}pK_{\mathrm{a}}^{\mathrm{CH}} = pK_{\mathrm{a}}^{\mathrm{CH}}(\mathrm{MeCN}) - pK_{\mathrm{a}}^{\mathrm{CH}}(\mathrm{DMSO})$$
$$= \log {}^{\mathrm{D}}\gamma_{\mathrm{C}^{-}}^{\mathrm{A}} + \log {}^{\mathrm{D}}\gamma_{\mathrm{H}^{+}}^{\mathrm{A}} - \log {}^{\mathrm{D}}\gamma_{\mathrm{CH}}^{\mathrm{A}}$$
(105)

Again, as is the case for the comparison between pure acetonitrile and aqueous acetonitrile, the dominant contribution to the solvent effect on the pK_a^{CH} value comes from the much weaker solvation of the proton in acetonitrile, i.e., $\log^{D} \gamma_{H^+}^{A} = 11.4$.^{195,196}

HYDROLYSIS OF IONIZABLE CARBENE COMPLEXES

Thermal reactions

The hydrolysis of Fischer carbene complexes such as **11**, **13**, **42**, **87** and **112** has been discussed in the section "Hydroxide ion and water as nucleophiles" and proceeds by a standard nucleophilic addition–elimination mechanism (equation 74). On the other hand, carbene complexes with ionizable α -carbons are hydrolyzed by a different mechanism which involves the deprotonated carbene complex as the key intermediate. This conclusion is based on a detailed kinetic investigation of the hydrolysis of **66**, **68**, **144** and **8** in 50% MeCN–50% water.^{201–204}



The organic products of the hydrolysis reactions were: acetaldehyde and methanol for **68**,²⁰¹ acetaldehyde and ethanol for **66**,²⁰¹ β -methoxystyrene for **144**,²⁰² and 2-hydroxytetrahydrofuran (**159**) in equilibrium with small amounts of 4-hydroxybutanal (**160**) for **8**.²⁰³





Fig. 9 Rate–pH profile of the hydrolysis of $(CO)_5M=C(OMe)CH_3$ (**68**) in 50% MeCN–50% water (v/v) at 25°C. (Figure taken from Ref. 201, by permission from the American Chemical Society.)

Figure 9 shows a typical rate-pH profile; the points below pH 8.5 represent a "water reaction," those above 8.5 an OH^- catalyzed reaction while the leveling off at pH > 12 is due to the ionization of the carbene complex. The hydrolysis is subject to general base catalysis; the points on the pH-rate profile between pH 4.8 and 11.3 were obtained by extrapolation of buffer plots to zero buffer concentration.

The pH dependence of the hydrolysis of all compounds studied is, in principle, consistent with the mechanism of Scheme 6 that applies to the alkoxyphenylcarbene complexes, and so are the products of the reactions of 68, 66 and 8. However, the products obtained in the hydrolysis of 144 and the fact that in basic solution the hydrolysis of all the compounds is subject to a substantial kinetic solvent isotope effect are inconsistent with Scheme 6, at least at pH > 8.5. The mechanism that accounts best for all experimental observations at pH > 8.5, including the isotope effect, is shown in Scheme 17 for the example of 68. It involves rapid deprotonation of **68** followed either by slow protonation of **135** with water $(k_2^{H_2O})$ or a buffer acid $(k_2^{BH}[BH])$ and subsequent rapid conversion of 161 to 162, or slow concerted water (k_{2c}) or buffer acid catalyzed $(k_{2c}^{BH}[BH])$ conversion of 135 to 162 (more on these two alternatives below). Complexation between (CO)₅Cr and the enol ether activates the latter toward basic hydrolysis which rapidly leads to the vinyl alcohol and tautomerization to the aldehyde. Control experiments demonstrated that the kind of complexation indicated by 162 indeed promotes rapid hydrolysis of the enol ether.²⁰¹ In the reactions of **144** complexation of the enol ether (β -methoxystyrene) appears to be weak, presumably because of steric crowding, and hence the reaction



 $(CO)_5CrOH^- + CH_3CH = O + CH_3OH$

Scheme 17

essentially stops at the β -methoxystyrene stage although small amounts of PhCH₂CH=O could be detected with **144b**. Incidentally, in the absence of water, e.g., when **68** or **8** is reacted with pyridine or other tertiary amines, hydrolysis of **162** is not possible and the reaction always stops at the enol ether stage. This process has been exploited for the synthesis of complex enol ethers from alkoxycarbene complexes.²³

Regarding the conversion of **135** into **162**, there are two possible pathways, stepwise or concerted (Scheme 17). The stepwise pathway entails rate-limiting protonation on the metal followed by rapid reductive elimination while the concerted pathway involves protonation on the carbene carbon which is simultaneous with bond cleavage between the metal and the carbene carbon. For the reactions with tertiary amines, e.g., pyridine, in the absence of water, the stepwise analog of Scheme 17, i.e., equation (106), has been advocated²³ although no experimental evidence has been cited to exclude a concerted pathway.



For the reactions in aqueous acetonitrile the concerted pathway is more plausible based on the following reasoning.

The rate constant for the conversion of **135** to **162** is 0.16 s^{-1} .²⁰¹ If the stepwise mechanism applies, this means $k_2^{\text{H}_2\text{O}} = 0.16 \text{ s}^{-1}$. If the concerted mechanism prevails, we have $k_{2c}^{\text{H}_2\text{O}} = 0.16 \text{ s}^{-1}$. If we assume that the pK_a^{MH} (metal proton) of **161** is comparable to that of **163**, i.e., $pK_a^{\text{MH}} \approx 0.7$,¹⁴⁷ one can estimate a k_{-2}^{OH} value based on equation (107)

$$k_{-2}^{\rm OH} = k_2^{\rm H_2O} \frac{K_{\rm a}^{\rm MH}}{K_{\rm w}}$$
(107)

where $K_{\rm w} = 6.46 \times 10^{-16} \,{\rm M}^2$ is the ionic product of water in 50% MeCN-50% water.¹⁷¹ One obtains $k_{-2}^{\rm OH} \approx 3.1 \times 10^{14} \,{\rm M}^{-1}$, a nonsensical value that is about three orders of magnitude larger than the rate constant of a diffusion controlled proton transfer. Hence the step-wise mechanism must be discarded, at least in aqueous acetonitrile. The same conclusion is reached using similar arguments for the buffer catalyzed pathway.²⁰¹

$$\begin{array}{c} H \\ I \\ (CO)_5 Cr - C - Ph \\ I \\ SCH_2 CH_2 OH \end{array}$$

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An interesting question is why the base catalyzed hydrolysis of ionizable carbene complexes proceeds by the mechanism of Scheme 17 instead of that of Scheme 11. Apparently Scheme 17 is energetically more favorable than Scheme 11. Detailed analysis indicates that for the nucleophilic mechanism to be competitive with Scheme 17 in the case of **68**, the rate constant for OH⁻ attack on **68** would have to be at least 3-fold higher than for OH⁻ attack on the phenyl analog (**11**).¹⁵⁷ This contrasts with the expectation that it is **11** which should be the more reactive carbene complex, due to the electron withdrawing inductive effect of the phenyl group. The potential π -donor effect of the phenyl group which would counteract its inductive effect is negligible because this group was found to have an orthogonal orientation both in the solid state²⁰⁵ and in solution.²⁰⁶ The notion that the phenyl group in **11** is electron withdrawing relative to the methyl group in **68** is also supported by ⁵³Cr NMR data.⁶

$$(CO)_5 Cr = C'_{Ph}^{OMe}$$

Light-induced reactions

The hydrolysis of **144a** and **144b** is catalyzed by light.²⁰² Accelerations of the order of 5- to 6-fold in the presence of high light intensities have been observed. A



Scheme 18

possible mechanism for this photochemical process is shown in Scheme 18.²⁰² This mechanism is consistent with the fact that PhCH=CHOCH₃ is the main organic product of the reaction, just as for the thermal reaction, and with a kinetic solvent isotope effect that is similar to that for the thermal process and suggests that reaction of **164** with water is rate limiting. The results did not, however, allow a distinction between the two pathways that lead to **166**. Note that the increased reactivity of **166** compared to **164** is consistent with a higher electron density induced by replacing one of the strongly electron withdrawing CO ligands by acetonitrile.

The hydrolysis of 146^{207} (isopropyl carbene) is also catalyzed by light although the effect is smaller than for 144a or 144b and the phenomenon was not studied in detail. On the other hand, no light-induced rate accelerations were observed for the hydrolysis of the cyclic carbene complexes 8^{203} and 141,²⁰⁴ or of the ethyl carbene complex 145.¹⁷⁴ It is unclear at this point what structural factors power the photochemical reaction shown in Scheme 18.

Acknowledgments

I gratefully acknowledge the tremendous contributions of all my co-workers whose names are cited in the references and who made it all possible. I also thank the National Science Foundation for its financial support (Grant CHE-0098553).

References

- 1. Dötz, K.H., Fischer, H., Hofmann, P., Kreissl, F.R., Schubert, U. and Weiss, K. (1983). *Transition Metal Carbene Complexes*. Verlag Chemie, Weinheim
- 2. Taylor, T.E. and Hall, M.B. (1984). J. Am. Chem. Soc. 106, 1576
- 3. Carter, E.A. and Goddard, W.A., III (1986). J. Am. Chem. Soc. 108, 4746
- 4. Crabtree, R.H. (1988). *The Organometallic Chemistry of Transition Metals*, Chapter 11. Wiley, New York
- 5. Fischer, E.O. and Maasböl, A. (1967). Chem. Ber. 100, 2445
- 6. Hafner, A., Hegedus, L.S., de Weck, G., Hawkins, B. and Dötz, K.H. (1988). J. Am. Chem. Soc. 110, 8413
- 7. Marynick, D.S. and Kirkpatrick, C.M. (1985). J. Am. Chem. Soc. 107, 1993
- 8. Dötz, K.H. (1984). Angew. Chem. Int. Ed. Engl. 13, 587
- 9. Gallop, M.A. and Roper, W.R. (1986). Adv. Organomet. Chem. 25, 121
- 10. Brookhart, M. and Studabaker, W.B. (1987). Chem. Rev. 87, 411
- 11. Schubert, U. (ed.) (1988). NATO ASI Series, Series C: Mathematical and Physical Sciences. Advances in Metal Carbene Chemistry. vol. 269
- Wulff, W.D. (1989). Advances in Metal-Organic Chemistry. Liebeskind, L.S. (ed.), vol. 1, p. 209–393. JAI, London
- 13. Dötz, K.H. (1990). New J. Chem. 14, 433
- Wulff, W.D. (1991). Comprehensive Organic Synthesis. Trost, R.H., Fleming, I. and Paquette, L.A. (eds), vol. 5, p. 1065–1113. Pergamon, Oxford
- 15. Reissig, H.-U. (1991). Organic Synthesis Highlights. Mulzer, J., Altenbach, H.-J., Braun, M., Krohn, K. and Reissig, H.-U. (eds), p. 168–191. VCH, Weinheim
- 16. Guerchais, V. (1994). Bull. Soc. Chim. Fr. 131, 803
- 17. Hegedus, L.S. (1995). Acc. Chem. Res. 28, 299
- Wulff, W.D. (1995). Comprehensive Organometallic Chemistry II. Abel, E.W., Stone, F.G.A., Wilkinson, G. and Hegedus, L.S. (eds), vol. 12, p. 469–547. Pergamon, New York
- 19. Harvey, D.F. and Sigano, D.M. (1996). Chem. Rev. 96, 271
- 20. de Meijere, A. (1996). Pure Appl. Chem. 68, 61
- 21. Aumann, R. and Nienaber, H. (1997). Adv. Organomet. Chem. 41, 163
- 22. Dörwald, F.Z. (1999). Metal Carbenes in Organic Synthesis. VCH, Weinheim
- 23. Hegedus, L.S. (1999). *Transition Metals in the Synthesis of Complex Organic Molecules* (2nd edn), Chapter 6. University Science Books, Sausalito, CA
- 24. Aumann, R. (2000). Eur. J. Org. Chem. 17
- 25. de Meijere, A., Schirmer, H. and Duetsch, M. (2000). Angew. Chem. Int. Ed. Engl. 39, 3964
- Casey, C.P. (1976). Transition Metal Organometallics in Organic Synthesis. Alper, H. (ed.), p. 190–223. Academic, New York
- Lattuada, L., Licandro, E., Maiorana, S. and Papagni, A. (1991). Organometallics 10, 807
- 28. Aumann, R. and Heinen, A. (1987). Chem. Ber. 120, 537
- Wulff, W.D., Anderson, B.A., Toole, A.J. and Xu, Y.-C. (1994). *Inorg. Chim. Acta* 220, 215
- 30. Xu, Y.-C. and Wulff, W.D. (1987). J. Org. Chem. 52, 3263
- 31. Fischer, E.O. and Massböl, A. (1968). J. Organomet. Chem. 12, P15
- 32. Fischer, E.O. and Plabst, W. (1974). Chem. Ber. 107, 3326
- 33. Casey, C.P. and Anderson, R.L. (1975). J. Chem. Soc., Chem. Commun. 895
- Doyle, M.P. (1995). Comprehensive Organometallic Chemistry II. Abel, E.W., Stone, F.G.A. and Wilkinson, G. (eds), vol. 12, p. 387–395. Pergamon, Oxford
- 35. Dötz, K.H. and Fischer, E.O. (1972). Chem. Ber. 105, 1356

- 36. Fischer, E.O. and Dötz, K.H. (1972). Chem. Ber. 105, 3966
- 37. Casey, C.P. and Burkhardt, T.I. (1974). J. Am. Chem. Soc. 96, 7808
- 38. Dötz, K.H. and Tomuschat, P. (1999). Chem. Soc. Rev. 28, 187
- Gut, H.-P., Welk, N., Link, U., Fischer, H. and Steiner, U.E. (2000). Organometallics 19, 2354
- Foley, H.C., Strubinger, L.M., Targos, T.S. and Geoffroy, G.L. (1983). J. Am. Chem. Soc. 105, 3064
- Collman, J.P., Hegedus, L.S., Norton, J.R. and Finke, R.G. (1987). *Principles and Applications of Organotransition Metal Chemistry*, Chapter 16. University Science Books, Mill Valley, CA
- 42. Schmidt, M.A., Miller, J.R. and Hegedus, L.S. (1991). J. Organomet. Chem. 413, 143
- 43. Hegedus, L.S. (1997). Tetrahedron 53, 4105
- 44. Casey, C.P. and Anderson, R.L. (1975). J. Chem. Soc., Chem. Commun. 895
- 45. Casey, C.P. and Cesa, M.C. (1982). Organometallics 1, 87
- 46. Werner, H. and Rascher, H. (1968). Helv. Chim. Acta 51, 1765
- 47. Fischer, H., Fischer, E.O., Kreiter, C.G. and Werner, H. (1974). Chem. Ber. 107, 2459
- 48. Fischer, H. In Ref. 1, p. 252
- 49. Fischer, H., Mühlemeier, J., Märkl, R. and Dötz, K.H. (1982). Chem. Ber. 115, 1355
- 50. Torrent, M., Duran, M. and Solà, M. (1998). Organometallics 17, 1492
- 51. Fischer, H. and Hofmann, P. (1999). Organometallics 18, 2590
- 52. Fischer, H., Motsch, A. and Kleine, W. (1978). Angew. Chem. Int. Ed. Engl. 17, 842
- 53. Fischer, H., Motsch, A., Märkl, R. and Ackermann, K. (1985). Organometallics 4, 726
- 54. Fischer, H. and Motsch, A. (1981). J. Organomet. Chem. 220, 301
- 55. Fischer, H., Fischer, E.O., Himmelreich, D., Cai, R., Schubert, U. and Ackermann, K. (1981). Chem. Ber. 114, 3220
- 56. Fischer, H., Fischer, E.O., Cai, R. and Himmelreich, D. (1983). Chem. Ber. 116, 1009
- 57. Fischer, H. (1980). J. Organomet. Chem. 195, 55
- 58. Fischer, H., Fischer, E.O. and Cai, R. (1982). Chem. Ber. 115, 2707
- 59. Fischer, E.O. and Fischer, H. (1974). Chem. Ber. 107, 657
- 60. Casey, C.P. and Shusterman, A.J. (1970). J. Mol. Catal. 8, 1
- 61. Casey, C.P., Scheck, D.M. and Shusterman, A.J. (1979). Fundam. Res. Homogenous 3, 141
- Casey, C.P., Shusterman, A.J., Vollendorf, N.W. and Haller, K.J. (1982). J. Am. Chem. Soc. 104, 2417
- 63. Dahlgren, R.M. and Zink, J.I. (1977). Inorg. Chem. 16, 3154
- 64. Edwards, B.H. and Rausch, M.D. (1981). J. Organomet. Chem. 210, 91
- Foley, H.C., Strubinger, L.M., Targos, T.S. and Geoffroy, G.L. (1983). J. Am. Chem. Soc. 105, 3064
- Bechara, J.N., Bell, S.E.J., McGarvey, J.J. and Rooney, J.J. (1986). J. Chem. Soc., Chem. Commun. 1785
- 67. Bell, S.E.J., Gordon, K.C. and McGarvey, J.J. (1988). J. Am. Chem. Soc. 110, 3107
- 68. Servaas, P.C., Stufkens, D.J. and Oskam, A. (1990). J. Organomet. Chem. 390, 61
- 69. Rooney, A.D., McGarvey, J.J. and Gordon, K.C. (1995). Organometallics 14, 107
- Rooney, A.D., McGarvey, J.J., Gordon, K.C., McNicholl, R.-A., Schubert, U. and Hepp, W. (1993). Organometallics 12, 1277
- Gut, H.-P., Welte, N., Link, U., Fischer, H. and Steiner, U.E. (2000). Organometallics 19, 2354
- 72. Klabunde, U. and Fischer, E.O. (1967). J. Am. Chem. Soc. 89, 7141
- 73. Connor, J.A. and Fischer, E.O. (1969). J. Chem. Soc. A 578
- 74. Fischer, E.O. and Kollmeier, H.-J. (1971). Chem. Ber. 104, 1339
- 75. Fischer, E.O. and Leupold, M. (1972). Chem. Ber. 105, 599
- 76. Fischer, E.O., Heckl, B. and Werner, H. (1971). J. Organomet. Chem. 28, 359

- 77. Fischer, E.O. and Aumann, R. (1968). Chem. Ber. 101, 963
- 78. Fischer, E.O. and Knauss, L. (1970). Chem. Ber. 103, 1262
- 79. Kreiter, C.G. (1968). Angew. Chem. Int. Ed. Engl. 7, 390
- 80. Fischer, E.O., Leupold, M., Kreiter, C.G. and Müller, J. (1972). Chem. Ber. 105, 150
- 81. Lam, C.T., Senoff, C.V. and Ward, J.E.H. (1974). J. Organomet. Chem. 70, 273
- 82. Aumann, R. and Schröder, J. (1990). Chem. Ber. 123, 2053
- 83. Burkhardt, T.J. and Casey, C.P. (1973). J. Am. Chem. Soc. 95, 5833
- 84. Fischer, E.O. and Riedmuller, S. (1976). Chem. Ber. 109, 3358
- 85. Fischer, E.O., Held, W. and Kreissl, F.R. (1977). Chem. Ber. 110, 3842
- Fischer, E.O., Held, W., Kreissl, F.R., Frank, A. and Haltner, G. (1977). *Chem. Ber.* 110, 656
- Casey, C.P., Burkhardt, T.J., Bunnell, C.A. and Calabrese, J.C. (1977). J. Am. Chem. Soc. 99, 2127
- Fischer, E.O., Kreis, G., Kreissl, F.R., Kreiter, C.G. and Müller, J. (1973). *Chem. Ber.* 106, 3910
- 89. Casey, C.P. and Brunsvold, W.P. (1977). Inorg. Chem. 16, 391
- 90. Bell, R.A., Chisholm, M.H., Couch, D.A. and Rankel, L.A. (1977). Inorg. Chem. 16, 677
- 91. Heckl, B., Werner, H. and Fischer, E.O. (1968). Angew. Chem. Int. Ed. Engl. 7, 817
- Werner, H., Fischer, E.O., Heckl, B. and Kreiter, C.G. (1971). J. Organomet. Chem. 28, 367
- 93. Steinmetz, A.L., Hershberger, S.A. and Angelici, R.J. (1984). Organometallics 3, 461
- 94. Pickering, R.A. and Angelici, R.J. (1981). Inorg. Chem. 20, 2977
- Kreissl, F.R., Fischer, E.O., Kreiter, C.G. and Weiss, K. (1973). Angew. Chem. Int. Ed. Engl. 12, 563
- 96. Kreissl, F.R. and Fischer, E.O. (1974). Chem. Ber. 107, 103
- 97. Bernasconi, C.F. and Stronach, M.W. (1993). J. Am. Chem. Soc. 115, 1341
- 98. Bernasconi, C.F., Whitesell, C. and Johnson, R.A. (2000). Tetrahedron 56, 4917
- 99. Satterthwait, A.C. and Jencks, W.P. (1974). J. Am. Chem. Soc. 96, 7018
- 100. Gresser, M.J. and Jencks, W.P. (1977). J. Am. Chem. Soc. 99, 6963
- 101. Yang, C.C. and Jencks, W.P. (1988). J. Am. Chem. Soc. 110, 2972
- 102. Bernasconi, C.F., Rossi, R.H. and Schmid, P. (1977). J. Am. Chem. Soc. 99, 4090
- 103. Bernasconi, C.F. (1978). Acc. Chem. Res. 11, 147
- 104. Terrier, F. (1991). Nucleophilic Aromatic Displacement, p. 55. VCH, New York
- 105. Bernasconi, C.F., Fassberg, J., Killion, R.B. and Rappoport, Z. (1990). J. Org. Chem. 55, 4568
- 106. Bernasconi, C.F., Leyes, A.E., Evantova, I. and Rappoport, Z. (1995). J. Am. Chem. Soc. 117, 1703
- 107. Eigen, M. (1964). Angew. Chem. Int. Ed. Engl. 3, 1
- 108. Ahrens, M.-L. and Maass, G. (1968). Angew. Chem. Int. Ed. Engl. 7, 818
- 109. Kreissl, F.R., Kreiter, C.G. and Fischer, E.O. (1972). Angew. Chem. Int. Ed. Engl. 11, 643
- 110. Kreissl, F.R. and Held, W. (1975). J. Organomet. Chem. 86, C10
- 111. Kreissl, F.R., Fischer, E.O., Kreiter, C.G. and Fischer, H. (1973). Chem. Ber. 106, 1262
- 112. Fischer, H. (1979). J. Organomet. Chem. 170, 309
- 113. Choi, H.S. and Sweigart, D.A. (1982). J. Organomet. Chem. 228, 249
- 114. Pickering, R.A. and Angelici, R.J. (1982). J. Organomet. Chem. 225, 253
- 115. Kreiter, C.G. (1968). Angew. Chem. Int. Ed. Engl. 7, 390
- 116. Schubert, U. and Fischer, E.O. (1975). Justus Liebigs Ann. Chem. 393
- Fischer, E.O., Schubert, U., Kalbfus, W. and Kreiter, C.G. (1975). Z. Anorg. Allg. Chem. 416, 135
- Bernasconi, C.F., Flores, F.X., Gandler, J.R. and Leyes, A.E. (1994). Organometallics 13, 2186

- 119. Marcus, R.A. (1965). J. Chem. Phys. 43, 679
- 120. Marcus, R.A. (1968). J. Phys. Chem. 72, 891
- 121. Bernasconi, C.F. (1987). Acc. Chem. Res. 20, 301
- 122. Bernasconi, C.F. (1992). Adv. Phys. Org. Chem. 27, 119
- 123. Bernasconi, C.F. (1992). Acc. Chem. Res. 25, 9
- 124. Bordwell, F.G. and Boyle, W.J., Jr. (1972). J. Am. Chem. Soc. 94, 3907
- 125. Leffler, J.E. and Grunwald, E. (1963). *Rates and Equilibria of Organic Reactions*, p. 156. Wiley, New York
- 126. Kresge, A.J. (1975). Acc. Chem. Res. 8, 354
- 127. Jencks, W.P. (1985). Chem. Rev. 85, 511
- 128. Bernasconi, C.F. (1989). Tetrahedron 45, 4017
- 129. Bernasconi, C.F. and Renfrow, R.A. (1987). J. Org. Chem. 52, 3035
- 130. Terrier, F., Millot, F. and Morel, G. (1976). J. Org. Chem. 41, 3892
- Bernasconi, C.F., Fassberg, J., Killion, R.B., Schuck, D.F. and Rappoport, Z. (1991). J. Am. Chem. Soc. 113, 4937
- 132. Bernasconi, C.F. and García-Río, L. (2000). J. Am. Chem. Soc. 122, 3821
- 133. Bernasconi, C.F., Flores, F.X. and Kittredge, K.W. (1998). J. Am. Chem. Soc. 120, 7983
- 134. Bernasconi, C.F., Kittredge, K.W. and Flores, F.X. (1999). J. Am. Chem. Soc. 121, 6630
- 135. Guthrie, J.P. (1978). J. Am. Chem. Soc. 100, 5892
- 136. Pearson, R.G. and Songstad, J. (1967). J. Am. Chem. Soc. 89, 1827
- 137. Pearson, R.G. (1969). Surv. Prog. Chem. 5, 1
- 138. Sander, E.G. and Jencks, W.P. (1968). J. Am. Chem. Soc. 90, 6154
- 139. Hine, J. (1975). Structural Effects on Equilibria in Organic Chemistry, p. 225. Wiley, New York
- 140. Hupe, D.J. and Jencks, W.P. (1977). J. Am. Chem. Soc. 99, 451
- 141. Jencks, W.P., Brant, S.R., Gandler, J.R., Fendrich, G. and Nakamura, C. (1982). J. Am. Chem. Soc. 104, 7054
- 142. Bernasconi, C.F. and Killion, R.B. (1988). J. Am. Chem. Soc. 110, 7506
- 143. Jencks, W.P., Haber, M.T., Herchlag, D. and Nazaretian, K.L. (1986). J. Am. Chem. Soc. **108**, 479
- 144. Richard, J.P. (1987). J. Chem. Soc., Chem. Commun. 1768
- 145. McClelland, R.A., Kanagasabapathy, V.M., Banait, N.S. and Steenken, S. (1992). J. Am. Chem. Soc. 114, 1816
- 146. Andraos, J. and Kresge, A.J. (1992). J. Am. Chem. Soc. 114, 5643
- 147. Bernasconi, C.F. and Ali, M. (1999). J. Am. Chem. Soc. 121, 11384
- 148. Kristjánsdóttir, S.S. and Norton, J.P. (1992). *Transition Metal Hydrides*. Dedieu, A. (ed.), p. 309. Verlag Chemie, New York
- 149. Hansch, C., Leo, A. and Taft, R.W. (1991). Chem. Rev. 91, 165
- 150. Unger, S.H. and Hansch, S.C. (1976). Prog. Phys. Org. Chem. 12, 9
- 151. Bernasconi, C.F. and Ali, M. (2001). Organometallics 20, 3383
- 152. Bernasconi, C.F., Ali, M. and Lu, F. (2000). J. Am. Chem. Soc. 122, 1352
- 153. Kirby, A.J. (1980). Adv. Phys. Org. Chem. 17, 183
- 154. Mandolini, L. (1986). Adv. Phys. Org. Chem. 22, 1
- 155. Aumann, R., Hinterding, P., Krüger, C. and Goddard, R. (1993). J. Organomet. Chem. 459, 145
- 156. Fischer, E.O., Kreiss, G. and Kreissl, F.R. (1973). J. Organomet. Chem. 56, C37
- 157. Bernasconi, C.F., Flores, F.X. and Kittredge, K.W. (1997). J. Am. Chem. Soc. 119, 2103
- 158. Jencks, W.P. (1972). J. Am. Chem. Soc. 94, 4731
- 159. Bernasconi, C.F. and Perez, G.S. (2000). J. Am. Chem. Soc. 122, 12441
- Bernasconi, C.F., Ketner, R.J., Brown, S.D., Chen, X. and Rappoport, Z. (1999). J. Org. Chem. 64, 8829
- 161. Fischer, H. and Dötz, K.H. (1980). Chem. Ber. 113, 193

- 162. Fischer, H. (1980). J. Organomet. Chem. 197, 303
- 163. Fischer, H. and Märkl, R. (1985). Chem. Ber. 118, 3683
- 164. Fischer, H. and Zeuner, S. (1987). J. Organomet. Chem. 327, 63
- 165. Fischer, H. (1981). J. Organomet. Chem. 222, 241
- 166. Kreiter, C.G. (1968). Angew. Chem. Int. Ed. Engl. 7, 390
- 167. Casey, C.P. and Anderson, R.L. (1974). J. Am. Chem. Soc. 96, 1230
- 168. Fickling, M.M., Fischer, A., Mann, B.R., Packer, J. and Vaughan, J. (1959). J. Am. Chem. Soc. 81, 4226
- 169. Gandler, J.R. and Bernasconi, C.F. (1989). Organometallics 8, 2282
- 170. Bernasconi, C.F. (1997). Chem. Soc. Rev. 26, 299
- 171. Bernasconi, C.F. and Sun, W. (1993). J. Am. Chem. Soc. 115, 12526
- 172. Bernasconi, C.F. and Sun, W. (1997). Organometallics 16, 1926
- 173. Bernasconi, C.F. and Leyes, A.E. (1997). J. Am. Chem. Soc. 119, 5169
- 174. Bernasconi, C.F., Sun, W., García-Río, L., Kin-Yan, and Kittredge, K. (1997). J. Am. Chem. Soc. 119, 5583
- 175. Bernasconi, C.F., Leyes, A.E. and García-Río, L. (1998). Organometallics 17, 4940
- 176. Bernasconi, C.F. and Ali, M. (1999). J. Am. Chem. Soc. 121, 3039
- 177. Bernasconi, C.F. and Ragains, M.L. (2001). J. Am. Chem. Soc. 123, 11890
- 178. Amyes, T.L. and Richard, J.P. (1996). J. Am. Chem. Soc. 118, 3129
- 179. Amyes, T.L. and Richard, J.P. (1992). J. Am. Chem. Soc. 114, 10257
- 180. Bernasconi, C. F.; Ragains, M. L. Unpublished results
- Bruice, P.Y. (2001). Organic Chemistry (3rd edn), p. 169. Prentice-Hall, Upper Saddle River, NJ
- 182. Jones, M., Jr. (2000). Organic Chemistry (2nd edn), p. 111. W.W. Norton, New York
- 183. Chesnut, D.B. and Bartolotti, L.J. (2000). Chem. Phys. 253, 1
- 184. Bernasconi, C.F. and Sun, W. (2002). J. Am. Chem. Soc. 124, 2299
- 185. Bernasconi, C. F.; Ali, M. Unpublished observations
- 186. Kresge, A.J. (1974). Can. J. Chem. 52, 1897
- 187. Bordwell, F.G. (1988). Acc. Chem. Res. 21, 456
- 188. Wulff, W.D. and Yang, D.C. (1983). J. Am. Chem. Soc. 105, 6726
- 189. Moore, E.J., Sullivan, J.M. and Norton, J.R. (1986). J. Am. Chem. Soc. 108, 2257
- 190. Fermin, M.C., Thiyagarajan, B. and Bruno, J.W. (1993). J. Am. Chem. Soc. 115, 974
- 191. Schwesinger, R. (1985). Chimia 39, 269
- 192. Schwesinger, R. and Schlemper, H. (1987). Angew. Chem. Int. Ed. Engl. 26, 1167
- 193. Parker, A.J. (1969). Chem. Rev. 69, 1
- 194. Bernasconi, C.F., Leyes, A.E., Ragains, M.L., Shi, Y., Wang, H. and Wulff, W.D. (1998). J. Am. Chem. Soc. 120, 8632
- 195. Kolthoff, I.M. and Chantooni, M.K. (1972). Anal. Chem. 44, 194
- 196. Koltfhoff, I.M. and Chantooni, M.K. (1972). J. Phys. Chem. 76, 2026
- 197. Anderson, R. L. Ph.D. Thesis, University of Wisconsin, Madison, WI, 1974
- 198. Casey, C.P. and Brunsvold, W.R. (1976). J. Organomet. Chem. 118, 309
- 199. Xu, Y.-C. and Wulff, W.D. (1987). J. Org. Chem. 52, 3263
- 200. Gandler, J.R. and Bernasconi, C.F. (1992). J. Am. Chem. Soc. 114, 631
- 201. Bernasconi, C.F., Flores, F.X. and Sun, W. (1995). J. Am. Chem. Soc. 117, 4875
- 202. Bernasconi, C.F. and Sun, W. (1995). Organometallics 14, 5615
- 203. Bernasconi, C.F. and Leyes, A.E. (1997). J. Chem. Soc., Perkin Trans. 2 1641
- 204. Bernasconi, C.F., Leyes, A.E. and García-Río, L. (1998). Organometallics 17, 4940
- 205. Mills, O.S. and Redhouse, A.D. (1968). J. Chem. Soc. A 642
- 206. Amin, S.R., Jayaprakash, K.N., Nandi, M., Sathe, K.M. and Sarkar, A. (1996). Organometallics 15, 3528
- 207. Bernasconi, C.F., García-Río, L., Sun, W., Yan, K. and Kittredge, K.W. (1997). J. Am. Chem. Soc. 119, 5583

Thermodynamic Stabilities of Carbocations

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Dedicated to Professor Edward M. Arnett

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1 Introduction

Small ions such as methylium (CH_3^+) , methanium (CH_5^+) and ethylium $(C_2H_5^+)^{1,2}$ are long-time favorites of experimental chemical physicists and theoretical chemists. Some of their work is reflected in this chapter. Furthermore, these and/or other cognate species such as vinylium $(C_2H_3^+)$ and cyclopropenylium $(c-C_3H_3^+)$ ions have been either detected or postulated to be significant constituents of the interstellar space³ and the stratospheres of the outer planets and some of their satellites^{4,5} as a result of the ionization of hydrocarbons.⁶ Even larger species have recently been considered.⁷ So, a great deal of experimental and computational information on their structure, molecular spectra and energetics is available and part of it is summarized here. On the other hand, many of these species are far too reactive to be experimentally observed in solution, let alone the solid state. At the other end of the "stability spectrum" we have species such as substituted triarylmethylium, cyclopropenylium and tropylium ions, which form authentic hydrocarbon salts. A series of these and other remarkable carbocations has recently been reviewed.⁸ The cyclopropenylium and tropylium ions are aromatic species with highly delocalized π -electron systems.⁹

A number of carbenium ions lies between these two extremes, including secondary and tertiary alkyl carbocations as well as protonated aromatic hydrocarbons, many of them identified in solution, and more recently, as relatively stable solid salts (some representative examples are examined here). The study of all these species has long been a very important part of physical organic chemistry. Their industrial importance has promoted a great deal of research, both experimental^{10–15} and theoretical.^{16–20}

This chapter essentially addresses the following topics:

- (1) The quantification of the concept of "stability". More precisely, thermodynamic criteria are applied in order to rank carbenium ions on the basis of their intrinsic stabilities, that is, stabilities in the absence of solvent. Both experimental and computational data are used.
- (2) The relationship between the intrinsic thermodynamic stability of these species and their reactivity in solution, from both the thermodynamic and the kinetic points of view.
- (3) Relevant structural features of these species, as revealed by experimental or computational methods data, are reviewed. Furthermore, quantum mechanical calculations at the highest possible levels (within our computational capabilities) were performed on most of these species. The scope of this chapter is limited to a relatively small number of ions that illustrate some representative examples of structural effects.

2 Quantitative thermodynamic criteria of stability in the gas phase

DEFINITIONS AND EXPERIMENTAL TECHNIQUES

We are concerned here with the thermodynamic stability of carbocations. For the sake of generality, we formally consider that they derive from a neutral molecule R–H, which can either lose a hydride or be protonated in the gas phase, reactions (1) and (2):

$$R-H(g) \rightarrow R^{+}(g) + H^{-}(g) \tag{1}$$

$$R-H(g) + H^{+}(g) \rightarrow RH_{2}^{+}(g)$$
⁽²⁾

A quantitative ranking of stabilities for any two of these species (e.g., R_1^+ and R_2^+) can be defined through the thermodynamic state functions for the hydride- or proton-

$$R_1H(g) + R_2^+(g) \rightarrow R_2H(g) + R_1^+(g)$$
 (3a)

$$R_1H(g) + R_2H_2^+(g) \to R_2H(g) + R_1H_2^+(g)$$
(4)

The standard enthalpies and Gibbs energy changes for these reactions at some temperature *T*, respectively, $\Delta_r H^{\circ}_T$ and $\Delta_r G^{\circ}_T$, involve the corresponding standard magnitudes for the various reactants and products.

For example, the standard enthalpy change for reaction (3a) at 298.15 K is given by equation (3b):

$$\Delta_{\rm r} H^{\circ}(3a) = \Delta_{\rm f} H^{\circ}{}_{\rm m}[{\rm R}_{2}{\rm H}(g)] + \Delta_{\rm f} H^{\circ}{}_{\rm m}[{\rm R}^{+}_{1}(g)] - \Delta_{\rm f} H^{\circ}{}_{\rm m}[{\rm R}_{1}{\rm H}(g)] - \Delta_{\rm f} H^{\circ}{}_{\rm m}[{\rm R}_{2}^{+}(g)]$$
(3b)

wherein the various $\Delta_f H^\circ_m$ terms are the standard enthalpies of formation of the various species involved in the reaction.

Experimental values of the standard enthalpies of formation of the neutral species, generally obtained by thermochemical or kinetic techniques, $^{23-25}$ are frequently available and there are some excellent critical compilations of these data²⁶⁻³¹ that can be eventually estimated empirically³²⁻³⁵ or computationally (see later).

Several methods exist that allow the determination of the standard enthalpies of formation of the ionic species. The reader is referred to two recent rigorous and detailed chapters by Lias and Bartmess³⁶ and Ervin.³⁷ The vast majority of the experimental data reported here are obtained by means of Fourier transform ion cyclotron resonance spectroscopy (FT ICR), high-pressure mass spectrometry (HPMS), selected ion flow tube (SIFT), and pulsed-field ionization (PFI) techniques, particularly pulsed-field ionization photoelectron photoion coincidence (PFI-PEPICO). All these experimental techniques have been examined quite recently, respectively, by Marshall,³⁸ Kebarle,³⁹ Böhme,⁴⁰ Ng⁴¹ and Baer.⁴² These chapters appear in a single (remarkable) issue of the *International Journal of Mass Spectrometry*. An excellent independent discussion of the thermochemical data of ions, with a careful survey of these and other experimental methods, is given in Ref. 37.

From these techniques the following raw experimental data are obtained:

(1) FT ICR, HPMS and SIFT yield the standard Gibbs energy change, $\Delta_r G^{\circ}(5a)$ for reaction (5a), the proton exchange between two bases, B₁ and B₂,

$$B_1H^+(g) + B_2(g) \to B_1(g) + B_2H^+(g) \qquad \Delta_r G^{\circ}(5a)$$
 (5a)

 $\Delta_{\rm r}G^{\circ}(5{\rm a})$ is given by the difference between $\Delta_{\rm r}G^{\circ}(5{\rm b})$ and $\Delta_{\rm r}G^{\circ}(5{\rm c})$ pertaining, respectively, to reactions (5b) and (5c):

$$B_2H^+(g) \to B_2(g) + H^+(g)$$
 (5b)

$$B_1 H^+(g) \rightarrow B_1(g) + H^+(g) \tag{5c}$$

The standard Gibbs energy change for reaction (6) is known as the gas-phase basicity of base B, GB(B):

$$BH^+(g) \to B(g) + H^+(g) \qquad GB(B) = \Delta_r G^{\circ}(6) \tag{6}$$

From this it follows that the unknown GB of a base, say B_1 , can be obtained from the experimental values of $\Delta_r G^{\circ}(5a)$ and $GB(B_2)$, B_2 being an appropriate reference base.^{43,44}

- (2) SIFT and HPMS experiments performed at different temperatures provide the standard enthalpy change for reaction (5a), $\Delta_r H^{\circ}(5a)$. As before, combination of this datum with $\Delta_r H^{\circ}(5b)$ leads to $\Delta_r H^{\circ}(5c)$, also known as the proton affinity of B₁, PA(B₁).^{43,44}
- (3) Ionization techniques are extremely important sources of thermochemical data for ions. Excellent chapters are given in Refs. 37, 41 and 42.

Here, we simply consider that they yield (*inter alia*) the adiabatic ionization energies (IE) and appearance energies (AE) defined as the standard energy changes at 0 K for reactions (7) and (8) (assuming for the latter that there is no potential barrier in the reaction coordinate and little or no kinetic shift):

$$\dot{R}(g) \to R^+(g) + e^- \qquad 0 \text{ K IE}_0 = \Delta_r E^\circ_0(7)$$
(7)

$$AB(g) \rightarrow A^{+}(g) + B(g) + e^{-} \qquad 0 \text{ K } AE_{0} = \Delta_{r} E^{\circ}_{0}(8)$$
(8)

Extracting $\Delta_f H^{\circ}_{m}(R^+)$ or $\Delta_f H^{\circ}_{m}(A^+)$ at finite temperatures from IE or AE, respectively, requires the corresponding $\Delta_f H^{\circ}_{m}$ values for the appropriate neutral species as well as their corresponding integrated heat capacities. The latter can be obtained from compilations (see above) or else computed by quantum chemical methods. Several conventions, clearly presented and compared in Refs. 36 and 37, exist for the determination of $\Delta_f H^{\circ}_m(e^-)$. Here, we follow the so-called "ion convention" because it is used in a number of important sources of thermochemical data for ions. In this convention, the enthalpy of formation of the electron is defined as equal to its integrated heat capacity at all temperatures. Note that as indicated by Bartmess,^{45,46} the most rigorous approach is to treat the electron as a fermion and to apply Fermi–Dirac statistics to compute its integrated heat capacity and entropy. We suggest that institutions such as IUPAC ought to officially recommend this treatment.

Directly determined experimental standard entropies for carbenium ions in the gas phase, S_{m}° , are seldom available. Those we present here are derived generally from the combination of experimental gas phase $\Delta_{r}G_{m}^{\circ}$ and $\Delta_{r}H_{m}^{\circ}$ values. We also report the data obtained by quantum mechanical methods. A careful discussion of these data is given in Ref. 47. When needed, S_{m}° values for neutral species were taken from compilations^{29,30,48} or computed.

THE SPECIFIC CASE OF CARBOCATIONS

Hydride, halide and proton exchanges

The quantitative ranking of stabilities can sometimes be obtained directly by HPMS or FT ICR. In both these cases the main condition is the absence of isomerization or other extraneous processes.

Carbocations can exchange hydride, as in reaction (3a),⁴⁹ or halide anions (mostly chlorides and bromides),⁵⁰ as in reaction (9):

$$R_2^+(g) + R_1 - X(g) \to R_1^+(g) + R_2 - X(g)$$
(9)

As we show below, exchanges of other species can be studied, at least formally. In all cases, the rankings defined through equations (3a) and (9) are linked through equation (10):

$$R_2 - H(g) + R_1 - X(g) \rightarrow R_2 - X(g) + R_1 - H$$
 (10)

This is an isodesmic process involving the neutral species only. In principle, $\Delta_r G^{\circ}(10)$ and $\Delta_r H^{\circ}(10)$ can be obtained experimentally. Otherwise, this "leaving group correction" can be obtained by quantum chemical calculations. Here, we use both methods.

Equilibrium proton exchange between ethylenic compounds is ideally suited whenever isomerization and/or other processes cannot compete with proton exchange. If contributions from the neutral species are very small, rankings obtained by this method and by chloride exchange are essentially identical. For example, the standard Gibbs energy changes for reactions (11) and (12) are, respectively,⁵¹ – 7.7 and – 7.5 kcal mol⁻¹:

$$C_6H_5CH(CH_3)Cl(g) + tert - C_4H_9^+(g) \rightarrow C_6H_5CH(CH_3)^+(g) + tert - C_4H_9Cl(g)$$
(11)

$$C_6H_5CH = CH_2(g) + tert - C_4H_9^+(g) \rightarrow C_6H_5CH(CH_3)^+(g) + (CH_3)_2C = CH_2(g)$$
 (12)

Other important examples are also known.⁵²

The dissociative proton attachment method

The construction of a quantitative scale of carbocation stabilities based on hydride or halide exchanges would require the systematic overlapping equilibration⁴⁴ of a large number of species (as in reactions (11) and (12)). This was the time-honored procedure for the construction of the gas-phase proton basicity scale.⁴³ In the present case, this is difficult to achieve because of the widely different stabilities involved and because of the possibility of rearrangement of the ions.

It has long been known that in the gas phase, R-X compounds (X = OH, halogen, SH, etc.) can react with protonated bases, BH⁺, to undergo a "dissociative proton

attachment process" (DPA), as shown in reaction (13):

$$BH^{+}(g) + R - X(g) \rightarrow B(g) + R^{+}(g) + HX(g)$$
(13)

Lias and Ausloos⁵³ have summarized earlier literature on these processes.

We have recently shown that by varying the strength of the bases (B), it is possible to determine the onset for reaction (13) fairly accurately.⁵⁴ This is conveniently done by means of FT ICR. Full experimental details are given in Refs. 54 and 55. Briefly stated, $BH^+(g)$ is formed by chemical ionization following the electron ionization of a mixture of B(g) and R-X(g) (in general, fragment ions from B and/or R-X act as proton sources) and then isolated by ion ejection techniques and allowed to react. The formation/absence of R⁺(g) is monitored for periods ranging from 1 to (sometimes) 60 s.

The dissociative proton transfer from BH⁺(g) to R–X under a pressure *P*, reaction (13), will take place spontaneously (assuming a negligible dissociation barrier) when $\Delta_r G^{P}(13) < 0$. It can be easily shown that $\Delta_r G^{P}(13)$ is given by equation (14):

$$\Delta_{\rm r} G^P(13) = \Delta_{\rm r} G^P(15) + \Delta_{\rm r} G^P(16) \tag{14}$$

wherein $\Delta_r G^P(15)$ and $\Delta_r G^P(16)$ pertain, respectively, to reactions (15) and (16):

$$H^{+}(g) + R - X(g) \rightarrow R^{+}(g) + HX(g)$$
(15)

$$BH^+(g) \to B(g) + H^+(g) \qquad \Delta_r G^{\circ}(16) = GB(B)$$
(16)

The experimentally determined onset for reaction (13) satisfies the condition $\Delta_r G^P(13) \approx 0$. It then follows that $\Delta_r G^P(15) \approx -\Delta_r G^P(16)$.

Let R_1 -X and R_2 -X, respectively, stand for two halides (or alcohols or other derivatives) and B_1 and B_2 two reference bases defining their respective DPA onsets. The position of equilibrium (17) as determined by $\Delta_r G^{\circ}(17)$ is a quantitative measure of the relative stabilities of $R_1^+(g)$ and $R_2^+(g)$:

$$R_1X(g) + R_2^+(g) \to R_1^+(g) + R_2X(g) \qquad \Delta_r G^{\circ}(17), \ \Delta_r G^{P}(17)$$
 (17)

It can be readily shown that $\Delta_r G^{\circ}(17) = \Delta_r G^{P}(17)$. Then, and from the discussion given above, equation (18) follows:

$$\Delta_{\rm r} G^{\circ}(17) \approx {\rm GB}({\rm B}_2) - {\rm GB}({\rm B}_1) \tag{18}$$

This expression reduces the determination of the relative stabilities of $R_1^+(g)$ and $R_2^+(g)$ through the formal exchange of the halide, hydroxide or other anions, reaction (17), to the actual determination of the DPA onsets for the corresponding neutral precursors, reaction (13).

These are bracketing experiments and their precision (estimated at *ca*. 2 kcal mol^{-1}) is lower than that achieved under "true" equilibrium conditions. They have, however, the following advantages: (1) ions are formed under extremely mild conditions (see later); (2) the acidity of the protonating species BH⁺(g) can be varied within very wide limits by an appropriate choice of B(g); (3) gas-phase basicities are available for hundreds of compounds.

Over the last several years, the DPA method has been applied to a number of systems. The following illustrates its scope and the reliability of the data derived therefrom.

- (1) We have applied DPA to *tert*-butyl and *sec*-butyl chlorides.^{55,56} As discussed in other sections, the difference in stabilities we obtained was in excellent agreement with that provided by other methods. The most important point, however, is that the DPA onset for *sec*-BuCl could be clearly determined, in spite of the fact that *sec*-Bu⁺ readily rearranges to *tert*-Bu⁺. This is explained by the fact that the DPA are *entropy-driven*, *endoergic reactions*. This makes DPA extremely valuable for the study of relatively unstable species.^{55,57,58} Examples of such processes are known.^{59–61} They are currently under study in our laboratories.
- (2) We have applied DPA to a set of halomethanes.⁶² A value of $201.9 \pm 2.0 \text{ kcal mol}^{-1}$ for $\Delta_{\rm f} H^{\circ}_{\rm m}({\rm CCl}_3^+)$ was obtained from that study.^{63,64} From the most recent $\Delta_{\rm f} H^{\circ}_{\rm m}$ and adiabatic ionization energy for the trichloromethyl radical, respectively, $17.0 \pm 0.6 \text{ kcal mol}^{-165}$ and $8.06 \pm 0.02 \text{ eV}$,⁶⁶ one gets $\Delta_{\rm f} H^{\circ}_{\rm m}({\rm CCl}_3^+) = 202.7 \pm 0.8 \text{ kcal mol}^{-1}$. Using instead the computed ionization energy (CEPA-1 level with triple- ζ equivalent basis set) of 7.990 eV,⁶⁷ $\Delta_{\rm f} H^{\circ}_{\rm m}({\rm CCl}_3^+)$ becomes $201.0 \pm 2.0 \text{ kcal mol}^{-1}$. Other examples exist showing that the accuracy of the thermodynamic data obtained by DPA is sufficient for the purposes of this study.
- (3) Consider reaction (17). For DPA results to be acceptable, one has to ensure that rankings of carbocation stabilities obtained by using different precursor species (i.e., different X) are self-consistent. For example, let us suppose we compare the stabilities of $R_1^+(g)$ and $R_2^+(g)$ through DPA studies of the respective chlorides and alcohols (X = Cl and X = OH). The corresponding experimental $\Delta_r G^{\circ}(17)$ values are noted, $\Delta_r G^{\circ}(17, X = Cl)$ and $\Delta_r G^{\circ}(17, X = OH)$. They must satisfy equation (19):

$$\Delta_{\mathbf{r}}G^{\circ}(17, \mathbf{X} = \mathbf{OH}) = \Delta_{\mathbf{r}}G^{\circ}(17, \mathbf{X} = \mathbf{CI}) + \Delta_{\mathbf{r}}G^{\circ}(20)$$
(19)

wherein $\Delta_r G^{\circ}(20)$ pertains to reaction (20), an isodesmic process involving the neutral precursors and having the meaning of a leaving group correction:

$$R_2Cl(g) + R_1OH(g) \rightarrow R_2OH(g) + R_1Cl(g)$$
⁽²⁰⁾

We have recently reported results of a systematic DPA study of bridgehead carbocations using as precursors alcohols, chlorides and bromides.⁵⁸ Figure 1 is a plot of $\Delta_r G^{\circ}(17, X = Br, \text{ calc. Cl})$, i.e., values of $\Delta_r G^{\circ}(17, X = Br)$ obtained from experimental $\Delta_r G^{\circ}(17, X = Cl)$ and corrected as indicated above, versus the experimental $\Delta_r G^{\circ}(17, X = Br)$ and $\Delta_r G^{\circ}(17, X = Br, \text{ calc. OH})$. It shows that the self-consistency is excellent.

The same was found to hold with a series of tertiary alkyl cations obtained from their chlorides and alcohols.⁶⁸

(4) Reaction (3a) is one of the key reference processes used here. $\Delta_r G^{\circ}(3a)$ can also be obtained computationally. In keeping with previous studies, $\Delta_r G^{\circ}(3a)$ is


Fig. 1 Self-consistency of DPA results. Correction of leaving group effects.

referred to 1-adamantyl cation (R_2^+). A most stringent test of the selfconsistency of the DPA method is the comparison of the $\Delta_r G^{\circ}(3a)$ values derived from experiment (upon the appropriate leaving group correction) with those obtained by purely computational methods (ranging from MP2(full)/6-31G(d) to G2(MP2) levels). Figure 2 is an example of the quality of the agreement.

3 Theoretical calculations

Enthalpies and Gibbs free energies for the experimental processes of the systems involved were evaluated using high-level *ab initio* calculations. The results reported herein are based on $G2^{69}$ calculations for the smallest systems (up to five heavy atoms) and $G2(MP2)^{70}$ for the intermediate ones (between six and eight heavy atoms). Larger systems were optimized at the MP2(full)/6-31G(d) level. Thermodynamic corrections were generally calculated at the RHF/6-31G(d) level, with the scaling used in the G-n family of calculations. Thus, the results obtained for all the molecules can be compared at the same level, if necessary, since the calculations at the lowest levels are embedded in the G2 and G2(MP2) methodologies. In the same way, calculations at lower levels can be upgraded easily when the necessary computational resources become available.

The G2 and G2(MP2) computational methods used here provide molecular energies with deviations from the experimental data similar, in many cases, to the



Fig. 2 Computed Gibbs energy changes for reaction (3a) versus DPA-based results.

accuracy of the latter. The reported average deviations are 1.21 and 1.52 kcal mol⁻¹ for the G2 and G2(MP2) calculations, respectively.^{69,70}

4 Uncertainties

From the inspection of reactions (3a), (4), (7) and (8), it follows immediately that the uncertainties on the standard enthalpies of formation of gaseous ions are determined by those affecting the thermochemical data for the relevant neutral species (molecules or free radicals) and the specific method used for ion generation. The uncertainty assigned to the standard enthalpy of formation of a gaseous ion is the root-sum-of-squares combination of the individual uncertainties of the various contributions.⁴³

Perusal of the data presented in the following sections would show that extremely accurate (better than 0.01 kcal mol⁻¹) ionization energies can be obtained with state-of-the-art optical spectroscopy and photoionization and electron ionization techniques.

The uncertainties affecting experimental *differences* in GB values as determined by equilibrium methods are frequently in the range 0.1-0.3 kcal mol⁻¹. Individual differences in PA values obtained by van't Hoff plots of equilibrium constants for processes such as proton or halide exchange reactions are accurate to within a 1-2 kcal mol⁻¹ range as indicated by Kebarle and co-workers.⁷¹ The usual multipleoverlap technique likely halves these uncertainties in the case of data originating in the same laboratory. As indicated in the discussion of reactions (5a) and (6), "absolute" GB and PA values must be anchored to some specific reference values. Overall, and unless warranted by other evidence, we take the accuracy of "absolute" GB and PA values as *ca*. 2 kcal mol⁻¹.⁴³ In many cases this is practically the main contributor to the uncertainty on $\Delta_f H^\circ_m$ values for ions. Note, however, that in some cases this is perhaps too conservative an estimate, in view of the excellent level of agreement that often exists between independent $\Delta_f H^\circ_m$ values obtained from ionization and equilibrium techniques.

DPA onsets are estimated to be known within 2 kcal mol⁻¹. These values must also be anchored to some reference. This leads to an estimated uncertainty of *ca*. 2.8 kcal mol⁻¹ on the formal $\Delta_f G^{\circ}_m(17)$ values. Furthermore, in order to extract $\Delta_f H^{\circ}_m$ values for ions, leaving group and entropy corrections are needed. With few exceptions, and because of the scarcity of experimental data, these corrections are determined computationally by means of isodesmic reactions.^{57,58,68} We estimate that the leaving group correction introduces an extra 1 kcal mol⁻¹ uncertainty. Unless otherwise stated, we anchor DPA onsets obtained from chlorides and alcohols to the onsets for the corresponding 1-adamantyl derivatives. We have recently introduced the DPA of methyl ethers as an appropriate tool for the study of very stable species (see later); their corresponding onsets are anchored to that for the methyl ether of tricyclopropyl carbinol.

Uncertainties on the experimental standard enthalpies of formation of neutral species vary widely. For most hydrocarbons and radicals they are in the 0.1-1.0 and 0.5-2.0 kcal mol⁻¹ ranges, respectively.

Very few standard entropies for ions relevant to this study are available that derive from purely experimental data. These data can be obtained, at least formally, from the combination of PA and GB values for processes such as reaction (2):

$$R-H(g) + H^{+}(g) \rightarrow RH_{2}^{+}(g) \qquad \Delta_{r}S^{\circ}(2)$$
(2)

The accuracy of $\Delta_r S^{\circ}(2)$ is estimated at 1–2.5 cal mol⁻¹ K⁻¹. $S^{\circ}[RH_2^+(g)]$ is even less accurate, because of the contribution from the uncertainty on $S^{\circ}[RH(g)]$.

A recent study⁴⁷ indicates that quantum mechanical calculations of moderate to high level are able to provide values of $\Delta_r S^{\circ}(2)$ accurate to within 0.3– 0.6 cal mol⁻¹ K⁻¹. As regards the standard entropies for individual ions, we estimate that the accuracy of the scaled HF/6-31G(d) values used in the G-n family calculations^{69,70} (see above) is about 2 cal mol⁻¹ K⁻¹, based on the differences between values computed at this level and experimental or high-level computational data.

The accuracy of the data we present is not homogeneous and it is rather low in some cases, largely because of the unavailability of thermodynamic state functions for neutral species. Obviously, the spirit of this work is quite different from that of the excellent databases from NIST. Thus, while we have striven to provide the "best" possible thermodynamic values, we have also considered that even a "lowresolution" study of the relationships between structure, intrinsic (gas phase) stability and solution reactivity of carbocations is bound to provide useful results.

5 Thermodynamics and structure of selected species

CARBOCATIONS C1-C4

Methylium ion (CH_3^+) (1)

Spectroscopic studies in the gas phase^{72–76} and very high level *ab initio* calculations⁷⁷ indicate that this species is an oblate symmetric top belonging to the D_{3h} molecular point group (Fig. 3). The experimentally estimated C–H distance is 1.087 Å, the computed value (1.089 Å) being in excellent agreement.

The standard enthalpy of formation given in Table 1 is based on the experimental value of the adiabatic ionization potential of the methyl radical⁷⁸ and has been adjusted⁷⁹ to a series of fundamental enthalpies of formation. Very recently,⁸⁰ the onset of the formation of CH₃⁺ from methane was determined quite accurately by means of the PFI-PEPICO technique. This datum, combined with other experimental results, yields the accurate 0 K dissociation energies (D_0) for CH₄ and CH₄⁺, D_0 (H–CH₃) = 103.479 ± 0.023 and D_0 (H–CH₃⁺) = 39.321 ± 0.092 kcal mol⁻¹.

Methanium ion (CH_5^+) (2)

Reliable experimental values for $PA(CH_4)$ and $GB(CH_4)$ are available, particularly from HPMS and flowing afterglow studies.^{81,82} When combined with the (rather accurate) data for the neutral species, they provide the thermodynamic data given in Table 1. The results of G2 *ab initio* calculations (Ref. 83 and this study) are also reported.

Within the Born–Oppenheimer approximation, computational studies at extremely high levels^{84,85} show that the global minimum, $2a(C_s)$, has a C_s symmetry and that it can be viewed as an H₂ molecule tightly bound to a methyl cation (three-center two-electron bonds). Two other structures, $2b(C_s)$ and $2c(C_v)$, respectively, of symmetries C_s and C_{2v} , are transition states. $2b(C_s)$ involves the rotation of the H₂ moiety and allows the exchange of the two hydrogen atoms.



Fig. 3 Structure of methylium ion, 1, optimized at the MP2(full)/6-31G(d) level.

| Cations | $\Delta_{\rm f} H^{\circ}{}_{\rm m}({ m g})^a$ | $S^{\circ}_{m}(g)^{b}$ |
|--|---|---|
| Methylium, CH_3^+ (1) | 261.83 ± 0.06^{c} | 44.59 ± 0.06^{d} |
| Methanium, CH_5^+ (2) | $\begin{array}{c} 218.0 \pm 2.0^{e} \\ 220.3 \pm 2.0^{g} \end{array}$ | 54.2 ± 1.2^{f} 53.8 ± 2.0^{h} |
| Ethylium, $C_2H_5^+$ (3) | $215.6 \pm 2.0^{i} 215.17 \pm 0.50^{k} 215.25 \pm 0.50^{m} 215.2 \pm 2.0^{n}$ | 55.2 ± 2.0^{i} 55.3 ± 2.0^{l} |
| Propan-2-ylium, <i>iso</i> - $C_3H_7^+$ (4) | $\begin{array}{r} 193.04 \pm 0.36^{o} \\ 192.2 \pm 2.0^{q} \\ 192.7 \pm 2.0^{s} \\ 194.4 \pm 2.0^{t} \end{array}$ | $ \begin{array}{r} 68.0 \pm 2.0^{p} \\ 66.23 \pm 0.24^{r} \end{array} $ |
| Butan-2-ylium (sec-butyl), sec- $C_4H_9^+$ (5) | $184.5 \pm 1.3^{u} 182.4 \pm 2.0^{w} 183.0 \pm 2.0^{y} 183.4 \pm 2.0^{z}$ | $69.8 \pm 2.0^{\nu}$ 71.2 ± 2.0^{x} |
| 2-Methylpropan-2-ylium (<i>tert</i> -butyl), <i>tert</i> - $C_4H_9^+$ (6) | $\begin{array}{c} 169.9 \pm 0.9^{aa} \\ 170.03 \pm 0.26^{ac} \\ 169.7 \pm 0.6^{ae} \\ 169.2 \pm 1.7^{af} \end{array}$ | $73.4 \pm 2.0^{ab} \\ 74.04 \pm 0.26^{ad}$ |

 Table 1
 Thermodynamic data for selected carbocations, C1-C4

^{*a*}In kcal mol⁻¹.

^{*b*}In cal mol⁻¹ K⁻¹.

 ${}^{c}\Delta_{f}H^{o}_{m}(g)$ from Ref. 79, based on the experimentally resolved threshold PE spectrum of CH₃ (Ref. 78) and vibrational data summarized in Chase, M. W., Jr. NIST-JANAF Thermochemical Tables, 4th edn (1998). J. Phys. Chem. Ref. Data, Monograph No. 9. The very high level calculation of the ionization energy of CH₃ (Ref. 77) agrees with experiment within 0.02 kcal mol⁻¹.

^dObtained in this work using the smoothed experimental vibrational frequencies and C-H bond length given in Ref. 77.

^{*e*}From the averaged experimental PA(CH₄) taken from Ref. 43 and $\Delta_{\rm f} H^{\circ}_{\rm m}$ (CH₄) = -17.80 ± 0.10 kcal mol⁻¹.²⁶

^fFrom the experimental $S^{\circ}_{m}(g)$ value for CH₄, 45.09 ± 0.10 cal mol⁻¹ K⁻¹ (Ref. 26) and the recommended experimental values⁴³ for PA and GB(CH₄). Uncertainties are estimated from the differences between the experimental values determined.⁸¹

^{*g*}From $\Delta_{f}H^{\circ}_{m}(CH_{4})$ and the computed (G2) PA(CH₄), 127.6 kcal mol⁻¹ (this work).

^hScaled, symmetry-corrected value from this work [HF/6-31G(d)].

^{*i*}From the experimental $PA(C_2H_4) = 162.6 \text{ kcal mol}^{-1}$ (Ref. 81) and $\Delta_f H^o_m(C_2H_4) = 12.54 \pm 0.25 \text{ kcal mol}^{-1}$.

^{*j*}From the experimental $GB(C_2H_4)$ and $PA(C_2H_4)$ from Refs. 43 and 81.

^{*k*}From the adiabatic ionization potential of ethyl radical, 8.117 \pm 0.008 eV determined by photoionization mass spectrometry⁹⁵ and $\Delta_f H^{\circ}_{m}(C_2H_5) = 28.35 \pm 0.41$ kcal mol⁻¹ from Ref. 96. Thermal corrections as in Ref. 95.

¹From the *ab initio* scheme by Radom and co-workers.⁴⁷

^{*m*}From PFI-PE spectroscopy and ion coincidence detection using multi-bunch synchrotron radiation.⁹⁷ ^{*n*}From the experimental $\Delta_{\rm f} H^{\circ}_{\rm m}({\rm C}_{2}{\rm H}_{4})$ and PA(C₂H₄) = 162.98 kcal mol⁻¹ computed at the G2 level.⁴⁷

^oPFI-PEPICO TOF experiments and experimental $\Delta_f H^o_m(g)$ for the various precursor species.¹⁰²

^{*p*}From the experimental S_{m}° , (Ref. 26) PA and GB for C₃H₆.

^{*q*}From the experimental PA of propene $(178.4 \text{ kcal mol}^{-1})^{81}$ and its $\Delta_{f}H^{\circ}_{m}(g)$ value, 4.879 kcal mol⁻¹.²⁶ ^{*r*}Ab initio computed value, from Ref. 47.

^sFrom the experimental $\Delta_{f}H^{\circ}_{m}(g)$ of $C_{3}H_{6}$ and its G2-computed PA.^{47,83}

^tComputed (CBSQ, atomization).⁹⁹

"From $\Delta_f H^o_m(tert-C_4H_9^+) = 169.9 \pm 0.9 \text{ kcal mol}^{-1}$ (see later) and the experimental enthalpy of isomerization $sec-C_4H_9^+ \rightarrow tert-C_4H_9^+$, $-14.55 \pm 0.50 \text{ kcal mol}^{-1}$.^{108,109} This value pertains to the formation of an equilibrating mixture of isomers I and II *in solution*.

^vScaled, symmetry-corrected value for structure **I**, computed at the MP2(full)/6-31G(d) level (this work). ^wFrom the experimental $\Delta_f H^{\circ}_m$ (Ref. 26) and $S^{\circ}_m (E-C_4H_8)^{29}$ and $GB(E-C_4H_8) = -172.5 \pm 1.1 \text{ kcal mol}^{-1}$ (Ref. 107) corrected with the computed S°_m values for structures **I** and **II**. This value pertains to the formation of an equilibrating mixture of isomers **I** and **II** *in the gas phase*.

^xScaled value for structure **II**, computed at the MP2(full)/6-31G(d) level (this work).

^yFrom the experimental $\Delta_f H^{\circ}_m(E-C_4H_8) = -2.58 \text{ kcal mol}^{-1}$ (Ref. 26) and the G2-computed PA(*E*-C_4H_8) to yield structure I (this work).

^zFrom the experimental $\Delta_t H^o_m (E-C_4H_8)$ and the G2-computed PA($E-C_4H_8$) to yield structure II (this work).

^{aa}From PEPICO studies.¹¹³

^{*ab*}From the experimental GB(*iso*-C₄H₈) = 185.4 kcal mol⁻¹ (Ref. 115) and PA(*iso*-C₄H₈).⁸¹

^{ac}From photoionization mass spectrometry.¹¹⁴

adComputed value.47

^{*ae*}From $\Delta_{\rm f} H^{\circ}_{\rm m}(iso-C_4H_8) = -4.29 \pm 0.26 \text{ kcal mol}^{-1}$, (Ref. 26) and PA(*iso*-C_4H_8) = 191.7 \pm 0.5 kcal mol}^{-1}.⁸¹

^{*df*}From the experimental $\Delta_f H^\circ_m$ and S°_m for *iso*-C₄H₈, from Ref. 26 and the calculated (G2) values for the PA and GB of *iso*-C₄H₈, respectively, 192.2 and 185.6 kcal mol^{-1.47,83}

 $2c(C_{\nu})$ allows the exchange between these hydrogens and those of the methyl tripod (Fig. 4).

These calculations also show that, in terms of electronic energies, structures $2\mathbf{b}(C_s)$ and $2\mathbf{c}(C_v)$ are some 0.1 and 0.8 kcal mol⁻¹ above structure $2\mathbf{a}(C_s)$. Inclusion of the vibrational contributions makes the barrier heights vanishingly small and raises some intriguing questions. In any case, these results suggest an extremely floppy structure and the possibility of easy scrambling of the hydrogen atoms.

Recently,^{86,87} CH₅⁺ has been the subject of *ab initio* integral path calculations, which include nuclear quantum effects such as zero-point motion and tunneling in full dimensionality and go beyond the harmonic approximation. These studies confirm the above results, in that while protons undergo large-amplitude pseudorotational motion and become scrambled and statistically equivalent, this motion is concerted and the situations in which the vibrating nuclear skeleton having an H₂ moiety attached to a CH₃ tripod are the most important contributors to the overall appearance of the cation.

At this point, however, the situation is far from being entirely clear, because of the following facts:

- (1) The experimental (gas phase) IR spectrum of the C–H stretching region of CH_5^+ has been obtained.⁸⁸ It portrays some 900 lines and has not yet been interpreted.
- (2) Mass spectrometric studies⁸⁹ seem to indicate that, in the absence of intermolecular collisions, CH_4D^+ and CD_4H^+ do not show any rearrangement.



Fig. 4 Possible structures (Born–Oppenheimer approximation), for methanium ion, 2, optimized at the MP2(full)/6-31G(d) level. 2a is the absolute minimum; 2b and 2c are transition states.



Fig. 5 Structure of ethylium ion, 3, optimized at the MP2(full)/6-31G(d) level.

This situation has prompted a lively debate⁹⁰ but still remains a "tough test for experiment and theory".⁹¹

Ethylium ion, ethyl cation $(C_2H_5^+)$ (3)

It has long been known^{92,93} that this ion has a non-classical, bridged $C_{2\nu}$ structure and that the classical C_s form is a saddle point on the potential energy surface of the C₂H₅⁺ manifold⁹⁴ (Fig. 5).

The rewardingly coincident $\Delta_{\rm f} H^{\circ}{}_{\rm m}(g)$ values for this ion, reported in Table 1, were obtained from three experimental sources: (1) the proton affinity of ethylene;⁴³, ⁸¹ (2) the ionization energy of the ethyl radical;⁹⁵ (3) the PFI-PE spectrum of C₂H₅Br,⁹⁶ combined with the $\Delta_{\rm f} H^{\circ}{}_{\rm m}(g)$ of the appropriate neutral^{26,97} species. $S^{\circ}{}_{\rm m}(g)$, as obtained from the experimental PA and GB values for C₂H₄ together with the experimental $S^{\circ}{}_{\rm m}(C_2H_4)$, is presented in Table 1 together with the computational value.

Propan-2-ylium ion, iso-propyl cation ($iso-C_3H_7^+$) (4)

According to theoretical calculations and experimental evidence in solution,^{22,98,99} this species has a *chiral* structure with C_2 symmetry but very low methyl rotation barriers (Fig. 6). Cation **4** has been studied in solution by NMR^{100,101} but no data seem to be available for this ion in the solid state.

 $\Delta_{\rm f} H^{\circ}_{\rm m}(4)$ has been obtained experimentally from the dissociative ionization onset for the 2-propyl chloride, bromide and iodide PFI-PEPICO TOF.¹⁰² The experimental value of the PA of propene⁸¹ leads to a very close result. These data are summarized in Table 1. The reader is referred to Ref. 102 for an extremely careful discussion on the experimental and calculated values of $\Delta_{\rm f} H^{\circ}_{\rm m}(4)$. Highlevel theoretical calculations using various methods,^{47,83,98,99} are also in good



Fig. 6 Structure of propan-2-ylium ion, 4, optimized at the MP2(full)/6-31G(d) level.

agreement with these results. They are presented in Table 1. $S^{\circ}_{m}(4)$ given in Table 1 is computed at the E3 level.⁴⁷

The classical structures of propan-1-ylium ion (1-propyl cation) are not minima on the potential energy surface of the $C_3H_7^+$ manifold.^{22,98} They are some 20 kcal mol⁻¹ above **4**, as determined computationally.⁹⁸ This result is in fair agreement with gas-phase photoionization experiments on the *n*-propyl radical by Schultz,¹⁰³ and Dyke and co-workers.¹⁰⁴ These structures are transition states involved in the hydrogen scrambling of **4** in superacidic media.^{22,105}

Butan-2-ylium ion, sec-butyl cation (sec- $C_4H_9^+$) (5)

Experimental evidence (essentially IR and NMR data) in solution at low temperatures, summarized in Ref. 106, as well as high-level *ab initio* calculations¹⁰⁶ indicate that this ion exists as a mixture of structures I and II, of, respectively, C_2 and C_1 point group symmetries (Fig. 7).

The GB of *E*-C₄H₈ has been determined experimentally by ICR, using bracketing techniques.¹⁰⁷ We have carried out G2 calculations providing computed GB values for *E*-C₄H₈ and leading to structures **I** and **II**. These calculations show: (1) that the thermodynamic stabilities *of both* structures are extremely close and, more precisely, that a gaseous sample of *sec*-C₄H₉⁺ in thermal equilibrium at 298.15 K is very nearly a 1:1 mixture of isomers **I** and **II**; (2) that these computed GB values are in extremely good agreement with experiment. We have combined the experimental GB, $\Delta_f H^{\circ}_m$ and S°_m for *E*-C₄H₈ and the computed S°_m values for structures **I** and **II** to derive the corresponding values for $\Delta_f H^{\circ}_m(5)$ (Table 1). Arnett and co-workers^{108,109} used low-temperature calorimetry in superacids to determine the standard enthalpy change for the isomerization *sec*-C₄H₉⁺ \rightarrow *tert*-C₄H₉⁺. This datum, combined with $\Delta_f H^{\circ}_m(tert$ -C₄H₉⁺), provides an "apparent" value (assuming negligible differential solvation effects) of $\Delta_f H^{\circ}_m(5)$ quite consistent with the independent results reported above for the gas phase.



Fig. 7 Isomeric structures I and II of butan-2-ylium ion, 5, optimized at the MP2(full)/6-31G(d) level.



Fig. 8 Structure of 2-methylpropan-2-ylium ion, 6, optimized at the MP2(full)/6-31G(d) level.

The primary cations $n-C_4H_9^+$ and $iso-C_4H_9^+$ are not on the minima on the PES of the $C_4H_9^+$ manifold.¹⁰⁶ The stability of these species can be estimated from vertical ionization potentials of appropriate substrates.¹¹⁰ The corresponding $\Delta_f H^\circ_m$ values are some 30 kcal mol⁻¹ more positive than $\Delta_f H^\circ_m (tert-C_4H_9^+)$.

2-Methylpropan-2-ylium ion, tert-butyl cation (tert- $C_4H_9^+$) (6)

Solid state nutation NMR spectroscopy¹¹¹ and X-ray diffraction,¹¹² show that this ion has a trigonal structure with 1.46 and 2.51 Å for central-methyl and C–C methyl–methyl distances (Fig. 8).

Remarkably coincident experimental values for $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{6})$ can be obtained from: (1) PEPICO studies on *tert*-butyl iodide;¹¹³ (2) appearance energies for the formation of **6** from isobutane, neopentane and *tert*-butyl iodide (threshold photoionization mass spectrometry);¹¹⁴ (3) the PA of *iso*-butene.⁸¹ They are presented in Table 1, together with $S^{\circ}_{\rm m}(\mathbf{6})$ determined from the experimental values of PA and GB(*iso*-C₄H₈),¹¹⁵ together with $\Delta_{\rm f} H^{\circ}_{\rm m}$ (*iso*-C₄H₈) and $S^{\circ}_{\rm m}(iso$ -C₄H₈).

G2 values for the PA and GB of *iso*-C₄H₈, particularly with entropies treated according to Ref. 47, lead to thermodynamic data in excellent agreement with experiment. Recent very high-level calculations¹¹⁶ on the *tert*-butyl radical also provide extremely consistent results. Our *ab initio* [MP2(full)/6-31G(d)] geometries are in excellent agreement with experimental data.

ALIPHATIC CARBOCATIONS WITH MORE THAN FOUR CARBON ATOMS

2-Methylbutan-2-ylium ion $(tert-C_5H_{11}^+)$ (7)

To our knowledge, there are no X-ray data on the structure of this ion. Its ¹³C NMR spectrum in solution has long been known.¹¹⁷

The experimental $\Delta_{\rm f} H^{\circ}_{\rm m}(7)$ and $S^{\circ}_{\rm m}(7)$ values reported in Table 2 were obtained from the experimental standard enthalpy and Gibbs energy changes for the hydride exchange between ions 6 and 7 (reaction (21)) as determined by pulsed HPMS:^{118,119}

$$iso-C_5H_{12}(g) + (6)(g) \rightarrow (7)(g) + iso-C_4H_{10}(g) \qquad \Delta_r G^{\circ}(21), \ \Delta_r H^{\circ}(21)$$
(21)

The structure of this ion, shown in Fig. 9, was fully optimized at the MP2(full)/6-31G(d) level (this work) and belongs to the C_s symmetry point group. Schleyer and coworkers¹²⁰ pointed out two important properties of this structure: (1) the lengthening of the C3–C4 bond to *ca*. 1.58 Å, indicative of hyperconjugation; (2) the reduction of the C4–C3–C2⁺ angle to 101.5°, showing distortion of the C–C hyperconjugating form toward bridging. This implies some "non-classical" character for **7**.

The computed ¹³C NMR shifts for this structure are in excellent agreement with experiment. This is not the case for other conformations, close in energy but not displaying this incipient bridging.¹²⁰

We have computed $\Delta_r G^{\circ}(21)$ and $\Delta_r H^{\circ}(21)$ at the G2(MP2) level. They are in excellent agreement with experiment and lead to $\Delta_f H^{\circ}_m(7)$ and $S^{\circ}_m(7)$ values, also given in Table 2.

2-Methylpentan-2-ylium ion (tert- $C_6H_{13}^+$) (8)

To our knowledge, no experimental structural data are available for this species. The experimental $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{8})$ and $S^{\circ}_{\rm m}(\mathbf{8})$ were obtained from the experimental standard enthalpy and Gibbs energy changes for the hydride exchange between ions **6** and **8**, reaction (22a), as determined by pulsed HPMS:^{119,121}

$$C_6H_{14}(g) + (6)(g) \rightarrow (8)(g) + iso-C_4H_{10}(g) \qquad \Delta_r G^{\circ}(22a)$$
 (22a)

Fully independent values of $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{8})$ and $S^{\circ}_{\rm m}(\mathbf{8})$, also reported in Table 2, were obtained as follows:

(1) We applied the DPA technique to 2-chloro-2-methylpentane and to other tertiary alkyl chlorides.⁶⁸ These data lead to $\Delta_r G^{\circ}(22b)$:

$$tert-C_6H_{13}Cl(g) + (\mathbf{6})(g) \rightarrow (\mathbf{8})(g) + tert-C_4H_9Cl(g) \qquad \Delta_r G^{\circ}(22b)$$
(22b)

Upon the appropriate leaving group corrections, this immediately provides $\Delta_r G^{\circ}(22a)$. This datum, corrected for the entropy terms, yields $\Delta_r H^{\circ}(22a)$, which in turn, upon combination with the experimental $\Delta_f H^{\circ}_m$ for 2-methylpentane, **6** and *iso*-C₄H₁₀(g) leads to a value of $\Delta_f H^{\circ}_m(\mathbf{8})$ in very good agreement with the results presented above.

(2) All the species involved in reaction (22a) were studied at the MP2/6-311G(d,p) level in Ref. 68; $\Delta_r H^{\circ}(22a)$ obtained from these calculations provides a value of $\Delta_f H^{\circ}_m(\mathbf{8})$ in satisfactory agreement with experiment.

| Cations | $\Delta_{\rm f} H^{\circ}{}_{\rm m}({ m g})^a$ | $S^{\circ}_{m}(g)^{b}$ |
|--|--|--|
| 2-Methylbutan-2-ylium ($tert$ -C ₅ H ⁺ ₁₁) (7) | 162.4 ± 2.0^{c} 164.1 ± 2.0^{d} | 83.3 ± 2.0^{c} 85.2 ± 2.0^{e} |
| 2-Methylpentan-2-ylium ($tert$ -C ₆ H ⁺ ₁₃) (8) | $\begin{array}{c} 156.2 \pm 2.0^{f} \\ 155.0 \pm 3.0^{h} \\ 154.3 \pm 2.0^{j} \end{array}$ | 92.2 ± 2.0^{g} 91.1 ± 2.0^{i} |
| 2,4-Dimethylpentan-2-ylium ($C_7H_{15}^+$) (9) | 147.4 ± 2.1^k 146.7 ± 2.0^m | 96.1 ± 2.0^{l} |
| 2,4,4-Trimethylpentan-2-ylium ($C_8H_{17}^+$) (10) | $\begin{array}{c} 139.8 \pm 2.1^n \\ 140.5 \pm 2.1^p \\ 139.2 \pm 2.1^q \end{array}$ | $101.0 \pm 2.0^{\circ}$ |
| 3-Ethyl-5,5-dimethylhexan-3-ylium $(C_{10}H_{21}^+)$ (11) | $128.3 \pm 2.1^{r} \\ 129.0 \pm 2.1^{t} \\ 126.8 \pm 2.0^{u}$ | 113.2 ± 2.0^{s} |
| 2,2,4,6,6-Pentamethylheptan-4-ylium ($C_{12}H_{25}^+$) (12) | 112.0 ± 2.1^{v} 113.3 \pm 2.1 ^x 110.8 \pm 2.0 ^y | 125.1 ± 2.0 ^w |

 Table 2
 Thermodynamic data for selected aliphatic carbocations with five and more carbon atoms

^{*a*}In kcal mol⁻¹.

^bIn cal mol⁻¹ K⁻¹.

^cFrom $\Delta_r H^{\circ}(21) = -2.6 \pm 1.0$ and $\Delta_r G^{\circ}(21) = -1.9 \pm 0.5$ kcal mol⁻¹, respectively; averages of data from Refs. 118 and 119. $\Delta_f H^{\circ}_{m}(C_5 H_{12}) = -36.73 \pm 0.14$ kcal mol⁻¹ (Ref. 43) and $S^{\circ}(C_5 H_{12})$, 82.2 cal mol⁻¹ K⁻¹.²⁹

^{*d*}From $\Delta_r H^{\circ}(21) = -0.93 \pm 1.0 \text{ kcal mol}^{-1}$ at the G2(MP2) level (this work).

^{*e*}Scaled HF/6-31G(d) value from the calculations in (d).

^fFrom $\Delta_r H^{\circ}(22a) = -4.2 \pm 1.0$ and $\Delta_r G^{\circ}(22a) = -4.1 \pm 0.5$ kcal mol⁻¹, respectively; averages of data from Refs. 119 and 121. $\Delta_f H^{\circ}_{m}(C_6H_{14}) = -41.66 \pm 0.25$ kcal mol⁻¹ (Ref. 43) and $S^{\circ}(C_6H_{14}) = 91.1$ cal mol⁻¹K⁻¹.²⁹

^{*g*}From the data in (f).

^hBased on $\Delta_r H^{\circ}(22a) = -5.4 \pm 2.8 \text{ kcal mol}^{-1}$ from DPA experiments⁶⁸ treated as indicated in the text. ⁱScaled HF/6-31G(d) value from the calculations in Ref. 68.

^{*j*}From the MP2/6-311G(d,p) value of $\Delta_r H^{\circ}(22a)$.⁶⁸

^kDetermined from the DPA results for chlorides⁶⁸ with the appropriate leaving group and entropy corrections indicated in the text.

¹Scaled, computed HF/6-31G(d) value from data in Ref. 68.

^mBased on the MP2/6-311G(d,p) data anchored, to the experimental $\Delta_f H^o{}_m[C_7H_{16}(g)]$, -48.30 ± 0.23 kcal mol⁻¹.²⁶

"Determined from the DPA results for chlorides⁶⁸ with the appropriate leaving group and entropy corrections indicated in the text.

^oScaled, computed HF/6-31G(d) value from data in Ref. 68.

 p As in (n) but based on the DPA of the corresponding alcohol.

 qBased on the MP2/6-311G(d,p) data anchored to the experimental $\Delta_f H^\circ_m[C_8H_{18}(g)], -53.57\pm0.32~kcal~mol^{-1}.^{26}$

⁷Determined from the DPA results for chlorides⁶⁸ with the appropriate leaving group and entropy corrections indicated in the text.

^sScaled, computed HF/6-31G(d) value from data in Ref. 68.

^tAs in (a) but based on the DPA of the corresponding alcohol.

"Based on the MP2/6-311G(d,p) data anchored to the experimental $\Delta_f H^o_m[C_{10}H_{22}(g)]$, $-62.72 \pm 0.50 \text{ kcal mol}^{-1}$.²⁸

 $^{\nu}$ Determined from the DPA results for chlorides⁶⁸ with the appropriate leaving group and entropy corrections indicated in the text.

^wScaled, computed HF/6-31G(d) value from data in Ref. 68.

^xAs in (v) but based on the DPA of the corresponding alcohol.

^yBased on the MP2/6-311G(d,p)//MP2/6-31G(d) data anchored to the experimental $\Delta_f H^{\circ}_{m}[C_{12}H_{26}(g)]$, -75.43 ± 0.50 kcal mol^{-1.28}

Notice in Fig. 10 that the C3–C4 bond is elongated as in the case of 7. This can also be taken as implying some contribution from hyperconjugation.¹²⁰

2,4-Dimethylpentan-2-ylium ($C_7H_{15}^+$) (9), 2,4,4-trimethylpentan-2-ylium ($C_8H_{17}^+$) (10), 3-ethyl-5,5-dimethylhexan-3-ylium ($C_{10}H_{21}^+$) (11) and 2,2,4,6,6-pentamethylheptan-4-ylium ($C_{12}H_{25}^+$) (12) ions

To our knowledge, no experimental structural information is available for these species (Chart 1).

Experimental thermodynamic information on these ions was obtained by DPA using chlorides and alcohols as substrates.⁶⁸ Treatment of the experimental data by



Fig. 9 Structure of 2-methylbutan-2-ylium ion, 7, optimized at the MP2(full)/6-31G(d) level.



Fig. 10 Structure of 2-methylpentan-2-ylium ion, **8**, optimized at the MP2(full)/6-31G(d) level.

the methods indicated above led to the $\Delta_{\rm f} H^{\circ}_{\rm m}$ values summarized in Table 2. Relevant structural features of these ions, as determined by *ab initio* methods shall be reported in due course. Here, we notice that the experimental $\Delta_{\rm f} H^{\circ}_{\rm m}$ values are in good agreement with the MP2/6-311G(d,p) data, as shown in Table 2.



Chart 1

CYCLIC SPECIES WITHOUT FORMAL π SYSTEMS

Cyclopropylium ion $(c-C_3H_5^+)$ (13)

In principle, the species of $C_{2\nu}$ symmetry,^{99,122} Fig. 11, can be formed in the gas phase by protonation of cyclopropene²¹ and by photoionization of the cyclopropyl radical.¹⁰⁴

These methods lead to rather different values of $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{13})$. Thus, using $\Delta_{\rm f} H^{\circ}_{\rm m}(c\text{-}\mathrm{C}_{3}\mathrm{H}_{4}) = 66.22 \pm 0.60 \text{ kcal mol}^{-1}$ (Ref. 27) and PA(*c*-C₃H₄) = -195.6 kcal mol⁻¹, (Ref. 43) we get $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{13}) \approx 236.3 \text{ kcal mol}^{-1}$. Combining the ionization energy of the cyclopropyl radical, $8.18 \pm 0.03 \text{ eV}$,¹⁰⁴ with $\Delta_{\rm f} H^{\circ}_{\rm m}(c\text{-}\mathrm{C}_{3}\mathrm{H}_{5}) = 63.7 \pm 2.7 \text{ kcal mol}^{-1}$ based on the experimental ICR data from Ref. 123, we obtain $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{13}) \approx 252.7 \text{ kcal mol}^{-1}$.

High-level *ab initio* calculations fully rationalize these discrepancies. Thus, it has been shown that the cyclic $C_{2\nu}$ structure is not a minimum on the potential energy surface of the c-C₃H₅⁺ manifold.^{124,125} We have confirmed at the MP2(full)/6-311 + G(d,p) level that this is indeed the case. As indicated by Glukhovtsev *et al.*¹²² the cyclic $C_{2\nu}$ structure appears as a shallow minimum at the HF/6-31G(d) level. We took advantage of this to estimate the PA of c-C₃H₄ at the standard G2 level. The value we obtain, 171.3 kcal mol⁻¹, puts $\Delta_f H^{\circ}_m(13)$ at about 260.6 kcal mol⁻¹. Notice that the meaning of the apparent PA(c-C₃H₄) has been recently re-interpreted.¹²⁶

All these facts are perfectly consistent with solution chemistry results showing the essentially barrierless electrocyclic disrotatory ring opening of cyclopropyl cations



Fig. 11 Structure of cyclopropylium ion, **13**, optimized at the HF/6-31G(d) level. This structure is not a minimum at correlated levels.

to yield allylic cations in solvolytic processes,¹²⁷ this being a classical example of a pericyclic reaction.¹²⁸ To our knowledge, only a single-substituted cyclopropylium ion has been observed by NMR (in SbF₅–SO₂ClF solution between -120 and -60° C), the 11-methyl-tricyclo[4.4.1.0^{1.6}]undec-11-yl cation.¹²⁹

Cyclobutylium $(c-C_4H_7^+)$ (14) and cyclopropylmethylium $(c-C_3H_5CH_2^+)$ (15) ions

Because of their relevance in physical organic chemistry a great deal of experimental information is available on these species, an excellent summary being given in Ref. 131. According to high-level calculations, ^{130,131} **14** has a puckered structure (symmetrical, non-classical bicyclobutonium, C_s point group), as shown in Fig. 12.

It is known experimentally¹³⁰ that in solution at low temperature, **14** is in equilibrium with the bisected (C_s symmetry point group) **15**, (Fig. 13), and that the interconversion barrier is small (*ca.* 2 kcal mol⁻¹).

Notice that the bisected structure of **15** is taken to reflect the stabilizing interaction $^{132-134}$ between the empty p orbital of the CH₂⁺ moiety and the Walsh orbital $^{135-139}$ of the cyclopropyl ring.

As regards the thermodynamic state functions for the equilibrating mixture of these two ions in the gas phase, we believe that the only piece of information available so far is the GB of cyclobutene.²⁶ This datum, combined with the experimental $S_{m}^{\circ}(c-C_{4}H_{6})$ and the computed $S_{m}^{\circ}(14)$ (this work) leads to $\Delta_{f}H_{m}^{\circ}(14)$ (Table 3).

Our G2 calculations provide PA and GB values for c-C₄H₆ to yield **14** which, once combined with the experimental thermodynamic data for c-C₄H₆, lead to the



Fig. 12 Structure of cyclobutylium ion, 14, optimized at the MP2(full)/6-31G(d) level.



Fig. 13 Structure of cyclopropylmethylium ion, 15, optimized at the MP2(full)/6-31G(d) level.

value of $\Delta_{\rm f} H^{\circ}_{\rm m}(14)$ reported in Table 3. As can be seen, the agreement with experiment is only moderately good.

High-level *ab initio* calculations^{130,131} have already indicated that species **14** and **15** have very close thermodynamic stabilities. Our own investigations show that, at the G2 level, $\Delta_r H^{\circ}(23)$ and $\Delta_r G^{\circ}(23)$ are, respectively, equal to -0.5 and $0.0 \text{ kcal mol}^{-1}$:

$$(15) \rightarrow (14) \qquad \Delta_{\rm r} H^{\circ}(23), \ \Delta_{\rm r} G^{\circ}(23) \tag{23}$$

That $\Delta_r G^{\circ}_{m}(23) \approx 0$ kcal mol⁻¹ is in remarkably good agreement with the results of solvolytic studies.^{130,140} This and $\Delta_r H^{\circ}_{m}(23) = -0.5$ kcal mol⁻¹ imply that in solution at -132° C, an equilibrating mixture of these cations contains about 72 and 28%, respectively, of isomers **14** and **15**, in very fair agreement with experiment.¹³⁰

It has just been reported¹⁴¹ that two different $C_4H_7^+$ ions can be generated in the gas phase from cyclobutyl and cyclopropylcarbinyl chlorides. When sampled shortly (*ca.* 10^{-6} s) after their formation and in the absence of excitation, they display different patterns of reactivity, quite consistent with the structures given above for **14** and **15**.

Cyclopentylium ion $(c-C_5H_9^+)$ (16)

This ion is known in solution at low temperature and has been studied in superacid media¹⁴² and even at 70 K in the solid state.¹⁴³

| Cations | $\Delta_{\rm f} H^{\circ}{}_{\rm m}({ m g})^a$ | $S^{\circ}_{m}(g)^{b}$ |
|---|--|------------------------|
| Cyclobutylium, c -C ₄ H ₇ ⁺ (14) | $216.3 \pm 2.2^{c} \\ 212.2 \pm 2.2^{e,f} \\ 212.0 \pm 2.3^{f,g}$ | 67.6 ± 2.0^{d} |
| Cyclopropylmethylium, c -C ₃ H ₅ CH ₂ ⁺ (15) | $\begin{array}{l} 218.4 \pm 2.2^{h} \\ 214.4 \pm 2.2^{f,i} \\ 213.0 \pm 2.3^{f,k} \end{array}$ | 69.5 ± 2.0^{j} |
| Cyclopentylium, c -C ₅ H ₉ ⁺ (16) | $\begin{array}{c} 193.1 \pm 2.0^{l} \\ 193.7 \pm 2.0^{m} \\ 191.9 \pm 2.0^{o} \end{array}$ | 69.7 ± 2.0^{n} |

 Table 3
 Thermodynamic data for cyclic carbocations C4 and C5

^{*a*}In kcal mol⁻¹.

 b In cal mol⁻¹ K⁻¹.

^cFrom the experimental GB(c-C₄H₆), 180.1 kcal mol⁻¹ (Ref. 21), $\Delta_{\rm f} H^{\circ}_{\rm m}(c$ -C₄H₆) = 37.5 ± 0.4 kcal mol⁻¹ (Ref. 26) and $S^{\circ}_{\rm m}(c$ -C₄H₆) = 62.6 ± 2.0 cal mol⁻¹K⁻¹ (Ref. 30) and the computed [MP2(FC)/6-311G(d,p)], scaled $S^{\circ}_{\rm m}$ (14), 67.6 ± 2.0 cal mol⁻¹K⁻¹.

^dComputed, scaled MP2(FC)/6-311G(d,p) value.

^eFrom the computed PA(*c*-C₄H₆) to yield **14**, 190.8 kcal mol⁻¹ and the experimental $\Delta_f H^{\circ}_{m}(c$ -C₄H₆).

^fThese values seem more reliable than those based on the experimental GB(c-C₄H₆).

^gG2-computed value.

^hFrom the experimental GB(*c*-C₄H₆), $\Delta_{f}H^{\circ}_{m}(c$ -C₄H₆) and $S^{\circ}_{m}(c$ -C₄H₆), and the computed [MP2(FC)/6-311G(d,p)], scaled $S^{\circ}_{m}(15)$, 69.5 ± 2.0 cal mol⁻¹ K⁻¹ and assuming $\Delta_{f}G^{\circ}_{m}(14) = \Delta_{f}G^{\circ}_{m}(15)$, on the basis of G2 calculations.

^{*i*}From the G2-computed PA(*c*-C₄H₆) to yield **15**, 190.3 kcal mol⁻¹ and the experimental $\Delta_t H^{\circ}_{m}(c$ -C₄H₆). ^{*j*}Computed, scaled MP2(FC)/6-311G(d,p) value.

^{*k*}G2-computed value.

¹From the experimental $\Delta_f H^{\circ}_{m}(c-C_5H_8) = -18.36 \pm 0.36 \text{ kcal mol}^{-1}$ (Ref. 26), $S^{\circ}_{m}(c-C_5H_8) = 69.2 \text{ cal mol}^{-1}\text{K}^{-1}$ (Ref. 30), $\Delta_r H^{\circ}_{m}(24)$ and $\Delta_r G^{\circ}_{m}(24)$ from Ref. 149 and the appropriate data for *iso*-C₃H₇⁺(g) (Table 1).

^{*m*}As above, but using data from Refs. 117, 147 and 148.

ⁿScaled, symmetry-corrected value from the calculations in (o).

^oAs above, using our computed G2(MP2) values of $\Delta_r H^{\circ}_{m}(24)$ and $\Delta_r G^{\circ}_{m}(24)$.

¹H and ¹³C NMR spectra show **16** to have a highly fluxional structure, this being a consequence of very rapid 1,2-hydride shifts.^{144,145}

Its structure in the gas phase, as obtained at various correlated *ab initio* levels¹⁴⁶ is twisted (C_2 symmetry), as shown in Fig. 14. This is important because: (1) It shows, as shown in Fig. 14, that the C(1)–C(2)–H_a and C(1)–C(2)–H_e angles are quite different and the C–H_a bond lengths are relatively large, as they fit to partial hydrogen bonding and to a hyperconjugative effect. The same holds for the hydrogens bound to C(5); (2) this structure is able to account for the experimental ¹³C NMR spectra of **16** in solution and in the solid state.

Experimental standard thermodynamic state functions for this ion in the gas phase can be obtained from the following sources: the GB of cyclopentene, an ICR



Fig. 14 Structure of cyclopentylium ion, 16, optimized at the MP2(full)/6-31G(d) level.

bracketing value from Ref. 107 and the standard Gibbs energy change and enthalpy for reaction $(24)^{121,148,149}$ as determined by HPMS:

$$iso-C_3H_7^+(g) + c-C_5H_{10}(g) \rightarrow C_3H_8(g) + c-C_5H_9^+(g)$$
 (24)

These thermodynamic data are collected in Table 3, together with the results of our G2(MP2) calculations. The agreement is reasonable.

For the purposes of linking the present results to those obtained by the DPA technique, the standard Gibbs energy change for reaction (25), a chloride ion exchange between a carbocation (R^+) (**16** in this case) and 1-adamantyl cation (1-Ad⁺), is necessary. We obtained this datum by combining the experimental $\Delta_r G^{\circ}(24)$ with the computed value of the standard Gibbs energy change for the chlorine exchange between 1-adamantyl chloride and cyclopentane, no experimental data being available for cyclopentyl chloride. The result is reported in Table 4:

$$1-\text{AdCl}(g) + R^+(g) \rightarrow 1-\text{Ad}^+(g) + \text{RCl}(g) \qquad \Delta_r G^\circ(25)$$
(25)

Notice that in some cases, tert-C₄H₉Cl has been used instead of 1-AdCl as a reference.

CYCLOPROPYL-SUBSTITUTED CARBOCATIONS

The stabilizing interactions between the empty p orbital in the methyl cation and the Walsh orbital of cyclopropane reported in the case of **15** are also present in other cognate ions. Some examples are as follows.

| Cation | $\Delta_{\rm r} G^{\circ}{}_{\rm m}(25)$ |
|---|--|
| Methylium (CH_3^+) (1) | -77.4 ± 2.8^{a} |
| Propan-2-ylium $(iso-C_3H_7^+)$ (4) | -22.7 ± 3.0^{a} |
| Butan-2-ylium (sec- $C_4H_9^+$) (5) | -18.9 ± 3.0^{a} |
| 2-Methylpropan-2-ylium (tert- $C_4H_9^+$) (6) | -6.0 ± 1.0^{b} |
| 2-Methylpentan-2-ylium $(tert-C_6H_{13}^+)$ (8) | -1.1 ± 2.8^{c} |
| 2.4-Dimethylpentan-2-vlium $(C_7H_{15}^+)$ (9) | 1.9 ± 2.8^{c} |
| 2.4.4-Trimethylpentan-2-vlium $(C_8H_{17}^+)$ (10) | 4.9 ± 2.8^{c} |
| 3-Ethyl-5.5-dimethylhexan-3-ylium ($C_{10}H_{21}^+$) (11) | 7.6 ± 2.8^{c} |
| $2.2.4.6.6$ -Pentamethylheptan-4-ylium (C ₁₂ H $_{25}^+$) (12) | $13.3 + 2.8^{\circ}$ |
| Cyclopropylmethylium $(c-C_2H_5-CH_2^+)$ (15) | $-7.1 + 2.0^{d}$ |
| Cyclopentylium $(c-C_{c}H_{c}^{+})$ (16) | -136 ± 2.5^{a} |
| 1-Cyclopropylethan-1-ylium $(c-C_2H_z-CHCH_z^+)$ (17) | $20 + 20^d$ |
| 2-Cyclopropylpropan-2-ylium $(c-C_2H_{c-C}(CH_2)^{+})$ (18) | $142 + 20^d$ |
| Dicyclopropylmethylium $[(c-C_0H_c)_2-CH^+]$ (19) | 12.0 ± 2.0^{d} |
| 1 1-Dicyclopropylethan-1-ylium $[(c-C_2H_2)_2CCH_2^+]$ (20) | 22.0 = 2.0 $22.1 + 2.0^{d}$ |
| Tricyclopropyletiai Γ ynum [($c = C_3 H_5$)/ $c = 13$] (20) | 22.1 = 2.0 $29.0 + 3.0^{\circ}$ |
| Norbornan-1-vlium $(1-Nh^+)$ (C_2H^+) (22) | $-27.3 \pm 2.8^{\circ}$ |
| Norbornan-2-ylium $(2 - Nb^+)$ $(C - H^+)$ (23) | -3.0 ± 1.0^{b} |
| Norbornan-2-ylium $(2-Nb^+)$ $(C_{2}H_{11}^+)$ (23) | $-222 + 28^{c}$ |
| Bicyclo[2,2,2]octan-1-ylium $(C_0H_{10}^+)$ (26) | $-74 + 28^{\circ}$ |
| Noradamantan-3-vlium $(C_{co}H_{co}^+)$ (27) | -174 ± 2.6 |
| $\Delta damantan_1-vlium (C_{10}H_{13}^+)$ (28) | 0.00 (by definition) |
| Adamantan-2-vlium ($C_{10}H_{+5}^{+}$) (20) | $-7.6 \pm 2.8^{\circ} (-11.3 \pm 3.0)^{\circ}$ |
| $\operatorname{Bicyclo[3,3,2]decan-1-vlium}(C_{10}H_{12}^{+})(30)$ | $10.4 + 3.3^{\circ}$ |
| Homoadamantan-3-vlium (C_1, H_2^+) (31) | $47 + 28^{c}$ |
| Homoadamantan-4-vlium (C_1, H_7^+) (32) | $-0.9 + 3.0^{\circ}$ |
| Bicyclo[3 3 3]undecan-1-ylium (C_1, H_{+}^+) (33) | $13.2 + 2.8^{\circ}$ |
| [1]Diadamantan-1-vlium ($C_{10}H_{\pi}^+$) (34) | 15.2 = 2.0 $16.7 + 2.8^{\circ}$ |
| C_{2} Cyclopropenvlium (C_{2} H ⁺ ₂) (38) | $52 + 20^{\circ}$ |
| 1 2-Dipropyleyclopropenylium $[c_{2}C_{2}H(C_{2}H_{2})^{+}]$ (40) | 3.2 = 2.0 $32.3 + 3.4^{c}$ |
| Tripropyleyclopropenylium $[c-C_2(C_2H_2)^+]$ (41) | 32.5 = 5.1 $41.5 + 3.4^{c}$ |
| Norbornen-7-vlium $(C-H_{2}^{+})$ (43) | 13 ± 28^{c} |
| Norbornadien-7-vlium $(C_7H_7^+)$ (44) | $69 + 28^{\circ}$ |
| Cyclohentatrienvlium (Tronvlium) $(C_2H_2^+)$ (45) | $29.8 + 2.0^d (32.3 + 3.4)^c$ |
| Benzylium $(C_2H_2^+)$ (46) | -58 ± 20^{b} |
| 4-Nitrobenzylium (4-NO ₂ -C ₂ H ₂ -CH ⁺) | -172 ± 20^{b} |
| 4 -Methovybenzylium (4 -MeO-C $_{c}$ H $_{c}$ CH ⁺) | $77 + 20^{b}$ |
| 1-Phenylethan-1-ylium ($C_0H_0^+$) (47) | 1.7 ± 2.0^{b} |
| $2 - [A - Methovynhenvl]nronan - 2 - vlium [A - MeO - C - H - C(CH -)^+]$ | 1.7 = 2.0 16.7 ± 2.0^{b} |
| $2-[4-Wethoxyphenyl]propan-2-ylium [4-Web-C_6f15C(CH_3)_2]$ $2-[3-Nitrophenyl]propan-2-ylium [3-NO-C-H_C(CH_3)_2]$ | $-32 + 20^{b}$ |
| 2-[5-1110pnon31]propan-2-ylium $(C_1H^+_{-1})$ (48) | 5.2 ± 2.0 6.4 ± 1.0^{b} |
| Diphenylmethanylium $(C_{10}H_{+}^{\pm})$ (40) | 13.4 ± 3.0^{f} |
| 1 1-Diphenvlethan 1-vlium (C_1, H^+) (50) | 13.7 ± 3.0^{f} |
| Triphenylmethanylium ($C_{14}H_{13}$) (50) | 25.5 ± 5.0 28 4 ± 3.0^{f} |
| riphonymiculallyllulli (C191115) (31) | 20.4 ± 3.0 |

Table 4 Standard Gibbs energy changes for reaction (25), $\Delta_r G^{\circ}_{m}(25)$

All values in kcal mol^{-1} . Specific details for each ion are given in the text.

^{*a*}From the $\Delta_f H^{\circ}_m(g)$ and $\mathcal{S}^{\circ}_m(g)$ values for ions, as reported in the various tables. Whenever a choice was possible, the most accurate value was used. Data for 1-AdCl and **28** are from Ref. 64. Data for other neutral species are either experimental or computed as indicated in the text.

^bDirect experimental value (equilibrium techniques).

^cFrom DPA experiments and, when necessary, the appropriate leaving group correction for chloride, (see text).

^dCalculated at the G2(MP2) level.

^eWith leaving group correction for OTs.

^{*f*}From the computed $\Delta_{f}H^{\circ}_{m}(g)^{155}$ and the leaving group correction for chloride.

1-Cyclopropylethan-1-ylium ion $(c-C_3H_5CHCH_3^+)$ (17)

Compound **17** has been obtained at low temperature in superacid media.¹⁵⁰ Both its ¹H and ¹³C NMR spectra are known.¹⁵¹

NMR evidence indicates that it has a bisected structure (C_s symmetry), the most stable isomer having the methyl group in position *trans* relative to the ring (see Fig. 15).

 $\Delta_r H^{\circ}_{m}(17)$ as reported in Table 5 was obtained from the experimental standard enthalpy of formation³¹ and the PA of vinylcyclopropane as given in Ref. 43, based on (ICR) GB data from Taft's group.

G2(MP2) calculations (this work) provide thermodynamic data also reported there. Interestingly, the computed $\Delta_r H^{\circ}_m$ value for gaseous vinylcyclopropane is in excellent agreement with the literature data.^{31,35} The structure for the *trans* form of the ion at the MP2(full)/6-31G(d) level is portrayed in Fig. 15. $\Delta_r G^{\circ}_m(25)$ is given in Table 4.

2-Cyclopropylpropan-2-ylium ion $(c-C_3H_5C(CH_3)_2^+)$ (18)

This ion has been generated in solution at low temperature in superacid medium.¹⁵⁰ The ¹H and ¹³C NMR spectra of this and cognate ions are reported and discussed in Refs. 151 and 152.

Theoretical calculations, such as those performed in this work, lead to the structure presented in Fig. 16, in which the plane defined by the carbons of the 2-propyl moiety is a perpendicular bisector of the cyclopropyl ring, as in the case of the 1-cyclopropylethyl cation. This is an important result, in full agreement with the NMR spectrum of the ion. Furthermore, rotation around the $C(1)-C(\alpha)$ bond involves a barrier of *ca.* 13.7 kcal mol^{-1.153} These structural features, as well as those presented by **15** and **17**, are consequences of the stabilizing interaction between the strained C–C bonds of the cyclopropane moiety and the formally empty p orbital of the alkyl moiety.^{135–140}

The value of $\Delta_{\rm f} H^{\circ}_{\rm m}(18)$ given in Table 5 was obtained from the experimental PA of 2-propenylcyclopropane. This value is that recommended in Ref. 43, based on scaled data from Taft's group and Ref. 21. We also report $\Delta_{\rm f} H^{\circ}_{\rm m}[c-C_3H_5C(CH_3)_2^+]$ from our G2(MP2) calculations. $\Delta_{\rm r} G^{\circ}_{\rm m}(25)$ is given in Table 4.

Dicyclopropylmethylium ion $[(c-C_3H_5)_2CH^+]$ (19)

The ¹H NMR spectrum of **19** in solution in superacid media was reported in 1970.¹⁵³ As far as we know, its structure in the solid state is not yet available. The X-ray



Fig. 15 Structure of 1-cyclopropylethan-1-ylium ion, 17, optimized at the MP2(full)/6-31G(d) level.

structure of the hexafluoroantimonate of hydroxydicyclopropylmethylium is known, however, ¹⁵⁴ and shows that both rings adopt an essentially "bisected" structure. The structure of **19**, optimized at the MP2(full)/6-31G(d) level, has C_2 symmetry (Fig. 17) and also shows this feature.

No thermodynamic data seem to be available for this species in the gas phase. The data reported in Table 5 were obtained computationally at the MP4(SDQ)/6-31G(d)//MP2(full)/6-31G(d) level.¹⁵⁵ The computed $\Delta_r G^{\circ}(25)$ is given in Table 4.

1,1-Dicyclopropylethan-1-ylium ion $[(c-C_3H_5)_2CCH_3^+]$ (20)

We are not aware of experimental structural data on this species. The structure we present here (Fig. 18) is that optimized at the MP2(full)/6-31G(d) level and has a C_s symmetry clearly showing the bisected positions of the cyclopropyl rings.

The value of $\Delta_{f}H^{\circ}_{m}[(c-C_{3}H_{5})_{2}CCH_{3}^{+}]$ given in Table 5 was obtained from the experimental PA of $[(c-C_{3}H_{5})_{2}C=CH_{2}]$ and the computed (this work) $\Delta_{f}H^{\circ}_{m}(g)$ for

| Cations | $\Delta_{\rm f} H^{\circ}{}_{\rm m}({ m g})^a$ | $S^{\circ}_{m}(g)^{b}$ |
|---|--|------------------------|
| 1-Cyclopropylethan-1-ylium $(c-C_3H_5CHCH_3^+)$ (17) | $\begin{array}{l} 201.1 \pm 2.1^{c} \\ 197.6 \pm 2.1^{e,f} \\ 194.5 \pm 2.1^{f,g} \end{array}$ | 77.6 ± 2.0^{d} |
| 2-Cyclopropylpropan-2-ylium $(c-C_3H_5C(CH_3)_2^+)$ (18) | 178.8 ± 2.1^{h} 180.0 ± 2.1^{i} 181.2 ± 2.1^{k} | 84.6 ± 2.0^{j} |
| Dicyclopropylmethylium ((c -C ₃ H ₅) ₂ CH ⁺) (19) | 217.5 ± 2.0^{l} | 82.0 ± 2.0^{m} |
| 1,1-Dicyclopropylethan-1-ylium ($(c-C_3H_5)_2CCH_3^+$) (20) | 199.3 ± 4.0^{n} | 92.9 ± 2.0^o |
| Tricyclopropylmethylium ((c -C ₃ H ₅) ₃ C ⁺) (21) | 232.0 ± 4.0^{p} | 97.8 ± 2.0^{q} |

 Table 5
 Thermodynamic data for selected cyclopropyl-containing carbocations

^{*a*}In kcal mol⁻¹.

^bIn cal mol⁻¹ K⁻¹. ^cFrom the experimental PA of vinylcyclopropane,⁴³ 195.1 \pm 2.0 kcal mol⁻¹ and $\Delta_{\rm f} H^{\circ}_{\rm m}(c-{\rm C}_{3}{\rm H}_{5}{\rm CHCH}_{2}) =$

 $30.40 \pm 0.30 \text{ kcal mol}^{-1.35}$

^dScaled HF/6-31G(d) value from this work.

^{*e*}From the computed (G2MP2) PA of c-C₃H₅CHCH₂, 199.0 kcal mol⁻¹ and the experimental $\Delta_f H^{\circ}_{m}(c$ -C₃H₅CHCH₂).

^fValues more reliable than those based on the experimental PA of vinylcyclopropane. See Ref. 126. ^gFrom the computed $\Delta_r H^\circ_m$ for the hydride exchange between **17** and **4** together with the experimental value of $\Delta_r H^\circ_m (c-C_3H_5CH_2CH_3)$.

^hFrom the experimental PA of 2-propenylcyclopropane,⁴³ 208.3 \pm 2.0 kcal mol⁻¹ and $\Delta_f H^{\circ}_{\text{m}}(c-C_3H_5\text{CCH}_2\text{CH}_3) = 21.4 \pm 0.5$ kcal mol⁻¹ from Refs. 31 and 35.

^{*i*}From the computed (G2MP2) PA of *c*-C₃H₅CCH₂ CH₃, 207.0 kcal mol⁻¹ and the experimental and $\Delta_f H^{\circ}_{m}(c$ -C₃H₅CCH₂CH₃).

^{*j*}Scaled HF/6-31G(d) value extracted from the G2(MP2) calculation.

^{*k*}From the computed [G2(MP2)] $\Delta_r H^{\circ}_{m}$ for the hydride exchange between 2-cyclopropyl-2-propyl cation and *tert*-butyl cation and the experimental $\Delta_t H^{\circ}_{m}$ values for all the other species.

¹Computed at the G2(MP2) level (this work).

^mScaled HF/6-31G(d) value from this work.

^{*n*}From the experimental PA[(*c*-C₃H₅)₂C=CH₂], 216.2 kcal mol⁻¹⁴³ and the computed $\Delta_f H^{\circ}_{m}[(c-C_3H_5)_2C=CH_2]$ (see text).

^oScaled, symmetry-corrected HF/6-31G(d) value from this work.

^{*p*}From the DPA onset of tricyclopropylcarbinol and the computed $\Delta_{f}H^{\circ}_{m}[(c-C_{3}H_{5})_{3}CH]$ (this work).

^qScaled, symmetry-corrected HF/6-31G(d) value from this work.

this species. The former is taken from Ref. 43 and is based on experimental ICR data from Taft's group and Ref. 21. $\Delta_f H^\circ_m[(c-C_3H_5)_2C=CH_2]$ was estimated through the combination of experimental $\Delta_f H^\circ_m$ (vinylcyclopropane, g) and the enthalpy of the isodesmic reaction (26) computed at the MP4(SDQ)/6-31G(d)//MP2(full)/6-31G(d) level.¹⁵⁶ It is difficult to assess the uncertainty of this value, but it might reach some 2–3 kcal mol⁻¹. The computed $\Delta_r G^\circ_m(25)$ is given in Table 4:

$$(c-C_3H_5)_2C=CH_2(g) + C_2H_4(g) \rightarrow 2c-C_3H_5CH=CH_2$$
 (26)



Fig. 16 Structure of 2-cyclopropylpropan-2-ylium ion, 18, optimized at the MP2(full)/6-31G(d) level.

Tricyclopropylmethylium ion $[(c-C_3H_5)_3C^+]$ (21)

This is a remarkably stable ion, already observed in 1955.¹⁵⁷ Its first ¹H NMR spectrum (in H_2SO_4 at room temperature!) was reported in 1962¹⁵⁸ and so were a little later, the spectra of a variety of cyclopropyl carbocations in superacid media.¹⁵⁹ Its ¹³C spectrum has been carefully analyzed.^{160,161}

The structure computed at the MP2(full)/6-31G(d) level (this work) and portrayed in Fig. 19 is propeller-shaped and has C_{3h} symmetry. It obviously minimizes internal repulsions between the rings while allowing for maximal simultaneous overlap between the empty p orbital of the central carbon and the Walsh orbitals of the rings. This situation is to be compared to that prevailing in the trityl cation, which is much less favorable.

No thermodynamic data for this ion in the gas phase seem to be available so far. We have carried out DPA studies of tricyclopropyl carbinol and its methyl ether. This provides $\Delta_f H^{\circ}_{m}(21)$. Unfortunately, the associated uncertainty is somewhat



Fig. 17 Structure of dicyclopropylmethylium ion, **19**, optimized at the MP2(full)/6-31G(d) level.

large because of the unavailability of experimental enthalpies of formation for tricyclopropylmethane and other relevant species. As in the previous case, no G2-type calculations could be performed on these species because of their size. Instead, the enthalpy of the isodesmic reaction (27) was computed at the MP4(SDQ)/6-31G(d)//MP2(full)/6-31G(d) level¹⁵⁶ and combined with the experimental value of $\Delta_{\rm f} H^{\circ}_{\rm m} (c-C_3 H_5 CH_3)$ to obtain $\Delta_{\rm f} H^{\circ}_{\rm m} [(c-C_3 H_5)_3 CH]$. The computed $\Delta_{\rm r} G^{\circ}_{\rm m}(25)$ is given in Table 4:

$$(c-C_3H_5)_3CH(g) + 2CH_4(g) \rightarrow 3c-C_3H_5CH_3(g)$$
 (27)

SECONDARY AND TERTIARY CARBOCATIONS DERIVED FROM CAGE HYDROCARBONS

Norbornan-1-ylium ion $(1-Nb^+)$ (C₇H₁₁⁺) (22)

To our knowledge, this highly strained $ion^{57,58,162,163}$ has not been observed in solution or in the solid state. It is known that in solution under stable ion



Fig. 18 Structure of 1,1-dicyclopropylethan-1-ylium ion, **20**, optimized at the MP2(full)/6-31G(d) level.

conditions, it is not observed and it rearranges to yield a 2-norbornyl cation.¹⁶⁴ No high-level *ab initio* studies seem to have been published by other workers on this species. Figure 20 portrays its optimized structure at the MP2(full)/6-31G(d) level.

The stability of this ion was experimentally estimated by DPA^{57,58} using as substrates the corresponding chloride and bromide. The experimental $\Delta_r G^{\circ}(25)$, is reported in Table 4.

The DPA onset for 1-NbCl, subject to leaving group and entropy corrections similar to those used for **6**, led to the value of $\Delta_f H^{\circ}(22)$ reported in Table 6. This table also presents $\Delta_f H^{\circ}(22)$ obtained at the G2(MP2) level.

Norbornan-2-ylium ion $(2-Nb^+)$ (C₇H⁺₁₁) (23)

Few species have raised so much interest and for such a long period of time as $23.^{165-170}$



Fig. 19 Structure of tricyclopropylmethylium ion, **21**, optimized at the MP2(full)/6-31G(d) level.

¹³C and ¹H NMR spectra of this ion in superacid media at low temperatures, ^{171,172} as well as in the solid state at 5 K, ¹⁷³ strongly indicate that this ion has the "non-classical" structure depicted in Fig. 21.

The IR¹⁷⁴ and ESCA¹⁷⁵ spectra of this ion at low temperature also support this contention.

High-level *ab initio* calculations in Ref. 176 and references therein agree with these results. Our own G2(MP2) data, based on this non-classical structure, lead to energetic values in excellent agreement with experiment (see Table 6). It is interesting that these concepts have been challenged very recently.¹⁷⁷

The experimental thermodynamic state functions presented in Table 6 have been obtained from the following sources: (1) the averaged, selected experimental values of PA and GB for norbornene;⁴³ (2) the experimental HPMS standard Gibbs energy change for the hydride exchange between *tert*-butyl and 2-norbornyl cations;¹⁴⁹ (3) the standard Gibbs energy change for the chloride exchange between *tert*-butyl and 2-norbornyl cations as determined from DPA results^{57,58} (see Table 4) and the appropriate leaving group and entropy corrections. All these results are extremely self-consistent.



Fig. 20 Structure of norbornan-1-ylium ion, 22, optimized at the MP2(full)/6-31G(d) level.

Norbornan-7-ylium ion $(7-Nb^+)$ (C₇H⁺₁₁) (24)

To our knowledge, this ion has not been observed in solution in superacid media although its IR spectrum, using the SbF₅ matrix technique at very low temperatures, has been obtained from various precursors, including 7-norbornyl chloride.^{178,179} These experiments, together with high-level *ab initio* calculations,^{179,180} indicate that **24** has the non-classical (C_1 symmetry) structure portrayed in Fig. 22.

7-Norbornyl chloride was subject to DPA treatment in the gas phase.¹⁸¹ As in the cases discussed above, its reaction onset was used to estimate $\Delta_r G^{\circ}(25)$ (Table 4). Also, applying the same kind of leaving group and entropy corrections, the value of $\Delta_f H^{\circ}_{m}(24)$ reported in Table 6 was obtained. $\Delta_f H^{\circ}_{m}(24)$ computed at the G2(MP2) level is also given in Table 6.

Cubylium ion $(C_8H_7^+)$ (25)

The stability of this ion (Chart 2) was obtained by direct bromide exchange with 3noradamantyl cation, prepared in turn by DPA of the 3-bromonoradamantane.^{57,58} For the sake of consistency with the treatment in Refs. 57 and 58, $\Delta_{\rm f} H^{\circ}_{\rm m}(25)$ reported in Table 6 was anchored to that for the 1-adamantyl cation, using for the

| Cations | $\Delta_{\mathrm{f}} H^{\mathrm{o}}{}_{\mathrm{m}}(\mathrm{g})^{a}$ | $S^{\circ}_{m}(g)^{b}$ |
|--|--|------------------------|
| Norbornan-1-ylium ($C_7H_{11}^+$) (22) | 208.7 ± 3.1^{c} 204.8 ± 2.0^{e} | 77.0 ± 2.0^{d} |
| Norbornan-2-ylium ($C_7H_{11}^+$) (23) | 186.1 ± 2.1^{f} 185.8 ± 2.1^{g} 186.3 ± 3.0^{i} 187.1 ± 2.0^{j} | 78.8 ± 2.0^{h} |
| Norbornan-7-ylium ($C_7H_{11}^+$) (24) | $205.6 \pm 3.0^{k} \\ 203.2 \pm 2.0^{l}$ | 76.4 ± 2.0^{m} |
| Cubylium $(C_8H_7^+)$ (25) | 360.2 ± 3.8^{n} 358.9 ± 3.2^{o} | 85.6 ± 2.0^{p} |
| Bicyclo[2.2.2]octan-1-ylium ($C_8H_{13}^+$) (26) | 174.2 ± 3.0^{q} 176.1 ± 3.0^{r} | 78.7 ± 2.0^{s} |
| Adamantan-1-ylium ($C_{10}H_{15}^+$) (28) | $162.6 \pm 3.0^{t} 164.1 \pm 3.2^{u} 162.9 \pm 3.2^{v} 160.7 \pm 3.7^{x} $ | 81.07 ± 2.0^{w} |
| Adamantan-2-ylium $(C_{10}H_{15}^+)$ (29) | 171 ± 3^{y} 171.9 ± 1.1^{z} 172.1 ± 1.1^{ab} | 84.45 ± 2.0^{aa} |
| Bicyclo[3.3.2]decan-1-ylium ($C_{10}H_{17}^+$) (30) | $\frac{160.5 \pm 3.5^{ac}}{161.6 \pm 3.0^{ad}}$ | 85.3 ± 2.0^{ae} |
| Bicyclo[3.3.3]undecan-1-ylium ($C_{11}H_{19}^+$) (33) | $\frac{161.7 \pm 3.0^{af}}{160.2 \pm 3.0^{ag}}$ | 86.6 ± 2.0^{ah} |

 Table 6
 Thermodynamic data for secondary and tertiary carbocations derived from cage hydrocarbons

^{*a*}In kcal mol⁻¹.

^{*b*}In cal mol⁻¹ K⁻¹.

^cFrom the DPA results on 1-NbCl and *tert*-BuCl with the computed leaving group and entropy corrections^{57,58} and the $\Delta_{\rm f} H^{\circ}_{\rm m}$ values for *tert*-C₄H₉Cl and *iso*-C₄H₁₀, respectively, -42.99 ± 0.50 and -32.25 kcal mol⁻¹.²⁶ $\Delta_{\rm f} H^{\circ}_{\rm m}$ (6) is from Table 1.

^eG2(MP2) computed value (this work).

^{*f*}From the averaged, selected experimental values of PA and GB for norbornene⁴³ and the experimental $\Delta_f H^{\circ}_{m}(C_7H_{10})$, 19.7 \pm 0.50 kcal mol⁻¹ (Ref. 26) and $S^{\circ}(C_7H_{10}) = 74.3 \pm 2.0$ cal mol⁻¹ K⁻¹ (Walsh, R. and Wells, J. M. (1976) *J. Chem. Thermodyn.* 8, 55).

^jAs in (f) but using the computed [G2(MP2)] PA and GB of norbornene.

^dScaled value at the HF/6-31G* level.

^gFrom the experimental HPMS¹⁴⁹ standard Gibbs energy change for the hydride exchange between *tert*butyl and 2-norbornyl cations with scaled entropy corrections for all species from our G2(MP2) calculations.

^hScaled value at the HF/6-31G* level.

^{*i*}From the standard Gibbs energy change for the chloride exchange between *tert*-butyl and 2-norbornyl cations as determined from DPA results^{57,58} and the appropriate leaving group and entropy corrections, the latter as in (g).

^{*k*}From the standard Gibbs energy change for the chloride exchange between *tert*-butyl and 7-norbornyl cations as determined from DPA results¹⁸¹ with the appropriate leaving group corrections (at the MP2(full)/6-31G(d) level). Entropy corrections are scaled, values at the HF/6-31G(d) level. Ancillary experimental data for the other species are from Tables 1 and 3. $\Delta_f H^{\circ}_{m}$ (norbornane) = 13.30 ± 0.50 kcal mol⁻¹.²⁶

^{*l*}G2(MP2) value.

^{*m*}The value of $S^{\circ}(24)$ is a scaled MP2(FU)/6-31G(d) value from this work.

^{*n*}Experimental value obtained as indicated in the text. Leaving group correction at the HF/6-31G(d) level. $\Delta_f H^{\circ}_{m}(adamantane) = -32.0 \pm 0.50 \text{ kcal mol}^{-1},^{26} \Delta_f H^{\circ}_{m}(cubane) = 148.7 \pm 1.0 \text{ kcal mol}^{-1},^{26} \Delta_f H^{\circ}_{m}(1-adamantyl) = -162.0 \pm 2.0 \text{ kcal mol}^{-1},^{64}$

^{*o*}Computed at the MP2/6-311G(d,p) level. Anchored to experimental data as indicated in the text. ^{*p*}Symmetry-corrected HF/6-31G(d) value.⁵⁷

^{*q*}From the standard Gibbs energy change for the chloride exchange between *tert*-butyl and bicyclo[2.2.2]octyl cations as determined from DPA results⁵⁷ and the appropriate leaving group corrections (at the HF/6-31G(d) level). Entropy corrections at the HF/6-31G(d) level. Ancillary experimental data for the other species are from Tables 1 and 3.

^{*r*}Computed values at the MP2/6-311G(d,p) level for the hydride exchange between *tert*-butyl and bicyclo[2.2.2]octyl cations and anchored to the experimental data for *tert*-butyl cation and *iso*-butane. Entropy corrections at the HF/6-31G(d) level.

^sSymmetry-corrected HF/6-31G(d) value.⁵⁷

^tFrom HPMS data¹⁸⁶ for hydride exchange between *tert*-Bu⁺ and 1-Ad⁺ together with data from Ref. 26 for the neutral species.

"From the same source, for the homologous chloride anion exchange, together with $\Delta_f H^{\circ}_{m}(tert-C_4H_9Cl) = 42.99 \pm 0.50 \text{ kcal mol}^{-1}$ (Ref. 26) and $\Delta_f H^{\circ}_{m}(1-\text{AdCl}) = -42.45 \pm 0.60 \text{ kcal mol}^{-1}$.⁶⁴

 $^{\nu}$ From the DPA and FT ICR equilibrium study of the same process with the entropy corrections (scaled, symmetry-corrected HF/6-31G(d) values) from Ref. 64.

^wScaled, symmetry-corrected HF/6-31G(d) value from Ref. 64.

^{*x*}Average of the G2(MP2) calculations for hydride and chloride exchanges, anchored to the experimental $\Delta_f H^o_{m}(g)$ values for the neutral species.

^yFrom Ref. 189.

^zFrom Ref. 190 and ancillary data from Ref. 192.

^{aa}Computed [HF/6-31G(d)], scaled value.

^{*ab*}G2(MP2) value anchored to experimental data for *iso*- $C_3H_7^+$ (Table 1) and *iso*- C_3H_7 (Ref. 26), see Ref. 192.

^{ac}From the standard Gibbs energy change for the chloride or hydroxyl exchanges between *tert*-butyl and bicyclo[3.3.2]decyl cations as determined from DPA results⁵⁷ and the appropriate leaving group corrections [at the HF/6-31G(d) level]. Entropy corrections are symmetry-corrected values at the HF/6-31G(d) level. Ancillary experimental data for the other species are from Tables 1 and 3. $\Delta_f H^{\circ}_m(g)$ for bicyclo[3.3.2]decane is -25.3 ± 0.50 kcal mol⁻¹.²⁶

^{*ad*}Computed values at the MP2/6-311G(d,p) level for the hydride exchange between *tert*-butyl and bicyclo[3.3.2]decyl cations and anchored to the experimental data for *tert*-butyl cation and *iso*-butane. Entropy values are symmetry-corrected at the HF/6-31G(d) level.

^{ae}Computed [HF/6-31G(d)] value.

^{*af*}From the averaged standard Gibbs energy changes for the chloride or hydroxyl exchanges between *tert*butyl and bicyclo[3.3.3]undecyl cations as determined from DPA results⁵⁷ and the appropriate leaving group corrections [at the HF/6-31G(d) level].⁵⁷ Entropy corrections are symmetry-corrected values at the HF/6-31G(d) level. Ancillary experimental data for the other species are from Tables 1 and 3. $\Delta_f H^{\circ}_m(g)$ for bicyclo[3.3.3]undecane is -21.2 ± 0.50 kcal mol⁻¹.²⁶

^{*ag*}Computed values at the MP2/6-311G(d,p) level for the hydride exchange between *tert*-butyl and bicyclo[3.3.3]undecyl cations and anchored to the experimental data for *tert*-butyl cation and *iso*-butane. Entropy values are symmetry-corrected at the HF/6-31G(d) level.

^{ah}Computed [HF/6-31G(d)] value.



Fig. 21 Structure of norbornan-2-ylium ion, 23, optimized at the MP2(full)/6-31G(d) level.

latter $\Delta_{\rm f} H^{\circ}_{\rm m} = 162.0 \pm 2.0 \text{ kcal mol}^{-1}.^{64}$ We also report the results computed at the MP2/6-311G(d,p) level.^{57,58}

Bicyclo[2.2.2]octan-1-ylium ion ($C_8H_{13}^+$) (26)

 $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{26})$ (see Chart 2) presented in Table 6 is based on the DPA study of the 1chlorobicyclo[2.2.2]octane⁵⁷ (see also Table 4). These results were shown to be in excellent agreement with those obtained from the corresponding bromide.⁵⁸ Experimental data were treated exactly as in the case of **25**. Computational results at the MP2/6-311G(d,p) level are also given in Table 6.

Noradamantan-3-ylium ion $(C_{10}H_{13}^+)$ (27)

The $\Delta_r G^{\circ}(25)$ value given in Table 4 was obtained from the DPA study of 3-noradamantyl bromide with the appropriate leaving group corrections^{57,58} (see Chart 2).



Fig. 22 Structure of norbornan-7-ylium ion, 24, optimized at the MP2(full)/6-31G(d) level.

Adamantan-1-ylium ion $(C_{10}H_{15}^+)$ (28)

This ion (see Chart 2) is of both historical and chemical importance. It was one of the first tertiary carbocations to be identified in solution in superacid media,¹⁸² studied by ¹H and ¹³C NMR in solution,¹⁸³ prepared as a stable solid salt,¹⁸⁴ and characterized by its IR spectrum in cryogenic SbF₅ matrices.¹⁸⁵ It was amongst the first ions to be studied by ICR through halide exchange reactions^{50,186} and also the first tertiary carbocation to undergo a systematic treatment by means of DPA.⁵⁴

 $\Delta_{\rm f} H^{\circ}{}_{\rm m}(28)$, presented in Table 6, is based on the most recent HPMS and FT ICR studies by hydride and halide exchanges^{58,60,187} (see Table 4). These results have been shown to be quite consistent with the G2(MP2) studies of both kinds of processes.⁶⁴ The value of $\Delta_{\rm f} H^{\circ}{}_{\rm m}(28)$ used here is 162.0 ± 2.0 kcal mol⁻¹, simultaneously consistent with these experimental and computational data.

Adamantan-2-ylium ion $(C_{10}H_{15}^+)$ (29)

As far as we know, this ion (Chart 2) has not been observed in solution or in the solid state. It has, however, been studied in the gas $phase^{186}$ and computationally at the MP2/6-31G(d)//6-31G(d) + ZPVE6-31G(d) level.¹⁸⁸



Chart 2

Thermodynamic experimental data for this ion comes from three sources: (1) its appearance energy from a variety of precursors;¹⁸⁹ (2) the adiabatic ionization energy of the 2-adamantyl radical;¹⁹⁰ (3) the DPA study of 2-adamantyl chloride and 2-adamantyl alcohol.¹⁹¹

On the basis of a G2(MP2) study of 1- and 2-adamantyl radicals and cations, ¹⁹² the results by Aubry *et al.*¹⁸⁹ and Kruppa and Beauchamp¹⁹⁰ seem quite consistent,¹⁹¹ they are also in very good agreement with DPA studies of 2-adamantanol and 2-adamantyl chloride. The results are summarized in Table 6 (see also Table 4).

A value for $\Delta_f H^{\circ}_{m}(29)$ of 171.9 \pm 2.0 kcal mol⁻¹ is simultaneously consistent with these experimental and computational data.

Bicyclo[3.3.2]*decan-1-ylium ion* ($C_{11}H_{17}^+$) (**30**)

The thermodynamic data for this ion presented in Table 6 were obtained from the DPA study of the corresponding alcohol and chloride.^{57,58} The average $\Delta_r G^{\circ}(25)$ is given in Table 4. Leaving group and entropy corrections were applied as in previous cases. Values obtained at the MP2/6-311G(d,p) level are reported in Table 4.

Homoadamantan-3-ylium ion $(C_{11}H_{17}^+)$ (31)

This ion (see Chart 2) was obtained from the DPA of the corresponding chloride and alcohol.^{57,58} The value of $\Delta_r G^{\circ}(25)$ given in Table 4 is the average of both results (with the appropriate leaving group correction).

Homoadamantan-4-ylium ion $(C_{11}H_{17}^+)$ (32)

This ion (Chart 2) was obtained from the DPA of the corresponding chloride.¹⁹³ The experimental $\Delta_r G^{\circ}(25)$ value is given in Table 4.

Bicyclo[3.3.3]undecan-1-ylium ion ($C_{10}H_{17}^+$) (33)

This ion was obtained from the DPA of the corresponding chloride and alcohol.^{57,58} The value of $\Delta_r G^{\circ}(25)$ given in Table 4 is the average of both highly consistent results (with the appropriate leaving group correction). $\Delta_f H^{\circ}_m(33)$ derived therefrom is given in Table 6 together with the results obtained at the MP2/6-311G(d,p) level.

Diadamantan-1-ylium ion $(C_{19}H_{27}^+)$ (34)

This ion was obtained from the DPA of the corresponding chloride and alcohol.⁵⁸ The value of $\Delta_r G^{\circ}(25)$ given in Table 4 is the average of both highly consistent results. Because of the size of the system, leaving group corrections were computed at the AM1 level.

CARBENIUM IONS WITH FORMAL π SYSTEMS

Vinylium ion $(C_2H_3^+)$ (35)

Experimental values for the standard enthalpy of formation of this ion, as reported in Table 7, are in fair agreement. They originate in: (1) the proton affinity of acetylene¹⁹⁴ obtained in SIFT experiments; and (2) the adiabatic ionization energy of the vinyl radical¹⁹⁵ combined with its standard enthalpy of formation.²⁵

We have obtained $\Delta_f H^{\circ}_{m}(g)$ values for $C_2H_3^+$ by combining the standard enthalpy of protonation of acetylene as computed at the G2 level (this work) with the experimental $\Delta_f H^{\circ}_{m}(C_2H_2)$. Also used was the computed (G2) ionization energy of vinyl radical together with the experimental value of $\Delta_f H^{\circ}_{m}(C_2H_3)$ to obtain $\Delta_f H^{\circ}_{m}(C_2H_3^+)$. These values are presented in Table 7. (see also Ref. 196).

Experimental spectroscopic studies in the millimeter wavelength region^{3,197,198} provided rotational constants and the inertial defect for this ion. These data show that the ground state of this species is planar and has a non-classical $C_{2\nu}$ structure (1, Fig. 23). Near-IR studies of $C_2H_3^+$ led to results^{199,200} also in agreement with this non-classical structure. Furthermore, the splitting of the spectral lines was interpreted in terms of the tunneling of protons between the unequivalent axial and apical positions. This tunneling takes place through the classical structure **2** (Fig. 23). From the most recent work,¹⁹⁸ the difference in stability between these structures has been estimated at 4.7 kcal mol⁻¹.

The computed difference in standard enthalpies of formation between structures **1** and **2** amounts to 4.3 kcal mol⁻¹. High-level structural studies²⁰¹ reveal a small upward tilting (0.3°) of the axial hydrogens with respect to the internuclear C–C

| Cations | $\Delta_{\mathrm{f}} H^{\circ}{}_{\mathrm{m}}(\mathrm{g})^{a}$ | $S^{\circ}_{m}(g)^{b}$ |
|---|--|------------------------|
| Vinylium $(C_2H_3^+)$ (35) | $266.6 \pm 3.2^{c} \\ 269.9 \pm 1.9^{d} \\ 265.7 \pm 2.5^{f} \\ 266.8 \pm 2.0^{g}$ | 52.4 ± 2.0^{e} |
| 2-Propargylium $(C_3H_3^+)$ (36) | $280.8 \pm 1.1^h \\ 279.6 \pm 2.0^j$ | 59.3 ± 2.0^{i} |
| 2-Propenylium, allyl $(C_3H_5^+)$ (37) | $228.9 \pm 0.6^{k} 226.0 \pm 2.0^{l} 228.7 \pm 2.0^{n}$ | 60.4 ± 2.0^{m} |

Table 7 Thermodynamic data for non-aromatic carbocations with formal π bonds

^{*a*}In kcal mol⁻¹.

^bIn cal mol⁻¹ K⁻¹.

^cFrom the experimental PA(C₂H₂), $153.3 \pm 3.2 \text{ kcal mol}^{-1}$ (Ref. 194) and $\Delta_f H^{\circ}_{m}(C_2H_2)$, 54.190 kcal mol⁻¹.²⁶

^dFrom the adiabatic ionization energy of vinyl radical, 8.59 ± 0.03 eV (Ref. 195) and $\Delta_f H^{\circ}_{\text{m}}(\text{C}_2\text{H}_3) = 71.5 \pm 1.5 \text{ kcal mol}^{-1}.^{25}$ See also Berkowitz, J., Ellison, G. B. and Gutman, D. (1994) *J. Phys. Chem.* 98, 2744 for a detailed discussion of the matter.

^eSymmetry-corrected, scaled value at the HF/6-31G(d) level (this work).

^fFrom the G2-computed ionization energy of vinyl radical (194.1 kcal mol⁻¹ at 0 K) and the experimental $\Delta_{f}H^{o}_{m}(C_{2}H_{3})$ (this work).

^{*g*}From the G2-computed PA(C₂H₂) (this work) (153.1 kcal mol⁻¹) and the experimental $\Delta_{f}H^{\circ}_{m}(C_{2}H_{2})$. ^{*h*}From the ionization energy of the propargyl radical²⁰⁷ and $\Delta_{f}H^{\circ}_{m}(C_{3}H_{3}) = 81.0 \pm 1.0$ kcal mol⁻¹ from Ref. 25. Thermal corrections from this work.

^{*i*}Scaled, corrected HF/6-31G(d) value from this work.

^jThis work, using the computed (G2) ionization energy of C₃H₃ (8.673 eV)²⁰⁷ and $\Delta_{\rm f} H^{\circ}_{\rm m}$ (C₃H₃) = 81.0 ± 1.0 kcal mol⁻¹.²⁵ Thermal corrections from this work.

^{*k*}From Ref. 212.

¹From the experimental $\Delta_t H^{\circ}_{m}(g)$ of allene, 45.6 ± 0.2 kcal mol⁻¹ (Ref. 27) and its experimental PA, 185.3 kcal mol⁻¹.¹⁹¹

^mHF/6-31G(d) value from this work, scaled and symmetry corrected.

^{*n*}This work, using the experimental $\Delta_f H^{\circ}_{m}(g)$ for allene and its G2-calculated PA (184.0 kcal mol⁻¹).

axis. Theory and computation on vinyl cations have been reviewed.²⁰² Studies on medium effects on acetylene and vinyl cation have been reported recently.²⁰³

Ab initio molecular dynamics^{204,205} calculations performed on vinyl cation confirm that the ground-state structure of $C_2H_3^+$ is best described as a quasi-planar bridged structure broadened significantly by anisotropic delocalization of the protons due to zero point motion.

2-Propargylium ion $(C_3H_3^+)$ (36)

To our knowledge, no direct experimental spectroscopic data are available for the propargyl cation. Very large scale *ab initio* calculations have been carried out on both this ion and on the propargyl radical.²⁰⁶ They lead to vibrational frequencies for the latter in excellent agreement with experiment and to an adiabatic ionization


Fig. 23 Structure of vinylium ion, 35, optimized at the MP2(full)/6-31G(d) level. Structure 1 is the absolute minimum. Structure 2 is a transition state.

potential for this radical (8.650 eV) pleasantly close to the experimental value (8.673 eV).^{207,208} $\Delta_{\rm f} H^{\circ}_{\rm m}(36)$ (Table 7) can be obtained experimentally from the standard enthalpy of formation of the propargyl radical²⁵ and the zero kinetic energy photoelectron spectrum (ZEKE) of this species.²⁰⁷

Recent calculations at the CBS-Q level⁹⁹ as well as our present G2 computations lead to values of $\Delta_{\rm f} H^{\circ}_{\rm m}(\mathbf{36})$ in excellent agreement with experiment (Table 7). They all confirm the $C_{2\nu}$ symmetry of **36** (Fig. 24).

2-Propenylium ion, allyl cation $(C_3H_5^+)$ (37)

According to careful *ab initio* calculations, this species is planar and has a $C_{2\nu}$ symmetry (Fig. 25).^{209,210} According to Ref. 210, this planarity seems to be enforced by the σ framework although the contribution from the π conjugation is also relevant.

Based on the adiabatic ionization energy of the allyl radical as determined by Houle and Beauchamp²¹¹ and on its own work, the Boulder group²¹² determined a value of $\Delta_f H^{\circ}_{m}(\mathbf{37})$ in remarkable agreement with that obtained much earlier by Lossing²¹³ by means of appearance energies. $\Delta_f H^{\circ}_{m}(\mathbf{37})$ can also be derived from the experimental GB of allene (upon appropriate correction of entropy effects; see Table 7) obtained by ICR, using a bracketing technique.²¹⁴ $\Delta_f H^{\circ}_{m}(\mathbf{37})$ computed at the CBS-Q level⁹⁹ is in very nice agreement with the datum from Ref. 212. So is our own G2 result (see Table 7).

TWO-ELECTRON AROMATIC AND HOMOAROMATIC IONS

Cyclopropenylium ion $(C_3H_3^+)$ (**38**)

This species, an "archetypal aromatic cation"¹²² has a planar, fully symmetrical structure (D_{3h} symmetry point group) (Fig. 26) as confirmed by the systematic



Fig. 24 Structure of 2-propargylium ion, 36, optimized at the MP2(full)/6-31G(d) level.



Fig. 25 Structure of 2-propenylium ion, 37, optimized at the MP2(full)/6-31G(d) level.



Fig. 26 Structure of cyclopropenylium ion, 38, optimized at the MP2(full)/6-31G(d) level.

experimental study of the IR and Raman spectra of some of its salts (in liquid sulfur dioxide solution and in polycrystalline state).²¹⁵

 $\Delta_{\rm f} H^{\circ}_{\rm m}(38)$ has been obtained experimentally through the determination of the appearance potential of this ion from a variety of neutral precursors including several molecules.²¹³ These studies lead to the values reported in Table 8.

No experimental value of $S^{\circ}_{m}(38)$ seems to be presently available. That given in Table 8 was calculated in this work.

Compound **38** has been the subject of a large number of theoretical studies, the most recent ones being listed in Refs. 216–218. The relevant C–C and C–H distances, as obtained at the MP4/6-311G 4^* + level,²¹⁶ are, respectively,

| Cations | $\Delta_{\rm f} {H^{\rm o}}_{\rm m}({ m g})^a$ | $S^{\circ}_{m}(g)^{b}$ |
|--|--|------------------------|
| Cyclopropenylium $(C_3H_3^+)$ (38) | $\begin{array}{c} 257.0 \pm 2.0^{c} \\ 256.7 \pm 2.0^{e} \\ 257.2 \pm 2.0^{f} \end{array}$ | 54.8 ± 2.0^{d} |
| Propylcyclopropenylium $(c-C_3H_2-C_3H_7^+)$ (39) | 222.5 ± 2.9^{g} | 85.43 ± 2.0^{h} |
| Cyclobutenylium $(C_4H_5^+)$ $(c-C_4H_5^+)$ (42) | 210.1 ± 2.9^{i} | 59.6 ± 2.0^{j} |
| Norbornen-7-ylium $(C_7H_9^+)$ (43) | 215.7 ± 3.3^k 215.2 ± 2.6^m | 74.2 ± 2.0^{l} |
| Norbornadien-7-ylium $(C_7H_7^+)$ (44) | 247.9 ± 3.8^n 249.9 ± 2.0^p | $71.4 \pm 2.0^{\circ}$ |
| Cycloheptatrienylium, tropylium $(C_7H_7^+)$ (45) | 211.4 ± 3.8^{q} 209.8 ± 2.0 ^s | 69.8 ± 2.0^{r} |

Table 8 Thermodynamic data for aromatic and homoaromatic two- and six- π -electron carbenium ions

^{*a*}In kcal mol⁻¹.

^bIn cal mol⁻¹ K⁻¹.

^cFrom appearance energies.²¹³

^dFrom the scaled, symmetry-corrected HF/6-31G* value taken from the G2 calculation (this work).

^eFrom G2 computations.⁹⁹

^fFrom CBS-Q calculations.⁹⁹

^jSymmetry-corrected HF/6-31G(d) value taken from the G2(MP2) calculation (this work).

^kObtained from the DPA study of the corresponding alcohol and chloride.¹⁸²

^gComputed at the G2(MP2) level (this work); see text.

^hSymmetry-corrected HF/6-31G(d) value taken from the G2(MP2) calculation (this work).

^{*i*}Computed at the G2 level (this work); see text.

¹Scaled HF/6-31G(d) value taken from the G2(MP2) calculation (this work).

^{*m*}Computed at the G2(MP2) level (this work).

ⁿObtained from the DPA study of the corresponding alcohol.¹⁸²

^oScaled HF/6-31G(d) value taken from the G2(MP2) calculation (this work).

^{*p*}Computed at the G2(MP2) level (this work).

^qFrom DPA studies.¹⁵⁶

^rScaled, symmetry-corrected HF/6-31G(d) value (this work).

^sComputational value from Ref. 242.

 1.363_2 and 1.079_5 Å. While the latter is close to that in benzene, the former is significantly shorter (C–C bond length of 1.397 Å for benzene). The various vibrational frequencies, including anharmonicity effects and Fermi resonance pairs for this ion and all its deuterated isotopomers, have been computed at extremely high levels and are in excellent agreement with experiment whenever a comparison has been possible.^{216,217}

The fact that the C–C bond lengths are shorter and the C–C stretching frequencies and the delocalization energies per π electron are higher for cyclopropenium than for benzene, is taken as an indication of "superaromaticity".^{122,215}

The $\Delta_{\rm f} H^{\circ}_{\rm m}(38)$ values computed using G2 methodology¹²² and Petersson's complete basis technique (CBS-Q)⁹⁹ are nicely consistent and in excellent agreement with experiment (see Table 8).

1-Propylcyclopropen-2-ylium ion $(c-C_3H_2C_3H_7^+)$ (**39**)

To our knowledge, no experimental structural data seem to be available for this ion. The value of $\Delta_f H^o_m(39)$ reported in Table 8 was obtained computationally at the G2(MP2) level. The fully optimized structure (of C_s symmetry) portrayed in Fig. 27 derives from these calculations.



Fig. 27 Structure of 1-propylcyclopropenylium ion, 39, optimized at the MP2(full)/6-31G(d) level.

1,2-Dipropylcyclopropen-3-ylium $[c-C_3H(C_3H_7)_2^+]$ (**40**) and tripropylcyclopropenylium $[c-C_3(C_3H_7)_3^+]$ (**41**) ions

Again, no structural data seem to be available for these species. Their optimized structures [MP2(full)/6-31G(d) level; this work], of C_s symmetry, are given in Figs. 28 and 29.

Compounds **40** and **41** were generated by DPA of the corresponding precursors,¹⁵⁶ namely the methyl ethers of 1,2-dipropylcyclopropen-3-ol and tripropylcyclopropen-1-ol. These onsets were anchored to that of the methyl ether of 1,1,1-tricyclopropylcarbinol. An independent onset for 1,1,1-tricyclopropylcarbinol is available so that the onsets of **40** and **41** can be ultimately referred to that of the 1-adamantyl cation. These results, upon the appropriate leaving group correction, provide the values of $\Delta_r G^{\circ}(25)$ given in Table 4.

Cyclobutenylium ion $(c-C_4H_5^+)$ (42)

In classical studies, Olah and co-workers²¹⁹ obtained this cation in solution and were able to show that at -110° C it has a puckered structure (Fig. 30) displaying true aromatic delocalization. Compound **42** is thus the simplest homoaromatic system.^{220–222}

This aromatic character was confirmed by an extensive series of high-level quantum mechanical studies (structure, energetics and NMR chemical shifts) by Schleyer and co-workers.²²³

G2 studies have been recently performed on 42 and cyclobutadiene.²²⁴

To our knowledge, there is no experimental information on the thermodynamics of this ion. The results we present in Table 8 are based on our own G2 calculations for reaction (28), the hydride exchange between cyclopropenylium and cyclobutenylium ions:

$$c - C_3 H_3^+(g) + c - C_4 H_6(g) \rightarrow c - C_3 H_4(g) + c - C_4 H_5^+(g)$$
 (28)

Norbornen-7-ylium ion $(C_7H_9^+)$ (43)

The discovery by Winstein, Woodward, and coworkers of a 10^{11} rate enhancement in the solvolysis of *anti*-7-norborneyl tosylate relative to that of 7-norbornyl tosylate²²⁵ led to extensive investigations of this ion.

The ${}^{1}\text{H}^{226,227}$ and ${}^{13}\text{C}$ NMR spectra of 43^{228} and some of its derivatives²²⁹ in solution at low temperature are known. These spectra as well as the X-ray structure of 7-phenyl-2,3-dimethyl-7-norbornenyl cation²³⁰ indicate that C(7) in 43 is "canted" towards the formal double bond, implying a significant degree of homoconjugation (Fig. 31).

Ab initio studies^{231,232} fully agree with these contentions and further, through computation of the ¹³C NMR shifts, lead to an entirely consistent image of this ion as a species of C_s symmetry.

Thermodynamic data for this species (see Table 8) were obtained from the DPA study of *anti*-7-norbornenyl chloride and *anti*-7-norbornenol¹⁸² with the appropriate



Fig. 28 Structure of 1,2-Dipropylcyclopropen-3-ylium ion, 40, optimized at the MP2(full)/6-31G(d) level.



Fig. 29 Structure of tripropylcyclopropenylium, 41, optimized at the MP2(full)/6-31G(d) level.



Fig. 30 Structure of cyclobutenylium ion, 42, optimized at the MP2(full)/6-31G(d) level.



Fig. 31 Structure of norbornen-7-ylium ion, 43, optimized at the MP2(full)/6-31G(d) level.

corrections. We present in Table 8 thermodynamic data for 43 obtained at the G2(MP2) level in this work. The results are extremely consistent. $\Delta_r G^{\circ}_{m}(25)$ is given in Table 4.

Norbornadien-7-ylium ion $(C_7H_7^+)$ (44)

7-Norbornadienyl chloride solvolyzes some 10^3 times faster than *anti*-7-norbornenyl chloride.²³³ This enormous neighboring group participation is associated with the intrinsic stability of **44**. This ion has also been studied by NMR in solution at low temperature.^{226,227} These experiments as well as *ab initio* studies^{231,232} indicate that the most stable structure of **44** corresponds to that presented in Fig. 32. This is a structure of C_s symmetry, the more symmetrical $C_{2\nu}$ structure being the transition state between the two C_s mirror images.²³⁴

Experimental thermodynamic data for 44 (see Table 8) were obtained from the DPA study of 7-norbornadienol.¹⁸² $\Delta_r G^{\circ}_m(25)$ is given in Table 4. Thermodynamic data for 44 obtained at the G2(MP2) level in this work are given in Table 8.

SIX-ELECTRON AROMATIC IONS

Cycloheptatrienylium ion (tropylium) $(C_7H_7^+)$ (45)

The synthesis and description of some important properties (including its very simple IR spectrum, a consequence of the D_{7h} symmetry) of this ion were carried out



Fig. 32 Structure of norbornadien-7-ylium ion, 44, optimized at the MP2(full)/6-31G(d) level.

in 1954 by Doering and Knox^{235} even though it was actually prepared unintentionally, but not identified, in 1881.²³⁶ Interest in this and other species originates in Hückel's classical studies²³⁷ ($4n + 2\pi$ -electron rule).

The remarkably simple ¹H and ¹³C NMR spectra of **45** in solution are known²³⁸, ²³⁹ as is its X-ray structure (in salts).^{240,241}

The thermodynamic data for this ion as reported in Table 8 were obtained from the following sources:

- (1) The DPA study of tropyl methyl ether, $C_7H_7OCH_3$, with the appropriate leaving group corrections.¹⁵⁶
- (2) The G2(MP2) results from Nicolaides and Radom.²⁴²

The experimental determination of $\Delta_{\rm f} H^{\circ}_{\rm m}(45)$ has long been marred, *inter alia*, by the interconversion of the molecular ions of toluene and cycloheptatriene prior to dissociation.^{243,244} The experimental adiabatic ionization energy of the tropyl radical,^{245,246} has been found to be in excellent agreement with the result of highlevel *ab initio* calculations,^{247,248} but the computed standard enthalpy of formation of the radical seems significantly different from the reported experimental value.^{247,248} It is thus rewarding to find that both DPA and computed values are in excellent agreement. The structure portrayed in Fig. 33 was optimized at the MP2(full)/6-31G(d) level in this work. The value of $\Delta_{\rm r} G^{\circ}(25)$ reported in Table 4 is from our DPA studies corrected with the corresponding leaving group corrections.

PHENYL-SUBSTITUTED CARBOCATIONS

Benzylium ion $(C_7H_7^+)$ (46)

To our knowledge, **46** has never been observed in solution under stable conditions, even at low temperature. Pulse radiolysis²⁴⁹ of benzyl chloride as well as flash photolysis²⁵⁰ of several derivatives in HFIP have allowed the observation of the electronic absorption spectra of benzyl and its 4-methyl and 4-methoxy derivatives. The ¹H and ¹³C NMR spectra of the 2,4,6-trimethylbenzyl cation and other more heavily substituted benzyl cations, however, have been studied at low temperature in superacid media.²⁵¹ In the gas phase, cold benzyl radical has been probed by two-color, resonant two-photon ionization techniques, thus providing very accurate vibrational frequencies below 650 cm⁻¹ for the benzyl cation.²⁵² Furthermore, the adiabatic ionization energy of benzyl radical and several isotopomers in the ground state were determined from their threshold photoionization spectra using resonant two-photon excitation and detection of electrons by pulsed field ionization.²⁵³ This information, combined with $\Delta_f H^{\circ}_m(C_6H_5CH_2)$ from Ref. 212 leads to the value of $\Delta_f H^{\circ}_m(\mathbf{46})$ reported in Table 9.

Data for this ion were also obtained through the study of hydride and chloride exchange with ions of known stability using both HPMS²⁵⁴ and FT ICR.^{51,52} These results, also reported in Tables 9 and 4, respectively, are rewardingly coincident. This is particularly important because it tends to indicate that the well-known benzyl/tropylium isomerization problem²⁴⁴ does not seem to affect significantly these results

The study of substituent effects on the stability of substituted benzyl cations^{51,52,}¹⁴⁹ is important as a source of thermodynamic information as well as a means to provide a conceptual link between the stabilities of a number of important carbocations. Mishima's study,⁵¹ in particular, shows that the stabilities in terms of standard Gibbs energy for 4-methoxy- and 4-nitro-substituted benzyl cations differ by some 24.9 kcal mol⁻¹. The former has a stability comparable to that of bicyclo[3.3.3]undecyl (manxyl) cation, while the latter is comparable to the *c*-pentyl cation. We present these two extreme cases in Table 4. The relative stabilities of other substituted benzyl cations are given in Refs. 50 and 51.

The results of high-level *ab initio* calculations on this $C_{2\nu}$ ion (Fig. 34) are also summarized in Table 9. G2-level computations lead to $\Delta_f H^o{}_m(g)$ values in excellent agreement with experiment.²⁴² Equally satisfactory are the results obtained at the G2(MP2,SVP) level for the ionization potential of benzyl radical.²⁴⁸

1-Phenylethan-1-ylium ion, α -phenethyl cation (C₈H₉⁺) (47)

As far as we are aware, 47 has not been observed as a stable species in solution, although flash photolysis²⁵⁰ of ethylbenzene and several of its derivatives in HFIP has allowed the detection of their electronic absorption spectra in the near-UV–visible region.



Fig. 33 Structure of cycloheptatrienylium (tropylium) ion, **45**, optimized at the MP2(full)/6-31G(d) level.

 $\Delta_{\rm f} H^{\circ}_{\rm m}(47)$ can be obtained from the experimental PA and GB of styrene. Over the years, a number of values has been obtained, essentially by HMPS and ICR. Here, we use the average value recommended in Ref. 43. Values are also available from the FT ICR study⁵¹ of chloride exchange between this ion and the *tert*-butyl cation (through the appropriate ladder of experimental values). As shown in Table 9, the results are extremely self-consistent.

We present in Fig. 35 the structure (C_s symmetry) optimized at the MP2(full)/6-31G(d) level (this work). Recently, a large computational study has been published on benzyl and larger carbocations.¹⁵⁵ It uses both the optimized force field MMP2 extended to carbocations and MP4sdq/6-31G(d)//MP2(full)/6-31G(d) calculations. It provides, *inter alia*, valuable estimates of standard heats of formation. In this case, the difference in stability between phenethyl and 4-methylbenzyl cations (1.7 by *ab initio* computations and 2.9 kcal mol⁻¹ by MMP2) is in good agreement with the experimental results in Ref. 51. Our own G2(MP2) results are presented in Table 9.

The experimental $\Delta_r G^{\circ}(25)$ for this ion is given in Table 4.

| Cations | $\Delta_{\mathrm{f}} H^{\circ}{}_{\mathrm{m}}(\mathrm{g})^{a}$ | $S^{\circ}_{m}(g)^{b}$ |
|---|--|------------------------|
| Benzylium ($C_7H_7^+$) (46) | 216.8 ± 0.6^{c} 216.8 ± 2.0^{e} | 74.6 ± 2.0^{d} |
| 1-Phenylethan-1-ylium $(C_8H_9^+)$ (47) | $\begin{array}{c} 200.2 \pm 2.0^{f} \\ 200.6 \pm 2.0^{g} \\ 198.7 \pm 2.0^{i} \end{array}$ | 84.7 ± 2.0^{h} |
| 2-Phenylpropan-2-ylium $(C_9H_{11}^+)$ (48) | 187.4 ± 2.0^{i} 187.2 ± 2.0^{k} | 93.1 ± 2.0^{l} |
| Diphenylmethanylium $(C_{13}H_{11}^+)$ (49) | 225.7 ± 2.0^{m} | 99.7 ± 2.0^{n} |
| 1,1-Diphenylethan-1-ylium $(C_{14}H_{13}^+)$ (50) | 212.7 ± 2.3^{o} 214.1 ± 2.0^{q} | $107.9. \pm .2.0^{p}$ |
| Triphenylmethanylium ($C_{19}H_{15}^+$) (51) | 243.7 ± 2.0^{r} | 123.8 ± 2.0^{s} |

Table 9 Thermodynamic data for phenyl-substituted carbocations

^{*a*}In kcal mol⁻¹.

^{*b*}In cal mol⁻¹ K⁻¹.

^cFrom Ref. 212. See text.

^dScaled, symmetry-corrected HF/6-31G(d) value (this work).

^eComputed (G2) value from Ref. 242.

^fFrom the averaged experimental GB(styrene), 193.4 kcal mol⁻¹ (Ref. 43), and the entropy correction from the same reference.

^gFrom the standard Gibbs energy change for the formal hydride exchange between α -phenethyl and *tert*butyl cations as determined by FT ICR in Ref. 51. Entropies corrected using the experimental values for *iso*-butene²⁶ and styrene²⁶ (82.48 cal mol⁻¹ K^{-1}) and our computed value for the phenethyl cation. ^hScaled HF/6-31G(d) value from this work.

^{*i*}Combining experimental values for $\Delta_f H^{\circ}_m$, C₆H₅-CH₂CH₃, *iso*-butane and **6** with the standard enthalpy for the hydride exchange between 6 and 47.

^jFrom the averaged experimental PA(α -methyl styrene), 206.6 kcal mol⁻¹ (Ref. 43) and $\Delta_f H^{\circ}_m$ $(C_9H_{10}) = 28.27 \pm 0.34 \text{ kcal mol}^{-1.26}$

^kComputed (MMP2) value from Ref. 155.

¹Scaled, symmetry-corrected value at the HF/6-31G(d) level (this work).

^mMMP2 value from Ref. 155.

"Scaled, symmetry-corrected value at the HF/6-31G(d) level (this work).

^oFrom the averaged experimental PA (1,1-diphenylethylene), 211.7 kcal mol⁻¹ (Ref. 43) and $\Delta_{\rm f} H^{\circ}_{\rm m}$ $(C_{14}H_{12}) = 58.7 \pm 1.1 \text{ kcal mol}^{-1}.^{26}$

^{*p*}Scaled, symmetry-corrected value at the HF/6-31G(d) level (this work).

^qMMP2 value from Ref. 155.

^{*r*}MMP2 value from Ref. 155.

^sScaled, symmetry-corrected value at the HF/6-31G(d) level (this work).

2-Phenylpropan-2-ylium ion, cumyl cation ($C_9H_{11}^+$) (48)

This ion as well as its derivatives have played a key role in the development of current concepts and quantitative treatment of substituent effects on organic reactivity.^{256,257}

The UV-visible absorption spectra of 48 and some 4-substituted derivatives have been obtained in HFIP solution using laser flash photolysis techniques.²⁵⁸ The ¹H



Fig. 34 Structure of benzylium ion, 46, optimized at the MP2(full)/6-31G(d) level.

NMR spectrum of this ion at low temperature and in superacid media was obtained in 1964^{259,260} and carefully analyzed in 1966.²⁶¹ The X-ray structure of its hexafluoroantimonate(V) salt was obtained much more recently²⁶² and it reveals significant benzylic delocalization.

The $\Delta_f H^{\circ}_{m}(g)$ value reported in Table 9 was obtained from the PA recommended in Ref. 43, based on ICR data from Taft's and Marshall's groups.

We present in Fig. 36 the $C_{2\nu}$ structure of **48**, obtained at the MP2(full)/6-31G(d) level (this work).

The experimental $\Delta_f H^{\circ}_{m}(48)$ is in quite good agreement with the computed MMP2 and MP4sdq/6-31G(d)//MP2(full)/6-31G* results from Ref. 155 (see Table 9).

We report in Table 4 the experimental $\Delta_r G^{\circ}(25)$ for this ion as given in Refs. 51 and 52. Also reported are values for 4-methoxy and 3-nitro cumyl cations, taken from the same references. These are extreme cases of substituent effects in these systems and span a range of stabilities somewhat smaller than that of the homologous benzyl derivatives (19.9 versus 24.3 kcal mol⁻¹). Still, this yields a stability for the 4-methoxy derivative quite close to that of trityl cation.



Fig. 35 Structure of 1-phenylethan-1-ylium ion, **47**, optimized at the MP2(full)/6-31G(d) level.

Diphenylmethanylium ion, benzhydryl cation $(C_{13}H_{11}^+)$ (49)

The ¹H NMR spectrum of this ion was reported in the early $1960s^{259,260}$ and analyzed a little later.²⁶¹

This ion and several ring-substituted derivatives have been generated by nanosecond laser flash photolysis in solution.²⁶³

As far as we know, 49 has not yet been studied in the solid state.

Direct experimental determinations of its thermodynamic state functions are still missing. Fortunately, a fairly reliable value for $\Delta_{\rm f} H^{\circ}_{\rm m}(49)$ is available thanks to the MMP2 and MP4sdq/6-31G(d)//Mp2(full)/6-31G(d) calculations in Ref. 155. This value is presented in Table 9. The value of $\Delta_{\rm r} G^{\circ}(25)$ given in Table 4 as well as the structure in Fig. 37 were obtained in this study.



Fig. 36 Structure of 2-phenylpropan-2-ylium ion, **48**, optimized at the MP2(full)/6-31G(d) level.

1,1-Diphenylethan-1-ylium ion $(C_{14}H_{13}^+)$ (50)

This ion is known in solution at low temperature in strongly acidic media and was generally studied together with benzhydryl cation.^{259–261}

Its UV-visible spectrum has been obtained by laser flash photolysis techniques.²⁵⁸ Interestingly, it has recently been generated and studied encapsulated in zeolites²⁶⁴ and at ambient temperature in superacid molten salts.²⁶⁵

The experimental $\Delta_f H^{\circ}_{m}(50)$ given in Table 9 was obtained from the GB of 1,1diphenylethene as determined by Taft's group and reported in Ref. 43. This value is in agreement with the computational result from Ref. 155.

We present in Table 4 the $\Delta_r G^{\circ}(25)$ obtained in this study and based on the experimental data indicated above. The structure optimized at the MP2(full)/6-31G(d) level portrayed in Fig. 38 was also obtained in this work.



Fig. 37 Structure of diphenylmethanylium ion, 49, optimized at the MP2(full)/6-31G(d) level.

Triphenylmethanylium ion, triphenyl methyl cation ($C_{19}H_{15}^+$) (51)

This species^{266–268} as well as the salt character of some of its derivatives have long been known.^{269,270} Structure **51** has been studied by UV–visible²⁷¹ and IR²⁷² spectroscopies. Its ¹H NMR spectrum in solution was reported in 1959²⁷³ and its features carefully discussed in 1966.²⁶¹ The ¹³C NMR spectrum in superacid solution was also reported in 1964.²⁷⁴ It has also been obtained in the solid state, trapped in a zeolite.²⁷⁵ The ¹³C NMR spectra of a number of other triaryl and heteroaryl ions have also been reported.²⁷⁶

All these techniques as well as the X-ray spectra of its salts²⁷⁷ agree on the twisting of the phenyl rings out of the central carbon plane by *ca*. 33°. Figure 39 presents the structure of this ion (symmetry group D_3) obtained at the MP2(full)/6-31G(d) level in this work. Our calculated structure agrees nicely with this result and with the MMP2 structure.¹⁵⁵

The $\Delta_{\rm f} H^{\circ}_{\rm m}(51)$ given in Table 9 was obtained computationally at the MMP2 level¹⁵⁵ and was used to estimate the value of $\Delta_{\rm r} G^{\circ}(25)$ given in Table 4.

6 Solution reactivity

Consider reactions (3a) and (4):

$$R_1H(g) + R_2^+(g) \rightarrow R_2H(g) + R_1^+(g)$$
 (3a)

$$R_1H(g) + R_2H_2^+(g) \rightarrow R_2H(g) + R_1H_2^+(g)$$
 (4)

For comparison purposes it seems useful to take the cumyl cation 48 as a reference ion, on account of the fact that it can be obtained through reactions (29) and (30) and



Fig. 38 Structure of 1,1-diphenylethan-1-ylium ion, 50, optimized at the MP2(full)/6-31G(d) level.

that in both cases, fairly reliable thermodynamic data are available for reactants and products:

$$C_6H_5CH(CH_3)_2(g) + CH_3^+(g) \rightarrow (48)(g) + CH_4(g) \qquad \Delta_r H^{\circ}(29)$$
 (29)

$$C_6H_5C(CH_3)CH_2(g) + CH_5^+(g) \rightarrow (48)(g) + CH_4(g) \qquad \Delta_r H^{\circ}(30)$$
 (30)

Using data given in the previous sections, we get $\Delta_r H^{\circ}(29) = -93.3 \pm 2.1$ and $\Delta_r H^{\circ}(30) = -76.8 \pm 2.2$ kcal mol⁻¹.

Let us now examine two rather extreme cases: the hydride exchange between tricyclopropylmethyl and cumyl cations and the proton exchange between guaiazulene²⁷⁸ and α -methylstyrene, respectively (reactions 31 and 32):

$$(c-C_{3}H_{5})_{3}C^{+}(g) + C_{6}H_{5}CH(CH_{3})_{2}(g) \rightarrow (48)(g) + (c-C_{3}H_{5})_{3}CH(g) \Delta_{r}H^{\circ}(31)$$
(31)

$$(g) + (g) + (g) \rightarrow (g) + [48](g) \Delta_r H^{\circ}(32)$$
(32)

The corresponding $\Delta_r H^{\circ}(31)$ and $\Delta_r H^{\circ}(32)$ are, respectively, -11.0 ± 4.1 and 28.4 ± 2.8 kcal mol⁻¹.²⁷⁹

These results show that the range of structural effects on the intrinsic (gas phase) stability of carbenium ions amounts to at least $110 \text{ kcal mol}^{-1}$. Let us see now how



Fig. 39 Structure of triphenylmethanylium ion, 51, optimized at the MP2(full)/6-31G(d) level.

the differences in intrinsic stability affect solution ("real world"?) reactivity. As in the previous section, thermodynamic criteria seem well suited for this purpose and so are kinetic data, through formally straightforward extra thermodynamic relationships.

THERMODYNAMIC PROPERTIES

The pK_{R^+} and pK_a scales

The use of reaction (33) in solution as a quantitative tool for the ranking of carbocation stability is at least half-a-century old:²⁸⁰

$$R-OH + H^+ \rightarrow R^+ + H_2O \tag{33}$$

The concept is extremely simple, as it takes as its starting point the equilibrium constant for this reaction, $K_{\rm R^+}^{157,281,282}$

$$K_{\rm R^+} = a_{\rm R^+} a_{\rm H_{2}O} / a_{\rm ROH} a_{\rm H^+} \tag{34}$$

Given two alcohols, respectively, R₁OH and R₂OH, the position of equilibrium (35)

$$\mathbf{R}_2^+ + \mathbf{R}_1 \mathbf{OH} \rightarrow \mathbf{R}_1^+ + \mathbf{R}_2 \mathbf{OH} \tag{35}$$

is simply given by K_{35} , equation (36):

$$K_{35} = K_{\rm R(1)^+} / K_{\rm R(2)^+} \tag{36}$$

or

$$\log K_{35} = pK_{R(1)^+} - pK_{R(2)^+}$$
(37)

Reaction (38) is also suitable for comparison purposes. Following the same procedure as above, $\log K_{38}$ is given by equation (39):

$$R_1H(aq) + R_2^+(aq) \rightarrow R_2H(aq) + R_1^+(aq)$$
 (38)

$$\log K_{38} = pK_a(R_1) - pK_a(R_2)$$
(39)

From an operational point of view, difficulties might arise from the fact that rather high acid concentrations and/or aqueous-organic solvents are often used in these studies, the reference state being the pure water solution. These shortcomings can be averted by the use of appropriate methods, recently reviewed by Cox in this series.²⁸³

In what follows we shall use both the pK_{R^+} and pK_a scales for the quantitative comparison of carbenium ions, without attempting any correction for aqueousorganic medium effects. All data are anchored to **48**. Taft and co-workers already compared gas phase and solution basicities of ethylenic and aromatic hydrocarbons.^{278,284} Now the pK_{R^+} scale has been widely extended^{285,286} through the determination of K_{R^+} from the reaction rate constants for the forward and reverse processes in reaction (33). This is a very important breakthrough. Also, a number of new data for other species in the gas phase has become available.

We can now carry out a comparison of solution and gas-phase rankings of stabilities, respectively, using $\Delta p K_{R^+}$ and $\Delta p K_a$ values; i.e., referred to processes leading to **48**. The reference processes in the gas phase are the chloride exchange between **48** and the relevant ion, i.e. reaction (40), and the proton exchange between α -methylstyrene and the relevant base, i.e. reaction (41). $\Delta_r G^\circ$ values for both reactions are given, respectively, by the differences in $\Delta_r G^\circ(25)$ for the corresponding chloride and cumyl chloride (data from Table 4) and the difference in GB values between the relevant base and α -methylstyrene. Formally, very similar processes are compared as they only differ in the leaving group:

$$C_6H_5C(Cl)(CH_3)_2(g) + R^+(g) \rightarrow (48)(g) + R - Cl(g) \qquad \Delta_r G^{\circ}(40)$$
 (40)

$$C_6H_5C(CH_2)CH_3(g) + RH_2^+(g) \rightarrow (48)(g) + R - H(g) \qquad \Delta_r G^{\circ}(41)$$
 (41)

Figure 40 is a joint plot of $\Delta p K_{R^+}$ and $\Delta p K_a$ (using data from Refs. 278, 284–287) against $\Delta_r G^{\circ}(40)$ and $\Delta_r G^{\circ}(41)$. Squares denote $\Delta p K_a / \Delta_r G^{\circ}(41)$ and open circles denote $\Delta p K_{R^+} / \Delta_r G^{\circ}(40)$. They define a straight line of very good statistical quality involving ranges of some 45 kcal mol⁻¹ and 27 log units, respectively. The level of

self-consistency of these results is both rewarding and surprising, on account of the variety of experimental methods involved. Conversion of log units to Gibbs energies leads to a slope of 0.77.

We take this result as implying that: (1) the ranking of intrinsic stabilities of these ions is faithfully reproduced in solution with a moderate attenuation (*ca.* 24%); (2) the ionic systems described by this line follow a constant pattern of solvation which we call "general" for the sake of simplicity. This does not mean that these effects are small in absolute value but rather, that *differential* solvation effects are likely to be small; and (3) the fact that the slope is smaller than unity suggests that solvation energetics are not exactly constant and that the less stable ions interact more strongly with the solvent. The correlation spans a range of stabilities going all the way from benzylium to tripropylcyclopropenylium ion. Table 4 shows that this range includes most tertiary alkyl cations, several stabilized secondary ones, a number of bridgehead carbocations and phenyl- and cyclopropyl-stabilized methyl cations. This correlation appears, therefore, as a very powerful and simple unifying factor.

Ions 38, 49–51 do not seem to follow the "general" pattern and were not included in the correlation. Interestingly, at least the gas-phase data for 38, the cyclopropenylium ion, seem quite reliable. This ion, however, is small and readily accessible to solvent molecules. Furthermore, *ab initio* calculations indicate that 38 is a strong hydrogen bond donor to water. It seems reasonable to attribute at least part of this enhanced stability in solution to hydrogen bonding to the solvent. A plot similar to Fig. 40 but involving standard Gibbs energy changes for hydroxide instead of chloride exchange is also nicely linear and, once again, 38 is deviant but 49–51 are now "well behaved". This is important, inasmuch as the pK_{R^+} is based on the protonation of alcohols and strongly suggests that these ions do not display any abnormal behavior in solution. It follows that the departures shown in Fig. 40 originate in the leaving group correction for the corresponding chlorides. At this point, unfortunately, the possibility for these corrections being affected by computational artifacts cannot be ruled out.

Ions with pK_{R^+} values of about 14 have recently been synthesized.²⁸⁸ Assuming that the linearity extends up to these ions, this would bring the range of structural effects on stabilities of carbenium ions to at least 120 kcal mol⁻¹.

Solution thermochemistry

The quantitative thermodynamic study of the ionization of appropriate precursors R-X (X = OH, halogen) by superacidic species (FSO₃H, SbF₅, FSO₃H–SbF₅ mixtures) in low nucleophilicity solvents [SO₂(1), SO₂ClF, CH₂Cl₂] at low temperatures was carried out during the 1970s and early 1980s by Arnett and co-workers.²⁸⁹ The basic concept behind this effort was "to provide the basis for a single stability scale in solution ranging from the relatively unstable secondary ions (e.g., isopropyl or cyclopentyl) to the more familiar triarylmethylcarbenium ions. If this were accomplished, future workers could use a scale to interpolate and include



Fig. 40 Plot of $\Delta p K_{R^+}$ and $\Delta p K_a$ (i.e., values relative to **48** and α -methylstyrene) versus $\Delta_r G^{\circ}(40)$ and $\Delta_r G^{\circ}(41)$. Open circles denote $\Delta p K_{R^+} / \Delta_r G^{\circ}(40)$ and full squares denote $\Delta p K_a / \Delta_r G^{\circ}(41)$. A = α -methylstyrene; B = hexamethylbenzene; C = 1,1-diphenylethylene; D = 1,1'-ethenylidenebis-[4-methyl-benzene]; E = 1,1-dicyclopropylethylene; F = azulene; G = guaiazulene.

almost any ion whose ionization energy can be related directly or indirectly to the gas phase scale, the pK_R scale, thermochemical scales or solvolysis rates".²⁹⁰

We present in Fig. 41 a plot of the ionization enthalpies of alcohols in a 1:1 mixture of SbF₅ and FSO₃H in SO₂ClF solution,²⁹⁰ ΔH°_{i} , against $\Delta_r G^{\circ}(25)$. The proportionality is very clear but the scatter is significant. More limited groups of ions yield better correlations, particularly in the case of the ionization of chlorides by SbF₅.²⁹¹ Entropic factors might play a role, as well as, in some cases, ion-pairing. Differential solvation effects in these systems are well known. Thus, adamantyl appears as less stable than the *tert*-butyl cation while the opposite occurs in the gas phase. We have recently shown computationally that at least part of this reversal can be largely attributed to solvation effects.⁶⁸ It was shown, much earlier, that the



Fig. 41 Enthalpies of ionization, ΔH°_{i} for alcohols with 1:1 mixtures of SbF₅ and FSO₃H in SO₂ClF solution versus $\Delta_r G^{\circ}(25)$.

ranking of basicities of alkylbenzenes was reversed on going from the gas phase to superacid solutions,²⁹² a fact also consistent with solvation effects.

As things stand now, a more complete understanding of the situation still requires considerable work.

REACTION KINETICS

It is generally accepted that S_N1 solvolysis rates for a series of similar compounds under similar conditions reflect directly the stability of their carbocationoid intermediates.^{166,293} However, the lack of reliable experimental data for stabilities of carbenium ions prevented verification of this hypothesis for a long time. Several approaches at various levels of sophistication were advanced to estimate these ion stabilities: their strain was estimated via empirical correlations with IR-stretching frequencies of the respective carbonyl groups, and non-bonded interactions from molecular models or, subsequently, by empirical force-field calculations.^{294–302} However, owing to the empirical nature of these methods, the significance of the results could not be truly assessed. More recently, Arnett *et al.*^{109,290} reported heats of ionization (ΔH_i) of alkyl chlorides to stable carbocations in SbF₅-solvent mixtures. Correlation of the heats of ionization with the respective free energies of activation for ethanolysis afforded a straight line over a range of 22 kcal mol⁻¹ for ΔH_i .

Initial attempts to establish an analogous correlation between solvolytic reactivity for a limited series of bridgehead halides and stability of the respective carbenium ions in the gas phase by ion cyclotron resonance (ICR) were met by only limited success owing to apparent rearrangement of some of the ions under the conditions of the ICR experiments.³⁰³ Although highly strained ions such as 1-norbornyl were particularly prone to undergo such rearrangements, the ICR measurements alone did not allow a distinction between unrearranged and rearranged ions. However, when ions were generated by DPA rather than on electron impact, they are not significantly vibrationally excited and, hence, are less prone to undergo rearrangement. As shown above for bridgehead derivatives, the experimental ion stabilities as determined by DPA and adjusted with the appropriate leaving group correction correlate well with the stabilities calculated by ab initio methods (MP2/6-311G**).^{57,58} Figure 42 shows the correlation between the rate constants for solvolysis of a series of representative bridgehead bromides under standard conditions (log k for tosylate solvolysis relative to 1-adamantyl in 80% ethanol at $(70^{\circ}C)^{304}$ versus ion stabilities $[\Delta_r G^{\circ}(25)]^{55}$

The correlation in Fig. 42 is given by $\log k = 0.427\Delta_r G^{\circ}(25) + 0.41$ ($r^2 = 0.986$, $\sigma = 0.75$, non-bridgehead ions excluded from the correlation). It spans *ca*. 50 kcal mol⁻¹ in $\Delta_r G^{\circ}(25)$. Conversion of $\log k$ in units of free energies of activation results in a slope of 0.67, in reasonable agreement with the slopes reported by Arnett (-0.89 in SO₂ClF, -1.02 in CH₂Cl₂ and -0.95 in SO₂). The lower value of the slope in the gas phase may be attributed to the absence of stabilization by solvent.

The bridgehead derivatives represent a mechanistically homogenous series of model compounds. Nucleophilic solvent participation (NSP) and E1-pathways upon solvolysis are excluded for skeletal reasons, and relative rate constants are independent of leaving group and solvent. This is not necessarily so in the general case of solvolysis. DPA has now been applied to halides and alcohols leading to carbenium ions representative of various classes, such as secondary aliphatic, strained bicyclic, homoaromatic, benzyl- and cyclopropyl-stabilized ions. The relative ion stabilities are reported in Table 4. The rate constants for solvolysis of the respective halides, *p*-toluenesulfonates or *p*-nitrobenzoates are available in the literature. For reasons of consistency with the plot shown in Fig. 42, they are extrapolated to 70°C and expressed relative to 1-adamantyl solvolysis under the same conditions, assuming relative rates to be independent of leaving groups and solvent, as in the bridgehead series. These assumptions are not always justified, however. For example, significant variations in the relative rates in different solvents occur in the case of benzylic derivatives, but no systematic study was undertaken in



Fig. 42 Plot of log *k* for bridgehead derivatives versus $\Delta_r G^{\circ}(25)$ (X = Br), open squares, $\Delta_r G^{\circ}(29)$ (X = Br, calculated Cl), diamonds, and $\Delta_r G^{\circ}(25)$ (X = Br, calculated OH), filled circles.

order to determine which data are most representative. In order to minimize complications owing to NSP, rate constants for acetolysis were used in preference to data from other solvent systems. Details will be reported in a forthcoming publication.³⁰⁵

As indicated earlier, the experimental or computed gas-phase data were subject whenever necessary to the appropriate leaving correction in order to generate a homogeneous set of Gibbs energy changes based on chloride exchange, $\Delta_r G^{\circ}(25)$ (summarized in Table 4). For some non-bridgehead derivatives, specific leaving group corrections for *p*-toluenesulfonates were applied in order to compare exactly the same systems in the gas phase and in solution.¹⁹¹

In Fig. 43, log k for 24 substrates versus the appropriate $\Delta_r G^{\circ}(25)$ values are represented. A subset of 18 substrates follow the correlation equation log $k = 0.422\Delta_r G^{\circ}(25) + 0.50$ ($r^2 = 0.9686$, $\sigma = 0.97$). This correlation spans over 50 kcal mol⁻¹ in $\Delta_r G^{\circ}(25)$. Most important is the fact that the slope is practically identical to that found for bridgehead derivatives (Fig. 42), while the quality of the fit is also excellent. While consistent behavior of bridgehead and bicyclic secondary derivatives such as 7-norbornyl (24), 2-adamantyl (29), or 4-homoadamantyl (32) upon solvolysis might be expected on the grounds of structural similarities and the absence of NSP, that of the other compounds of this series is surprising. The stability



Fig. 43 log *k* for 24 substrates relative to 1-adamantyl chloride (computed as tosylates in 80% ethanol at 70°C), versus $\Delta_r G^{\circ}(25)$.

of the bicyclic secondary ions is mainly determined by strain effects. The 2norbornyl cation (23) is stabilized by σ -participation, and the 7-norbornenyl (43) and 7-norbornadienyl (44) cations are homoaromatic and bishomoaromatic, respectively. In the cyclopropylcarbinyl cations 17-21 stabilization occurs through the Walsh orbitals of the cyclopropane rings, while the phenyl-substituted cations 46-51 profit from resonance stabilization. Despite these different causes for ion stability, the correlation suggests ion stability as by far the predominant factor determining solvolytic reactivity of secondary and tertiary derivatives. At the same time, the correlation confirms the qualitative concepts of strain and anchimeric participation, which have been developed over the years in the context of solvolysis.

Out of the 24 compounds represented in Fig. 43, there are six which fall outside the correlation. The low rate of triphenylmethyl chloride (**51**) in comparison to that of diphenylmethyl (**49**) or 1,1-diphenylethyl chloride (**50**) has been known for more than 35 years³⁰⁶ but, to our knowledge, has not been rationalized. However, if log *k* is correlated with the pK_{R^+} values of the corresponding alcohols in condensed phase, the corresponding cations, **49–51**, are well behaved (exactly as discussed in the previous section), so that their deviations in Fig. 43 must be attributed to a solvation effect. The deviations observed with the 2-propyl (**4**), 2-butyl (**5**), and *tert*-butyl (**6**) derivatives are more significant. The rate constants for these compounds, and also that of cyclopentyl (**16**) refer to tosylate acetolysis and are corrected for NSP.³⁰³ This NSP correction brings **16** on to the correlation line, but the other compounds react still significantly faster than predicted on the grounds of the stability of the respective carbenium ions. According to the definition used here,³⁰³ NSP is defined as the rate enhancement experienced by a substrate, relative to 1-adamantyl, upon solvent change from trifluoroacetic acid to the solvent of interest. Solvents other than trifluoroacetic acid may be used as reference, but this does not fundamentally change the problem. NSP occurs in solvolysis reactions that lead to small, undelocalized carbenium ions. As shown by Takeuchi and colleagues⁶⁸ for tertiary solvolysis, NSP vanishes upon an increase of steric crowding around the reaction center, and upon increased stabilization of the positive charge by alkyl substitution. Even in the most limiting solvent HFIP, tert-butyl derivatives react still significantly faster than predicted on the grounds of the stability of the tert-butyl cation. In other words, NSP alone does not account for the total of the rate enhancement of tert-butyl derivatives, and the same phenomenon occurs with 2-propyl and 2-butyl. Although the transition state for solvolysis is close to the carbenium ion with respect to structure and energy, less charge is developed owing to the proximity of the leaving group and also to solvation. Apparently, these stabilizing effects are more important in the case of reactions leading to small ions such as 2-propyl or tert-butyl where charge is strongly localized, than to larger ions, where charge may be dispersed over a large number of atoms. For this reason, Arnett's heats of ionization of halides in the condensed phase are a more realistic model for the solvolysis reaction than is ion stability in the gas phase because they contain contributions for solvation and, although to some unknown degree, stabilization by the counter-ion.²⁹⁰ However, as Figs. 42 and 43 show, for ions having a sufficiently large number of atoms, the gasphase stability of the carbenium ions is reflected to the same degree in the transition state for solvolysis over a very large range of reactivities, and the discrepancies between ion stabilities as determined in solution and in the gas phase are of minor significance considering the overall consistency of the data over the large range of ion stabilities.

7 Conclusion

This "low resolution" overview of the gas-phase stabilities of a variety of carbenium ions shows that they are by no means exotic data unrelated to solution reactivity. Quite on the contrary, they are the fundamental factor determining the relative stabilities of the same species in solution (as determined by the pK_R and pK_a scales, for example). Furthermore, the ranking of solvolysis rates of compounds R-X is largely dependent on the intrinsic stability of R^+ . While this was already known in the case of bridgehead carbocations, we see now that this applies also to nonbridgehead tertiary cations as well as to a variety of secondary cations, even sometimes in cases wherein the access of solvent to the reaction center is not hindered. Intrinsic stability of carbocations thus appears as a very powerful unifying factor.

A number of significant problems obviously remains to be solved:

(1) It seems highly desirable to extend both the accuracy and the size of the data base. This implies the need for more and more accurate *experimental* data.

- (2) The very existence of a remarkably simple and extensive relationship between thermodynamic and kinetic reactivity in solution and intrinsic stability is intriguing, on account of the variety of stabilities, structures and exposures to solvent involved. This raises important questions about, *inter alia*, the role of C-H···solvent hydrogen bonding interactions of carbocations in solution as well as charge delocalization effects within hydrocarbon frameworks of the carbocations.
- (3) The recent development of important quantitative empirical treatments of electrophilicity–nucleophilicity in solution^{307,308} calls for an investigation of the relationship between intrinsic stability and solution electrophilicity.

Acknowledgments

This work was supported by grants PB96-0927-C02-01 and BQU2000-1497 of the Spanish DGES, 20-52581.97 and 2000-058801.99 of the Swiss National Science Foundation. Valuable assistance from M. Jean-Claude Rossier is greatly appreciated. Interesting discussions with Profs J. F. Liebman and H. Mayr are acknowledged.

References

- 1. Regarding the IUPAC nomenclature for these species, see Powell, W.H. (1993). *Pure Appl. Chem.* **65**, 1357
- Regarding the IUPAC nomenclature for general physical organic chemistry, see Müller, P. (1994). Pure Appl. Chem. 66, 1077
- 3. Glassgold, A.E., Omont, A. and Guelin, M. (1992). Astrophys. J. 396, 115
- Banaszkiewicz, M., Lara, L.M., Rodrigo, R., López-Moreno, J. and Molina-Cuberos, G.J. (2000). *Icarus* 147, 386
- 5. Anicich, V.G., Milligan, D.B., Fairley, D.A. and McEwan, M.J. (2000). Icarus 146, 118
- 6. Encrenaz, T.H., Drossart, P., Feuchtgruber, H., Lellouch, E., Bezard, B., Fouchet, T. and Atreya, S.K. (1999). *Planet. Space Sci.* **47**, 1225
- 7. Thaddeus, P., McCarthy, M.C., Travers, M.J., Gottlieb, C.A. and Chen, W. (1998). Faraday Discuss. 109, 121
- 8. Okamoto, K., Takeuchi, K. and Kitagawa, T. (1995). Adv. Phys. Org. Chem. 30, 173
- Tanaka, T., Kitagawa, T., Komatsu, K. and Takeuchi, K. (1997). J. Am. Chem. Soc. 119, 9313
- Asensi, M.A., Corma, A., Martínez, A. and Martínez, C. (1996). Prepr. Am. Chem. Soc. Div. Pet. Chem. 41, 692
- 11. Corma, A., Martínez, A. and Martínez, C. (1996). J. Catal. 164, 422
- 12. Corma, A. and García, H. (1997). Catal. Today 38, 257
- 13. Corma, A. and García, H. (1998). Top. Catal. 6, 127
- Corma, A. and Martínez, A. (1998). NATO ASI Series 3. 44. Kluwer Academic Publishers, Dordrecht, Holland
- 15. Corma, A. and García, H. (2000). Dalton 1381
- 16. Farcasiu, D. and Lukinskas, P. (1998). J. Phys. Chem. A 102, 10436
- 17. Farcasiu, D. and Hancu, D. (1999). J. Am. Chem. Soc. 121, 7173
- 18. Farcasiu, D. and Lukinskas, P. (2000). J. Phys. Chem. A 104, 1944

- 19. Farcasiu, D., Norton, S.H. and Hancu, D. (2000). J. Am. Chem. Soc. 122, 668
- 20. Boronat, M., Viruela, P. and Corma, A. (2000). Phys. Chem. Chem. Phys. 2, 3327
- Aue, D.H. and Bowers, M.T. (1979). Gas Phase Ion Chemistry. Bowers, M.T. (ed.), vol. 2, Chapter 1. Academic Press, New York
- 22. Raghavachari, K., Whiteside, R.A., Pople, J.A. and Schleyer, P.v.R. (1981). J. Am. Chem. Soc. 103, 5649
- Pilhaja, K. (1987). Thermochemical Methods in the Structural Analysis of Organic Compounds. *Molecular Structure and Energetics. Physical Measurements*. Liebman, J.F. and Greenberg, A. (eds), 2, Chapter 5. VCH Publishers, New York
- 24. Chickos, J.S. (1987). Heats of Sublimation. *Molecular Structure and Energetics*. *Physical Measurements*. Liebman, J.L. and Greenberg, A. (eds), **2**, Chapter 3. VCH Publishers, New York
- 25. Tsang, W. (1996). *Heats of Formation of Organic Free Radicals by Kinetic Methods in Energetics of Organic Free Radicals*. Martinho Simões, J.A., Greenberg, A. and Liebman, J.F. (eds), . Blackie Academic and Professional, London
- Afeefy, H.Y., Liebman, J.F. and Stein, S.E. (2000). Neutral Thermochemical Data in NIST Chemistry WebBook. *NIST Standard Reference Database Number 69*. Mallard, W.G. and Linstrom, P.J. (eds), (http://webbook.nist.gov). National Institute of Standards and Technology, Gaithersburg, MD
- 27. Pedley, J.B., Naylor, R.D. and Kirby, S.P. (eds) (1986). *Thermochemical Data of Organic Compounds* (2nd ed). Chapman & Hall, New York
- Pedley, J.B. (ed.), (1994). *Thermochemical Data and Structures of Organic Compounds*. vol. 1. Thermodynamics Research Center, College Station, TX, USA
- 29. *TRC Thermodynamic Tables*. Thermodynamic Research Center, The Texas A&M University System, USA. These tables are being updated continuously
- Dorofeeva, O.V., Gurvich, L.V. and Jorish, S.V. (1986). J. Phys. Chem. Ref. Data 15, 438
- 31. Liebman, J.F. and Greenberg, A. (1989). Chem. Rev. 89, 1225
- 32. Domalski, E.S. and Hearing, E.D. (1990). J. Phys. Chem. Ref. Data 19, 881
- 33. Domalski, E.S. and Hearing, E.D. (1996). J. Phys. Chem. Ref. Data 25, 1
- Domalsky, E.S. (ed.), (1993). NIST Estimation of the Thermodynamic Properties for Organic Compounds at 298.15 K—Compounds Containing the Elements C,H,N,O,S, and Halogens. National Institute of Standards and Technology, Washington, DC
- 35. Roth, W.R., Adamczak, O., Breuckmann, R., Lennartz, H.-W. and Boese, R. (1991). *Chem. Ber.* **124**, 2499
- 36. Lias, S.G. and Bartmess, J.E. (2000). NIST Chemistry WebBook. NIST Standard Reference Database Number 69. Mallard, W.G. and Linstrom, P.J. (eds), (http:// webbook.nist.gov). National Institute of Standards and Technology, Gaithersburg, MD, USA
- 37. Ervin, K.M. (2001). Chem. Rev. 101, 391
- 38. Marshall, A.G. (2000). Int. J. Mass Spectrom. 200, 331
- 39. Kebarle, P. (2000). Int. J. Mass Spectrom. 200, 313
- 40. Böhme, D.K. (2000). Int. J. Mass Spectrom. 200, 97
- 41. Ng, C.Y. (2000). Int. J. Mass Spectrom. 200, 357
- 42. Baer, T. (2000). Int. J. Mass Spectrom. 200, 443
- 43. Hunter, E.P.L. and Lias, S.G. (1998). J. Phys. Chem. Ref. Data 27, 413
- 44. Abboud, J.-L.M. and Notario, R. (1999). Energetics of Stable Molecules and Reactive Intermediates. *NATO Science Series*. Minas da Piedade, M.E. (ed.), **535**. Kluwer Academic Publishers, Dordrecht
- 45. Bartmess, J.E. (1994). J. Phys. Chem. 98, 6420
- 46. Bartmess, J.E. (1995). J. Phys. Chem. 99, 6755
- 47. East, A.L.L., Smith, B.J. and Radom, L. (1997). J. Am. Chem. Soc. 119, 9014

- Yungman, V.S. (2000). Entropy and Heat Capacity of Organic Compounds in NIST Chemistry WebBook. *NIST Standard Reference Database Number 69*. Mallard, W.G. and Linstrom, P.J. (eds), (http://webbook.nist.gov). National Institute of Standards and Technology, Gaithersburg, MD
- 49. Solomon, J.J. and Field, F.H. (1973). J. Am. Chem. Soc. 95, 4483
- 50. Staley, R.H., Wieting, R.D. and Beauchamp, J.L. (1977). J. Am. Chem. Soc. 99, 5964
- 51. Mishima, M., Arima, K., Inoue, H., Usui, S., Fujio, M. and Tsuno, Y. (1995). Bull. Chem. Soc. Jpn 68, 3199
- Chuchani, G., Mishima, M., Notario, R. and Abboud, J.-L.M. (1999). Advances in Quantitative Structure-Property Relationships. Charton, M. and Charton, B.I. (eds), 2. JAI Press, Standford, Conn
- 53. Lias, S.G. and Ausloos, P. (1977). Int. J. Mass Spectrom. Ion Phys. 23, 273
- Abboud, J.-L.M., Notario, R., Ballesteros, E., Herreros, M., Mó, O., Yáñez, M., Elguero, J., Boyer, G. and Claramunt, R. (1994). J. Am. Chem. Soc. 116, 2486
- Abboud, J.-L.M., Castaño, O., Herreros, M., Leito, I., Notario, R. and Sak, K. (1998). J. Org. Chem. 63, 8995
- 56. Abboud, J. -L. M.; Leito, I.; Kaljurand, I.; Notario, R. Unpublished results
- 57. Abboud, J.-L.M., Castaño, O., Della, E.W., Herreros, M., Müller, P., Notario, R. and Rossier, J.C. (1997). *J. Am. Chem. Soc.* **119**, 2262
- Abboud, J.-L.M., Herreros, M., Notario, R., Lomas, J.S., Mareda, J., Müller, P., Notario, R. and Rossier, J.C. (1999). *J. Org. Chem.* 64, 6401
- 59. Meot-Ner, M. (1987). NATO ASI Series C 193, 383
- 60. Meot-Ner, M. (1991). J. Phys. Chem. 95, 6580
- 61. Irikura, K.K. (1999). J. Am. Chem. Soc. 121, 7689
- 62. Abboud, J.-L.M., Castaño, O., Elguero, J., Herreros, M., Jagerovic, N., Notario, R. and Sak, K. (1998). *Int. J. Mass Spectrom. Ion Proc.* **175**, 35
- 63. Using the updated GB values from Ref. 43 and $\Delta_f H^{\circ}_{m}$ for 1-adamantyl cation from Ref. 64.
- Flores, H., Dávalos, J.Z., Abboud, J.-L.M., Castaño, O., Gomperts, R., Jiménez, P., Notario, R. and Roux, M.V. (1999). J. Phys. Chem. A 103, 7555
- 65. Hudgens, J.W., Johnson, R.D., Tsai, B.P. and Kafafi, S.A. (1990). J. Am. Chem. Soc. 112, 5763
- 66. Robles, E.S.J. and Chen, P. (1994). J. Phys. Chem. 98, 6919
- 67. Horn, M. and Botschwina, P. (1994). Chem. Phys. Lett. 228, 259
- 68. Takeuchi, K., Takasuka, M., Shiba, E., Kinoshita, T., Okazaki, T., Abboud, J.-L.M., Notario, R. and Castaño, O. (2000). J. Am. Chem. Soc. **122**, 7351
- 69. Curtiss, L.A. and Pople, J.A. (1991). J. Chem. Phys. 94, 7221
- 70. Curtiss, L.A., Raghavachari, K. and Pople, J.A. (1993). J. Chem. Phys. 98, 1293
- 71. Sharma, R.B., Sen Sharma, D.K., Hiraoka, K. and Kebarle, P. (1985). *J. Am. Chem. Soc.* **107**, 3747
- 72. Jacox, M.E. (1994). J. Phys. Chem. Ref. Data, Monograph No. 3 124
- 73. Koenig, T., Balle, T. and Snell, W. (1975). J. Am. Chem. Soc. 97, 662
- Dyke, J., Jonathan, N., Lee, E. and Morris, A. (1976). J. Chem. Soc. Faraday Trans. 2 72, 1385
- Crofton, M.W., Kreiner, W.A., Jagod, J.F., Rehfuss, G.D. and Oka, T. (1985). J. Chem. Phys. 83, 3702
- Crofton, M.W., Jagod, J.F., Rehfuss, G.D., Kreiner, W.A. and Oka, T. (1988). J. Chem. Phys. 88, 666
- 77. Dixon, D.A., Feller, D. and Peterson, K.A. (1997). J. Phys. Chem. A 101, 9405
- 78. Blush, J.A., Chen, P., Wiedmann, R.T. and White, M.G. (1993). J. Chem. Phys. 98, 3557
- 79. Ruscic, B., Litorja, M. and Asher, R.L. (1999). J. Phys. Chem. A 103, 8625

- Weitzel, K.-M., Malow, M., Jarvis, G.K., Baer, T., Song, Y. and Ng, C.Y. (1999). J. Chem. Phys. 111, 8267
- 81. Szulejko, J.E. and McMahon, T.B. (1993). J. Am. Chem. Soc. 115, 7839
- 82. Hemsworth, R.S., Rundle, H.W., Böhme, D.K., Schiff, H.I., Dunkin, D.B. and Fehsenfeld, F.C. (1973). J. Chem. Phys. 59, 61
- 83. Smith, B.J. and Radom, L. (1993). J. Am. Chem. Soc. 115, 4885
- Schreiner, P.R., Kim, S.-J., Schaefer, H.F., III and Schleyer, P.v.R. (1993). J. Chem. Phys. 99, 3716
- 85. Müller, H., Kutzelnigg, W. and Klopper, W. (1997). J. Chem. Phys. 106, 1863
- 86. Marx, D. and Parrinello, M. (1995). Nature 375, 216
- 87. Marx, D. and Parrinello, M. (1999). Science 284, 59
- 88. White, E.T., Tang, J. and Oka, T. (1999). Science 284, 135
- Heck, A.J.R., de Konig, L.J. and Nibbering, N.M.M. (1991). J. Am. Soc. Mass Spectrom. 2, 453
- Kramer, G.M., Oka, T., White, E.T., Marx, D. and Parrinello, M. (1999). Science 286, 1051a
- 91. Schreiner, P.R. (2000). Angew. Chem. Int. Ed. 39, 3239
- 92. Hariharan, P.C., Lathan, W.A. and Pople, J.A. (1972). Chem. Phys. Lett. 4, 385
- 93. Zurawski, B., Ahlrichs, R. and Kutzelnigg, W. (1973). Chem. Phys. Lett. 21, 309
- 94. Perera, S.A., Bartlett, R.J. and Schleyer, P.v.R. (1995). J. Am. Chem. Soc. 117, 8476 and references therein
- 95. Ruscic, B., Berkowitz, J., Curtiss, L.A. and Pople, J.A. (1989). J. Chem. Phys. 91, 114
- 96. Baer, T., Song, Y., Liu, J., Chen, W. and Ng, Y. (2000). Faraday Discuss. 115, 137
- 97. Brouard, M., Lightfoot, P.D. and Pilling, M.J. (1986). J. Phys. Chem. 90, 445
- 98. Koch, W., Liu, B. and Schleyer, P.v.R. (1989). J. Am. Chem. Soc. 111, 3479
- 99. Jursic, B.S. (1999). J. Mol. Struct (Theochem) 491, 193
- 100. Saunders, M., Hagen, E.L. and Rosenfeld, J. (1968). J. Am. Chem. Soc. 90, 6881
- 101. Saunders, M., Vogel, P., Hagen, E.L. and Rosenfeld, J. (1973). Acc. Chem. Res. 6, 53
- 102. Baer, T., Song, Y., Ng, C.Y., Liu, J. and Chen, W. (2000). J. Phys. Chem. A 104, 1959
- 103. Schultz, J.C., Houle, F.A. and Beauchamp, J.L. (1984). J. Am. Chem. Soc. 106, 3917
- 104. Dyke, J., Ellis, A., Jonathan, N. and Morris, A. (1985). J. Chem. Soc. Faraday Trans. 2 81, 1573
- 105. Saunders, M., Vogel, P., Hagen, L. and Rosenfeld, J. (1973). Acc. Chem Res. 6, 53
- 106. Sieber, S., Buzek, P., Schleyer, P.v.R., Koch, W. and de M. Carneiro, J.W. (1993). J. Am. Chem. Soc. 115, 259
- 107. Lias, S.G., Shold, D.M. and Ausloos, P. (1980). J. Am. Chem. Soc. 102, 2540
- 108. Bittner, E.W., Arnett, E.M. and Saunders, M. (1976). J. Am. Chem. Soc. 98, 3734
- 109. Arnett, E.M. and Petro, C. (1978). J. Am. Chem. Soc. 100, 5408
- 110. Lossing, F.P. and Semeluk, G.P. (1970). Can. J. Chem. 48, 955
- 111. Yannoni, C.S., Kendrick, R.D., Myhre, P.C., Bebout, D.C. and Peterson, B.L. (1989). J. Am. Chem. Soc. 111, 6440
- 112. Hollenstein, S. and Laube, T. (1993). J. Am. Chem. Soc. 115, 7240
- 113. Keister, J.W., Riley, J.S. and Baer, T. (1993). J. Am. Chem. Soc. 115, 12613
- 114. Traeger, J.C. (1996). Rapid Commun. Mass Spectrom. 10, 119
- 115. Value from Prof. Taft's group as anchored to other data in Ref. 43.
- 116. Smith, B.J. and Radom, L. (1998). J. Phys. Chem. A 102, 10787
- 117. Olah, G.A. and Donovan, D.J. (1978). J. Am. Chem. Soc. 100, 5163
- 118. Solomon, J.J., Meot-Ner, M. and Field, F.H. (1974). J. Am. Chem. Soc. 96, 3727
- 119. Goren, A. and Munson, B. (1976). J. Phys. Chem. 80, 2848
- 120. Schleyer, P.v.R., de M. Carneiro, J.W., Koch, W. and Forsyth, D.A. (1991). J. Am. Chem. Soc. 113, 3990
- 121. Solomon, J.J. and Field, F.H. (1975). J. Am. Chem. Soc. 97, 2625

THERMODYNAMIC STABILITIES OF CARBENIUM IONS

- 122. Glukhovtsev, M.N., Laiter, S. and Pross, A. (1996). J. Phys. Chem. 100, 17801
- 123. DeFrees, D.J., McIver, R.T., Jr. and Hehre, W.J. (1980). J. Am. Chem. Soc. 102, 3334
- 124. Taylor, C.A., Zerner, M.C. and Ramsey, B. (1986). J. Organomet. Chem. 317
- 125. Schleyer, P.v.R. and Bermer, M. (1988). J. Org. Chem. 53, 2362
- 126. Aue, D.H. (1997). In *Dicoordinated Carbocations and Related Species*. Rappoport, Z. and Stang, P.J. (eds), . Wiley, New York
- 127. Carey, F.A. and Sundberg, R.J. (2000). Advanced Organic Chemistry, Part A (4th ed), p. 617. Plenum Press, New York
- 128. Woodward, R.B. and Hoffmann, R. (1970). *The Conservation of Orbital Symmetry*. Academic Press, New York
- 129. Olah, G.A., Liang, G., Ledlie, D.B. and Costopoulos, M.G. (1977). J. Am. Chem. Soc. 99, 4196
- 130. Saunders, M., Laidig, K.E., Wiberg, K.B. and Schleyer, P.v.R. (1988). J. Am. Chem. Soc. 110, 7652
- 131. Koch, W., Liu, B. and DeFrees, D.J. (1988). J. Am. Chem. Soc. 110, 7325
- 132. Hoffmann, R. (1964). J. Chem. Phys. 40, 2480
- 133. Hehre, W.J., Radom, L., Schleyer, P.v.R. and Pople, J.A. (1986). *Ab Initio Molecular Orbital Theory*, p. 390. Wiley, New York
- 134. Jorgensen, W.L. and Salem, L. (1973). *The Organic Chemist's Book of Orbitals*, p. 35. Academic Press, New York
- 135. Walsh, A.D. (1947). Nature 159, 712
- 136. Walsh, A.D. (1949). Trans. Faraday Soc. 45, 179
- 137. de Meijere, A. (1979). Angew. Chem. Int. Ed. Engl. 18, 809
- 138. Sugden, T.M. (1947). Nature (London) 160, 367
- 139. Honegger, M., Heilbronner, E. and Schwenger, A. (1982). Nouv. J. Chim. 6, 519
- 140. Saunders, M. and Siehl, H.U. (1980). J. Am. Chem. Soc. 102, 6868
- 141. Holman, R.W., Plocica, J., Blair, L., Giblin, D. and Gross, M.L. (2001). J. Phys. Org. Chem. 14, 17
- 142. Olah, G.A. and White, A.M. (1969). J. Am. Chem. Soc. 91, 3954
- 143. Myrhe, P.C., Kruger, J.D., Hammond, B.L., Lok, S.M., Yannoni, C.S., Macho, V., Limbach, H.H. and Vieth, H.M. (1984). J. Am. Chem. Soc. 106, 6079
- 144. Saunders, M. and Vogel, P. (1971). J. Am. Chem. Soc. 93, 2559
- 145. Saunders, M. and Kates, M.R. (1978). J. Am. Chem. Soc. 100, 7082
- 146. Schleyer, P.v.R., de M. Carneiro, J.W., Koch, W. and Raghavachari, K. (1989). J. Am. Chem. Soc. 111, 5475
- 147. Solomon, J.J. and Field, F.H. (1976). J. Am. Chem. Soc. 98, 1567
- 148. Meot-Ner, M., Solomon, J.J. and Field, F.H. (1976). J. Am. Chem. Soc. 98, 1025
- 149. Sharma, R.B., Sen Sharma, D.K., Hiraoka, K. and Kebarle, P. (1985). J. Am. Chem. Soc. 107, 3747
- 150. Olah, G.A., Kelly, D.P., Jeuell, G.L. and Porter, R.D. (1970). J. Am. Chem. Soc. 92, 2544
- 151. Falkenberg-Andersen, C., Ranganayakulu, K., Schmitz, L.R. and Sorensen, T.S. (1984). *J. Am. Chem. Soc.* **106**, 178
- 152. Olah, G.A. and Liang, G. (1973). J. Am. Chem. Soc. 95, 3792
- 153. Kabakoff, D.S. and Namanworth, E.J. (1970). J. Am. Chem. Soc. 92, 3234
- 154. Childs, R.F., Faggiani, R., Lock, C.J.L., Mahendran, M. and Zweep, S.D. (1986). J. Am. Chem. Soc. 108, 1962
- 155. Reindl, B., Clark, T. and Schleyer, P.v.R. (1998). J. Phys. Chem. A 102, 8953
- 156. Abboud, J. -L. M.; Alkorta, I.; Dávalos, J.; Müller, P.; Quintanilla, E.; Rossier, J.-C.; Submitted for publication
- 157. Deno, N.C., Jaruzelski, J.J. and Schriesheim, A. (1955). J. Am. Chem. Soc. 77, 3044
- 158. Deno, N.C., Richey, H.G., Jr., Liu, J.D., Hodge, J.D., Houser, J.J. and Wisotsky, M.J. (1962). J. Am. Chem. Soc. 84, 1962

- 159. Pittman, C.U., Jr. and Olah, G.A. (1965). J. Am. Chem. Soc. 87, 5123
- 160. Olah, G.A., Prakash, R.V., Rasul, G. and Prakash, G.K.S. (1999). J. Am. Chem. Soc. 121, 9994
- 161. Schindler, M. (1987). J. Am. Chem. Soc. 109, 1020
- 162. Müller, P. and Mareda, J. (1995). J. Phys. Org. Chem. 8, 507
- 163. Müller, P. and Mareda, J. (1990). *Cage Hydrocarbons*. Olah, G.A. (ed.), Chapter 6. Wiley, New York
- 164. Schleyer, P.v.R., Fort, R.C., Watts, W.E., Comisarow, M.B. and Olah, G.A. (1964). J. Am. Chem. Soc. 86, 5679
- 165. Winstein, S. and Trifan, D.S. (1949). J. Am. Chem. Soc. 71, 2953
- 166. Schleyer, P.v.R. (1964). J. Am. Chem. Soc. 86, 1856
- 167. Brown, H.C. (1977). The Nonclassical Ion Problem. Plenum Press, New York
- 168. Grob, C.A. (1983). Acc. Chem. Res. 16, 426
- 169. Olah, G.A., Surya Prakash, G.K. and Saunders, M. (1983). Acc. Chem. Res. 16, 440
- 170. Brown, H.C. (1983). Acc. Chem. Res. 16, 432
- 171. Olah, G.A., Prakash, G.K.S., Arvaghi, M. and Anet, F.A. (1985). J. Am. Chem. Soc. 107, 3747
- 172. Jarret, R.M. and Saunders, M. (1987). J. Am. Chem. Soc. 109, 3365
- 173. Yannoni, C.S., Macho, V. and Myhre, P.C. (1982). J. Am. Chem. Soc. 104, 7380
- 174. Koch, W., Liu, B., DeFrees, D.J., Sunko, D.E. and Vancik, H. (1990). *Angew. Chem. Int. Ed. Engl.* **29**, 183
- 175. Johnson, S.A. and Clark, D.T. (1988). J. Am. Chem. Soc. 110, 4112
- 176. Perera, S.A. and Bartlett, R.J. (1996). J. Am. Chem. Soc. 118, 7849
- 177. Werstiuk, N.H. and Muchall, H.M. (2000). J. Phys. Chem. A 104, 2054
- 178. Mesić, M., Sunko, D.E. and Vančik, H. (1994). J. Chem. Soc. Perkin Trans. 2, 1135
- 179. Sieber, S., Schleyer, P.v.R., Vančik, H., Mesić, M. and Sunko, D.E. (1993). Angew. Chem. Int. Ed. Engl. 32, 1604
- 180. Schleyer, P.v.R. and Sieber, S. (1993). Angew. Chem. Int. Ed. Engl. 32, 1606
- 181. Abboud, J.-L. M.; Alkorta, I.; Dávalos, J.; Müller, P.; Rossier, J.-C. Submitted for publication
- 182. Schleyer, P.v.R., Fort, R.C., Jr., Watts, W.E., Comisarow, M.B. and Olah, G.A. (1964). J. Am. Chem. Soc. 86, 4195
- 183. Olah, G.A., Prakash, G.K.S., Shih, J.G., Krishnamurty, V.V., Mateescu, G.D., Liang, G., Sipos, G., Buss, V., Gund, T.M. and Schleyer, P.v.R. (1985). J. Am. Chem. Soc. 107, 2764
- 184. Olah, G.A., Svoboda, J.J. and Ku, A.T. (1973). Synthesis 492
- 185. Vančik, H., Percac, K. and Sunko, D.E. (1990). J. Am. Chem. Soc. 112, 7418
- 186. Houriet, R. and Schwarz, H. (1979). Angew Chem. Int. Ed. Engl. 91, 1018
- 187. Sharma, R.B., Sen Sharma, D.K., Hiraoka, K. and Kebarle, P. (1982). J. Am. Chem. Soc. 104, 7105
- 188. Dutler, R., Rauk, A., Sorensen, T.S. and Whitworth, S.M. (1989). J. Am. Chem. Soc. 111, 9024
- 189. Aubry, C., Holmes, J.L. and Walton, C.W. (1998). J. Phys. Chem. A 102, 1389
- 190. Kruppa, G.H. and Beauchamp, J.L. (1986). J. Am. Chem. Soc. 108, 2162
- 191. Abboud, J.-L.M., Castaño, O., Dávalos, J.Z., Jiménez, P., Gomperts, R. and Roux, M.V. (2002). J. Org. Chem. 67, 1057
- 192. Abboud, J.-L.M., Castaño, O., Dávalos, J.Z. and Gomperts, R. (2001). Chem. Phys. Lett. 337, 327
- 193. Abboud, J.-L. M.; Alkorta, I.; Dávalos, J.; Müller, P.; Rossier, J. C. Submitted for publication
- 194. Smith, D., Adams, N.G. and Ferguson, E.E. (1984). Int. J. Mass Spectrom. Ion Proc. 61, 15

- 195. Berkowitz, J., Mayhew, C.A. and Rušcic, B. (1988). J. Chem. Phys. 88, 7396
- 196. Glukhovtsev, M.N. and Bach, R.D. (1998). Chem. Phys. Lett. 286, 51
- 197. Bogey, M., Cordonnier, M., Demuynck, C. and Destombes, J.L. (1992). Astrophys. J. 399, L103
- 198. Bogey, M., Bolvin, H., Cordonnier, M., Demuynck, C., Destombes, J.L., Escribano, R. and Gómez, P.C. (1994). Can. J. Phys. 72, 967
- 199. Crofton, M.W., Jagod, M.-F., Rehfuss, B.D. and Oka, T. (1989). J. Chem. Phys. 91, 5139
- 200. Gabrys, C.M., Uy, D., Jagod, M.-F. and Oka, T. (1995). J. Phys. Chem. 99, 15611
- 201. Lammerstma, K. and Ohwada, T. (1996). J. Am. Chem. Soc. 118, 7247
- 202. Apeloig, Y. and Muller, T. (1997). In *Dicoordinated Carbocations and Related Species*. Rappoport, Z. and Stang, P.J. (eds), . Wiley, New York
- 203. Bagno, A. and Modena, G. (1999). Eur. J. Org. Chem. 2893
- 204. Tse, J.S., Klug, D.D. and Laasonen, K. (1995). Phys. Rev. Lett. 74, 876
- 205. Marx, D. and Parrinello, M. (1996). Science 271, 179
- 206. Botschwina, P., Oswald, R., Flügge, J. and Horn, M. (1995). Z. Phys. Chem. 188, 29
- 207. Gilbert, T., Pfab, R., Fischer, I. and Chen, P. (2000). J. Chem. Phys. 112, 2575
- 208. Harkless, J.A.W. and Lester, A.W., Jr. (2000). J. Chem. Phys. 113, 2680
- 209. Gobbi, A. and Frenking, G. (1994). J. Am. Chem. Soc. 110, 9275
- 210. Mo, Y., Lin, Z., Wu, W. and Zhang, Q. (1996). J. Phys. Chem. 100, 6469
- 211. Houle, F.A. and Beauchamp, J.L. (1978). J. Am. Chem. Soc. 100, 3290
- 212. Ellison, G.B., Davico, G.E., Bierbaum, V.M. and DePuy, C.H. (1996). Int. J. Mass. Spectrom. Ion Proc. 156, 109
- 213. Lossing, F.P. (1972). Can. J. Chem. 50, 3973
- 214. Value reported in Ref. 43 as "Unpublished results by Ausloos, P., and Lias, S. G."
- 215. Craig, N.C., Pranata, J., Reinganum, S.J., Sprague, J.R. and Stevens, P.S. (1986). *J. Am. Chem. Soc.* **108**, 4378. This study includes all the possible deuterated isotopomers.
- 216. Xie, Y. and Boggs, J.E. (1989). J. Chem. Phys. 90, 4320
- 217. Lee, T.J., Willetts, A., Gaw, J.F. and Handy, N.C. (1989). J. Chem. Phys. 90, 4330
- 218. Salcedo, R. and Olvera, C. (1999). J. Mol. Struct (Theochem) 460, 221
- 219. Olah, G.A., Staral, J.S., Spear, R.J. and Liang, G. (1975). J. Am. Chem. Soc. 97, 5489
- 220. Haddon, R.C. (1975). J. Am. Chem. Soc. 97, 3608 and references therein
- 221. Krygowski, T.M., Cyrañski, M.K., Czarnocki, Z., Häfelinger, G. and Katritzky, A.R. (2000). *Tetrahedron* 56, 1783. This important report contains a wealth of references to previous studies on aromaticity and cognate subjects
- 222. Minkin, V.I., Glukhovtsev, M.N. and Simkin, B.Ya. (1994). Aromaticity, and Antiaromaticity. Electronic and Stuctural Effects, Chapter 6. Wiley, New York
- Sieber, S., Schleyer, P.v.R., Otto, A.H., Gauss, J., Reichel, F. and Cremer, D. (1993). J. Phys. Org. Chem. 6, 445
- 224. Maksic, Z.B., Kovacevic, B. and Lesar, A. (2000). Chem. Phys. 253, 59
- 225. Winstein, S., Shatavsky, M., Norton, C. and Woodward, R.B. (1955). J. Am. Chem. Soc. 77, 4183
- 226. Richey, H.G., Jr. and Lustgarten, R.K. (1966). J. Am. Chem. Soc. 88, 3136
- 227. Brookhart, M., Diaz, A. and Winstein, S. (1966). J. Am. Chem. Soc. 88, 3135
- 228. Olah, G.A., Liang, G., Mateescu, G.D. and Riemenschneider, J.L. (1973). J. Am. Chem. Soc. 95, 8698
- 229. Olah, G.A. and Liang, G. (1975). J. Am. Chem. Soc. 97, 6803
- 230. Laube, T. (1989). J. Am. Chem. Soc. 111, 9224
- 231. Bremer, M., Schötz, K., Schleyer, P.v.R., Fleischer, U., Schindler, M., Kutzelnigg, W., Koch, W. and Pulay, P. (1989). Angew. Chem. Int. Ed. Engl. 28, 1042
- 232. Schulman, J.M., Disch, R.L., Schleyer, P.v.R., Bühl, M., Bremer, M. and Koch, W. (1992). J. Am. Chem. Soc. 114, 7897
- 233. Winstein, S. and Ordronneau, C. (1960). J. Am. Chem. Soc. 82, 2084

- 234. Vogel, P. (1985). Carbocation Chemistry, p. 256. Elsevier, Amsterdam
- 235. Doering, W.v.E. and Knox, L.H. (1954). J. Am. Chem. Soc. 76, 3203
- 236. Merling, G. (1881). Chem. Ber. 24, 3108
- 237. Hückel, E.Z. (1931). Z. Phys. 70, 204
- 238. Spiesecke, H. and Schneider, W.G. (1961). Tetrahedron Lett. 468
- 239. Garratt, P.J. (1986). Aromaticity. Wiley, New York
- 240. Engel, P., Keller, U.M., Bigler, P. and Neunschwander, M. (1976). *Helv. Chim. Acta* **59**, 2344
- 241. Takahashi, Y., Sankararaman, S. and Kochi, J.K. (1989). J. Am. Chem. Soc. 111, 2954
- 242. Nicolaides, A. and Radom, L. (1996). J. Am. Chem. Soc. 118, 10561
- 243. Moon, J.H., Choe, J.C. and Kim, M.S. (2000). J. Phys. Chem. A 104, 458
- 244. Lifshitz, C. (1994). Acc. Chem. Res. 27, 138
- 245. Elder, F.A. and Parr, A.C. (1969). J. Chem Phys. 50, 1027
- 246. Thrush, B.A. and Zwonenik, J.J. (1963). Discuss. Faraday Soc. 35, 196
- 247. Lee, E.P.F. and Wright, T.G. (1998). J. Phys. Chem. A 102, 4007
- 248. Smith, B.J. and Hall, N.E. (1997). Chem. Phys. Lett. 279, 165
- 249. Fujisaki, N., Comte, P. and Gaüman, T. (1993). J. Chem. Soc. Chem. Commun. 848
- 250. McClelland, R.A., Chan, C., Cozens, F., Modro, A. and Steenken, S. (1991). Angew. Chem. Int. Ed. Engl. 30, 1337
- 251. Olah, G.A., Shamma, T., Burrichter, A., Rasul, G. and Surya Prakash, G.K. (1997). J. Am. Chem. Soc. 119, 12923
- 252. Eiden, G.C., Weinhold, F. and Weisshaar, J.C. (1991). J. Chem. Phys. 95, 8665
- 253. Eiden, G.C., Lu, K.-T., Badenhoop, J., Weinhold, F. and Weisshaar, J.C. (1996). *J. Chem. Phys.* **104**, 8886
- 254. Sen Sharma, D.K. and Kebarle, P. (1981). Can. J. Chem. 59, 159
- 255. Smith, B.J. and Hall, N.E. (1997). Chem. Phys. Lett. 279, 165
- 256. Brown, H.C. and Okamoto, Y. (1958). J. Am. Chem. Soc. 80, 4979
- 257. Takeuchi, K., Takasuka, M., Ohga, Y. and Okazaki, T. (1999). J. Org. Chem. 64, 2375
- Cozens, F.L., Kanagasabapathy, V.M., McClelland, R.A. and Steenken, S. (1999). Can. J. Chem. 77, 2069
- 259. Olah, G.A. (1964). J. Am. Chem. Soc. 86, 932
- 260. Farnum, D.G. (1964). J. Am. Chem. Soc. 86, 934
- 261. Farnum, D.G. (1966). J. Am. Chem. Soc. 88, 2970
- 262. Laube, T., Olah, G.A. and Bau, R. (1997). J. Am. Chem. Soc. 119, 3087
- 263. McClelland, R.A., Kanagasabapathy, V.M., Banait, N.S. and Steenken, S. (1989). J. Am. Chem. Soc. 111, 3966
- 264. Ma, M. and Johnson, K.E. (1995). J. Am. Chem. Soc. 117, 1508
- 265. Pitchumani, K., Lakshminarasimhan, P.H., Prevost, N., Corbin, D.R. and Ramamurthy, V. (1997). Chem. Commun. 181
- 266. Norris, F.F. and Sanders, W.W. (1901). J. Am. Chem. Soc. 25, 54
- 267. Kehrmann, F. and Wentzel, F. (1901). Ber. Dtsch. Chem. Ges. 34, 3815
- 268. Gomberg, M. (1901). J. Am. Chem. Soc. 25, 328
- 269. v. Baeyer, A. (1905). Ber. Dtsch. Chem. Ges. 38, 569
- 270. This was confirmed by cryoscopy:Hantzsch, A. (1907). Z. Phys. Chem. 61, 257
- 271. Ferguson, L.N. (1948). Chem. Rev. 43, 385
- 272. Sharp, D.W.A. and Sheppard, N. (1957). J. Chem. Soc. 674
- 273. Moodie, R.B., Connor, T.M. and Stewart, R. (1959). Can. J. Chem. 37, 1402
- 274. Olah, G.A., Baker, E.B. and Comisarow, M.B. (1964). J. Am. Chem. Soc. 86, 1265
- 275. Tao, T. and Maciel, G.E. (1995). J. Am. Chem. Soc. 117, 12889
- 276. Avendaño, C., de Diego, C. and Elguero, J. (1990). Magn. Reson. Chem. 28, 1011
- 277. Gomes da Mesquita, A.H., MacGillvary, C.H. and Eriks, H. (1965). *Acta Crystallogr*. **18**, 437

- 278. Wolf, J.F., Abboud, J.-L.M. and Taft, R.W. (1977). J. Org. Chem. 42, 3316
- 279. Using scaled data from Ref. 43 and data for neutrals from Ref. 26
- 280. Gold, V. and Hawes, B.W.V. (1951). J. Chem. Soc. 2102
- 281. Deno, N.C. and Schriesheim, A. (1954). J. Am. Chem. Soc. 77, 3051
- 282. Arnett, E.M. and Bushick, R.D. (1964). J. Am. Chem. Soc. 86, 1564
- 283. Cox, R.A. (2000). Adv. Phys. Org. Chem. 35, 1
- 284. Wolf, J.F., Harch, P. and Taft, R.W. (1975). J. Am. Chem. Soc. 97, 2904
- 285. Amyes, T.L., Richard, J.P. and Novak, M. (1992). J. Am. Chem. Soc. 114, 8032
- 286. Richard, J.P., Amyes, T.L., Lin, S.-S., O'Donoghue, A., Toteva, M.M., Tsuji, Y. and Williams, K.B. (2000). Adv. Phys. Org. Chem. 35, 67
- 287. Hine, J. (1975). Structural Effects on Equilibria in Organic Chemistry, Chapter 7. Wiley, New York
- 288. Ito, S., Morita, N. and Asao, T. (1994). Tetrahedron Lett. 35, 751
- 289. Arnett, E.M. (1999). *J. Chem. Thermodyn.* **31**, 711. This recent paper contains a number of references to the vast and fundamental work of this author in the field of solution thermochemistry of carbenium ions
- 290. Arnett, E.M. and Hofelich, T.C. (1983). J. Am. Chem. Soc. 105, 2889
- 291. Arnett, E.M. and Pienta, N.J. (1980). J. Am. Chem. Soc. 102, 3328
- 292. Arnett, E.M. and Abboud, J.-L.M. (1975). J. Am. Chem. Soc. 97, 3865
- 293. Foote, C.S. (1964). J. Am. Chem. Soc. 86, 1853
- 294. Bingham, C.R. and Schleyer, P.v.R. (1971). J. Am. Chem. Soc. 93, 3189
- 295. Parker, W., Trauter, R.L., Watt, C.I.F., Chang, L.W.K. and Schleyer, P.v.R. (1974). *J. Am. Chem. Soc.* **96**, 7121
- 296. Schneider, H.J. and Thomas, F. (1980). J. Am. Chem. Soc. 102, 1424
- 297. Schneider, H.J., Schmidt, G. and Thomas, F. (1983). J. Am. Chem. Soc. 105, 3356
- 298. Müller, P., Blanc, J. and Perlberger, J.C. (1982). Helv. Chim. Acta 65, 1418
- 299. Müller, P., Mareda, J. and Milin, D. (1995). J. Phys. Org. Chem. 8, 507
- 300. Smith, M.R. and Harris, J.M. (1978). J. Org. Chem. 43, 3588
- 301. Farcasiu, D. (1978). J. Org. Chem. 43, 3878
- 302. Lenoir, D. and Frank, R.M. (1981). Chem. Ber. 114, 3336
- 303. Müller, P., Milin, D., Feng, W.Q., Houriet, R. and Della, E.W. (1992). J. Am. Chem. Soc. 114, 6169
- 304. Bentley, T.W. and Roberts, K. (1985). J. Org. Chem. 50, 5852
- 305. Müller, P.; Abboud, J.-L. M.; Rossier, J.-C. Unpublished
- 306. Brown, H.C. and Rei, M.-H. (1964). J. Am. Chem. Soc. 86, 5008
- 307. Mayr, H., Bug, T., Gotta, M.F., Hering, N., Irrgang, B., Janker, B., Kempf, B., Loos, R., Offial, A.R., Remennikov, G. and Schimmel, H. (2001). J. Am. Chem. Soc. **123**, 9500
- 308. Mayr, H. and Patz, M. (1994). Angew. Chem. Int. Ed. Engl. 33, 938
Transition State Analysis Using Multiple Kinetic Isotope Effects: Mechanisms of Enzymatic and Non-enzymatic Glycoside Hydrolysis and Transfer

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1 Introduction

This chapter addresses both the principles and practice of transition state $(TS^{\#1})$ analysis and recent advances in our understanding of hydrolysis and glycosyl transfer reactions of *N*- and *O*-glycosides. By transition state analysis, we mean specifically the use of multiple (or families of) kinetic isotope effects (KIEs) to define transition states in atomic detail. In the best cases, it has been possible to determine the structure of the transition state, a species that exists for less than a single bond vibration (*ca.* 10^{-13} s), with accuracy that rivals that of X-ray crystallography of stable molecules. These studies have elucidated both the chemistry of these reactions and the strategies used by enzymes to promote catalysis. The use of TS analysis to build quantitative, atomic resolution transition state models is still a relatively new technique. In this chapter, we describe the techniques of TS analysis as applied to glycoside chemistry and recent results that have emerged from these studies.

GLYCOSIDE CHEMISTRY

There is a long history of exploration of *O*- and *N*-glycoside chemistry by physical organic chemists.^{1–5} Biologists have long been interested in systems involving glycoside chemistry, such as cell–cell contacts and signaling; nucleic acid synthesis, repair and recycling; glycoprotein maturation, folding, and degradation; intracellular signaling; microbial (viral, bacterial and fungal) pathogenicity; apoptosis; cancer; lysosomal storage diseases; and even wood degradation.^{6–8} Recent advances in chemical and enzymatic synthesis and characterization (e.g., see Ref. 9) have made complex oligosaccharides experimentally accessible. The division of glycoside chemistry into pyranosides and furanosides coincides well with the division between *O*-glycosides (pyranosides), and *N*-glycosides (furanosides). The notable exception is the study of cationic *N*-glucosides by Bennet and coworkers, which have shed light on both classes of sugars.

We discuss in this chapter how TS analysis allows chemists to ask sophisticated and subtle questions of enzymatic and non-enzymatic glycoside hydrolysis and transfer reactions. It is now becoming possible to measure routinely (if perhaps not

^{#1} α_n , the intrinsic KIE on rate constant k_n ; ψ U, pseudouridine; ADP, adenosine 5'-diphosphate; AMP, adenosine 5'-monophosphate; ATP, adenosine 5'-triphosphate; BOVA, bond order vibrational analysis; CMP-NeuAc, cytidine monophosphate glycoside of *N*-acetylneuraminic acid; DFT, density functional theory, DTA, diphtheria toxin A-chain; EIE, equilibrium isotope effect; IE, isotope effect; immucillin H, (1*S*)-1-(9-deazahypozanthin-9-yl)-1,4-dideoxy-1,4-imino-D-ribitol; IU-NH, inosine –uridine-specific nucleoside hydrolase; KIE, kinetic isotope effect; MP2, second order Moller–Plesset perturbation theory; NAD⁺, oxidized nicotinamide adenine dinucleotide; n_{LG} , n_{Nu} , Pauling bond order to the leaving group or nucleophile; OPRTase, orotate phosphoribosyltransferase; pAPIR, *p*-aminophenyl iminoribitol; PNP, purine nucleoside phosphorylase; RTA, ricin toxin A-chain; SDKIE, solvent deuterium KIE; TS, transition state; UDG, uracil DNA glycosylase; UMP, uridine 5'-monophosphate.



Fig. 1 Simultaneous general acid and general base catalysis of glycoside hydrolysis.

yet easily) experimental KIEs and from them deduce in detail the structure and other essential features of the transition state.

ENZYMATIC VERSUS NON-ENZYMATIC TRANSITION STATES

Enzymes catalyze reactions by binding to transition states, lowering their energy. However, the transition state to which the enzyme binds is not necessarily the same as for the non-enzymatic reaction. TS analysis has shown that various enzymes can give different TS structures for the same reaction, which are in turn different from their non-enzymatic counterparts. Enzymes often do not merely stabilize the transition state; they change it. This is a reflection of the fact that the environment of an enzyme active site is different from aqueous solution. One crucial difference between an enzyme active site and aqueous solution is that the former can be viewed as a pre-organized "supersolvent".¹⁰ Using the example of O-glycoside hydrolysis, there would be catalytic advantage to protonating the alkoxide leaving group and deprotonating the water nucleophile simultaneously (Fig. 1), but such a transition state is highly unlikely in solution. The probability of these events happening simultaneously is vanishingly small because of the high entropic cost of forming a tetramolecular transition state (glycoside, nucleophile, general acid, general base), and the unlikelihood of having solvent molecules acting as both a general acid and a general base in a single solution. In an enzyme, however, suitably positioned functionalities in the active site can act as general acids and bases simultaneously. The entropic cost of transition state formation is mitigated by: (1) the need to form only a termolecular transition state (glycoside, nucleophile, enzyme); (2) the use of binding energy with the substrates to compensate for the entropic cost.

Retaining and inverting *O*-glycosidases make use of a general acid/general base pair comprised of a carboxylic acid and carboxylate of aspartic acid or glutamic acid side chains (see also Fig. 21). These side chains have similar pK_a values in solution; but the micro-environments of enzyme active sites allow each pK_a to be tuned so that each side chain is in the correct protonation state. This makes it possible for a carboxylate and a carboxylic acid to exist in the active site simultaneously.

Enzymes do not merely stabilize transition states, but can alter their structures, even whole reaction pathways to achieve catalysis. We can probe the potential energy surface of a reaction directly through computations or indirectly through experimental studies, but TS analysis can show how the potential energy surface itself is modified to promote catalysis.

THE PRACTICE OF TS ANALYSIS

Overview of TS analysis

TS analysis involves a number of steps. It is necessary that the kinetic mechanism of the reaction be thoroughly characterized. Conditions must be found where the experimental KIEs reflect the chemical steps of the reaction rather than binding events or conformational changes. Isotopically labeled substrates are then synthesized and KIEs measured.

Advances in measurement. Recent emphasis on biologically relevant systems has created special challenges and unique solutions to the challenges of KIE measurement. Synthesis of complex molecules labeled at a variety of positions has been made possible by the routine availability of recombinant enzymes to build up complex biosynthetic pathways in a test tube. The "traditional" techniques of measuring KIEs by isotope ratio mass spectrometry¹¹ or scintillation counting of radiolabeled compounds,¹² are now being complemented by methods such as wholemolecule mass spectrometry,¹³⁻¹⁷ short-lived isotopes¹⁸ and novel spectrophotometric techniques.¹⁹⁻²¹ These new methods hold the promise of greater versatility than isotope ratio mass spectrometry and both easier synthesis and faster measurements than radiolabeling techniques. One approach that holds great promise is the determination of KIEs on natural abundance isotopes (i.e. unlabeled compounds) using NMR to measure relative isotopic abundances in residual substrate after the reaction has been run to 90-99% completion.^{22,23} By eliminating the need to synthesize labeled compounds and offering the promise of measuring KIEs on every non-exchangeable nucleus in a single reaction, the technique of Singleton and coworkers holds great promise.

Advances in interpretation. Experimental KIEs are interpreted using quantum mechanical and/or bond order vibrational analysis (BOVA) approaches to yield the experimental transition state. In systems that are well understood, the accuracy of experimental TS structures derived using a BOVA "unified model" rivals X-ray crystallography of stable molecules. The largest advance in KIE interpretation since the description of BOVA by Sims and Lewis^{24,25} has been the increase in computational power available to the average chemist. Previously, vibrational models were reduced to a minimal number of atoms to make them computationally tractable. Today, a desktop computer can perform BOVA calculations with hundreds of atoms. The accessibility of post-Hartree–Fock calculations to the average chemist has made *ab initio* calculations a routine part of TS analysis, and made possible the structure interpolation approach to BOVA.

If possible, one of the transition states solved should be the non-enzymatic analogue of the enzymatic reaction. The non-enzymatic reaction acts as a reference for which there is frequently a variety of other experimental evidence to guide KIE interpretation and help validate the experimental transition state. In addition, the difference between the transition state for the enzymatic and non-enzymatic reactions reflects the interaction between the substrate's intrinsic reaction pathway^{#2} and the enzyme's use of binding energy to lower the energetic barrier to the transition state.

The theoretical and practical challenges of TS analysis remain formidable, but the increase in our understanding of chemical systems from each experimental transition state, and the impossibility of achieving these results by any other technique, fully justify the effort.

TS analysis as part of a research program

As powerful as TS analysis is, no single experimental technique can provide all the information required to understand an enzymatic system in detail. TS analysis is part of a suite of techniques that are necessary to characterize the structure and catalytic activity of enzymes.

A recent publication on purine nucleoside phosphorylase (PNP)²⁶ represents the end result of a systematic program of TS analysis^{27,28} combined with protein dynamic,²⁹ structural³⁰ and functional analyses^{31,32} that have yielded profound insights into the catalytic mechanisms of this enzyme, as well as powerful inhibitors with therapeutic potential.^{32,33} TS analysis demonstrated that the reaction proceeded with a highly dissociative^{#3} $A_N D_N$ mechanism, passing through an oxocarbenium ion-like (but still bimolecular) transition state. Cycles of inhibitor design and testing, as well as theoretical prediction of inhibition constants, resulted in a PNP inhibitor, immucillin H, with a dissociation constant of 23 pM. This inhibitor showed third-ofsites reactivity, consistent with being a good TS mimic, and was shown through solvent-exchange experiments to decrease protein dynamics. TS analysis, combined with the crystal structures of PNP immucillin H cocrystals, as well as cocrystals of a Michaelis complex analogue (substituting sulfate for phosphate) and the reaction products, along with docking of the experimental TS structure, resulted in convincing evidence for an "electrophile migration" mechanism. This is distinct from a more conventional view of nucleophilic substitutions proceeding through nucleophile approach to the electrophilic center concerted with leaving group departure. It is hard to imagine how these results could have been achieved without the use of TS analysis.

^{#2} This is not to say that the non-enzymatic transition state is the "intrinsic" transition state. It will reflect interactions of the molecule with solvent water, salts and other solutes. The intrinsic reaction *in vacuo* is generally not experimentally accessible for biologically relevant molecules.

 $^{^{\}frac{4}{3}}$ Several meanings of the word "dissociative" have been used. We use it to mean that the sum of nucleophile and leaving group bond orders at an $A_N D_N$ transition state are less than the corresponding bond orders in the reactants or products.

TS ANALYSIS AND INHIBITOR DESIGN

From a biological perspective, one of the primary motivations for studying enzymatic mechanisms is inhibitor design. The relevance of the transition state to inhibitor design stems from the application of TS theory to catalysis, which states that enzymes catalyze reactions by binding tightly to the transition state in preference to any other species, thereby stabilizing it.^{34,35} Enzymes typically bind reactants and products with equilibrium dissociation constants in the $10^{-3}-10^{-6}$ M range. Tighter binding would interfere with product release at the end of the catalytic cycle. In contrast, the inferred equilibrium dissociation constant of the enzyme TS complex for the reaction catalyzed by orotidine-5'-phosphate decarboxylase is 10^{-24} M.³⁶ Exploiting the enzyme's affinity for the transition state by synthesizing TS analogues as inhibitors, or in other ways designing mechanism-based inhibitors that rely on the catalytic powers of the enzyme to effect inhibition (β -lactam antibiotics being the classic example) is an effective means to inhibit enzymes.

Resistance is an additional consideration in the design of inhibitors as antimicrobial compounds. With the advent of vancomycin resistance, once thought to be the single antibiotic against which resistance would never arise,³⁷ it is now clear that resistance will eventually arise to every antibiotic. However, it is possible to consider designing TS analogues that are "resistance resistant antibiotics". An enzyme cannot lose affinity for its cognate transition state without also losing its catalytic ability. Thus, resistance to TS analogues must arise through specificity against remote parts of the inhibitor molecule. As long as the target enzyme has catalytic activity, it should be possible in principle to modify the TS analogue to counter changes in the enzyme's specificity.

MECHANISM NOMENCLATURE

According to IUPAC nomenclature, ${}^{38-40}$ a reaction mechanism is divided into elementary steps, with A_N representing a nucleophilic addition and D_N representing a nucleophilic dissociation. A bimolecular (S_N2) reaction is represented as A_ND_N. D_N + A_N and D_N * A_N mechanisms are stepwise (S_N1), with a discrete intermediate formed between leaving group departure and nucleophile approach. A D_N + A_N reaction has a cationic intermediate that is sufficiently long-lived for the leaving group to diffuse away into solution. A D_N * A_N mechanism involves an intermediate that is too short-lived to separate diffusionally from the leaving group. In an enzymecatalyzed reaction, the lifetime *per se* of the intermediate may not be relevant if the leaving group is bound to the enzyme and cannot diffuse away.

Oxocarbenium ions do not generally have a long enough lifetime in solution for diffusive separation of the leaving group. Thus, in the case of enzymatic reactions where an oxocarbenium ion intermediate is formed, it will not be possible for it to diffusionally separate from the enzyme and leaving group, so the mechanistic description $D_N * A_N$ will be used by default for stepwise enzymatic reactions. From the point of view of KIEs, $D_N + A_N$ and $D_N * A_N$ mechanisms are indistinguishable.



(b) Different rate-limiting and first irreversible steps



Fig. 2 Free energy profiles for enzyme-catalyzed reactions. The rate-limiting, partially irreversible and first irreversible steps are labeled for each reaction profile: (a) the simplest case where the rate-limiting and first irreversible steps are the same; (b) the first irreversible step occurring before the rate-limiting step; (c) a high-energy intermediate where its formation is rate-limiting and partially irreversible, while its breakdown is the first (fully) irreversible step; (d) external commitment to catalysis, where formation of the non-covalent Michaelis complex is partially irreversible, and k_3 is the first irreversible step. Each reaction involves association of free enzyme and a single substrate (E + S) to form a non-covalent Michaelis complex (E·S), which then undergoes two chemical steps to first form an intermediate complex (E·I), then the enzyme-product complex (E·P) before dissociation to give free enzyme and product (E + P). In the kinetic mechanisms, irreversible steps are indicated by a single headed arrow, and the reverse rate constants in parentheses.

In stepwise reactions, the double dagger symbol, ‡, is used to indicate the ratelimiting step when it is known.

Some authors have referred to highly dissociative $A_N D_N$ reactions as being " S_N 1-like". If the transition state is bimolecular, with both the nucleophile and the leaving group in the reaction coordinate, then the reaction has an $A_N D_N$ mechanism, regardless of the extents of bond loss or formation at the transition state. In this chapter, we will refer to highly dissociative $A_N D_N$ transition states as being "oxocarbenium ion-like" rather than " S_N 1-like".

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KINETICALLY SIGNIFICANT STEPS: IRREVERSIBLE STEPS AND RATE-LIMITING STEPS
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When performing TS analysis, it is crucially important to know which step(s) of the reaction are being probed by the experimental KIEs. The meaning of KIEs is

different depending on whether they are measured by the competitive or noncompetitive (direct) methods (see below).

Competitive KIEs reflect steps up to and including the first irreversible step of the reaction. In contrast, non-competitive KIEs reflect the rate-limiting step(s). In the simplest case, one step in the kinetic mechanism is both the rate-limiting and the first irreversible step (Fig. 2a). In enzymatic reactions, however, it is common for different steps in the kinetic mechanism to correspond to the rate-limiting step and the first irreversible step (e.g., Fig. 2b). A classic example is the hydrolysis of pnitrophenyl acetate by chymotrypsin.⁴¹ In this reaction, the acylation step (k_3) to form the covalent acetyl-chymotrypsin intermediate is much faster than deacylation (k_5) , but is effectively irreversible because the other product, *p*-nitrophenol, can diffuse away into solution. Under these conditions, the first irreversible step is different from the rate-limiting step. This type of situation is relatively common, and is observed, e.g., in enzymes where product release is the rate-limiting step, but the chemical step is effectively irreversible. Competitive KIEs measured on this reaction would include no contribution from the rate-limiting step and would reflect the reaction only up to the transition state of the first irreversible step. In contrast, non-competitive KIEs would reflect only the rate-limiting step and would include no contribution from the acylation step.

Just as reactions may have more than one partially rate-limiting step, similarly, they may have more than one partially irreversible step.^{#4} One common example in both enzymatic and non-enzymatic reactions is where a high-energy intermediate is formed that can then partition in both the forward (k_5) and reverse (k_4) directions (Fig. 2c). In this example, intermediate formation (k_3) is rate-limiting and partially irreversible, while its breakdown (k_5) is the first (fully) irreversible step. The degree of irreversibility of a given step is determined by the partitioning of the subsequent species forward or backward. In the previous examples (Fig. 2a and b), the energetic barriers after the E-I intermediate ensure it partitions overwhelmingly in the forward direction. In Fig. 2c, steps k_3 and k_5 are both partially irreversible, and competitive KIEs for this reaction would include contributions from both steps. With two different steps contributing to the experimental KIEs, it is likely that they would be uninterpretable for transition state analysis (but see the ricin-catalyzed reactions in Section 4). In contrast, non-competitive KIEs on this reaction would reflect only k_3 , the rate-limiting step.

We use the expression "kinetically significant step" throughout this chapter. Its meaning is context dependent. When the rate-limiting step is under consideration, then if one step is much slower than all others, it is the only kinetically significant step (k_3 in Fig. 2a). If the rates of two steps are similar to each other and much slower than all others, then both are partially rate-limiting and both are kinetically significant. In another context "kinetically significant" will refer to the irreversibility of steps. If every step before the first irreversible step is freely reversible and

^{#4} The expression "partially irreversible" is grammatically suspect, but the alternatives sacrifice clarity for correctness.

therefore at equilibrium (Fig. 2a and b), then it is the only kinetically significant step. If, however, an intermediate is formed that partitions significantly in both directions, then there are two partially irreversible steps (k_3 and k_5 in Fig. 2c), and both steps are kinetically significant. The partitioning ratio (i.e. k_5 versus k_4) determines the degree of irreversibility of formation of an intermediate.

The term "commitment to catalysis"⁴² is unique to KIE studies on enzymes and not widely used in the chemical literature. "External commitment to catalysis" refers to the partitioning of the first intermediate of the reaction, the non-covalent enzyme substrate (E·S) intermediate. Some enzymes are such efficient catalysts that nearly every molecule of substrate that binds proceeds forward to products (i.e. $k_3 \gg k_2$). Under these conditions, the first irreversible step of the reaction is substrate binding. If competitive isotope effects are measured, they will only reflect binding isotope effects, and have no contribution from the chemical steps of the reaction. In less extreme cases (Fig. 2d), both substrate binding (k_1) and the chemical step (k_3) are partially irreversible. "Internal commitment to catalysis" simply refers to the partitioning of any intermediate enzyme-bound species.

2 TS analysis: principles and procedures

THE MEANING OF KIES IN ENZYMATIC REACTIONS

Competitive versus non-competitive KIEs. KIE measurements can be made in a competitive or non-competitive manner.¹² In the non-competitive (or direct) methodology, kinetic constants are determined with the labeled and unlabeled compound in separate experiments, and the ratios of rate constants are compared to give the experimental KIEs. Non-competitive KIEs are generally not sufficiently accurate for transition state analysis, although recently the groups of Huskey and Toney have described methods for accurate measurement of non-competitive KIEs.^{19,20} The advantage of non-competitive KIEs is that it is possible in principle to measure isotope effects on every kinetic parameter (e.g., k_{cat}/K_M , k_{cat} , and K_M) individually.

It is generally necessary to measure competitive KIEs to achieve the confidence intervals of ± 0.002 to ± 0.005 needed to accurately describe a transition state. In competitive measurements, labeled and unlabeled reactants are combined in a single reaction mixture and allowed to react as competitive substrates in the enzymatic reactions (or as competitive reactants in non-enzymatic reactions). If there is an isotope effect at the labeled position, the faster reacting isotope will become depleted in the reactant and enriched in the product and the isotope ratio (unlabeled:labeled) will change. It is this change in isotopic composition that is measured to determine KIEs.

Competitive KIEs in enzymatic reactions are isotope effects on the second-order rate constant k_{cat}/K_{M} , also called the "specificity constant".³⁵ The value of k_{cat}/K_{M} reflects, in the simplest case, the difference between the energy of the free substrate

in solution and the transition state of the *first irreversible step* of the reaction (e.g., Fig. 2a and b).⁴³ After the first irreversible step, the reaction is committed to continuing forward and no further isotopic discrimination can occur. This assumes that all steps before the irreversible step are freely reversible and therefore at equilibrium. If this is not true and there are several partially irreversible steps, each one will contribute to the observed KIE (Fig. 2c and d).

Multiple kinetically significant steps. If a reaction has several steps that are kinetically significant (partially rate-limiting or partially irreversible), then an incremental change in any one of those steps will not be fully reflected in the macroscopic rate constant for the reaction. The practical upshot is that an isotope effect on one microscopic rate constant, k_n , will be reflected in the macroscopic rate constant *only* if k_n is the only kinetically significant step. If there is more than one kinetically significant step, the observed KIE will be less than the KIE on k_n . In the jargon of the field, the KIE on k_n would be "suppressed by kinetic complexity".

Intrinsic KIEs. The phrase "intrinsic KIEs" is often used. This refers to the situation where only chemical steps are kinetically significant, i.e. irreversible or rate-limiting, depending on the context. As the goal of TS analysis is to understand the chemical mechanisms of catalysis, the goal is always to measure intrinsic KIEs that do not contain contributions from non-chemical, isotopically insensitive steps.

Commitment to catalysis. "External commitment to catalysis", or often just "commitment to catalysis" occurs when an enzyme is so efficient that most of the Michaelis complex (E·S) partitions forward to undergo catalysis. When this occurs, substrate association is the first irreversible step, or partially irreversible step. The classic example of this is catalase, where the reaction is close to diffusion ratelimited, and $k_{cat}/K_{\rm M} = 4 \times 10^7 \, {\rm s}^{-1}$.³⁵ It is possible to test for commitment to catalysis using the isotope-trapping methodology of Rose.⁴⁴ If it is not close to zero, conditions may be modified to achieve this, such as using alternate substrates, adjusting reaction conditions, or using mutant enzymes. When commitment to catalysis is close to zero, this implies that substrate association is fully reversible and at equilibrium. The observable KIEs are those of the chemical step(s). If conditions are found where commitment to catalysis is moderate, it can be quantitated and the experimental KIEs corrected numerically,^{44,45} although this correction is only approximate as it does not take into account binding isotope effects (IEs) on association.

Conformational change. Another situation where a non-chemical step can contribute to the experimental KIEs is if a conformational change, such as a loop movement in the protein, is kinetically significant. Kinetically significant conformational changes are not rare in enzymology in general, and have (apparently) been observed in at least four TS analysis studies.^{46–49} Again, if possible, conditions are found where the conformational change is no longer kinetically significant.

Intermediates. Another complication is when an intermediate is formed that partitions to a significant extent both forward and backward. The observable KIEs will be a function of the intermediate partitioning.⁴³ Again, it may be possible to

adjust reaction conditions such that the intermediate partitions predominantly in one direction.

For non-enzymatic reactions, some of the same concerns apply. Kinetically significant substrate binding is not a concern, and conformational change is much less likely to be kinetically significant, however, partitioning of intermediates may have to be considered. In particular, glucopyranosides^{50–53} and sialylpyrano-sides^{54,55} have been shown to proceed through several simultaneous pathways, including direct nucleophilic displacement, formation of intimate ion pairs (or ion-neutral complexes) and solvent-separated ion pairs.

In spite of all these potential complications, it has proven possible in the majority of cases examined to generate experimental KIEs that reflect the chemical steps of the enzymatic reaction. Thus, one may embark on a TS analysis study with a good expectation of obtaining an experimental transition state.

THE REACTION COORDINATE AND THE STRUCTURE OF THE TRANSITION STATE

KIEs on the reaction coordinate. All the experimental and theoretical evidence gathered to date has supported the view that nucleophilic substitutions on glycosides proceed through transition states on the border between $A_N D_N$ and $D_N * A_N$. In conventional physical organic experiments, it is often difficult to distinguish between mechanisms for reactions that are very close to the borderline. In isotope effect studies, however, it is possible to unambiguously make this distinction. In some highly dissociative $A_N D_N$ transition states, the residual bond order^{#5} to the leaving group and the incipient bond order to the incoming nucleophile are so low (<0.05) that it seems they should have little or no observable effect on the KIEs. The question then arises whether these groups are present in the transition state at all. The reaction coordinate^{#6} motion makes a contribution to the primary carbon KIE of \geq 1.01 in addition to the zero point energy and excited state contributions of other vibrational modes. As such, there is a significant difference in calculated isotope effects for the same structure if either the nucleophile or leaving group is left out of the reaction coordinate. A_ND_N reactions are traditionally expected to give primary carbon KIEs in the range of ${}^{13}C$ KIE = 1.03–1.08 or ${}^{14}C$ KIE = 1.06–1.16, while $D_N * A_N$ mechanisms would give ¹³C KIE = 1.00-1.01 or ¹⁴C KIE = 1.00-1.02.⁵⁷ More recent calculations on N-(deoxy-)ribofuranosides have indicated approximate limits of ¹³C KIE = 1.013 - 1.04 or ¹⁴C KIE = 1.025 - 1.075 for A_ND_N mechanisms, and ¹³C KIE = 1.008-1.010 or ¹⁴C KIE = 1.015-1.018 for $D_N * A_N$ mechanisms.⁵⁸ As discussed in Section 4, examples have been found of both highly

^{#5} The Pauling bond order between atoms *i* and *j*, n_{ij} , is $n_{ij} = e^{(r_1 - r_{ij})/0.3}$, where r_{ij} is the bond length between atoms *i* and *j* and r_1 is the bond length for a single bond between atoms of elements *i* and *j*.^{24,56} ^{#6} In this chapter, "reaction coordinate" refers specifically to the vibrational normal mode at the transition state that has an imaginary frequency. Displacement of atoms along this normal mode in either direction results in a decrease in energy and movement of the molecule either forward toward products, or back towards reactants. In an A_ND_N mechanism, the reaction coordinate involves coordinated departure of the leaving group and approach of the nucleophile.

dissociative $A_N D_N$ and true stepwise $D_N * A_N$ reactions for both enzymatic and nonenzymatic reactions.

The structure of the transition state. TS analysis tends to emphasize enzyme– substrate contacts that promote catalysis, and on exploiting these interactions in designing inhibitors. The focus therefore tends to be on the structure of the transition state. However, an experimental transition state necessarily includes information on the reaction coordinate. Even though the focus of TS analyses tends to be on structure, there is more information about catalysis inherently available from the TS determination process.

VERY LOW BOND ORDERS

Glycoside hydrolysis and transfer reactions with $A_N D_N$ mechanisms are highly dissociative, with leaving group and nucleophile bond orders often in the range of 0.005–0.05. Because KIEs are a roughly logarithmic function of bond order,^{#7} a bond order of 0.005 is different than 0.01 from a vibrational point of view, even though it may not be clear what this means structurally. The concept of Pauling bond order arises from empirical observations of molecular structure (e.g., see Ref. 56), which necessarily involved studying bonds in stable molecules. The validity of the concept of bond order for very weak bonds has not been verified experimentally. Some workers have suggested that the usual factor of c = 0.3 used in calculating the Pauling bond order ($n_{ij} = e^{(r_1 - r_{ij})/c}$) should be replaced by $c \ge 0.6$ for bond orders less than 1.^{59,60} The inclusion of leaving groups and nucleophiles with very low bond orders in the TS structure is necessary because of their contribution to the A_ND_N reaction coordinate KIE; however, the structural interpretation of such low bond orders should be approached with caution.

OBSERVABLE KIES FOR STEPWISE REACTIONS

In an $A_N D_N$ mechanism only one chemical step is kinetically significant, the concerted approach of the nucleophile and departure of the leaving group. In stepwise mechanisms, either step, or both, may be kinetically significant and contribute to the observable KIEs. In a $D_N * A_N$ mechanism, the irreversible step can be either leaving group dissociation $(D_N^{\pm} * A_N)$ or nucleophilic attack $(D_N * A_N^{\pm})$; or, if the oxocarbenium ion intermediate partitions in both directions, both steps will be partially irreversible and contribute to the observable KIEs. As competitive KIEs always reflect k_{cat}/K_M , the observable KIEs for a $D_N * A_N$ kinetic mechanism (equation 1) will reflect changes in this kinetic parameter (equation 2), and are given by equation (3), assuming that $k_2 \gg k_3^{58}$

$$\mathbf{E} + \mathbf{S} \stackrel{k_1}{\underset{k_2}{\longleftrightarrow}} \mathbf{E} \cdot \mathbf{S} \stackrel{k_3}{\underset{k_4}{\longleftrightarrow}} [\mathbf{E} \cdot \mathbf{O}] \stackrel{k_5}{\longrightarrow} \mathbf{E} \cdot \mathbf{P} \stackrel{k_7}{\underset{k_8}{\longleftrightarrow}} \mathbf{E} + \mathbf{P}$$
(1)

^{#7} Unpublished data.

$$\frac{k_{\text{cat}}}{K_{\text{M}}} = \frac{k_1 k_3 k_5}{k_2 k_4 \left(1 + \frac{k_5}{k_4} \left(1 + \frac{k_3}{k_2}\right)\right)} \tag{2}$$

$$\text{KIE} = \frac{\frac{\alpha_1 \alpha_3}{\alpha_2} \left(\frac{\alpha_5}{\alpha_4} + \frac{k_5}{k_4}\right)}{1 + \frac{k_5}{k_4}} \tag{3}$$

where α_n is the intrinsic KIE on step *n*.

The observable KIEs depend on the partitioning of the oxocarbenium ion intermediate, i.e. k_5/k_4 (equation 3). If the intermediate partitions completely forward to products, then k_3 is the first irreversible step, $k_5 \gg k_4$, and KIE = $\alpha_1 \alpha_3 / \alpha_2$. If it is in equilibrium with the reactant, then k_5 is the first irreversible step, $k_4 \gg k_5$, and KIE = $(\alpha_1 \alpha_3 \alpha_5)/(\alpha_2 \alpha_4)$.

FROM EXPERIMENTAL KIEs TO TRANSITION STATES

The physical basis of IEs has been described in several excellent reviews.^{57,61,62} IEs are vibrational phenomena. They reflect the change in vibrational environment of an atom between the initial and final state (reactant and transition state for KIEs, reactant and product for equilibrium isotope effects (EIEs)). If there is a change in the structure of the molecule between the initial and final states, there will be a corresponding change in force constants (\approx bond strengths) in the bonds making up the molecule. These cause changes in the vibrational normal modes of the molecule, which in turn result in IEs. In deducing a transition state from experimental KIEs, one is, in essence, deducing the (experimentally inaccessible) vibrational spectrum of the transition state.

Electronic structure methods

Using electronic structure calculations, it is possible to calculate the structures and vibrational frequencies of reactant and TS molecules. From these, one may calculate IEs. Some authors have suggested it is necessary to use *ab initio* methods at the post-Hartree–Fock level to obtain vibrational frequencies accurate enough for IE calculations.^{63,64} Calculations at these levels are computationally expensive, so the "cheapest" available methods are generally used, namely MP2 or density functional theory (DFT) methods. At this level of theory, vibrational frequency calculations can be quite accurate, with overall root mean square errors as low as 34 cm⁻¹.⁶⁵ Because the frequencies calculated by quantum mechanical methods for a given structure are generally quite accurate, the factor limiting the accuracy of computed IEs is the accuracy of the structure. Thus, one may presume that if calculated and experimental KIEs agree well, then the calculated TS structure is accurate.

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Fig. 3 Reaction space for hydrolysis of a glycoside. In this More O'Ferrall–Jencks diagram, the axes are leaving group (n_{LG}) and nucleophile (n_{Nu}) bond orders. The positions of the reactant glycoside (lower left) and the product (top right) in reaction space are shown. In a pure, stepwise $D_N + A_N$ (S_N 1) mechanism, a discrete oxocarbenium ion intermediate is formed by dissociation of the leaving group (top left) before the nucleophile approaches in a separate step. A concerted synchronous $A_N D_N$ (S_N 2) mechanism would follow the dashed diagonal line, with approach of the nucleophile precisely equal to loss of the bond order to the leaving group. A dissociative $A_N D_N$ mechanism would pass through a transition state closer to the top left corner. Each point on the graph represents a trial TS structure generated by the structure interpolation method for which KIEs are calculated.

accurate, partly because of inherent limitations in the computational models, but mostly because solvents and enzymes change transition states. If calculated and experimental KIEs cannot be made to match, there is nothing more that can be done using electronic structure methods. Frequency calculations on quantum mechanical models are valid only at stationary points, i.e. energetic minima or saddle points; vibrational frequencies calculated for arbitrary non-optimized structures, or ones optimized with constraints, have no physical meaning.

Computational models that combine a high level quantum mechanical model of the reaction site surrounded by a layer of lower level semi-empirical or molecular mechanical atoms in the surrounding solvent or enzyme environment will one day make electronic structure calculations of transition states in the condensed phase routine, but these are still some distance in the future. At present, if KIEs calculated from electronic structure models do not match the experimental KIEs, it is necessary to use BOVA to find the transition state.



Fig. 4 Match of calculated to experimental KIEs in reaction space for the diphtheria toxincatalyzed hydrolysis of NAD⁺ (see Ref. 17): (bottom right panel) contour plot of calculated 1^{-15} N KIEs throughout reaction space; (bottom middle) expansion of the dissociative section of reaction space for 1^{-15} N KIEs. The shading shows the areas in each reaction space plot where the calculated KIEs match the experimental KIEs exactly (dark gray) or within the 95% confidence interval (light gray). The open circle shows the location of the experimentally determined transition state in reaction space.

Bond order vibrational analysis (BOVA) and structure interpolation

Using BOVA, one may calculate KIEs for any pair of reactant and TS structures. There is no requirement to use structures that are stationary points according to electronic structure calculations. One creates a vibrational model that treats the reactant and TS molecules as perfect harmonic oscillators with atoms (point masses) connected together with bonds (massless springs) with defined force constants. If the vibrational model and the structure of the reactant are accurate, then the TS model that gives calculated KIEs matching the experimental ones is considered the experimental transition state.

The *vibrational model* is (1) the set of internal coordinates controlling molecular vibrations, i.e. a list of bond stretches and different kinds of bends (bond angle, out-of-plane, linear, and torsional); (2) the force constants associated with each internal coordinate; and (3) coupling constants between internal coordinates to generate the reaction coordinate. KIEs are calculated from vibrational frequencies, which are in turn calculated using the inputted structures and the vibrational model. Vibrational models have been discussed in greater detail elsewhere.^{24,43,56,66} The programs available for BOVA are BEBOVIB^{24,25} and VIBIE.^{67,68}

Structure interpolation. In the past, TS structures for BOVA analysis were generated by the "*ad hoc*" method whereby one searches for the transition state by adjusting the TS structure in an iterative process until the calculated KIEs match the experimental ones. Structural parameters expected to contribute to the KIEs are adjusted and the KIEs recalculated. This approach may not find the best or only solution, and may include a solution that is chemically unreasonable. Enzyme-

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produced effects may not be recognized since it is not clear what the expected KIEs are. Also, the modeling process is extremely time consuming. In order to address the limitations of the *ad hoc* method, the structure interpolation approach to BOVA was developed.^{43,69} An algorithm describes the structure of a molecule as a function of its position in reaction space (Fig. 3) and generates trial TS structures throughout reaction space. The interpolation method relies on electronic structure calculations, X-ray crystallographic structures and other experimental evidence. In the interpolation algorithm, all geometrical parameters are varied simultaneously in a systematic way. Using BOVA, predicted KIEs are then calculated for each isotopic label for every trial TS structure. The predicted KIEs are plotted as a contour diagram for each isotopic label (Fig. 4, bottom right) and the areas where the calculated and experimental KIEs match are plotted for each isotopic label. Ideally, there will be one unique point in reaction space where the predicted and experimental KIEs for every isotopic label match each other. This is the transition state.

This approach has a number of advantages over the *ad hoc* approach, including both greater speed and greater accuracy. A further advantage of the structure interpolation approach, especially with a unified model (see below), is that a model is created of how structure, and therefore KIEs, change throughout reaction space. Any deviation from that model, such as might be caused by an enzyme, will be recognized by the deviation from the model.

Unified models. If experimental KIEs are available for several related reactions, it is possible to construct a "unified model" of the reaction to give the highest possible accuracy TS structures.⁴³ A unified model differs from any other TS analysis only in that a single vibrational model and structure interpolation model is used to determine transition states for several different reactions of a given type. For example, the transition states of four hydrolytic and two adenosine 5'-diphosphate (ADP)-ribosylation reactions of oxidized nicotinamide adenine dinucleotide (NAD⁺) were determined using a unified model. The fact that this model worked for all these different reactions lent support to the correctness of each step in the process.

Differences between modeling approaches. One consistent difference between transition states determined by the *ad hoc* and structure interpolation methods is that the residual leaving group bond order is lower in TS structures determined by structure interpolation. This is a result of the fact that with structure interpolation, the structure of the leaving group ring is varied throughout reaction space along with the ribosyl ring. The increase in bond orders within the ring in response to the loss of bond order in the *N*-glycosidic bond is more accurately reflected. It therefore takes a greater extent of bond breakage to the leaving group to result in the same loosening of the vibrational environment of the departing atom. The practical upshot is that a TS structure is obtained using structure interpolation that has lower bond order to the leaving group ($n_{\rm LG}$) and higher bond order to the nucleophile ($n_{\rm Nu}$). The transition state for cholera toxin-catalyzed NAD⁺ hydrolysis was reported to have $n_{\rm LG} = 0.09$, and $n_{\rm Nu} = 0.005$ by *ad hoc* TS modeling,⁷⁰ but $n_{\rm LG} = 0.02$, and $n_{\rm Nu} = 0.02$ using structure interpolation that accounts for changes in the nicotinamide ring.¹⁷

Similarly, inosine hydrolysis catalyzed by inosine–uridine-specific nucleoside hydrolase (IU-NH) was modeled by *ad hoc* methods with $n_{LG} = 0.19$ and $n_{Nu} = 0.005$,⁷¹ but has been reanalyzed recently using structure interpolation, which gives a TS structure with $n_{LG} = 0.05$ and $n_{Nu} = 0.06$ (unpublished results). It is worth emphasizing that these are small changes in structure, reflecting refinement of KIE interpretation over the years.

Because the process of TS analysis by BOVA has evolved between early studies cited in this chapter and more recent ones, the amount of detail available for different TS structures varies, which will be reflected in the discussion of some of these studies.

3 TS analysis: results and recent developments

THE NATURE OF THE OXOCARBENIUM ION

Whatever the exact mechanism of a particular glycoside hydrolysis or transfer reaction, nucleophilic substitution at the glycosidic bond involves the sugar becoming either an oxocarbenium ion intermediate or passing through a transition state that is highly oxocarbenium ion-like. Catalytic strategies that stabilize a discrete oxocarbenium ion will also likely be effective for an oxocarbenium ion-like $A_N D_N$ transition state. For this reason, we will discuss interactions as stabilizing an oxocarbenium ion, even when it is known that a particular reaction has an $A_N D_N$ mechanism and does not form a discrete intermediate.

Understanding catalytic mechanisms and designing inhibitors requires an understanding of the structure and charge distribution of the oxocarbenium ion. The resonance structures of an oxocarbenium ion imply a molecule that is somewhere between a carbocation (1a) and an oxonium ion (1b).



Information on the nature of the oxocarbenium ion from a variety of experimental and computational approaches has indicated that the oxonium-like resonance form **1b** predominates.^{72–77} Evidence emerging from TS analysis supports this model. In TS structures derived from both *ad hoc*^{27,28,70,71,78–81} and structure interpolation techniques, ^{17,47,69} and supported by electronic structure calculations, essentially all the loss in bond order upon leaving group departure is compensated by increases in the C1'-O4' and C1'-C2' bond orders. In NAD⁺ hydrolysis reactions, the total bond



Fig. 5 Structures of the presumed oxocarbenium ion of glucose and the protonated forms of deoxynojirimycin and isofagomine, potent inhibitors of many β -glycosidases.

order to C1' ($\sum n_{C1'}$) was higher in the transition state than in the reactant; the loss of bond order to the leaving group was compensated by an even larger increase in the bond orders between C1' and the other ribosyl ring atoms C2', O4' and H1'.

There is evidence that at least some enzymes specifically stabilize the oxocarbenium ion portion of the TS molecule to promote catalysis.⁸² The question of how enzymes may stabilize the oxocarbenium ion has generally focused on the change in charge distribution and sometimes on the positive charge that is presumed to accumulate on the ring oxygen. The fact that iminosugar compounds such as deoxynojirimycin (Fig. 5) are effective glycosidase inhibitors^{31,83–93} would tend to support this view. There are several lines of evidence, however, that point to other interactions. Electronic structure calculations indicate that the ring oxygen of the oxocarbenium ion is not positively charged, and experimental evidence supports this conclusion. Electronic structure calculations indicate that the ring oxygen in a glycoside bears significant negative charge, -0.33 in 2 (Fig. 6a, Table 1). When the glycosidic bond is broken and an oxocarbenium ion formed, π -bonding develops between C1' and O4'. The charge on O4' increases more than any other atom in the molecule, but only becomes -0.03 in the oxocarbenium ion. Assigning charges to

| Atom | Charge $[q(e^{-})]^a$ | | Bond | Pauling bond order $(n_{ij})^b$ | |
|------|-----------------------|-------|---------|---------------------------------|------|
| | 2 | 3 | | 2 | 3 |
| C1′ | 0.04 | 0.22 | C1′-O4′ | 0.97 | 1.72 |
| O4′ | -0.33 | -0.03 | C1'-C2' | 1.00 | 1.09 |
| H1' | 0.14 | 0.26 | C1'-H1' | 0.95 | 0.99 |
| O1′ | -0.29 | _ | C1′-O1 | 1.06 | _ |

 Table 1
 Selected atomic charges and bond orders in compounds 2 and 3 (see also Fig. 6)

^aMulliken atomic charges on atoms in model compounds.

 ${}^{b}n_{ij} = e^{(r_1 - r_{ij})/0.3}$, where r_{ij} is the bond length between atoms *i* and *j*, and r_1 is the bond length for a single bond.^{24,56} Single bond lengths (r_1): C-H = 1.09 Å, C-C = 1.526 Å, C-O = 1.41 Å.



Fig. 6 Changes in charge distribution on forming an oxocarbenium ion. (a) Model compounds for glycoside (2) and oxocarbenium ion (3). (b) Stick models of 2 and 3. (c) Areas of positive electrostatic potential (electron deficient) are shown in gray shading. (d) Areas of negative electrostatic potential (electron rich) are shown in gray shading. Because 2 is neutral and 3 is positively charged, the electrostatic potentials are plotted on different scales, with the shading showing electrostatic potential relative to the average for each molecule. Optimizations were performed using a hybrid DFT method with Becke's exchange

functional²²⁰ and Perdew and Wang's correlation functional,²²¹ with the 6-31 + G^{**} basis set (RB3PW91/6-31 + G^{**}) using Gaussian-98.²²² The isodensity surface is plotted at 0.002 e/bohr³.

atoms in molecules is somewhat arbitrary; however, different methods give essentially the same result, as do calculations of electrostatic potential (Fig. 6d). The exact results for these calculations will depend on the identity of the oxocarbenium ion and its environment. The important result is that the ring oxygen does not accumulate significant positive charge in the oxocarbenium ion. This conclusion is supported by previous calculations at the semi-empirical level.^{73,75} This implies that an enzyme active site with a negative charge located adjacent to the ring oxygen in the oxocarbenium ion would not be catalytically effective.

In a recent study, Withers and coworkers concluded that the reactivity of *O*-glycopyranosides is well explained by a Kirkwood–Westheimer model of field effects with all of the charge generated in the oxocarbenium ion being transferred to the ring oxygen.⁷⁷ Their analysis is consistent with the model of oxocarbenium ion structure discussed above because the Kirkwood–Westheimer model requires that the charge at the ring oxygen increases; it does not necessarily require that the charge becomes positive.

The computational results are supported by four lines of experimental evidence.

- (1) Aza-sugar inhibitors such as isofagomine, where the nitrogen atom is situated in a position analogous to the anomeric carbon, are extremely potent inhibitors of β -glycosidases, and often better inhibitors than the iminosugar inhibitors like deoxynojirimycin where the nitrogen atom is located at a position analogous to the ring oxygen.^{94–99}
- (2) The pH versus K_i profiles of iminosugar inhibitors indicate that in at least some cases it is the neutral, unprotonated form that binds the enzyme, while in other cases it is the protonated form.^{84,100} Some enzymes that bind the protonated form also bind with similar inhibition constants to compounds methylated at the ring nitrogen and therefore bearing a permanent positive charge. This demonstrates that there is not a tight, direct interaction between the enzyme and a positively charged amino group.
- (3) The X-ray crystallographic structures of enzymes complexed with iminosugar inhibitors do not show strong interactions between the ring nitrogen and groups on the enzyme.^{26,101,102} As the inhibitors are nearly isosteric with the substrate molecules, this can be taken as an argument against stabilization of the oxocarbenium ion through contacts with the ring oxygen.
- (4) Further evidence comes from a model of the DTA·TS complex for NAD⁺ hydrolysis based on the X-ray crystal structure of DTA·NAD⁺.^{17,103} In the complex, the positive charge that would accumulate on the anomeric carbon at the transition state was located directly adjacent to the carboxylate of a glutamate residue that is completely conserved in the bacterial toxin sequences.

The nearest possible protein contact to the ring oxygen was with the hydroxyl group of Tyr65, with an oxygen–oxygen distance of 3.1 Å. There were no negative charges located near the ring oxygen. The same features were observed in the docking of the TS structure of pertussis toxin-catalyzed ADP-ribosylation of protein $G_{i\alpha 1}$ with the enzyme.⁴⁷ Thus, the structures of enzyme TS models also support the idea that positive charge accumulates on the anomeric carbon rather than on the ring oxygen in an oxocarbenium ion.

Taken together, the evidence above suggests that the oxocarbenium ion intermediate or oxocarbenium ion-like transition state has significant C=O double bond character from charge delocalization/resonance stabilization, but that the positive charge accumulates on the anomeric carbon, C1', in spite of the fact that the oxygen atom is formally trivalent. Because of extensive π -bonding stabilization from the ring oxygen, the positive charge on C1' is much less than it would be in an unstabilized carbocation.

Intermediate lifetime. The lifetime of the enzyme oxocarbenium ion complex in $D_N * A_N$ mechanisms cannot be determined by KIE studies. It must exist for at least a few bond vibrations (*ca.* 10^{-12} s), but there is no upper limit on its lifetime aside from k_{cat} .

NON-CHEMICAL STEPS

It is crucial in performing TS analysis to know exactly which step of the reaction the experimental KIEs reflect. Using isotope-trapping experiments, it is possible to demonstrate whether formation of the Michaelis complex, E·S, is kinetically significant, and if necessary, to find conditions where it is not. However, internal steps^{#8} can also complicate the interpretation of KIEs. These can include, but are not limited to (1) establishment of equilibria between different enzyme-bound intermediates, (2) isotopically insensitive steps, such as conformational changes in the enzyme or substrate, or (3) substrate channeling.

There are at least four instances where non-chemical steps were manifested in KIEs, although in each case the nature of the non-chemical step was not clear. In the sialyltransfer reaction catalyzed by $\alpha(2 \rightarrow 6)$ sialyltransferase using cytidine monophosphate glycoside of *N*-acetylneuraminic acid (CMP-NeuAc) as the glycosyl donor (equation 4), Michaelis complex formation was kinetically significant,¹⁰⁴ but even after compensating numerically for this, the KIEs were small and inconsistent with any mechanism with only chemical steps being kinetically significant. Later experiments using an alternate substrate, uridine 5'-monophosphate (UMP)-NeuAc, yielded KIEs consistent with an oxocarbenium ion(-like) transition state.⁴⁸ This is consistent with a non-chemical step being kinetically significant in the reaction with CMP-NeuAc. A related result is that there

^{#8} An internal step is any step that occurs between formation of the Michael complex, E·S, and dissociation of the enzyme-product complex, E·P.



Fig. 7 Chemical and kinetic mechanism of RTA-catalyzed RNA depurination. In the proposed mechanism for RNA hydrolysis (solid line), free enzyme (E) and RNA substrate (S) combine to form a Michaelis complex (E·S), which proceeds to form a discrete oxocarbenium ion intermediate ([E·O]). This intermediate undergoes an irreversible, isotopically insensitive non-chemical step. The nature of the step is not known, but the oxocarbenium ion intermediate ultimately breaks down to form the depurinated product complex (E·P) before dissociation to free enzyme and products (E + P). The free energy profile is shown for DNA depurination (dashed line), which is proposed to proceed through a stepwise $D_N * A_N$ mechanism with only chemical steps being kinetically significant. The free energy profiles are purely schematic.

was a large solvent deuterium KIE (SDKIE) with CMP-NeuAc (${}^{D_2O}V_{max} = 2.6$, ${}^{D_2O}(V_{max}/K_M)_{CMP-NeuAc} = 1.3$), but not UMP-NeuAc (${}^{D_2O}V_{max} = 0.97$, ${}^{D_2O}(V_{max}/K_M)_{UMP-NeuAc} = 1.1$). This implies that the chemical step does not involve a proton transfer, but that the kinetically significant step with CMP-NeuAc does.



In the ADP-ribosylation of protein $G_{i\alpha 1}$ by pertussis toxin, the 1'-³H KIE was both pH- and temperature-dependent.⁴⁷ The 1'-³H KIE was examined simply because it

was large and therefore changes were easily observed. The decrease in 1'-³H KIE at temperatures <20°C or pH < 7.5 was interpreted as indicating that some isotopically insensitive step had become at least partially irreversible. For example, if a conformational step after the chemical step became slower under non-optimal conditions, then the lifetime of the enzyme-nicotinamide·ADP-ribosyl-G_{ia1} complex might become long enough for some of the complex to undergo the reverse reaction to enzyme·NAD⁺·G_{ia1}. In this case, enzyme·nicotinamide·ADP-ribosyl-G_{ia1} would partition both backward and forward, and the chemical step would be only partially irreversible. It is equally possible that the kinetically significant isotopically insensitive step occurred before the chemical one.

In the depurination of RNA catalyzed by ricin toxin A-chain (RTA), the reaction mechanism involved formation of equilibrium population of an oxocarbenium ion intermediate followed by an isotopically insensitive irreversible step (Fig. 7).⁴⁶ The experimental KIEs were most consistent with the calculated EIEs for oxocarbenium ion formation. This is what would be expected for a mechanism involving freely reversible oxocarbenium ion formation followed by an isotopically insensitive step.

In the yeast α -glucosidase-catalyzed hydrolysis of 4-nitrophenyl- α -D-glucopyranoside,⁴⁹ the primary leaving group ¹⁸O KIE was unity, 1.001, and the α -secondary ²H KIE was small, 1.01. However, the α - and β -secondary KIEs for hydrolysis of α -D-glucopyranosyl 4-bromoisoquinolinium bromide were large, 1.21 and 1.13, respectively. This implied that a partially irreversible conformational change prevented KIEs of the chemical step(s) from being observed for 4-nitrophenyl- α -Dglucopyranoside where the dominant reactant conformation is the ⁴C₁ chair. Intrinsic KIEs were determined for α -D-glucopyranosyl 4-bromoisoquinolinium bromide where the reactant conformation is ¹S₃ skew or twist. It was therefore assumed that the skew conformation more closely resembles the transition state, whereas compounds in chair conformations must undergo partially irreversible conformational changes prior to the chemical steps.

TRANSITION STATE IMBALANCE

One contribution of TS analysis studies to understanding catalysis has been the observation of TS imbalance effects in the ADP-ribosylation of protein $G_{i\alpha 1}$ by pertussis toxin.⁴⁷ This observation has previously been discussed only in passing.^{43, 69}

In most chemical reactions, several processes must occur between reactants and products. The main processes are the bonds that must be made or broken in the overall reaction. In addition to the main processes, several other processes may occur, including solvation/desolvation, charge delocalization or resonance stabilization. In its most general form, TS imbalance (also known as the Principle of Non-perfect Synchronization)^{105,106} simply states that not all these processes occur synchronously.



Fig. 8 TS imbalance effects on C1–N1 bond breakage in an *N*-glycoside analogue. (a) The structure of the model compound (4) was fully optimized using a hybrid DFT method, then the C–N bond was held fixed at increasing bond lengths as the rest of the molecule was reoptimized. Compound **5** was optimized in the absence of the leaving group. The Pauling bond order of the C1–O4 bond (n_{C1-O4}) in the resulting structures is plotted against the (fixed) Pauling bond order of the C1–N1 bond (n_{C1-N1}). The dotted line shows the linear relationship between n_{C1-O4} and n_{C1-N1} that would be expected in the absence of TS imbalance. The solid line is the curve fitted to the equation $\Delta n_{C1-O4} = \chi (n_{C1-N1})^m$, where the optimal value of the exponent *m* was found to be 1.7. Optimizations were performed as in Fig. 6. (b) Molecular orbital rationalization of TS imbalance. See text for details.

An archetypal example of TS imbalance is the nitroalkane anomaly.^{107,108} The rate (k_1) of deprotonation of arylnitroalkanes is more sensitive to substituents on the aryl ring than the equilibrium constant $(K_1 = k_{-1}/k_1)$, as reflected in an unusually large value of the Brønsted parameter, α_{CH} (equation 5). In addition, $\beta > 1.0$ for k_1 and $\beta < 0$ for k_{-1} . These data have been interpreted as showing that

while there is strong charge delocalization in the product arylnitronate anion, charge delocalization is not well developed at the transition state even though the main process, C–H bond breakage, is almost complete. The nitroalkane anomaly represents a common form of TS imbalance where, when there is an increase in charge delocalization or resonance stabilization between the reactants and products, it is less advanced at the transition state relative to the main processes than in the products.^{#9} The converse is also true; charge localization or loss of resonance stabilization tends to be more advanced in the transition state relative to the main processes than in the products.

TS imbalance has been studied in detail in contexts such as proton transfer,^{105,106, 109,111} but was unexpected (by us) in glycoside chemistry. However, the experimental KIEs for ADP-ribosylation of protein $G_{i\alpha 1}$ by pertussis toxin could not be successfully interpreted using the structure interpolation approach while assuming that variation of the ribosyl ring structure was fully synchronous with the main processes, namely breaking the leaving group bond and forming the nucleophile bond.

Electronic structure calculations on a model (4) were used to investigate TS imbalance effects on oxocarbenium ion (5) formation, where the main process is breaking the C–N bond (Fig. 8). In these calculations, the structure of 4 was fully optimized, then the length of the C1–N1 bond was fixed at increasing values and the rest of the molecule re-optimized. The extent of charge delocalization/resonance stabilization was followed by plotting the Pauling bond order between C1 and O4, n_{C1-O4} , as a function of the fixed bond order between C1 and N1, n_{C1-N1} . This was the bond that underwent the largest change between 4 ($n_{C1-O4} = 0.99$) and 5 ($n_{C1-O4} = 1.55$). The increase in n_{C1-O4} lagged behind the decrease in n_{C1-N1} . This was expected by analogy to the nitroalkane anomaly, where resonance stabilization lags the main process.

According to Kresge's model,¹⁰⁷ the lag in resonance stabilization can be modeled as $\Delta n_{\rm C1-O4} = \chi (\Delta n_{\rm C1-N1})^m$, where m = 2 and χ is some proportionality constant. Application of this equation to proton transfers in aqueous solution suggests values of *m* between 2 and 3.¹⁰⁵ In the current study, the best fit was obtained with m = 1.7. Calculations were performed on oxocarbenium ion

^{#9} The language used to describe TS imbalance (e.g., "lag in charge delocalization", or "resonance stabilization developing late") has tended to imply a kinetic effect, namely that certain processes such as charge delocalization occur too slowly to keep up with the main processes. As shown by Bernasconi and Wenzel^{109,110} and in the results described here, it is a thermodynamic effect. In structures where resonance stabilization "lags" the main processes, these are the most thermodynamically favorable extents of resonance stabilization.

formation with a variety of leaving groups and substituents, and TS imbalance was observed in every system examined, with the extent of TS imbalance depending on the identity of the leaving group and cation being formed (data not shown). TS imbalance was previously studied computationally for the carbon-to-carbon proton transfer between acetaldehyde and its enolate anion.^{109,110} Using atomic charge rather than bond order to measure the extent of TS imbalance, values of m = 1.1-2.3 were observed. It was suggested TS imbalance arises, at least in part, as a result of positively charged protons helping to stabilize and localize the negative charge on the carbon atoms (equation 6). The fact that TS imbalance was also observed in oxocarbenium ion formation, where such an effect could not be operative, implies that the structural manifestations of TS imbalance are universal phenomena whenever there is rehybridization of atoms and changes delocalization/resonance stabilization.^{#10}



Although the physical basis of TS imbalance is not well understood, one intuitive explanation of the current case is that in **4** the ring oxygen is a good π -bond donor because of its lone pair electrons but C1 is a very poor π -bond acceptor because it is maintained in an sp³-hybridized state by its bond to N1. There is not a concomitant increase in n_{C1-O4} as the C1–N1 bond is broken because C1 remains a poor π -bond acceptor in the continued presence of the C1–N1 bond. It is only as the C1–N1 bond becomes almost completely broken that C1 becomes a good π -bond acceptor and n_{C1-O4} increases sharply (Fig. 8).

This explanation is complementary with the "fraction of a fraction" explanation advanced by Kresge,¹⁰⁷ which says that development of resonance stabilization will, at most, be proportional to the loss of C1'–N1 bond order. However, the rehybridization of C1' from tetrahedral to planar is not complete, resulting in only partial resonance stabilization due to poorer electronic overlap. Thus, resonance stabilization is only a fraction of the fraction of C1'–N1 bond breakage. These explanations are also consistent with the idea that TS imbalance is correlated with changes in atom hybridization, which is in turn consistent with the results of calculations on the dissociation of methyl nicotinamide (equation 7). The sum of total bond orders to N1 ($\sum n_{N1}$) was a linear function of the main process, i.e. loss of the N1–C1 bond, as was the sum of N1–C bond orders within the ring (data not shown). The essential difference between this and the oxocarbenium ion example is

 $^{^{\#10}}$ There is some question whether the unusual reactivity properties manifested in the nitroalkane anomaly would occur in the gas phase, 108 which is a separate issue.



Fig. 9 Evidence for TS imbalance in the ADP-ribosylation of protein $G_{i\alpha 1}$. The shading shows the areas in reaction space where the calculated KIEs match the experimental KIEs exactly (dark gray) or within the standard deviation (light gray) for the indicated isotopic label. The asterisk indicates the experimentally determined transition state. Isotopic labels are as shown in Fig. 4. The results are shown for calculations done using the structure interpolation approach (a) without TS imbalance and (b) with TS imbalance. This figure is based on Fig. 4 of Ref. 47 with minor modifications of the vibrational model.

that there is no change in hybridization on the nitrogen atom N1; it is sp^2 -hybridized in both the reactant and product structures.



As mentioned above, it was necessary to include TS imbalance effects in the TS analysis of the ADP-ribosylation of protein $G_{i\alpha l}$. Once they were included, the calculated and experimental KIEs at 1-¹⁵N, 1'-¹⁴C, and 2'-³H matched very well (Fig. 9), though the match of 1'-³H was poor in both the cases (see Section 4). The differences between the structure interpolation models that included or neglected TS imbalance were small, with differences in bond length of $\Delta r_{Cl'-O4'} = -0.028$ Å, and $\Delta r_{Cl'-C2'} = -0.012$ Å, $\Delta r_{Cl'-Hl'} = 0.006$ Å. That changes this small could be detected illustrates an important property of using unified models in structure interpolation, namely that they can detect deviations from a model with great

sensitivity. It was possible to detect TS imbalance only because of the deviation from the unified model that had been successful in the TS analysis of four NAD⁺ hydrolysis reactions. It was not detected using the *ad hoc* method.⁸¹ Secondly, the smallness of these differences show that differences in structure can be determined more sensitively than the absolute structure. TS structures are not accurate to within 0.01 Å because they are based on reference structures derived from crystal structures and calculations, both of which have >0.01 Å errors in atom position. However, deviations from the unified model developed for NAD⁺ reactions were detected with great sensitivity.

What is the significance of TS imbalance to the reactivity of glycosides? Probably little. Bernasconi has suggested that resonance stabilization is associated with barriers to intrinsic reaction rates.¹⁰⁵ This can be rationalized in terms of π -bonding electrons being present in the path of the incoming nucleophile, increasing the barrier to nucleophilic attack. It has been shown that there is a significant intrinsic barrier to nucleophilic addition to some carbon cations,¹¹² and that the extent of resonance stabilization correlates with a reduction of intrinsic rates; however, Richard has shown that the intrinsic barriers for nucleophilic attack on oxocarbenium ions are much smaller than other types of cations.¹¹³ This implies that the barrier to the highly favorable addition of thiolate to a ribooxocarbenium ion will be negligible. The greatest significance of the observation of TS imbalance may be to illustrate the accuracy needed in both KIE measurements and interpretation to derive all the available mechanistic detail for reactions.

SECONDARY KIEs

A secondary KIE is any KIE on an atom not involved in the bond making or breaking steps of a reaction. Secondary KIEs are designated in terms of the number of bonds separating the isotopic label from the site where chemistry occurs; thus, in aldoses, the 1'-³H KIE is an α -secondary KIE, the 2'-³H KIE is a β -secondary KIE. Because secondary KIEs are generally smaller in magnitude than primary KIEs, it is most commonly hydron^{#11} KIEs that are measured. However, secondary ¹³C, ¹⁴C, ¹⁵N, and ¹⁸O KIEs have been measured, and are large in some cases.^{17,50,54,69}

Binding isotope effects on forming a Michaelis complex can be significant, 15,114,115 which could lead to the question of whether KIEs can be interpreted in terms of transition states if there are isotope effects on forming the enzyme substrate complex. Isotope effects on a given step contribute to the observable KIEs only if that step is kinetically significant. For KIEs on k_{cat}/K_{M} , only

^{#11} An unspecified isotope of hydrogen, ¹H, ²H, or ³H.

the irreversible step(s) contribute to the observable KIEs, whereas for KIEs on k_{cat} (or V_{max}) only the rate-limiting step(s) are relevant.^{#12}

α -Secondary hydron KIEs

Early on, *a*-secondary hydron KIEs were used to diagnose the mechanisms of glycosidic reactions, and there are many reactions for which that was the only KIE measured. ²H KIEs on the order of 1.08–1.17 were almost invariably observed. Instances of small KIEs can now likely be attributed to kinetically significant substrate binding (commitment to catalysis). Large α -secondary hydron KIEs are a sign of either a highly dissociative $A_N D_N$ or a $D_N * A_N$ mechanism because the rehybridization of the anomeric carbon toward sp^2 and the decreased steric crowding around C1' gives increased freedom in the out-of-plane bending modes of H1'.^{116,117} In a synchronous $A_N D_N$ mechanism, an inverse KIE at H1' would be expected.¹¹⁸ α -Secondary hydron KIEs have been studied in detail both experimentally and theoretically,^{116,118–122} and a great deal of that knowledge has been summarized by Matsson and Westaway.¹¹⁷ Until recently, there was debate about what determines the magnitude of α -secondary hydron KIEs. It has become clear that while there is an inverse contribution from the shortening of the C1'-H1' bond length in an oxocarbenium ion(-like) transition state, this contribution is similar in different transition states and the factor that determines the size of the KIE in a given reaction is the out-of-plane bending modes. These can vary from inverse to large and normal, depending on the TS structure.

Beyond these general trends, however, the detailed interpretation of α -secondary hydron KIEs in glycosidic reactions has proven problematic. As a specific example, the 1'-³H KIEs for pertussis toxin-catalyzed reactions were 1.21 ± 0.01 for NAD⁺ hydrolysis,⁸⁰ and 1.20 ± 0.01 for the ADP-ribosylation of protein $G_{i\alpha 1}$.⁴⁷ The experimental 1'-³H KIEs were within experimental error of each other in spite of the fact that the transition states were different, with the ADP-ribosylation transition state being more synchronous than the hydrolysis reaction (n_{LG} and n_{Nu} were 0.05 and 0.001 for hydrolysis, and 0.11 and 0.09 for ADP-ribosylation, respectively). Based on the BOVA vibrational model used in modeling NAD⁺ transition states, the 1'-³H KIE for the ADP-ribosylation was predicted to be 1.055, which is in general

^{#12} This can be rationalized as follows. Consider an enzymatic reaction that displays ideal Michaelis– Menten kinetics, i.e. equilibrium formation of a Michaelis complex, followed by an irreversible chemical step to form products. Further assume that Michaelis complex formation involves one specific enzyme-substrate contact that causes a ²H EIE. If that contact persists unchanged in the transition state, then there is no further isotope effect on the chemical step and the observable KIE will be equal to the EIE. In other words, if the specific enzyme-substrate contact is the same in the transition state as in the Michaelis complex, this will be reflected in the observable KIE for that label being equal to the EIE of Michaelis complex formation. If, on the other hand, the specific enzyme-substrate contact is removed in the transition state, then there will be an isotope effect on the chemical step that is opposite and equal in magnitude to the EIE on Michaelis complex formation. The observable KIE will be unity, accurately reflecting the lack of the specific enzyme-substrate contact at the transition state.



 α -glucopyranose

β-glucopyranose

Fig. 10 EIEs on the anomeric equilibrium of α -and β -glucopyranose. The reported standard errors for all measurements were ± 0.003 to ± 0.005 . The EIE at H6 was for the dideutero compound.

agreement with previous computational studies.^{116,118} Other experimental 1'-³H KIEs have proven similarly difficult to interpret (unpublished data).

One possible complicating factor in understanding these α -secondary hydron KIEs is that hydron KIEs in general are relatively sensitive to non-covalent and remote interactions. As reviewed by Wade,¹²³ non-covalent hydron KIEs have been demonstrated in a wide variety of contexts, including isotope effects on affinity/ dissociation constants, chromatographic behavior, even smell. Lewis and Schramm recently demonstrated ²H EIEs on the anomeric equilibrium between α - and β -Dglucose in solution (Fig. 10).¹²⁴ Binding KIEs or EIEs have been measured for a number of enzyme substrate interactions, including PNP adenosine,¹²⁵ lactate dehydrogenase·NAD⁺,¹²⁶ lactate dehydrogenase·NADH·oxamate,¹⁵ and phosphoenolpyruvate carboxylase-phosphoenolpyruvate.¹¹⁵ Shifts in peak positions in isotope-edited vibrational spectra of enzyme·ligand complexes provide further evidence of changes in the vibrational environments of atoms upon binding enzymes (e.g., see Refs. 127-131). Isotope effects are vibrational effects, and so observing vibrational frequency shifts strongly implies that there are binding EIEs.^{#13} Given the fact that stabilization of the oxocarbenium ion would likely occur through interactions near the anomeric carbon (see above), it would be expected that there would be contributions to the 1'-³H KIE from direct enzyme TS contacts.

β -Secondary hydron KIEs and hyperconjugation

While detailed interpretation of α -secondary hydron KIEs has been problematic, β -secondary (2'-³H) KIEs have proven useful in both diagnosing the oxocarbenium ion character of glycosidic reaction transition states, and providing information on the ring conformation.

^{#13} An isotope effect reflects the sum of changes in all the bond stretching and bending forces experienced by an atom, so it is not possible to determine an isotope effect from the shift of a single peak in a vibrational spectrum.

Strong hyperconjugation is common. In pentofuranosyl oxocarbenium ions, the combination of strong double-bonding character in the C1'-O4' bond and the 5-membered ring constrains the ring to have two energetically favorable envelope conformations, 3'-endo (³E) or 3'-exo (E₃) (e.g., 3'-endo-**6** or 3'-exo-**6**). In the 3'-exo conformation, the C2'-H2' bond is nearly eclipsed with the developing empty p-orbital on C1', with a dihedral angle of 10°. This allows hyperconjugation, with π -bonding between C1' and C2', and a concomitant loss of bond order in the C2'-H2' σ -bond. In the 3'-endo conformer, this dihedral angle increases to 52°. Because of the strong angular dependence of hyperconjugation, this results in a much smaller extent of hyperconjugation.^{117,132} Thus, a large β -secondary hydron KIE in riboside reactions implies both a highly oxocarbenium ion-like structure and a 3'-exo ring conformation. In the vast majority of riboside reactions examined, the β -secondary hydron KIE has been large, >1.1 for ³H, >1.065 for ²H, indicating that a majority of reactions pass through the transition state in a 3'-exo conformation.

A 3'-endo conformation with RTA. To date, the sole exception is RTA-catalyzed RNA hydrolysis, where 2'-³H KIE = 1.012^{46} (see Section 4). RNA depurination by RTA appears to be unique among the reactions investigated so far, in that the β secondary ³H KIE was small, 1.012 ± 0.005 .⁴⁶ As the other experimental KIEs for this reaction indicated a discrete oxocarbenium ion intermediate, the small 2'-³H KIE was interpreted as indicating a 3'-endo conformation. This is supported by two lines of evidence. Calculated 2^{-3} H KIEs for compound **6** were 1.18 for the 3'-exo conformer, and 0.994 for the 3'-endo conformer. The latter KIE is close to the experimental one. In NMR¹³³ and X-ray¹³⁴ RNA stem-loop structures, the ribose ring conformation in the reactive adenosine residue was either 3'-endo or an unusual 2'-exo conformation. Conversion of the 3'-endo/2'-exo reactant to a 3'-endo oxocarbenium ion could be accomplished without significant adjustment of the RNA backbone, while conversion to a 3'-exo oxocarbenium ion would have required large changes. Thus, the unusual 3'-endo conformation in the oxocarbenium ion intermediate appears to have been imposed on the tertiary structure of the RNA backbone. It is also possible that other external factors, such as enzyme hydrogen bonding to the 2'-hydroxyl could have enforced a 3'-endo conformation. The fact that this is the only known case of a ribooxocarbenium ion in a 3'-endo conformation and that it appears to be in response to external factors (viz. the RNA backbone structure) underscores the preference for the 3'-exo conformation in unconstrained structures.





Fig. 11 Hyperconjugative stabilization of the oxocarbenium ion of 2-deoxy sugars. Using the empirically derived angular dependence of hyperconjugation, where hyperconjugation $\propto \exp(\cos^2\theta)$, and θ is the dihedral angle defined by p-orbital-C1'-C2'-H2'S,¹³² the total amount of hyperconjugative stabilization is plotted as a function of the angle θ .

2-Deoxy sugars. Maximal hyperconjugation is achieved for 2-hydroxy sugars with the C2-H2 bond eclipsed with the developing p-orbital. In 2-deoxy sugars, both C2-H2 bonds are capable of hyperconjugation, and maximum total hyperconjugation is achieved when both C2-H2 bonds are at a dihedral angle of $\approx 30^{\circ}$ to the C1' p-orbital (Fig. 11). Stereospecific 2'-³H KIEs for hydrolysis of the *N*-glycosidic bond of deoxyuridine in DNA by uracil DNA glycosylase (UDG) were large and equal to each other within experimental error: $2'S^{-2}H$ KIE = 1.10 ± 0.01, $2'R^{-2}H$ KIE = 1.11 ± 0.01.¹³⁵ Stereospecific KIEs for the RTA-catalyzed depurination of a DNA substrate were also large and similar to each other, $2'S^{-3}H$ $KIE = 1.117 \pm 0.002$, $2'R^{-3}H$ $KIE = 1.146 \pm 0.008$.⁵⁸ Using the Swain-Schaad¹³⁶ relationship to convert these to ²H KIEs gives $2'S^{-2}H$ KIE = 1.08, $2'R^{-2}H$ KIE = 1.10, similar to the UDG reaction. Dideutero β -secondary ²H KIEs for reactions involving CMP-NeuAc and its derivatives were 1.218-1.28.48,54 To achieve such large KIEs, it is necessary that both individual ²H KIEs be large, 1.10-1.13, assuming they are equal. This implies that there is a strong hyperconjugative stabilization of the oxocarbenium ion(-like) structures for these reactions.

While the large β -secondary KIEs do indicate hyperconjugation in 2-deoxy sugars, they are not helpful in determining the ring conformation of an oxocarbenium ion. In the optimized structures of deoxy-ribooxocarbenium ion



Fig. 12 Conformations for a glycopyranosyl oxocarbenium ion. Hydroxyl groups have been omitted for clarity. The coplanar atoms are indicated with asterisks.

model compounds, the 3'-exo and 3'-endo conformers would both have strong hyperconjugative stabilization and similar β -secondary KIEs for each 2'-hydron.⁵⁸

Hexopyranosides. In hexopyranosides, the 6-membered rings lend more conformational variability than pentofuranosides. Four possible ring conformations exist that can accommodate the requisite four coplanar atoms of an oxocarbenium ion (Fig. 12).

For glucopyranosides the $B_{2.5}$ boat conformation would have the hydron nearly perpendicular to the incipient cation p-orbital of the anomeric carbon, resulting in no hyperconjugation and possibly an inverse inductive KIE. In the ^{2,5}B boat, and ³H₄ and ⁴H₃ half-chair conformations, the C2-H2 bond would be almost eclipsed with the incipient p-orbital, leading to large β -secondary KIEs. However, the ${}^{3}H_{4}$ conformation is sterically unfavorable for a glucopyranosyl oxocarbenium ion. The β -secondary ²H KIE values for the yeast α -glucosidase-catalyzed hydrolysis of α -Dglucopyranosyl 4-bromoisoquinolinium bromide,⁴⁹ α -D-glucopyranosyl pyridinium bromide¹³⁷ and α -D-glucopyranosyl isoquinolinium bromide¹³⁷ were 1.13, 1.12 and 1.11, respectively (see Section 4). These are similar in magnitude to (deoxy)ribonucleoside reactions, indicating that significant hyperconjugation occurs at the transition state of both groups of reactions. As with pentofuranoses, β -secondary KIEs help determine the hexopyranose ring conformation. Because of the greater conformational flexibility of pyranosides there are two conformations which would allow for strong hyperconjugation: ${}^{2,5}B$ and ${}^{4}H_{3}$. The preponderance of transition states exhibiting hyperconjugative stabilization emphasizes the importance of this effect in stabilizing an oxocarbenium ion(-like) transition state.

Remote secondary hydron KIEs

Large hydron KIEs have been observed in enzymatic reactions at sites far removed from where chemistry is occurring. In enzymatic hydrolysis and transfer reactions of *N*-ribosides, it has become a common pattern to observe an inverse 4'-³H KIE and a normal 5'-³H KIE. These KIEs are not intrinsic to the reaction as the KIEs at these positions are negligible in the non-enzymatic reactions.^{69,138,139} Examples of such enzymatic KIEs include inosine hydrolysis by IU-NH from *Crithidia fasciculata*, where 4'-³H KIE = 0.992, 5'-³H KIE = 1.051^{71} and NAD⁺ hydrolysis by diphtheria toxin A-chain (DTA), where 4'-³H KIE = 0.990, 5'-³H KIE = 1.032.¹⁷



Fig. 13 Proposed TS structure for IU-NH versus crystal structure of inhibitor cocrystallized bound at the active site of IU-NH.

In the IU-NH reaction, several explanations of the large, normal 5'-³H KIE were considered, including the possibility of the 5'-hydroxymethyl group adopting a conformation where the hydroxyl group is positioned above the ribosyl ring, located where it can stabilize the positive charge on the oxocarbenium ion-like transition state through electrostatic interactions (Fig. 13). This model was supported by the determination of the X-ray structure of IU-NH cocrystallized with a transition state analogue that had the 5'-hydroxymethyl group in exactly the proposed position.¹⁰¹ Based on the cocrystal structure of IU-NH *p*-aminophenyl iminoribitol (pAPIR), the large 5'-³H KIE may be attributed to the effects of the strong hydrogen bond with the carboxylate residue of Glu166 as well as distortion of the $\angle C4'-C5'-O5'$ bond angle due to electrostatic interactions between the lone pair electrons of O5' and the electron deficient oxocarbenium ion. This bond angle distortion leads to decreases in the bond-bending force constants for $\angle O5'-C5'-H5'$, $\angle C4'-C5'-H5'$ and $\angle H5'-C5'-H5'$, which in turn leads to normal KIEs.

In NAD⁺ hydrolysis, the extended conformation of NAD⁺ in the active site cleft precludes the enzymatic 5'-³H KIE arising from the same factors. However, model calculations showed that increasing or decreasing the $\angle C4' - C5' - O5'$ bond angle by 15° from optimal would give a calculated 5'-³H KIE that matched the experimental KIE, while protonation of the adjacent phosphate residue gave a calculated 5'-³H KIE of 1.049. Thus, the observed 5'-³H KIE may arise from some combination of geometric distortion and/or protonation or strong hydrogen bonding with the proximal phosphate residue.

Bruner and Horenstein observed a very large inverse ζ -secondary ³H KIE = 0.944 in a reaction catalyzed by $\alpha(2 \rightarrow 6)$ sialyltransferase (equation 4).⁴⁸ This result demonstrates that, unlike non-enzymatic reactions, there is not necessarily any distance dependence on secondary KIEs in enzymatic reactions; enzymes commonly use binding interactions remote from the reaction site to promote catalysis.

Although it is not always possible to assign the specific sources of remote hydron KIEs, the observation of remote KIEs that are significantly different from the non-

enzymatic reactions is a sign that the enzyme is using binding energy at remote sites to promote catalysis.

CATALYTIC STRATEGIES

Opposite catalytic strategies?

One striking observation about the reactions catalyzed by RTA and pertussis toxin is that the enzymes appear to use "opposite" catalytic strategies. The analogous non-enzymatic reactions, hydrolysis of adenosine 5'-monophosphate (AMP)⁷⁸ and NAD⁺,⁶⁹ both have highly dissociative $A_N D_N$ mechanisms with oxocarbenium ion-like transition states. In the ADP-ribosylation of protein $G_{i\alpha 1}$ by pertussis toxin, the $A_N D_N$ mechanism is more synchronous, with more nucleophile participation and lower oxocarbenium ion character than the non-enzymatic reaction.^{#14} In contrast, the RTA-catalyzed depurination of RNA⁴⁶ and DNA⁵⁸ substrates proceeds through stepwise $D_N * A_N$ mechanisms where the enzyme stabilizes an oxocarbenium ion to the point that it becomes a discrete intermediate. This is illustrated with a hypothetical free energy surface for each reaction (Fig. 14).

A consideration of the evolutionary pressures acting on each enzyme may provide a rationale for why different strategies may be effective. The role of pertussis toxin in vivo is to ADP-ribosylate protein $G_{i\alpha l}$. There is no evolutionary advantage to pertussis toxin hydrolyzing NAD⁺, as the cellular pool of NAD⁺ is too large for a single enzyme molecule to damage a cell by depleting NAD⁺. However, a single toxin molecule can ADP-ribosylate enough protein $G_{i\alpha l}$ in a cell to cause a physiological effect. As such, there is evolutionary advantage for the enzyme to bind NAD⁺ but not cause any reaction until protein $G_{i\alpha l}$ has bound to the enzyme NAD⁺ complex. Pertussis toxin uses binding energy from interaction with the second substrate, protein $G_{i\alpha l}$, to increase the nucleophile participation at the transition state. In the case of RTA, its role is to depurinate and thereby inactivate 28S ribosomal RNA, which is needed for protein biosynthesis. The identity of the incoming nucleophile is unimportant; however, it is convenient that a good nucleophile, water, is present at a concentration of 55 M. It is therefore an evolutionarily effective strategy for RTA to catalyze the formation of a highly reactive oxocarbenium ion. As a ribooxocarbenium ion is too reactive to have a finite lifetime in solution, an enzyme-bound oxocarbenium ion will likely also be highly reactive, undergoing nucleophilic attack as soon as a water molecule diffuses into position. For RTA there is no evolutionary advantage to being selective about the nucleophile; therefore forming a highly reactive oxocarbenium ion is evolutionarily effective. It has been estimated that a single molecule of RTA can kill a cell.

^{#14} The ADP-ribosyl transfer reaction has a sulfur nucleophile, as compared with water in the nonenzymatic reaction. Given the very weak dependence on nucleophile ability^{140,141} of this reaction, the nonenzymatic transition state with a sulfur nucleophile would be expected to be similar to that with water.


Fig. 14 Hypothetical free energy surfaces for the glycoside hydrolysis and transfer reactions, as inferred from TS analysis. Compared to the highly dissociative $A_N D_N$ transition state for the non-enzymatic reaction (middle), RTA (left) and pertussis toxin (right) appear to use opposite catalytic strategies in that they increase and decrease, respectively, the amount of oxocarbenium ion character in the ribosyl ring at the transition state.

Evolutionary pressures have constrained pertussis toxin to a mechanism with significant nucleophile participation while RTA has evolved a catalytic strategy with an oxocarbenium ion intermediate that reacts quickly with nearby water molecules. By this logic, other ADP-ribosylating toxins, such as cholera and diphtheria toxins, would be expected to proceed through relatively synchronous transition states. Nucleoside hydrolases (enzymes that hydrolyze nucleosides to ribose and a purine or pyrimidine base) could use mechanisms with oxocarbenium ion intermediates, though the transition states characterized to date have been $A_N D_N$ (see below).

It is striking that the only enzymes characterized to date that form an oxocarbenium ion intermediate, RTA and UDG (see below), are enzymes that act on nucleic acids and which have extended binding sites encompassing several nucleotide residues. It is possible that only these enzymes can form enough enzyme-substrate contacts through their extended binding sites to pay for the energetic cost of forming highly reactive oxocarbenium ion intermediates.

Arguments that invoke evolutionary forces are distinct, of course, from questions of how different catalytic strategies are effected through specific enzyme substrate interactions. Because very detailed pictures of transition states have not previously been available, these kinds of questions are only now beginning to be addressed.

Catalytic strategies from crystal structures

The combination of protein crystallography and TS analysis has provided important insight into enzyme function not available from either technique in isolation. One especially important lesson is that enzyme structures change dramatically when binding good TS analogue inhibitors. This has led to a virtuous cycle of TS analysis and TS analogue design leading to good inhibitors, which when cocrystallized with their target enzymes lead to further insights into enzyme TS interactions, and can then lead to further rounds of inhibitor design.

The structures discussed in this section are IU-NH, PNP, the DNA repair enzyme UDG, and DTA, an ADP-ribosylating toxin. These reactions have substantially different catalytic requirements and the enzyme mechanisms reflect this. Purine leaving groups can be activated for departure by protonation, as seen with PNP. Pyrimidines cannot be activated by protonation, but can leave as anions. UDG specifically stabilizes the uracil anion, promoting its departure. The nicotinamide ring of NAD⁺ is cationic, but becomes neutral upon departure, so stabilizing positive or negative charges will not be useful in this case. IU-NH, a non-specific



Fig. 15 A schematic showing the essential features of transition state stabilization by IU-NH, based on the cocrystal structure of an IU-NH-TS analogue complex. The 3'-exo ribosyl ring conformation allows simultaneous favorable interactions between the lone pair electrons of the 5'-hydroxyl oxygen and the positive charge on the oxocarbenium ion-like ribosyl ring, while also forming a bidendate interaction with the active site Ca^{2+} ion. Ca^{2+} binds and orients the substrate, as well as helping to activate the nucleophile water. Asp10 acts as a general base to deprotonate the nucleophile after the transition state. There are few direct interactions with the leaving group, consistent with the experimental evidence that IU-NH operates primarily through interactions with the ribosyl ring.

nucleoside hydrolase, appears to function primarily through stabilization of the oxocarbenium ion-like portion of the transition state, lessening its dependence on leaving group activation.

PNP, IU-NH and UDG were cocrystallized with inhibitors, while DTA was cocrystallized with a substrate, NAD⁺. The structures of unliganded enzymes have been only minimally useful because without a substrate or strong inhibitor molecule bound, it is difficult to know what enzyme TS contacts are likely to be present. This problem arises partly because of the large number of possible interactions between enzyme and substrate, and is compounded by the fact that the structures of enzyme active sites change upon substrate or inhibitor binding. With IU-NH, side chain motions of greater than 10 Å were observed upon binding a strong inhibitor.

IU-nucleoside hydrolase (IU-NH). The experimental TS structure for IU-NHcatalyzed inosine hydrolysis was used as the basis for designing TS analogue inhibitors, one of which was cocrystallized with the enzyme.¹⁰¹ IU-NH is relatively non-specific, hydrolyzing all the naturally occurring purine and pyrimidine nucleosides with good efficiency. Given the lack of specificity for the leaving group and the differing catalytic requirements of purine and pyrimidine leaving groups, it might be expected that IU-NH operates primarily through oxocarbenium ion stabilization and/or nucleophile activation. In support of this view is the fact that IU-NH efficiently hydrolyses *p*-nitrophenyl- β -D-ribofuranoside,⁸² unlike PNP and other nucleoside hydrolases tested. Also, it is inhibited by a wide variety of iminosugar inhibitors, with little specificity for parts of the inhibitor corresponding to the substrate's leaving group.¹⁴² This is consistent with the small number of contacts in the crystal structure between the enzyme and "leaving group" atoms in the inhibitor. It would appear that the catalytic activity of IU-NH arises predominantly through interactions in the ribosyl ring, a view that is supported by the fact that 2'-, 3'-, and 5'-deoxynucleosides are poor substrates and deoxyiminoribitols are similarly poor inhibitors. This view of the catalytic mechanism (Fig. 15) is supported by features of the NH-IU·pAPIR complex. The inhibitor pAPIR was bound in a 3'-exo conformation, the conformation predicted by TS analysis,⁷¹ but which is uncommon in crystal structures of ribosides.¹⁴³ The carboxylate moiety of Glu166 would stabilize an oxocarbenium ion by hydrogen bonding to the 5'-hydroxyl group, orienting it such that the lone pair electrons of oxygen can electrostatically stabilize the positive charge on the ribosyl ring at the transition state. A Ca^{2+} ion bound in the active site was observed in the cocrystal structure that had been assigned as K^+ in the unliganded enzyme. Ca^{2+} was subsequently shown to be required for activity. The Ca^{2+} had an octahedral coordination sphere with ligands supplied by acidic side chains, and the 2'- and 3'hydroxyl groups of the inhibitor provided a bidentate ligand. A strong interaction of Ca^{2+} with the 2'-hydroxyl could lead to strong polarization of the C2'-O2' bond, or even deprotonation of the O2'H through a catalytic base. The polarized bond or oxyanion would help inductively stabilize the oxocarbenium ion. A water molecule is the last calcium ligand and is ideally located to act as the nucleophile. It is believed that Ca²⁺ helps to activate the nucleophilic water and facilitate its deprotonation, with Asp10 acting as a general base. Thus Ca²⁺ is proposed to have a role in substrate binding and orientation as well as nucleophile activation and/or possibly interaction with the 2'-hydroxyl.

Purine nucleoside phosphorylase (PNP). In contrast to IU-NH, PNP appears to use extensive contacts with the purine ring to promote catalysis and relatively few contacts with the ribosyl ring. The crystal structure of PNP has been determined in a complex with an iminosugar inhibitor,²⁶ immucillin H, which was developed based on TS analyses of PNP-catalyzed hydrolysis²⁷ and arsenolysis²⁸ reactions. TS analysis revealed that the enzyme catalyzes a dissociative $A_N D_N$ mechanism. The crystal structure was compared with the structures determined by Ealick and coworkers³⁰ of PNP in a Michaelis complex analogue, PNP-inosine-sulfate, and a product complex, PNP· α -D-ribose-1-phosphate-hypoxanthine.

There were numerous enzyme-ligand contacts with the purine ring and the phosphate (or sulfate), but relatively few with the oxocarbenium ion (Fig. 16). The picture that emerges from the PNP structures is a mechanism whereby the leaving group purine and the nucleophile phosphate are tightly bound and relatively immobile, being fixed by many enzyme-ligand contacts. The electrophile ribosyl ring, which makes fewer contacts with the enzyme, moves from the immobile leaving group to the immobile nucleophile – an electrophile migration.



Fig. 16 A schematic mechanism of PNP-catalyzed inosine phosphorolysis showing electrophile migration. This mechanism is based on cocrystal structures of PNP and TS analyses of inosine hydrolysis and arsenolysis. A large number of direct and water-mediated enzyme-substrate contacts hold the leaving group hypoxanthine and the nucleophile phosphate almost immobile while the ribosyl ring (the electrophile), because of the small number of contacts with the enzyme, is able to migrate from leaving group to nucleophile.

The distance of electrophilic migration can be estimated from the distances between the purine ring and phosphate or sulfate in the substrate analogue (5.6 Å) or product (5.2 Å) complexes, as well as the PNP immucillin H phosphate complex (4.8 Å). These distances are less than the 6 Å that is estimated to be the minimum amount of space needed for a discrete oxocarbenium ion intermediate to develop.²⁶

Of the three direct contacts between the ribosyl ring and PNP residues, two are hydrogen bonds to the 2'- and 3'-hydroxyls that likely do not contribute directly to stabilization of an oxocarbenium ion. The hydrogen bond between the His257 side chain and the 5'-hydroxyl is the only direct contact that would stabilize the oxocarbenium ion, by orienting the lone pair electrons on O5' to allow electrostatic stabilization of the positive charge. The other apparent factor in stabilizing the oxocarbenium ion is the negatively charged phosphate anion located directly beneath the ribosyl ring. The combination of the lone pair electrons of O5' and the phosphate anion creates an electron-rich "sandwich" to stabilize an oxocarbenium ion. Phosphate makes a modest contribution to oxocarbenium ion stabilization, with k_{cat} being only 100-fold higher for phosphorolysis than hydrolysis.³¹ This relatively modest oxocarbenium ion stabilization is further illustrated by the fact that pnitrophenyl- β -D-ribofuranoside is a very poor substrate for phosphorolysis.⁸² A major driving force for catalysis, then, appears to be stabilization of the purine ring leaving group. The crystal structure of PNP-imucillin H showed direct hydrogenbonding interactions with N7, O6 and N1H of the purine ring analogue, as well as protein-mediated water contacts that appear to be the source of the proton at N7. Protonation at N7 helps to make the hypoxanthine ring in the substrate a better leaving group by making the otherwise anionic leaving group neutral. Catalysis through leaving group stabilization is further supported by the extremely tight binding of product hypoxanthine, with $K_d = 1.3$ pM.



2'-deoxyuridine residue

Fig. 17 (a) Pseudouridine, (b) Pseudouridine residue from UDG-inhibitor cocrystal structure, PDB entry: 1EMH. (c) Orbital interactions arising from the proposed coupled anomeric/ σ - π_{Arom} interactions.¹⁴⁷ The localized lone pair electrons on N1 are a consequence of the proposed interactions, but not explicitly discussed in the original proposal. (d) The ion pair intermediate formed in the D_N * A_N mechanism proposed for UDG.

Uracil DNA glycosylase (UDG). UDG catalyzes the first step in the removal of uridine residues from DNA. These arise from spontaneous deamination of cytidine residues or misincorporation of UTP into DNA. TS analysis supported a stepwise $D_N * A_N$ mechanism.¹³⁵ UDG stabilizes the anionic pyrimidine leaving group, as evidenced by the fact that it lowers the pK_a of N1 by 3.4 units relative to solution.^{144, 145} There is a carboxylate side chain, Asp64, that is located adjacent to the α -face of the ribosyl ring. It is proposed that this stabilizes an oxocarbenium ion electrostatically as well as assisting nucleophilic attack by water, either through orienting it or by acting as a general base. Recent computational results¹⁴⁶ provide additional support for the experimental evidence of a $D_N * A_N$ mechanism.

Another interesting feature of the active site is evident in the structure of UDG cocrystallized with a pseudouridine-containing oligo-DNA inhibitor (Fig. 17).¹⁴⁷ The pseudouridine residue is distorted about C1' in the cocrystal structure, toward a trigonal geometry, as opposed to the planar geometry seen in other pseudouridine structures. It was proposed that this distortion represents ground state destabilization that promotes catalysis through coupled anomeric and $\sigma - \pi_{Arom}^{148}$ stereoelectronic effects (Fig. 17c). In this proposed mechanism, the anomeric effect (interaction of the empty p-orbital of the ribosyl ring oxygen with the σ^* -orbital of C1') leads to weakening of the *N*-glycosidic bond and strengthening of the C1'-O4' bond. Coupled with this effect, the observed distortion around N1 is proposed to facilitate

Table 2Effects of distortion by UDG

| Bond | Expected change in bond length ^a | Pseudouridine bond lengths (Å) | | Change as expected? | Bond | 2'-Deoxyuridine bond lengths (Å) | | Change as expected? |
|----------------|---|-----------------------------------|---|---------------------|----------------|-------------------------------------|------------------------|---------------------|
| | | Distorted ^b | Undistorted ^c | | | Distorted ^d | Optimized ^e | |
| C1'-C1 | Increase | 1.55 | 1.52 ± 0.01 | Yes | C1′-N1 | 1.53 | 1.48 | Yes |
| C1'-O4' | Decrease | 1.42 | 1.41 ± 0.01 | f | C1′-O4′ | 1.43 | 1.41 | No |
| C1–C2 C2–O2 | Increase Decrease | 1.39 1.22 | $\begin{array}{c} 1.43 \pm 0.01 \\ 1.23 \pm 0.01 \end{array}$ | No _ f | N1-C2 C2-O2 | 1.42 1.22 | 1.39 1.22 | Yes No |

^aExpected changes in bond lengths due to coupled anomeric/ $\sigma - \pi_{Arom}$ effects relative to undistorted pseudouridine.

^bBond lengths from X-ray crystallographic structure, Protein Databank (PDB) entry 1EMH.

^cAverage of bond lengths in four pseudouridine residues from PDB entries 1EHZ and 1EVV.

^dDistorted 2'-deoxyuridine had hydrogen atoms fully optimized and all bond lengths optimized, but with heavy atom bond angles constrained to match the distorted pseudouridine from the PDB entry 1EMH.

^eFully optimized 2'-deoxyuridine. Optimization was as in Fig. 6.

^fChange in bond lengths is too small to be measured.

an interaction between the σ -molecular orbital of the C1'-N1 σ -bond and π electrons from the uridine ring (illustrated with the π -orbital of C2–O2). One consequence of this proposed mechanism, however, is that the aromatic delocalization of lone pair electrons from N1 must be lost because there cannot be simultaneous interaction of the ring π -electrons with both the σ -orbital and the lone pair electrons of N1. While this explanation may be reasonable *a priori*, the distorted structure of pseudouridine did not display changes in bond lengths relative to four undistorted pseudouridine residues that would support the proposed coupled anomeric/ $\sigma - \pi_{Arom}$ interactions (Table 2).^{149,150} Comparing four bond lengths between distorted and undistorted pseudouridine residues, the C1-C1' bond length increased as expected, but the other bond lengths either did not change significantly or changed opposite to the direction expected. To determine if these putative stereoelectronic effects were operative in uridine rather than pseudouridine, electronic structure optimizations were performed on uridine, and uridine distorted to have the same bond angles as the distorted pseudouridine residue. As might be expected, the C1'-N1' bond in the distorted structure was longer, but other bond length changes were inconsistent with coupled anomeric/ σ - π_{Arom} effects. Thus, while the observed distortion of pseudouridine may indicate a ground state destabilization strategy by UDG, the evidence does not support coupled stereoelectronic effects. However, distortion about N1 would likely aid cleavage of the *N*-glycosidic bond through the conventional orbital interpretation, where the lone pair electrons of N1 remain conjugated with the aromatic π -electrons, and the sp²-hybridized σ -bond to C1' is cleaved.

The importance of His187 in stabilizing the uracil anion in the transition state or intermediate has been demonstrated by NMR and Raman spectroscopy, which showed a 34 cm^{-1} decrease in the C2–O2 bond stretch frequency in the enzyme substrate complex, in agreement with the presence of a strong hydrogen bond to stabilize a uracil anion.^{144,145,151} There was little change in the uracil ring vibrational modes, which is evidence against extensive ring distortion.

Diphtheria toxin A-chain (DTA). Several crystal structures of diphtheria toxin are available,^{152–156} including the cocrystal structure with substrate NAD⁺ bound in the active site.¹⁰³ Only minor side chain adjustments were necessary to accommodate the TS structure in place of NAD⁺. The salient features of the DTA·TS interactions include a very deep, form-fitting hydrophobic pocket in which the nicotinamide ring binds. The low dielectric environment of this pocket would lower the energy difference between the reactant and transition state by increasing the energy of the positively charged nicotinamide ring relative to the neutral form at the transition state, a case of ground state destabilization. The side chain of Glu148, has been shown by photoaffinity labeling¹⁵⁷ and mutagenesis¹⁵⁸ to be crucial for catalysis, and this glutamate side chain is conserved in all bacterial ADP-ribosylating toxin sequences. It is located directly adjacent to where positive charge accumulates at the anomeric carbon of the ribosyl ring in the transition state, where it could stabilize the formation of the oxocarbenium ion-like transition state (Fig. 18). It may also be involved in nucleophile orientation and/or activation in hydrolysis or ADPribosylation of elongation factor 2.



Fig. 18 A schematic of $DTA\cdot NAD^+$ interactions in the hydrolysis reaction. Left: in the Michaelis complex, the positive charge due to the quaternary nitrogen is delocalized throughout the nicotinamide ring and somewhat into the ribosyl ring. The ring oxygen possesses a large negative electrostatic potential. Right: at the transition state for NAD^+ hydrolysis, the large extent of C–N bond breakage renders the nicotinamide neutral and hydrophobic. It can now interact more favorably with the surrounding hydrophobic residues. The positive charge has now been localized on the ribosyl ring. It is stabilized by the invariant glutamate residue which is directly adjacent to the site where the positive charge accumulates at the transition state. This glutamate may also have a role in orienting and/or polarizing the water nucleophile.



Fig. 19 Double displacement mechanism for a retaining α -glucosidase, A \rightarrow B glycosylation, C \rightarrow D deglycosylation.

4 Specific reactions

GLUCOSIDE AND XYLOSIDE HYDROLYSIS AND GLUCOSIDASES

Glycosyl transferases and *O*-glycosidases modify glycoproteins and are important for polysaccharide catabolism. Inhibitors of these enzymes are potential drugs in the treatment of cancer, viral infection and diabetes. There are two mechanistic classes of glycosidases, retaining and inverting.^{4,159,160}

Retaining glycosidases operate via a double displacement mechanism (Fig. 19), although the identity of the intermediate formed is the subject of some controversy. Mechanistic studies have attempted to address the nature of the intermediate, whether a full-fledged oxocarbenium ion is formed, or a covalent glycosyl–enzyme intermediate. To this end, mechanistic insight on the ring conformation at the transition state and the degree of nucleophilic assistance can be established from these studies.

Non-enzymatic hydrolysis

KIEs were measured for the acid-catalyzed hydrolysis of methyl α - (7 α) and β -D-glucopyranoside (7 β),⁵⁰ and methyl α - (8 α) and β -D-xylopyranoside (8 β),¹⁶¹ the spontaneous hydrolysis of α - (9 α) and β -D-glucopyranosyl fluoride (9 β)¹⁶² and the spontaneous hydrolysis of α -D-glucopyranosyl 4-bromoisoquinolinium bromide (12).¹³⁷ TS analyses of enzymatic hydrolyses have been completed on fluoro α -D-glucopyranoside (9 α),¹⁶³ α -D-glucopyranosyl pyridinium bromide (10),¹³⁷ and α -D-glucopyranosyl isoquinolinium bromide (11).¹³⁷



O-Glucopyranosides and xylopyranosides. Bennet and Sinnott studied acidcatalyzed hydrolysis of methyl α - and β -D-glucopyranosides, 7α and 7β , using a family of KIEs to determine transition state information on glycoside hydrolysis

| Compound | KIE | | | | | | | |
|----------------|------------------------------------|-----------------------------------|-------------------------|--|----------------------|--|--|--|
| | α -Secondary ² H | β -Secondary ² H | Primary ¹³ C | Leaving group primary ¹⁸ O (or ¹⁵ N) | Ring ¹⁸ O | | | |
| $7 \alpha^{a}$ | 1.137 ± 0.007 | 1.073 ± 0.003 | 1.007 ± 0.001 | 1.026 ± 0.001 | 0.996 ± 0.001 | | | |
| $7\beta^{a}$ | 1.089 ± 0.006 | 1.045 ± 0.004 | 1.011 ± 0.002 | 1.024 ± 0.001 | 0.991 ± 0.002 | | | |
| $8\alpha^b$ | 1.128 ± 0.004 | 1.088 ± 0.008 | 1.006 ± 0.001 | 1.023 ± 0.002 | 0.983 ± 0.001 | | | |
| $8\beta^b$ | 1.098 ± 0.005 | 1.042 ± 0.004 | 1.006 ± 0.003 | 1.023 ± 0.003 | 0.978 ± 0.001 | | | |
| $9\alpha^c$ | 1.142 ± 0.008 | 1.065 ± 0.008 | 1.032 ± 0.003 | _ | 0.984 ± 0.005 | | | |
| $9\beta^{c,d}$ | 1.078 ± 0.001 | 1.027 ± 0.007 | 1.016 ± 0.002 | _ | 0.985 ± 0.005 | | | |
| 12^{e} | 1.188 ± 0.009 | 1.094 ± 0.010 | 1.005 ± 0.002 | $1.015 \pm 0.004 \ (^{15}N)$ | - | | | |

 Table 3 Experimental KIEs for spontaneous and acid-catalyzed hydrolysis of glucopyranosides

All kinetic runs were completed at 80°C unless stated otherwise.

^{*a*}From Ref. 50.

^bFrom Ref. 161.

^cFrom Ref. 162.

^{*d*}Data collected at 50°C and corrected to 80°C assuming $T \times \ln(\text{KIE}) = \text{constant.}$ ^{*e*}From Ref. 137. (Table 3).⁵⁰ Based on the small primary ¹³C KIEs for both anomers, the authors concluded that the acid-catalyzed hydrolyses occurred via a $D_N + A_N$ mechanism. The leaving group ¹⁸O KIEs suggested that the C–O bond to the aglycon was largely broken at the transition state, and the large α -secondary ²H KIE was consistent with formation of an oxocarbenium ion. The β -secondary ²H and ring ¹⁸O KIEs were used to determine the dihedral angle of the hydron at C2 and the incipient p-orbital at C1. These angles were used in combination with molecular mechanics to determine the ring conformation at the transition state. The large β -secondary ²H for the α -anomer was consistent with a flattened ring. The β -anomer conformation was more chair-like, i.e. more similar to the reactant state. One of the major conclusions was that their results were not consistent with the antiperiplanar lone pair hypothesis.¹⁶⁴ The authors concluded that methyl α -glucopyranoside was hydrolyzed through a skewed or boat conformation. The antiperiplanar lone pair hypothesis would predict the opposite.

Not surprisingly, most of the KIEs for the acid-catalyzed hydrolysis of the methyl xylopyranosides 8α and 8β (Table 3) were similar to those for the corresponding glucopyranosides 7α and 7β . There are differences, however, in the ring ¹⁸O KIEs. The larger inverse KIEs for the xylopyranosides indicated more resonance stabilization of the incipient oxocarbenium ion. Another difference was in the β -secondary ²H KIEs of the α -anomer, where the larger value for the xylose derivative suggested a TS with greater hyperconjugation, more closely resembling a ⁴H₃ half-chair or ^{2,5}B boat conformation as expected for a compound without the sterically encumbering C5 hydroxymethyl group. The more inverse ring ¹⁸O KIE and larger β -secondary KIEs suggest greater charge delocalization and resonance stabilization in xylopyranosides than glucopyranosides.

N-Glucopyranosides. KIEs for non-enzymatic reactions are useful as a reference for comparison with enzymatic KIEs. Owing to the low rates for the uncatalyzed hydrolysis of **10** and **11** ($t_{1/2} \approx 33$ h for **10** at 80°C), TS analysis for the spontaneous hydrolysis of **12** was conducted (Table 3) for comparison with the yeast α -glucosidase-catalyzed reaction on all three compounds.¹³⁷ The small primary anomeric ¹³C KIE was consistent with a D_N + A_N mechanism. The β -secondary ²H KIE for **12** was larger than that for **7** α , indicating stronger hyperconjugation. The leaving group primary ¹⁵N KIE of 1.015 suggested significant C–N bond cleavage.^{#15}

Glucopyranosyl fluorides. The larger primary ¹³C KIEs for 9α and 9β (Table 3) are consistent with an $A_N D_N$ mechanism.¹⁶² The authors used BOVA and the previously solved TS structure for the enzyme-catalyzed hydrolysis of $9\alpha^{163}$ as a

^{#15} The theoretical maximum KIE for breaking a C–N bond was previously calculated to be 1.044.¹⁶⁵ This calculation, however, neglects the fact that bond orders within the leaving group ring increase to compensate for the loss of the *N*-glycosidic bond, lowering the ¹⁵N KIE. The limits of ¹⁵N KIEs for nicotinamide and adenine departure have been calculated previously to be 1.034 and 1.024, respectively.^{17,58} It would be necessary to calculate the maximum possible value for an isoquinolinium bromide leaving group to determine the extent of bond breakage that a ¹⁵N KIE of 1.015 represents.

starting point to solve the TS structure for the non-enzymatic hydrolysis. The experimental transition states had a ${}^{4}C_{1}$ chair conformation, rather than a boat, halfchair or skew conformation as described in the studies by Bennet and Sinnott⁵⁰ and Huang *et al.*¹³⁷ Overall, the KIEs for the β -anomer (9β) were similar to those for the methyl glucoside (7β). The major difference was the larger primary anomeric ${}^{13}C$ KIE for 9β , implying an A_ND_N mechanism. It will take quantitative analysis of the experimental KIEs to make a definite choice between a bimolecular or stepwise mechanism.

Taken together, these four studies indicate that non-enzymatic hydrolysis of α -glucopyranosides with neutral leaving groups occurs via $D_N + A_N$ type mechanisms, whereas α -glucopyranosides with anionic leaving groups operate via $A_N D_N$ mechanisms. The α -secondary ²H KIE value measured for the spontaneous hydrolysis of compound **12** is larger than that reported for the acid-catalyzed hydrolysis of methyl α -D-glucopyranoside.⁵⁰ The $A_N D_N$ reaction of α -D-glucopyranosyl fluoride (9α) with azide ion gave an α -secondary ²H KIE value of 1.154 (corrected to 80° C).¹⁶² Although these values were suggestive of dissociative reactions, the fact that the value for 9α lies between those for methyl α -D-glucopyranoside (7α) and compound **12** is further evidence that α -secondary ²H KIEs cannot be used to differentiate between $D_N + A_N$ and $A_N D_N$ reactions at acetal centers.^{50,51}

Enzymatic hydrolysis

KIEs on V_{max} were investigated for the yeast α -glucosidase-catalyzed hydrolysis of 4-nitrophenyl- α -D-glucopyranoside and compounds **10–12** (Table 4).^{49,137} It was determined that 4-nitrophenyl- α -D-glucopyranoside had a partially rate-limiting conformational change, whereas for the pyridinium type compounds 10 and 12, only chemical steps were rate-limiting. This difference was attributed to the fact that 4nitrophenyl- α -D-glucopyranoside exists in a ${}^{4}C_{1}$ conformation which must undergo a conformational change to a skew or twist conformation before the chemical steps, unlike 10 and 12 which have skewed conformations in solution and require only small conformational changes before chemistry can occur. In a related study, large primary ¹³C KIEs for the yeast α -glucosidase-catalyzed hydrolysis of 10 and 11 (Table 4) demonstrated that both reactions occurred via A_ND_N mechanisms with a small degree of nucleophilic assistance. These results are consistent with the double displacement mechanism proposed for retaining glycosidases^{4,159,160} and transsialidase. These results are in contrast to the sugar beet α -glucosidase-catalyzed hydrolysis of the α -D-glucopyranosyl fluoride (9α) where the primary ¹³C KIE was 1.007,¹⁶² more consistent with a $D_N * A_N$ mechanism.¹⁶³

The β -secondary ²H KIEs for yeast α -glucosidase-catalyzed hydrolysis of **10** and **11** are suggestive of substantial overlap of a C–H(D) bond with the p-orbital at the transition state, consistent with the glycosyl ring approaching a ^{2,5}B boat conformation. Hosie and Sinnott had reported a similar value for the β -secondary ²H KIE on **10** as well as a large value for **12**.⁴⁹

| 1 | | cat cat ivi | | 15 | | | | |
|---------------|-------------------------|------------------------------------|-----------------------------------|-------------------------|---------------------------------------|--|--|--|
| Compound | KIE | | | | | | | |
| | Туре | α -Secondary ² H | β -Secondary ² H | Primary ¹³ C | Leaving group primary ¹⁵ N | | | |
| $9\alpha^{a}$ | $k_{\rm cat}/K_{\rm M}$ | 1.103 ± 0.009 | 1.027^{b} | 1.007^{c} | _ | | | |
| 10^d | k _{cat} | _ | 1.115 ± 0.006 | 1.028 ± 0.006 | 1.019 ± 0.007 | | | |
| 11^{d} | $k_{\rm cat}$ | _ | 1.106 ± 0.009 | 1.027 ± 0.005 | 0.985 ± 0.005 | | | |
| 12^{e} | $k_{\rm cat}$ | 1.21 ± 0.03 | 1.13 ± 0.02 | _ | - | | | |

Table 4 Experimental KIEs on k_{cat} or k_{cat}/K_{M} for the enzymatic hydrolysis of glucopyranosides

^{*a*}From Ref. 163.

^bFrom ²H KIEs calculated from ³H KIEs based on the Swain–Schaad relationship. ^cFrom ¹³C KIE calculated from ¹⁴C KIE based on the Swain–Schaad relationship.⁵⁷

^{*d*}From Ref. 137.

^eFrom Ref. 49.

The primary ¹⁵N KIEs for the spontaneous hydrolysis of **12** (1.018, corrected to 25°C) and the yeast α -glucosidase-catalyzed hydrolysis of **10** (1.019) suggested transition states with partial C–N bond cleavage. Similar results were found in a study by Scheuring and Schramm on pertussis toxin-catalyzed hydrolysis of NAD⁺.⁸⁰ Based on the family of KIEs (primary ¹⁵N KIE value of 1.021), semi-empirical calculations and BOVA, these authors concluded that the reaction transition state had partial C–N bond breakage with a weak nucleophilic component from the incoming water molecule.

The most notable feature of the KIEs for yeast α -glucosidase-catalyzed hydrolysis of **11** was the inverse primary ¹⁵N KIE of 0.985. No other primary ¹⁵N KIE for glycoside hydrolysis measured to date has been inverse. The β -secondary ²H KIE for **11** was large, indicating an oxocarbenium ion-like transition state, while the primary ¹³C was too large for a stepwise mechanism. Thus, the sugar ring KIEs indicated an $A_N D_N$ mechanism, inconsistent with the inverse primary ¹⁵N KIE. Any loss of C–N bond order in forming an $A_N D_N$ transition state should create a looser vibrational environment for the leaving group nitrogen and therefore a normal KIE. Protonation of ¹⁵N isoquinoline gives an inverse EIE of (1/1.0216 ± 0.0005) = 0.979.¹³⁷ Protonation of the (aromatic) ring nitrogen before or during C–N bond breakage would give an inverse contribution to the KIE, but this does not seem chemically reasonable. The source of the inverse ¹⁵N KIE is not yet understood. Lower than expected primary ¹⁵N KIEs have been observed for PNP- and RTA-catalyzed reactions, although never an inverse KIE.

These studies indicate that for enzyme-catalyzed reactions, 9α (with an anionic leaving group) reacts via a $D_N * A_N$ mechanism, whereas α -glucopyranosides with neutral leaving groups (10 and 11) operate via $A_N D_N$ mechanisms. These results are opposite to what was found for the spontaneous hydrolysis reactions. The enzymatic reactions could also indicate differences in the enzyme mechanism, regardless of the leaving group.

SIALIDASES AND SIALYLTRANSFERASES

There is considerable interest in sialyltransferase reactions because of their importance in carbohydrate–protein interactions, such as selectin-sialyl Lewis X, which are important for inflammation and metastasis.^{166–169} Sialyltransferases catalyze the transfer of *N*-acetylneuraminic acid from cytidine monophosphate (CMP) to an acceptor hydroxyl group of a glycoprotein, glycolipid or oligosaccharide with inversion of anomeric (C2) configuration (Fig. 20a). *Trans*-sialidases catalyze the transfer of α -D-*N*-acetylneuraminic acid from host glycoconjugates to acceptors on the parasitic cell surface with retention of anomeric configuration (Fig. 20b).¹⁷⁰ Sialylation camouflages the parasites, allowing them to evade the host immune system. Inhibitors of *trans*-sialidases may be useful for treating parasitic infections.¹⁷¹



Fig. 20 (a) Inverting reaction catalyzed by a sialyltransferase. (b) Retaining reaction catalyzed by a *trans*-sialidase.

Sialidases such as the influenza neuraminidase are retaining enzymes that catalyze the hydrolysis of α -D-N-acetylneuraminic acids from oligosaccharides or glycolipids and glycoproteins. Inhibitors of these enzymes are of great interest since it has been shown that neuraminidase activity can correlate with virulence.⁴

Acid-catalyzed hydrolysis

Horenstein and Bruner performed TS analysis on the acid-catalyzed solvolysis of CMP- β -D-N-acetylneuraminidine **13** (Table 5).



For the acid-catalyzed solvolysis reaction, the large β -secondary ²H and small anomeric primary ¹³C KIEs were suggestive of a late transition state that is dissociative in nature. Taken into account with methanolysis data, it was determined that the reaction occurs via a D_N + A_N mechanism with the formation of an oxocarbenium ion intermediate with finite lifetime, stabilized electrostatically by the carboxylate ion at C1. This allowed for capture by solvent from

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| KIE | IE <u>13</u> | | 14 | 15 | | 16 | |
|--|---|---|-----------------------------------|-----------------------------------|-----------------------------------|---|-----------------------------------|
| | Solvolysis ^a | Transfer ^{b,c} | Transfer ^{d,e} | Solvolysis ^e | Transfer ^e | Solvolysis ^e | Transfer ^{c,e} |
| β -Secondary ² H | 1.276 ± 0.008 | 1.044 ± 0.007 | 1.218 ± 0.010^d | 1.113 ± 0.012 | 1.053 ± 0.010 | - | 1.060 ± 0.008 |
| α -Secondary ¹ C Primary carbon | $1.013 \pm 0.004^{\circ}$ $1.030 \pm 0.005 (^{14}C)$ | 0.998 ± 0.004 $1.000 \pm 0.004 (^{14}C)$ | -1.028 ± 0.010 (¹⁴ C) | -1.016 ± 0.011 (¹³ C) | -1.021 ± 0.014 (¹³ C) | $^{-}$ 1.015 ± 0.008 (¹³ C) | $^{-}$ 1.032 ± 0.008 (13 C) |
| Remote ³ H | 1.002 ± 0.010 | 0.984 ± 0.007 | 0.944 ± 0.010 | - | - | - | - |

Table 5 Competitive experimental KIEs for sialoside solvolysis and on k_{cat}/K_M for enzyme-catalyzed transfer

^{*a*}From Ref. 54.

^bFrom Ref. 104.

^cKIEs corrected for commitment to catalysis.

^{*d*}From Ref. 48.

^eFrom Ref. 170.

either face of the oxocarbenium ion.⁵⁴ This is in contrast to the acid-catalyzed solvolysis KIEs for the sialyl-lactose compound (**15**) and the sialyl-galactose compound (**16**) (Table 5),¹⁷⁰ which had large primary ¹³C KIEs, suggestive of $A_N D_N$ mechanisms.

Enzymatic transfer

The transfer reaction catalyzed by rat liver $\alpha(2 \rightarrow 6)$ sialyltransferase using **13** and *N*-acetyllactosamine as the substrates displayed kinetically significant substrate binding.¹⁰⁴ Even after the observed KIEs for **13** were corrected, they were still too low for any mechanism involving only chemical steps, suggesting that a non-chemical step was kinetically significant (Table 5).

TS analysis was performed using the UMP analogue, 14, and *N*-acetyllactosamine as substrates. Compound 14 was a less efficient substrate, resulting in an increased barrier for the chemical step and allowing intrinsic KIEs to be determined. The β secondary ²H KIE for 14 was similar to the solvolysis reaction of 13, and 4.5-fold larger than that for enzymatic transfer with 13.⁴⁸ The anomeric primary ¹³C KIE for 14 was also similar to that determined for the solvolysis reaction of 13. Taken together, the KIEs for the enzyme-catalyzed reaction indicate a stepwise reaction, following a D_N * A_N mechanism. A large inverse KIE at H9 was found for 14, as discussed in Section 3.

Using a *trans*-sialidase from *Trypanosoma cruzi*, TS analysis was performed on the transfer of *N*-acetylneuraminic acid from the tri- and disaccharides **15** and **16**, respectively, to lactose.¹⁷⁰ Small β -secondary dideuterium KIEs and large anomeric primary ¹³C KIEs were observed for both compounds (Table 5). The large primary effects were believed to indicate an A_ND_N mechanism and could be evidence for a covalent enzyme-intermediate. The low β -secondary effects relative to solvolysis were rationalized by the authors to indicate differences in TS charge development rather than being a consequence of sugar ring conformation change.

BACTERIAL ADP-RIBOSYLATING TOXINS AND NON-ENZYMATIC NAD⁺ HYDROLYSIS

Certain bacteria exert toxic effects on their hosts through the action of ADPribosylation reactions where the nicotinamide ring of NAD⁺ is displaced and ADPribose is attached to some host cell protein (equation 8). DTA ADP-ribosylates diphthamide, a post-translationally modified residue on elongation factor 2, which is required for protein biosynthesis. Blocking protein biosynthesis causes cell death. Cholera and pertussis toxins ADP-ribosylate cellular G-proteins at specific Arg and Cys residues, respectively. In the absence of the physiological targets, all three enzymes will slowly hydrolyze NAD⁺ to ADP-ribose and nicotinamide. TS analysis has been performed on NAD⁺ hydrolysis catalyzed by cholera toxin,⁷⁰ DTA,¹⁷ and





TS analysis of NAD⁺ hydrolysis. The hydrolysis reactions of NAD⁺ all proceeded through highly dissociative A_ND_N transition states. The transition state for the pHindependent solvolytic hydrolysis of NAD⁺ had $n_{LG} = 0.02$ and $n_{Nu} = 0.005$. Assignment of the non-enzymatic mechanism as A_ND_N was based on a variety of experimental evidence in addition to the experimental KIEs. The large negative Brønsted coefficient, $\beta_{LG} = -0.9$, for non-enzymatic NAD⁺ hydrolysis¹⁷² was consistent with either a highly asynchronous $A_N D_N$ mechanism, or a $D_N\ast A_N$ mechanism with formation of a discrete oxocarbenium ion intermediate. In the analysis of Jencks,^{173,174} the rate-limiting step, and therefore the TS structure, depends on the stability of the oxocarbenium ion. If the oxocarbenium ion is stable enough to have a finite existence in solution, then a $D_N * A_N$ mechanism is possible. Chemical evidence in the literature rules this out for NAD⁺ hydrolysis. If an oxocarbenium ion were formed, the product distribution would be expected to be sensitive to nucleophile strength, and a mixture of inversion and retention of configuration at the anomeric carbon would be expected. In NAD⁺ reactions, the product distribution depends on the molar ratios of methanol and water rather than their nucleophilicities,¹⁴¹ and the methanolysis product is predominantly the α anomer (22:1, $\alpha:\beta$),¹⁷⁵ indicating that A_ND_N predominates. In addition, the 330-fold decrease in the rate of hydrolysis of a riboside analogue with a bulky substituent that prevents "backside solvation"¹⁷⁶ is further evidence for pre-association of the nucleophile and against a D_N * A_N mechanism. Finally, attempts to match the experimental KIEs using BOVA to a $D_N * A_N$ mechanism with either leaving group departure or nucleophile approach as the first irreversible step did not yield reasonable candidate TS structures.

The TS structures initially reported for the cholera and pertussis toxin-catalyzed hydrolysis reactions had leaving group bond orders, n_{LG} , of 0.09 and 0.1, respectively, and very low nucleophile bond orders, n_{Nu} , of 0.005 and 0.001. It was later shown using structure interpolation that it is necessary to account for increases in the N1–C2 and N1–C6 bond orders in the nicotinamide ring that partially compensate for the loss of C1'–N1 bond order. After accounting for changes in the nicotinamide ring structure, the TS structures were essentially the same as for DTA, which had $n_{LG} = 0.02$ and $n_{Nu} = 0.03$. It should be emphasized

that these changes are small and reflect a refinement of KIE interpretation methods. The enzymatic hydrolysis reactions form highly dissociative $A_N D_N$ transition states very similar to the non-enzymatic reaction. The increase in nucleophile bond order at the transition state relative to the non-enzymatic reaction may be a consequence of the invariant glutamic acid residue in the active sites that may help to orient and/or activate the water nucleophile. The experimental TS structure was docked with the DTA structure (see Section 3).

TS analysis of ADP-ribosylation by pertussis toxin. TS analyses have been performed on the ADP-ribosylation of a model peptide⁸¹ and protein $G_{i\alpha 1}^{47}$ by pertussis toxin. The KIEs for these reactions were identical within experimental error, meaning that the transition states were identical even though the specificity constant, k_{cat}/K_M , with protein $G_{i\alpha 1}$ was 800-fold higher than with the peptide. The ADP-ribosylation reactions were significantly more synchronous, $n_{LG} = 0.11$, $n_{\rm Nn} = 0.09$, than the hydrolysis reactions. This indicated that the enzyme uses binding energy to increase the nucleophile participation at the transition state. The fact that the transition states were the same for the peptide and the protein substrates implies that all the structural determinants needed to define the structure of the transition state are present in the 20-mer peptide substrate. The additional enzyme substrate interactions with protein $G_{i\alpha l}$ as the second substrate, which are necessarily remote from the site of catalysis, increase the catalytic efficiency 800fold without changing the transition state. The ADP-ribosylation of protein $G_{i\alpha 1}$ was performed using catalytic amounts of β - and γ -subunits, which formed an $\alpha\beta\gamma$ trimer. Thus, the experimental KIEs were measured for the TS complex of pertussis toxin·NAD⁺· $G_{\alpha\beta\gamma}$, a pentamolecular complex with a combined molecular mass of 100 kDa, possibly the most complex system examined to date by TS analysis. In spite of the size and complexity of the system, it was possible to measure accurately KIEs that reflected the chemical steps of the reaction.

Considering that a thiolate anion is $10^7 - 10^9$ -fold more nucleophilic than a neutral thiol, ^{177,178} the nucleophile in this reaction will be the thiolate anion of Cys351 of protein G_{ia1}. The inverse SDKIE of 0.65 was originally cited as evidence for a pre-equilibrium deprotonation of Cys351.⁴⁷ However, it is not greatly different from the SDKIEs measured for the enzymatic hydrolysis reactions, i.e. 0.73 for pertussis toxin, 0.82 for cholera toxin. SDKIEs of this magnitude can arise from simple viscosity effects.¹⁷⁹ Given the pK_a = 8.37 of an unperturbed cysteine residue in solution, at physiological pH 7.4 a significant fraction would be deprotonated and nucleophilic.

As with DTA, the experimental transition state was docked with the X-ray crystallographic structure of pertussis toxin, along with a model of the reactive peptide from protein $G_{i\alpha 1}$.¹⁸⁰ As in DTA, there is a carboxylate residue, Glu129, located directly adjacent to the incipient positive charge in the oxocarbenium ion-like transition state. His35, which has been shown to be important to catalysis and has been proposed to act as a general base to activate the nucleophile,¹⁸¹ is located near the pyrophosphate of NAD⁺ and not in a position to act as a general base. His35 is structurally homologous to His21 of DTA, which also interacts with the

pyrophosphate group, and His44 of cholera toxin, which had also been advanced as a candidate general base.⁷⁰

TS analysis studies, combined with structure and enzyme kinetic studies, have suggested some solutions to Cordes' dilemma,¹⁸² which was originally stated in relation to NAD⁺ glycohydrolase reactions. To rephrase the dilemma: (1) because the leaving group nicotinamide is cationic, general acid catalysis cannot be important; (2) because there is little nucleophile participation at the transition state, general base activation of the nucleophile cannot be important; (3) because both the reactant and transition state are positively charged, electrostatic stabilization is unlikely to be important, and therefore; (4) catalysis by distortion may be a resolution to this dilemma. Points (1) and (2) are true; however, point (4) was undermined by the lack of obvious distortion in the $DTA \cdot NAD^+$ complex.¹⁰³ As discussed above, the distribution of positive charge changes dramatically between reactant and the transition state, making it possible for enzymes to stabilize positive charge in a different location in the transition state than in the reactant. Also, hydrophobic interactions with the nicotinamide ring will tend to stabilize the transition state (and products) over the reactant NAD⁺, which bears significant positive charge delocalized around the nicotinamide ring. Finally, ADP-ribosylating toxins use binding energy from the protein substrates to stabilize transition states through as-yet unknown mechanisms, as evidenced by the 800-fold increase in k_{cat}/K_{M} between the peptide substrate of pertussis toxin and protein $G_{i\alpha 1}$.

NUCLEOSIDE AND AMP HYDROLASES

IU-nucleoside hydrolase (IU-NH)

IU-NH is among the most thoroughly characterized *N*-glycosidases. In addition to kinetic characterization^{82,183} and TS analysis,^{16,71} the electrostatic potential surfaces of substrate, transition state and products have been analyzed and used as a template for inhibitor design.¹⁸⁴ Inhibitors have been synthesized and characterized.^{100,142, 185–188} The crystal structure of IU-NH has been determined in the unliganded form^{189,190} and as a cocrystal with a potent inhibitor.¹⁰¹ In addition to kinetic characterization of inhibitors, theoretical studies were used to rationalize and then predict the inhibitory activity of different compounds.^{191–194}

The experimental KIEs⁷¹ were qualitatively similar to other hydrolysis reactions. The primary leaving group 9-¹⁵N KIE = 1.026 ± 0.004 indicated extensive C1'-N9 bond breakage at the transition state. The α - and β -secondary ³H KIEs indicated an oxocarbenium ion-like transition state, 1'-³H KIE = 1.150 ± 0.006 , 2'-³H KIE = 1.161 ± 0.003 , while the primary carbon KIE was too large for a stepwise mechanism, 1'-¹⁴C KIE = 1.044 ± 0.004 . The large remote δ -secondary ³H KIE, 5'-³H KIE = 1.051 ± 0.003 , was interpreted as being due to an enzyme-induced conformation of the C5' hydroxymethyl group that can help stabilize the oxocarbenium ion-like A_ND_N transition state (see Section 3).

| Isotopic label ^a | Acid catalyzed | AMP nucleosidase ^b | | Mutant AMP nucleosidase ^c | |
|---|--|--|--|--------------------------------------|-------------------|
| | | No ATP | 0.5 mM ATP | No ATP | 0.5 mM ATP |
| 1′- ¹⁴ C | 1.044 ± 0.003 | 1.035 ± 0.002 | 1.032 ± 0.002 | 1.041 ± 0.006 | 1.041 ± 0.003 |
| 9- ¹⁵ N 1/ ³ H | 1.030 ± 0.002 1.216 ± 0.004 | 1.030 ± 0.003 1.069 ± 0.003 | 1.025 ± 0.002 1.047 ± 0.002 | 1.034 ± 0.002 | 1.021 ± 0.002 |
| $2'^{-2}H$ | 1.077 ± 0.002 | 1.061 ± 0.002 | 1.043 ± 0.002 | | |
| 5'- ³ H | 1.006 ± 0.002 | 1.007 ± 0.003 | 1.006 ± 0.002 | | |

Table 6 Competitive experimental KIEs for AMP hydrolysis

KIEs are on k_{cat}/K_{M} for enzymatic reactions.



^bFrom Ref. 139. ^{*c*}From Ref. 79.

TS analysis revealed a dissociative $A_N D_N$ transition state with relatively high residual bond order to the leaving group adenine, $n_{LG} = 0.19$, and very weak bond order to the water nucleophile, $n_{Nu} = 0.005$. However, a reassessment of the experimental KIEs using the structure interpolation approach has indicated a more "balanced" transition state, with $n_{LG} = 0.05$, and $n_{Nu} = 0.06$ (unpublished results; see also Section 2). As discussed above, IU-NH appears to operate primarily through stabilization of the ribosyl portion of the oxocarbenium ion-like transition state, involving little interaction with the leaving group.

AMP nucleosidase and acid-catalyzed AMP hydrolysis

The experimental KIEs for acid-catalyzed AMP hydrolysis indicate an $A_N D_N$ mechanism. The 9-¹⁵N KIE = 1.030 is less than the semi-classical limit of 1.044⁶¹, ¹⁶⁵ because the increase in bond orders within the adenine ring compensate for the loss of the C1'-N9 bond. Protonation at N7 in the acid-catalyzed reaction makes the incipient anionic leaving group neutral, and helps to transfer π -bonding electrons into the bonds with N9.

The AMP nucleosidase reaction was studied by TS analysis with and without adenosine 5'-triphosphate (ATP) as an allosteric activator (Table 6).^{78,79,139,195} An allosteric activator activates the enzyme by binding at a location remote from the catalytic site. The presence of 0.5 mM ATP increases k_{cat} by 200-fold with minimal effect on $K_{\rm M}$. The KIEs for hydrolysis by AMP nucleosidase are similar to those for the non-enzymatic reaction, implying a similar mechanism. In the presence of the allosteric activator ATP, most of the experimental KIEs decreased in magnitude. This could be taken as a sign of non-chemical kinetically significant steps; however, the KIEs would be expected to decrease by the same proportion if that was true. In fact, the secondary hydron KIEs decrease more than the heavy atom KIEs, and the remote 5'-³H KIE did not decrease. The lower 9-¹⁵N KIE in the presence of ATP implies a lower extent of C1'-N9 bond breakage at the transition state, i.e. the enzyme stabilizes the leaving group more strongly and thereby achieves the transition state earlier in the C-N bond breakage.

A mutant enzyme was isolated and characterized kinetically.⁷⁹ This mutant was not sequenced; it is simply referred to as a V_{max} mutant because the value of k_{cat} was decreased by 50-fold while there was little change in other kinetic parameters, including K_{M} , activation by ATP, and inhibition by formycin 5'-phosphate. The larger 9-¹⁵N KIE = 1.034 with the unactivated mutant enzyme may be explained either by an even greater extent of C–N bond breakage at the transition state, or a lack of protonation at N7. Allosteric activation by ATP still increased by the same extent, 200-fold, and still caused a decrease in the 9-¹⁵N KIE. In this case, it may indicate protonation at N7 and/or a smaller extent of C–N bond breakage at the transition state.

| Isotopic label ^a | Experimental KIE | | | | |
|-----------------------------|---------------------------------------|-----------------------------|--|--|--|
| | Steady state arsenolysis ^b | Pre-steady state hydrolysis | | | |
| $1'^{-14}$ C | 1.026 ± 0.006 | 1.045 ± 0.005 | | | |
| 9- ¹⁵ N | 1.010 ± 0.005 | 1.000 ± 0.005 | | | |
| 1′- ³ H | 1.141 ± 0.004 | 1.151 ± 0.004 | | | |
| 2'- ³ H | 1.152 ± 0.003 | 1.145 ± 0.003 | | | |

Table 7 Experimental KIEs on k_{cat}/K_{M} for PNP-catalyzed reactions with inosine





^bFrom Ref. 28. Experimental KIEs were corrected for commitment to catalysis, $C_{\rm f} = 0.19$. ^cFrom Ref. 27.

PHOSPHORYLASE AND PYROPHOSPHORYLASE REACTIONS

Purine nucleoside phosphorylase (PNP)

Bovine PNP is involved in the mammalian purine salvage pathway, catalyzing phosphorolysis of inosine, guanosine or their 2'-deoxy analogues (equation 9). It is a trimeric enzyme with three identical subunits. It displays third-of-sites activity, with only one subunit active at any time.³¹ KIEs were measured for phosphorolysis of inosine by bovine spleen PNP.²⁸ The 1'-³H KIE = 1.047 ± 0.002 for the overall reaction was equal to the previously measured binding KIE = 1.047 ± 0.02 for formation of the Michaelis complex, PNP-phosphate-inosine.¹²⁵ This, along with primary 1'-¹⁴C KIE = 1.000 ± 0.001 for the overall reaction, indicated non-chemical kinetically significant steps. The commitment to catalysis (ratio of k_3/k_2 in Fig. 2) for phosphorolysis of inosine was later determined to be 2.1.²⁷

TS analysis was performed for two PNP-catalyzed reactions, arsenolysis of inosine using arsenate as a phosphate analogue, and a pre-steady state TS analysis of PNP-catalyzed inosine hydrolysis. The ribosyl ring KIEs (Table 7) were typical of those observed previously for other reactions of *N*-ribosides, indicating an A_ND_N mechanism for hydrolysis and either an A_ND_N or $D_N * A_N$ for arsenolysis. The



primary ¹⁵N KIEs, however, are unusually low.

TS analysis of inosine arsenolysis. TS analysis was performed on the arsenolysis of inosine under steady state conditions. The primary, $1'^{-14}$ C KIE was near the lower limit (1.025–1.029) calculated for an A_ND_N mechanism with adenosine⁵⁸ and could indicate either an A_ND_N or D_N * A_N mechanism. The large α - and β -secondary ³H KIEs indicated a transition state with high oxocarbenium ion character. Analysis of X-ray crystal structures of PNP demonstrates that there is not enough space in the enzyme active site between the leaving group purine and the phosphate nucleophile to allow formation of a stable oxocarbenium ion (see Section 3). Thus, the most likely mechanism for arsenolysis is an extremely dissociative A_ND_N mechanism. The primary, 9-¹⁵N KIE = 1.010 was lower than expected, as discussed below.

TS analysis of inosine hydrolysis. Experimental KIEs were determined for inosine hydrolysis under pre-steady state conditions because of the exceptionally slow dissociation of hypoxanthine (estimated $K_d = 1.3$ pM), which is rate-limiting under steady state conditions. Hypoxanthine binds to the other two PNP subunits with much lower affinity. KIEs in the ribosyl ring were typical of an $A_N D_N$ mechanism. When the reaction was run in 20% methanol, the product ratio was 85:15 1-methylribose:ribose. This is close to the ratio expected based on the relative nucleophilicities of MeOH and water, indicating that there is significant participation of the nucleophile in the reaction coordinate. As with the arsenolysis, the primary, 9-¹⁵N KIE of 1.000 was anomalous. These KIEs were initially rationalized as indicating an internal equilibrium formation of ribose and hypoxanthine; however, this is inconsistent with what is now known about the observable KIEs for stepwise reactions.⁵⁸

Mechanism, and the source of unusual KIEs. The primary 9-¹⁵N KIEs for the PNP-catalyzed reactions were anomalously low compared both to calculations and other experimental leaving group primary ¹⁵N KIEs. The expected KIE for complete *N*-glycoside bond breakage would be > 1.026, the KIE for inosine hydrolysis by IU-NH. Anomalously low 9-¹⁵N KIEs may arise from the strong enzyme leaving group interactions observed in cocrystal structures of PNP with substrate, product, and inhibitor complexes. For many reactions discussed in this chapter, the primary KIEs could be modeled by BOVA or quantum mechanical analyses and were apparently

"unperturbed". In the case of PNP, it is possible that strong leaving group activation by the enzyme affects the vibrational environment of N9, making it more vibrationally constrained. This could occur through interactions observed in the X-ray crystal structures, including protonation at N7 and hydrogen bond donation through the exocyclic oxygen. These would tend to reduce the KIE at N9. If this were occurring, it might be expected that the anomaly would be correlated with the tightness of hypoxanthine binding. The affinity of PNP for hypoxanthine has been estimated under steady state conditions in the presence ($K_d = 1.7 \ \mu M$) and absence $(K_d = 1.3 \text{ pM})$ of ribose-1-phosphate. The K_d of PNP with hypoxanthine in the presence of ribose-1-arsenate is not known, but pre-steady state experiments showed that there was a burst of hypoxanthine formation in the arsenolysis reaction, followed by a 10-fold slower steady state rate, with $k_{cat} = 2 \text{ s}^{-1}$. If rate-limiting hypoxanthine dissociation and diffusion-limited binding of hypoxanthine are assumed, then a value of $K_d = 5$ nM is estimated. Thus, the size of the anomaly in the 9-15N KIE (1.010 for arsenolysis, 1.000 for hydrolysis) correlates with tightness of the enzyme interaction with hypoxanthine ($K_d = 5 \text{ nM}$ and 1.3 pM, respectively).

A mechanism for arsenolysis involving a highly dissociative $A_N D_N$ transition state, with an anomalously low 9-¹⁵N KIE arising from enzyme activation of the leaving group is consistent with the pre-steady state and steady state kinetic experiments, with the crystal structures, and with the arsenolysis KIEs. In this model, the reaction proceeds through electrophile migration, where there are many enzyme interactions with the leaving group and nucleophile, but few with the ribosyl ring.

A variant on electrophile migration is suggested for the hydrolysis reaction, based on several lines of evidence: (1) PNP can form exceptionally tight interactions with hypoxanthine ($K_d = 1.3 \text{ pM}$); (2) there are many contacts with the phosphate nucleophile in the crystal structures which orient and activate anionic phosphate through its negatively charged oxygen atoms and increase the reaction rate 100-fold; (3) these contacts would not activate water or methanol nucleophiles, which are much smaller and neutral; (4) the paucity of enzyme ribosyl contacts argues against strong stabilization of the oxocarbenium ion; and (5) the larger 1'-¹⁴C KIE = 1.045 for inosine hydrolysis indicates a more synchronous A_ND_N transition state than arsenolysis, and therefore a smaller distance between leaving group and nucleophile atoms.

Thus, the water (or methanol) nucleophile is poorly situated and chemically incompatible with the nucleophile activation normally catalyzed by PNP, so nucleophilic attack will not be enzyme activated. Nonetheless, the large amount of 1-methylribose formed by PNP indicates that nucleophile participation is important. Taken together, this evidence suggests a mechanism where strong PNP·hypox-anthine binding activates the leaving group toward nucleophilic displacement and creates a vibrationally constrained environment for N9. This activated *N*-glycosidic bond is susceptible to attack from solvent water or methanol, which is not activated by the enzyme. The mechanism is dissociative $A_N D_N$, though more synchronous than the arsenolysis reaction.

Phosphoribosyl transfer reactions

Experimental KIEs have been determined by two groups for the orotate phosphoribosyltransferase (OPRTase)-catalyzed reaction (equation 10)^{196,197} and several other phosphoribosyltransferases.¹⁹⁷ Unfortunately, in the latter study, the primary, 1'-14C KIEs were measured using 5'-3H as a remote label, with the assumption that there was no KIE at H5'. It is now known that this is generally not true, and is specifically not true in the case of OPRTase, where $5'^{-3}H$ KIE = 1.028 ± 0.008 . KIEs were measured for OPRTase using pyrophosphate, the normal nucleophile, and phosphonoacetic acid, an alternate substrate. Phosphonoacetic acid was used because KIEs measured using pyrophosphate were very small, suggesting non-chemical kinetically significant steps. The ribosyl ring KIEs for OPRTase indicated a dissociative A_ND_N transition state. The secondary ³H KIEs suggested an oxocarbenium ion-like transition state, with 1'-³H $KIE = 1.200 \pm 0.007$ and 2'-³H $KIE = 1.140 \pm 0.007$, while the primary, 1'-¹⁴C $KIE = 1.040 \pm 0.004$ was too large for a $D_N * A_N$ mechanism. Interpretation of the primary ¹⁵N KIE was complicated by the fact that the substrate was labeled at two positions, N1 and N3. The 1.3^{-15} N KIE = 1.006 ± 0.005 was smaller than expected for a dissociative $A_N D_N$ mechanism. It was possible to rationalize the observed 1,3⁻¹⁵N KIE by invoking strong interactions with the orotate ring leading to an inverse KIE at N3. As with PNP, the structure of the transition state was modelled by docking the substrate at the active site cleft of OPRTase determined by X-ray crystallographic analysis^{198,199} shows many interactions between the enzyme and the leaving group orotate ring, and between the enzyme and the pyrophosphate nucleophile, but few direct contacts with the ribosyl ring. One may postulate that, as with PNP, the enzyme operates through an electrophile migration mechanism that proceeds through a highly dissociative A_ND_N transition state.

The crystal structure of hypoxanthine phosphoribosyltransferase has been solved in a ternary complex of the enzyme with the substrate 5-phosphoribosyl 1pyrophosphate (PRPP) and an analogue of hypoxanthine with a carbon atom in place of N9.²⁰⁰ As with PNP and OPRTase, these structures showed many contacts with the leaving group and nucleophile, but few with the ribosyl ring. Again, an electrophile migration mechanism may be operative here.



DNA AND RNA

Uracil DNA glycosylase (UDG)

UDG removes uridine residues from DNA. These arise from spontaneous deamination of cytidine residues, and from misincorporation of UTP into DNA. A wide variety of experimental approaches has been used to study UDG, including the TS analysis reported by Werner and Stivers^{135,144,145,151,201–203} (see also Section 3).

UDG has been shown to exhibit diffusion-limited binding with good substrates;²⁰³ however, selection of a single-stranded trinucleotide substrate avoided kinetically significant substrate association. Experimental KIEs were determined using a novel dual-label technique whereby a non-radioactive isotopic label of interest, e.g. 1'-¹³C (Fig. 17d), was placed in the ribosyl ring and a radiolabel, 2-¹⁴C, was placed in the uracil ring to act as a reporter on the label of interest. A tritium label in the uracil ring, 5-³H, was used as a reporter on the light isotope at the position of interest, 1'-¹²C in this example. Isotope ratios, and hence KIEs, where measured by using ion exchange chromatography to isolate the uracil product from the DNA substrate after reaction.

The experimental 1'-¹³C KIE was 1.010 ± 0.009 and the 1'-²H KIE was 1.20 ± 0.02 . The β -secondary ²H KIEs were large and nearly equal, 2'S-²H KIE = 1.10 ± 0.01 , $2'R^{-2}$ H KIE = 1.11 ± 0.01 , indicative both of high oxocarbenium ion character and of strong hyperconjugation. The authors compared the experimental $1'^{-13}C$ and $1'^{-3}H$ KIEs with all available primary and α -secondary KIEs for similar glycolysis and glycosyl transfer reactions. They concluded that the $1'^{-13}$ C and $1'^{-2}$ H KIEs, taken together, likely support a $D_N * A_N$ mechanism, although a highly dissociative A_ND_N mechanism is also possible. The large secondary ²H KIE is indicative of an oxocarbenium ion(-like) structure, as is the small 1'-¹³C KIE. The reported standard deviation for the 1'-¹³C KIE is somewhat large, 0.009; however, using the reported standard deviation and the number of independent determinations, 14, it is possible to estimate the 95% confidence interval as 0.005. The 1'-¹³C is near the calculated values for $D_N * A_N$ hydrolysis of adenosine, 1.008-1.009, but it is not unambiguously smaller than the calculated lower limit of 1.013-1.015#16 for an A_ND_N transition state.⁵⁸ The inhibition of UDG by DNA containing a 2'-deoxyuridine analogue with a methylene bridge between the deoxyribosyl and uracil moieties (17) provides some support for a discrete ion pair intermediate, though K_d for the inhibitor (160 nM) was not compared with the corresponding equilibrium dissociation constant, K_s , for the analogous substrate DNA.²⁰⁴ The methylene bridge between the rings would be expected to be too long to mimic an A_ND_N transition state, but could mimic an oxocarbenium/uracil ion pair.

^{#16} It is possible to estimate a ¹³C KIE from a ¹⁴C KIE using the approximate relationship ¹⁴C KIE = (13 C KIE)^{1.89,61} The reported estimate of 1'-1⁴C KIE for D_N * A_N mechanisms was 1.015–1.018, and the lower limits for an A_ND_N mechanism were 1.025–1.029.



Fig. 21 (a) Depurination reaction catalyzed by RTA. Stem-loop DNA oligonucleotides are also substrates. (b) Substrate oligonucleotides, with reactive (deoxy-)adenosine residue in bold. (c) Inhibitors of RTA.



TS analysis, in conjunction with complementary experimental approaches, has shown that UDG catalyzes uridine hydrolysis through an oxocarbenium ion(-like) transition state with strong leaving group activation through stabilization of the uracil anion. The relative energetic importance of oxocarbenium ion stabilization is not known, but it has been noted that there is an acidic residue, Asp64, located near the α -face of the oxocarbenium ion, where it may act to stabilize the positive charge electrostatically, and may also assist the water nucleophile by orienting it, and possibly acting as a general base.

Ricin toxin A-chain (RTA)

General. Ricin is a cytotoxic protein from castor beans.²⁰⁵ The catalytic domain, RTA, catalyzes the hydrolytic depurination of a single adenosine residue, A4324, of 28S rat ribosomal RNA.^{206,207} This single depurination stops protein biosynthesis

| Isotopic label ^a | RNA hydrolysis (A | -10) ^b | DNA depurination (dA-10) ^c | | |
|-------------------------------|-------------------|-----------------------------|---------------------------------------|-----------------------------|--|
| | Experimental KIE | Calculated EIE ^d | Experimental KIE | Calculated KIE ^e | |
| $1'^{-14}C$ | 0.993 ± 0.004 | 1.007 | 1.015 ± 0.001 | 1.018 | |
| 9- ¹⁵ N | 1.016 ± 0.005 | 1.024 | 1.023 ± 0.004 | 1.024 | |
| $7-^{15}N$ | 0.981 ± 0.008 | 0.985 | - | - | |
| $1'-{}^{3}H$ | 1.163 ± 0.009 | 1.288 | 1.187 ± 0.008 | 1.368 | |
| $2'-{}^{3}H$ or $2'S-{}^{3}H$ | 1.012 ± 0.005 | 0.994 | 1.117 ± 0.002 | 1.140 | |
| 2'R- ³ H | - | _ | 1.146 ± 0.008 | 1.535 ^f | |

Table 8 Experimental and calculated KIEs on k_{cat}/K_{M} for ricin-catalyzed depurinations



а

^bFrom Ref. 46.

^cFrom Ref. 58.

^dEIEs for conversion of reactant to oxocarbenium ion plus adenine (see text).

^{*e*}Calculated KIEs for $D_N * A_N^{\ddagger}$ mechanism (i.e. $k_4 \gg k_5$). The calculated KIEs for a $D_N^{\ddagger} * A_N$ mechanism (i.e. $k_4 \ll k_5$), which are similar, were also reported.⁵⁸

^fThis extremely large KIE is probably an artefact of the computational methods used.

and causes cell death. With a reported $k_{cat} = 1777 \text{ min}^{-1}$ on intact ribosomes, a single RTA molecule is sufficient to kill a cell. The minimal substrate of RTA is an RNA or DNA oligonucleotide with a stem of at least three base pairs, and a GAGA stem-loop structure^{208–210} (Fig. 21a). RTA has poor activity against small stem-loop substrates at physiological pH, but k_{cat} increases at low pH to 4 min⁻¹ for substrate A-10, and 214 min⁻¹ for A-14 at pH 4.1, within an order of magnitude of k_{cat} against ribosomes.

KIEs were determined for a stem-loop RNA substrate A- 10^{46} and an analogous DNA substrate, dA- 10^{58} (Table 8). In both these cases, hydrolysis reaction proceeded through a stepwise $D_N * A_N$ mechanism. Isotope-trapping experiments showed that substrate binding was not kinetically significant; however, the experimental KIEs with A-10 as substrate were inconsistent with any mechanism where only chemical steps were kinetically significant, implying equilibrium formation of an RTA·ox-ocarbenium ion-adenine complex, followed by an isotopically insensitive step. In contrast, the KIEs for dA-10 were consistent with a $D_N * A_N$ mechanism.

TS analysis of a DNA substrate. The experimental KIEs for dA-10, in particular the small primary 1'-¹⁴C KIE = 1.015 ± 0.001, were not consistent with an A_ND_N

mechanism. This is smaller than the calculated minimum KIE for an $A_N D_N$ mechanism, 1.025 when calculated by BOVA, and 1.029 by electronic structure methods. The large primary ¹⁵N KIE indicated a large extent of C–N bond breakage at the transition state, and the α - and β -secondary ³H KIEs were also large, indicating high oxocarbenium ion character. Taken together, these KIEs supported a $D_N * A_N$ mechanism.

The observable KIEs (see Section 3) were calculated using *ab initio* electronic structure methods for model compounds of the RTA-catalyzed depurination of dA-10 (Table 8). The calculated KIEs for the two mechanistic extremes, $k_4 \gg k_5$ or $k_5 \gg k_4$, were very similar to each other and it was not possible to distinguish between these two possibilities from the experimental KIEs.

TS analysis of an RNA substrate. Interpretation of the KIEs for A-10 hydrolysis was not straightforward. It was possible to make the following mechanistic deductions only because the KIEs for the DNA reaction were consistent with a $D_N * A_N$ mechanism with only chemical steps being kinetically significant. Having the DNA reaction as a benchmark made it possible to interpret the unusual KIEs for the RNA reaction. The experimental KIEs were not consistent with a mechanism where only chemical steps were kinetically significant, and isotope-trapping measurements of commitment to catalysis demonstrated that substrate binding was not kinetically significant. Instead, the KIEs indicated an oxocarbenium ion intermediate in equilibrium with reactant RNA, and with the first irreversible step being isotopically insensitive, occurring after oxocarbenium ion formation (Fig. 7). The primary, $1'^{-14}$ C KIE of 0.993 \pm 0.004, was even lower than for dA-10 and inconsistent with a $D_N * A_N$ mechanism. The primary 9-¹⁵N KIE of 1.016 ± 0.005 was less than expected for complete C-N bond breakage, 1.024, but still indicated a large amount of bond breakage. The α -secondary KIE indicated significant oxocarbenium ion character. These experimental KIEs, in particular the primary ¹⁴C KIE, were the most consistent with the EIEs of forming an oxocarbenium ion. The observable KIEs will be equal to the calculated EIEs of oxocarbenium ion formation if the next step in the reaction, k_5 , is the first irreversible one, and it is isotopically insensitive, i.e. $\alpha_5 = 1$. In other words, if $k_4 \gg k_5$ and $\alpha_5 = 1$, then $\text{KIE}_{\text{observable}} = \text{EIE}_{\text{oxocarbenium}} = (\alpha_1 \alpha_3) / (\alpha_2 \alpha_4)$. The nature of the isotopically insensitive step k_5 is not known, but could be, e.g. a protein conformational change or diffusion of water into position to perform nucleophilic attack on the oxocarbenium ion. In either case, an isotopic label on the substrate molecule would have no effect on the rate of this step. As seen in Fig. 7, the next step is nucleophilic attack by water. Because the first irreversible step has already been passed, this step is not kinetically significant for k_{cat}/K_{M} and will not be reflected in the observable KIEs.

Leaving group activation. There is evidence that RTA activates the adenine leaving group for departure by protonation. The large inverse 7-¹⁵N KIE was evidence that N7 is protonated in the RTA·oxocarbenium ion-adenine complex. The pH profile of RTA activity against A-10 gave evidence for protonation of N1. The pK_a value for N1 of a non-base-paired adenine is near 4.²¹¹ The k_{cat}/K_M versus pH

profile of RTA-catalyzed hydrolysis of A-10 is bell-shaped, with maximal activity around pH 4. Activity decreases as pH increases, with a pH-dependence consistent with two groups with $pK_a = 4.0^{212}$ The pH profile of RTA with 28S RNA in intact ribosomes has not been reported, but must be different from A-10 because the two ionizations in the descending limb would imply a 10^7 -fold decrease in k_{cat}/K_M at the physiological pH of 7.4. The pH profile of A-10 hydrolysis is consistent with N1 of the susceptible adenine being one of the groups with $pK_a = 4.0$. This is close to the unperturbed pK_a of an adenine base, suggesting that RTA is contributing little energy to protonate the substrate at N1. In contrast, it was proposed that the N1 atom of A4234 in ribosomes is protonated by the enzyme to promote catalysis at physiological pH. Thus, the enzyme uses binding energy from interaction with 28S RNA that is unavailable in interactions with A-10 RNA to promote protonation of N1. The energetic importance of protonation at both N1 and N7 was demonstrated by calculating the energetics of C1'-N9 bond dissociation in the model compound (18).⁴⁶ The bond dissociation energy was approximately 5.5 kcal/mol for N1H,N7H-18, as opposed to 68 and 55 kcal/mol, for N1H-18 and N7H-18, respectively.



Importance of 2'-hydroxyl group. The importance of the 2'-hydroxyl group to catalysis can be examined via the kinetic constants for RNA versus DNA substrates, A-10 versus dA-10. The value of k_{cat}/K_{M} for A-10 was 5.7-fold higher than dA-10. Adenosine undergoes acid-catalyzed hydrolysis 650-fold slower than 2'-deoxyadenosine,²¹³ similar to the difference in rates of acid-catalyzed hydrolysis between other 2-hydroxy- and 2-deoxy-N- and O-glycosides² as well as uncatalyzed Oglycoside hydrolysis.²¹⁴ Given the structural similarity between the highly oxocarbenium ion-like A_ND_N transition states and the corresponding oxocarbenium ions for 2-hydroxy and 2-deoxy sugars, this difference in reaction rates provides an estimate of the free energy differences between 2-hydroxy- and 2-deoxyoxocarbenium ions. The combination of a 5.7-fold higher value of k_{cat}/K_M with the 650-fold lower reactivity means that RTA accelerates the reaction of A-10 by 3700-fold relative the expectation based on its intrinsic reactivity. This means that the enzyme achieves 4.8 kcal/mol of stabilization of the transition state of the RNA reaction from the presence of the 2'-hydroxyl groups. Because every nucleotide residue of A-10 contains a 2'-hydroxyl, it is not possible to attribute this increase in k_{cat}/K_{M} to a single group. However, Orita et al.²¹⁵ reported that a stem-loop RNA substrate with a 2'-deoxyadenosine (dA) residue only at the susceptible position,

GdAGA, of the molecule had 26-fold higher "activity" than the all-RNA analogue. This report does not make it clear whether "activity" corresponds to k_{cat} or k_{cat}/K_{M} , and reactions were performed at pH 7.4, so direct comparison with the kinetic constants of A-10 and dA-10 are difficult. Nonetheless, the effect of an A to dA substitution at the susceptible position was between the 650-fold increase expected based on reactivity and the 5.7-fold decrease between A-10 and dA-10 implying contributions from the 2'-hydroxyls both at the susceptible position and at other positions. The 4.8 kcal/mol of stabilization achieved by the enzyme, compared to the 3.8 kcal/mol intrinsically higher activation energy for RNA hydrolysis, implies that more than enough energy is derived from enzymatic interactions with the 2-hydroxyl groups to stabilize the inherently less stable 2-hydroxy-ribooxocarbenium ion.

Inhibitor design. The results of recent inhibitor design studies²¹⁶ have provided support for the proposed $D_N * A_N$ mechanism. In this study, an iminosugar inhibitor, IA-10 (Fig. 21c), gave only modest inhibition of RTA, $K_i = 0.6 \mu M$, as did an inhibitor without the purine ring analogue, 1N-14, $K_i = 0.5 \mu M$. However, 50-fold stronger inhibition was observed with 1N-14 in the presence of adenine, which allowed formation of an RTA·1N-14·adenine complex with $K_i = 0.012 \mu M$. Tight binding was expected for the combination of 1N-14 and adenine by analogy to the RTA·oxocarbenium ion·adenine complex, where the distance between the oxocarbenium ion and adenine may be too large to be bridged by the C–C bond in IA-10. More potent inhibition may be possible by combining features of 1N-14 and adenine with a covalent linkage to reduce the entropy loss that must be compensated in forming a ternary complex.^{217,218}

5 Conclusions and future directions

In this chapter, we have attempted to provide a summary of the use of TS analysis in understanding glycoside chemistry, as well as an overview of recent developments in the field. TS analysis is only one tool in any comprehensive analysis of reactivity and catalysis, but an extremely powerful tool.

THE IMMEDIATE FUTURE

The immediate future is relatively clear. Continued advances in instrumental techniques, particularly in mass spectrometry and NMR, will make it possible to measure increasingly accurate and precise KIEs on increasingly small amounts of material. At the same time, continued growth in computational power and in the methods of KIE interpretation will make TS analysis an increasingly powerful tool. Currently, one major drawback is that it is too time consuming at present for application in the pharmaceutical industry. TS analysis will have to become much faster to see wide application outside of academia.

CRUCIAL QUESTIONS

It has become abundantly clear that glycoside hydrolyses and transfers are on the borderline between highly dissociative $A_N D_N$ transition states and $D_N * A_N$ stepwise mechanisms. In both cases, the sugar ring takes on cationic character which can be exploited by enzymes to promote catalysis and by chemists to design inhibitors. What other questions can TS analysis address in the study of glycoside chemistry?

- (1) Conformation of the sugar ring. Glycosidases and glycosyl transferases can enforce particular sugar ring conformations and there can be strong conformational effects on glycoside reactivity, such as those mediated through the anomeric effect and hyperconjugation. TS analysis can provide information on sugar ring conformations through secondary hydron KIEs. In addition, knowledge of sugar ring conformation can provide indirect information on the direction of the backbone chain in oligosaccharide or oligonucleotide substrates.
- (2) Oxocarbenium ion stabilization. Even though all glycoside reactions appear to proceed through oxocarbenium ion(-like) transition states or intermediates, not all enzymes interact strongly with the sugar part of the substrate. This point will not easily be addressed solely with KIE studies, but it is an important issue in catalysis.
- (3) *Leaving group activation.* Related to point 2, those enzymes that do not interact strongly with the sugar ring must achieve catalysis through interactions with the leaving group and nucleophile. In order to clarify the role of leaving group activation, remote KIEs have been measured, such as 7-¹⁵N KIEs in purines. Further study is needed to understand the mechanisms of leaving group activation.
- (4) Concerted $(A_N D_N)$ versus stepwise $(D_N * A_N)$ mechanisms. If an enzyme uses an $A_N D_N$ mechanism, then the position of the leaving group relative to the glycosyl ring is well defined in the TS structure. However, if a discrete oxocarbenium ion intermediate is formed, then it is possible that the leaving group has moved a large distance and/or has rotated relative to the glycosyl ring in this intermediate. This will have implications for inhibitor design, where compounds with extra atoms inserted between the sugar ring analogue and the leaving group analogue may be effective.^{204,219}
- (5) *Enzyme-substrate interactions.* While TS analysis can provide a detailed TS structure, and it is possible to dock that structure into an enzyme active site, it is not possible to determine by inspection or computation the energetics of individual enzyme-substrate contacts. An added complication is the fact that active sites generally change structure upon binding to the transition state and throughout the catalytic cycle. Further experimental studies, in combination with computational analyses of enzyme-substrate interactions are needed to understand the energetics of catalysis.

As the power of TS analysis increases, the questions that can be posed will evolve, becoming more subtle and more powerful.

Acknowledgments

The authors thank Professor Vern Schramm for his long-term and continuing support, and for helpful discussions. We also thank Drs Andy Bennet and Jim Stivers for sharing their results before publication. The writing of this chapter was supported by the Canadian Institutes of Health Research (CIHR) and the Natural Sciences and Engineering Research Council (NSERC).

References

- 1. BeMiller, J.N. (1967). Adv. Carbohydr. Chem. Biochem. 22, 25
- 2. Capon, B. (1969). Chem. Rev. 69, 407
- 3. Cordes, E.H. and Bull, H.G. (1974). Chem. Rev. 74, 581
- 4. Sinnott, M.L. (1990). Chem. Rev. 90, 1171
- 5. Zechel, D.L. and Withers, S.G. (2000). Acc. Chem. Res. 33, 11
- 6. Rowe, P.M. (1999). Lancet 354, 402
- 7. Dwek, R.A. (1996). Chem. Rev. 96, 683
- 8. Robyt, J.F. (1998). Essentials of Carbohydrate Chemistry. Springer, New York
- 9. Hecht, S.M. (1999). *Bioorganic Chemistry: Carbohydrates*. Oxford University Press, New York
- 10. Warshel, A. (1978). Proc. Natl Acad. Sci. USA 75, 5250
- Weiss, P.M. (1991). Enzyme Mechanism from Isotope Effects. Cook, P.F. (ed.), p. 291. CRC Press, Boca Raton, FL
- 12. Parkin, D.W. (1991). *Enzyme Mechanism from Isotope Effects*. Cook, P.F. (ed.), p. 269. CRC Press, Boca Raton, FL
- 13. Rosenberg, S. and Kirsch, J.F. (1979). Anal. Chem. 51, 1375
- 14. Bahnson, B.J. and Anderson, V.E. (1991). Biochemistry 30, 5894
- 15. Gawlita, E., Paneth, P. and Anderson, V.E. (1995). Biochemistry 34, 6050
- 16. Kline, P.C., Rezaee, M. and Lee, T.A. (1999). Anal. Biochem. 275, 6
- 17. Berti, P.J., Blanke, S.R. and Schramm, V.L. (1997). J. Am. Chem. Soc. 119, 12079
- Matsson, O., Axelsson, S., Hussenius, A. and Ryberg, P. (1999). Acta Chem. Scand. 53, 670
- 19. Xue, H., Wu, X. and Huskey, W.P. (1996). J. Am. Chem. Soc. 118, 5804
- 20. Zhou, X.Z. and Toney, M.D. (1998). J. Am. Chem. Soc. 120, 13282
- 21. Rosenberg, S. and Kirsch, J.F. (1979). Anal. Chem. 51, 1379
- 22. Singleton, D.A. and Thomas, A.A. (1995). J. Am. Chem. Soc. 117, 9357
- Singleton, D.A., Merrigan, S.R., Kim, B.J., Beak, P., Phillips, L.M. and Lee, J.K. (2000). J. Am. Chem. Soc. 122, 3296
- Sims, L.B. and Lewis, D.E. (1984). Isotope Effects: Recent Developments in Theory and Experiment, Buncel, E. and Leeds, C.C. (eds), vol. 6, p. 161. Elsevier, New York
- 25. Sims, L. B.; Burton, G. W.; Lewis, D. E. (1997). BEBOVIB-IV, QCPE No. 337. Quantum Chemistry Program Exchange, Department of Chemistry, University of Indiana, Bloomington, IN

- Fedorov, A., Shi, W., Kicska, G., Fedorov, E., Tyler, P.C., Furneaux, R.H., Hanson, J.C., Gainsford, G.J., Larese, J.Z., Schramm, V.L. and Almo, S.C. (2001). *Biochemistry* 40, 853
- 27. Kline, P.C. and Schramm, V.L. (1995). Biochemistry 34, 1153
- 28. Kline, P.C. and Schramm, V.L. (1993). Biochemistry 32, 13212
- 29. Wang, F., Miles, R.W., Kicska, G., Nieves, E., Schramm, V.L. and Angeletti, R.H. (2000). *Protein Sci.* 9, 1660
- Mao, C., Cook, W.J., Zhou, M., Federov, A.A., Almo, S.C. and Ealick, S.E. (1998). Biochemistry 37, 7135
- 31. Kline, P.C. and Schramm, V.L. (1992). Biochemistry 31, 5964
- Miles, R.W., Tyler, P.C., Furneaux, R.H., Bagdassarian, C.K. and Schramm, V.L. (1998). *Biochemistry* 37, 8615
- Kicska, G.A., Long, L., Horig, H., Fairchild, C., Tyler, P.C., Furneaux, R.H., Schramm, V.L. and Kaufman, H.L. (2001). *Proc. Natl Acad. Sci. USA* 98, 4593
- 34. Pauling, L. (1946). Chem. Eng. News 24, 1375
- 35. Fersht, A.R. (1985). Enzyme Structure and Mechanism. W.H. Freeman, New York
- 36. Radzicka, A. and Wolfenden, R. (1995). Science 267, 90
- 37. Levy, S.B. (1998). Sci. Am. 278, 46
- 38. Guthrie, R.D. and Jencks, W.P. (1989). Acc. Chem. Res. 22, 343
- 39. Commission on Physical Organic Chemistry (1989). Pure Appl. Chem. 61, 23
- 40. Commission on Physical Organic Chemistry (1989). Pure Appl. Chem. 61, 57
- 41. Gutfreund, H. and Sturtevant, J.M. (1956). Biochem. J. 63, 656
- 42. Cleland, W.W. (1982). Methods Enzymol. 87, 625
- 43. Berti, P.J. (1999). Methods Enzymol. 308, 355
- 44. Rose, I.W. (1980). Methods Enzymol. 64, 47
- 45. Northrop, D.B. (1981). Annu. Rev. Biochem. 50, 103
- 46. Chen, X.-Y., Berti, P.J. and Schramm, V.L. (2000). J. Am. Chem. Soc. 122, 1609
- 47. Scheuring, J., Berti, P.J. and Schramm, V.L. (1998). Biochemistry 37, 2748
- 48. Bruner, M. and Horenstein, B.A. (2000). Biochemistry 39, 2261
- 49. Hosie, L. and Sinnott, M.L. (1985). Biochem. J. 226, 437
- 50. Bennet, A.J. and Sinnott, M.L. (1986). J. Am. Chem. Soc. 108, 7287
- 51. Huang, X., Surry, C., Hiebert, T. and Bennet, A.J. (1995). J. Am. Chem. Soc. 117, 10614
- 52. Zhu, J. and Bennet, A.J. (1998). J. Am. Chem. Soc. 120, 3887
- 53. Zhu, J. and Bennet, A.J. (2000). J. Org. Chem. 65, 4423
- 54. Horenstein, B.A. and Bruner, M. (1996). J. Am. Chem. Soc. 118, 10371
- 55. Horenstein, B.A. and Bruner, M. (1998). J. Am. Chem. Soc. 120, 1357
- 56. Johnston, H.S. (1966). Gas Phase Reaction Rate Theory. Ronald Press, New York
- 57. Melander, L. and Saunders, W.H., Jr. (1980). *Reaction Rates of Isotopic Molecules*. Wiley, New York
- 58. Chen, X.-Y., Berti, P.J. and Schramm, V.L. (2000). J. Am. Chem. Soc. 122, 6527
- 59. Houk, K.N., Gustafson, S.M. and Black, K.A. (1992). J. Am. Chem. Soc. 114, 8565
- 60. Wilkie, J. and Williams, I.H. (1995). J. Chem. Soc., Perkin Trans. 2 1559
- 61. Huskey, W.P. (1991). *Enzyme Mechanism from Isotope Effects*, Cook, P.F. (ed.), p. 37. CRC Press, Boca Raton, FL
- 62. Suhnel, J. and Schowen, R.L. (1991). *Enzyme Mechanism from Isotope Effects*, Cook, P.F. (ed.), p. 3. CRC Press, Boca Raton, FL
- 63. Glad, S.S. and Jensen, F. (1996). J. Phys. Chem. 100, 16892
- 64. Scott, A.P. and Radom, L. (1996). J. Phys. Chem. 100, 16502
- 65. Wong, M.W. (1996). Chem. Phys. Lett. 256, 391
- 66. Burton, G.W., Sims, L.B., Wilson, J.C. and Fry, A. (1977). J. Am. Chem. Soc. 99, 3371
- 67. Huskey, W.P. (1996). J. Am. Chem. Soc. 118, 1663
- 68. Casamassina, T.E. and Huskey, W.P. (1993). J. Am. Chem. Soc. 115, 14

- 69. Berti, P.J. and Schramm, V.L. (1997). J. Am. Chem. Soc. 119, 12069
- 70. Rising, K.A. and Schramm, V.L. (1997). J. Am. Chem. Soc. 119, 27
- Horenstein, B.A., Parkin, D.W., Estupinan, B. and Schramm, V.L. (1991). *Biochemistry* 30, 10788
- 72. Andrews, C.W., Bowen, J.P. and Fraser-Reid, B. (1989). J. Chem. Soc., Chem. Commun. 1913
- 73. Winkler, D.A. and Holan, G. (1989). J. Med. Chem. 32, 2084
- 74. Andrews, C.W., Fraser-Reid, B. and Bowen, J.P. (1991). J. Am. Chem. Soc. 113, 8293
- Kajimoto, T., Liu, K.K.C., Pederson, R.L., Zhong, Z., Ichikawa, Y., Porco, J.A., Jr. and Wong, C.-H. (1991). *J. Med. Chem.* **113**, 6187
- 76. Smith, B.J. (1997). J. Am. Chem. Soc. 119, 2699
- 77. Namchuk, M.N., McCarter, J.D., Becalski, A., Andrews, T. and Withers, S.G. (2000). J. Am. Chem. Soc. **122**, 1270
- 78. Mentch, F., Parkin, D.W. and Schramm, V.L. (1987). Biochemistry 26, 921
- Parkin, D.W., Mentch, F., Banks, G.A., Horenstein, B.A. and Schramm, V.L. (1991). Biochemistry 30, 4586
- 80. Scheuring, J. and Schramm, V.L. (1997). Biochemistry 36, 4526
- 81. Scheuring, J. and Schramm, V.L. (1997). Biochemistry 36, 8215
- Mazzella, L.J., Parkin, D.W., Tyler, P.C., Furneaux, R.H. and Schramm, V.L. (1996). J. Am. Chem. Soc. 118, 2111
- 83. Inouye, S., Tsuruoka, T., Ito, T. and Niida, T. (1968). Tetrahedron 23, 2125
- 84. Legler, G. (1990). Adv. Carbohydr. Chem. Biochem. 48, 319
- Jeong, J.H., Murray, B.W., Takayama, S. and Wong, C.-H. (1996). J. Am. Chem. Soc. 118, 4227
- 86. Hoos, R., Huixin, J., Vasella, A. and Weiss, P. (1996). Helv. Chim. Acta 79, 1757
- Yagi, M., Kouno, T., Aoyagi, Y. and Murai, H. (1976). Nippon Nogei Kagaku Kaishi 50, 571
- 88. Legler, G. and Julich, E. (1984). Carbohydr. Res. 128, 61
- Evans, S.V., Fellows, L.E., Shing, T.K.M. and Fleet, G.W.J. (1985). *Phytochemistry* 24, 1953
- 90. Legler, G. and Pohl, S. (1986). Carbohydr. Res. 155, 119
- Winchester, B., Barker, C., Baines, S., Jacob, G.S., Namgoong, S.K. and Fleet, G. (1990). *Biochem. J.* 265, 277
- Cenci di Bello, I., Dorling, P., Fellows, L. and Winchester, B. (1984). FEBS Lett. 176, 61
- 93. Hughes, A.B. and Rudge, A.J. (1994). Nat. Prod. Rep. 135
- Jespersen, T.M., Dong, W., Sierks, M.R., Skrydstrup, T., Lundt, I. and Bols, M. (1994). Angew. Chem. Int. Ed. Engl. 33, 1778
- 95. Ichikawa, Y. and Igarashi, Y. (1995). Tetrahedron Lett. 36, 4585
- 96. Igarashi, Y., Ichikawa, M. and Ichikawa, Y. (1996). Tetrahedron Lett. 37, 2707
- 97. Bols, M., Hazell, R.G. and Thomsen, I.B. (1997). Chem. Eur. J. 3, 940
- 98. Bols, M. (1998). Acc. Chem. Res. 31, 1
- 99. Liu, H., Liang, X., Shoel, H., Bülow, A. and Bols, M. (2001). J. Am. Chem. Soc. 123, 5116
- 100. Parkin, D.W. and Schramm, V.L. (1995). Biochemistry 34, 13961
- 101. Degano, M., Almo, S.C., Sacchettini, J.C. and Schramm, V.L. (1998). *Biochemistry* **37**, 6277
- 102. Shi, W., Li, C.M., Tyler, P.C., Furneaux, R.H., Cahill, S.M., Girvin, M.E., Grubmeyer, C., Schramm, V.L. and Almo, S.C. (1999). *Biochemistry* 38, 9872
- 103. Bell, C.E. and Eisenberg, D. (1996). Biochemistry 35, 1137
- 104. Bruner, M. and Horenstein, B.A. (1998). Biochemistry 37, 289
- 105. Bernasconi, C.F. (1992). Adv. Phys. Org. Chem. 27, 119
- 106. Bernasconi, C.F. (1992). Acc. Chem. Res. 25, 9
- 107. Kresge, A.J. (1974). Can. J. Chem. 52, 1897
- 108. Yamataka, H., Mustanir, and Mishima, M. (1999). J. Am. Chem. Soc. 121, 10223
- 109. Bernasconi, C.F. and Wenzel, P.J. (1994). J. Am. Chem. Soc. 116, 5405
- 110. Bernasconi, C.F. and Wenzel, P.J. (1996). J. Am. Chem. Soc. 118, 10494
- 111. Bernasconi, C.F. (1985). Tetrahedron 41, 3219
- 112. Richard, J.P. (1995). Tetrahedron 51, 1535
- 113. Richard, J.P., Williams, K.B. and Amyes, T.L. (1999). J. Am. Chem. Soc. 121, 8403
- 114. Rakowski, K. and Paneth, P. (1996). J. Mol. Struct. 378, 35
- 115. Gawlita, E., Caldwell, W.S., Oleary, M.H., Paneth, P. and Anderson, V.E. (1995). Biochemistry 34, 2577
- 116. Poirier, R.A., Wang, Y. and Westaway, K.C. (1994). J. Am. Chem. Soc. 116, 2526
- 117. Matsson, O. and Westaway, K.C. (1998). Adv. Phys. Org. Chem. 31, 143
- 118. Glad, S.S. and Jensen, F. (1997). J. Am. Chem. Soc. 119, 227
- 119. Westaway, K.C. (1975). Tetrahedron Lett. 48, 4229
- 120. Westaway, K.C., VanPham, T. and Fang, Y.R. (1997). J. Am. Chem. Soc. 119, 3670
- 121. Barnes, J.A. and Williams, I.H. (1993). J. Chem. Soc., Chem. Commun. 1286
- 122. Koerner, T., Westaway, K.C., Poirier, R.A. and Wang, Y. (2000). Can. J. Chem. 78, 1067
- 123. Wade, D. (1999). Chem. Biol. Interact. 117, 191
- 124. Lewis, B.E. and Schramm, V.L. (2001). J. Am. Chem. Soc. 123, 1327
- 125. Stein, R.L., Romero, R., Bull, H.G. and Cordes, E.H. (1978). J. Am. Chem. Soc. 100, 6249
- 126. LaReau, R.D., Wan, W. and Anderson, V.E. (1989). Biochemistry 28, 3619
- 127. Gawlita, E., Lantz, M., Paneth, P., Bell, A.F., Tonge, P.J. and Anderson, V.E. (2000). *J. Am. Chem. Soc.* **122**, 11660
- 128. D'Ordine, R.L., Bahnson, B.J., Tonge, P.J., Carey, P.R. and Anderson, V.E. (1994). Biochemistry 33, 14733
- 129. Bell, A.F., Wu, J., Feng, Y. and Tonge, P.J. (2001). Biochemistry 40, 1725
- 130. Cheng, H., Sukal, S., Deng, H., Leyh, T.S. and Callender, R. (2001). *Biochemistry* 40, 4035
- 131. Deng, H., Wang, J.H., Callender, R.H., Grammer, J.C. and Yount, R.G. (1998). *Biochemistry* **37**, 10972
- 132. Sunko, D.E., Szele, I. and Hehre, W.J. (1977). J. Am. Chem. Soc. 99, 5000
- 133. Jucker, F.M., Heus, H.A., Yip, P.F., Moors, E.H. and Pardi, A. (1996). *J. Mol. Biol.* **264**, 968
- 134. Correll, C.C., Munishkin, A., Chan, Y.L., Ren, Z., Wool, I.G. and Steitz, T.A. (1998). Proc. Natl Acad. Sci. USA 95, 13436
- 135. Werner, R.M. and Stivers, J.T. (2000). Biochemistry 39, 14054
- 136. Stern, M.J. and Vogel, P.C. (1971). J. Am. Chem. Soc. 93, 4664
- 137. Huang, X.C., Tanaka, K.S.E. and Bennet, A.J. (1997). J. Am. Chem. Soc. 119, 11147
- 138. Parkin, D.W., Leung, H.B. and Schramm, V.L. (1984). J. Biol. Chem. 259, 9411
- 139. Parkin, D.W. and Schramm, V.L. (1987). Biochemistry 26, 913
- 140. Tarnus, C., Muller, H.M. and Schuber, F. (1988). Bioorg. Chem. 16, 38
- 141. Johnson, R.W., Marschner, T.M. and Oppenheimer, N.J. (1988). J. Am. Chem. Soc. 110, 2257
- 142. Miles, R.W., Tyler, P.C., Evans, G.B., Furneaux, R.H., Parkin, D.W. and Schramm, V.L. (1999). *Biochemistry* **38**, 13147
- 143. Moodie, S.L. and Thornton, J.M. (1993). Nucleic Acids Res. 21, 1369
- 144. Drohat, A.C. and Stivers, J.T. (2000). Biochemistry 39, 11865
- 145. Drohat, A.C. and Stivers, J.T. (2000). J. Am. Chem. Soc. 122, 1840
- 146. Dinner, A.R., Blackburn, G.M. and Karplus, M. (2001). Nature 413, 752

- 147. Parikh, S.S., Walcher, G., Jones, G.D., Slupphaug, G., Krokan, H.E., Blackburn, G.M. and Tainer, J.A. (2000). *Proc. Natl Acad. Sci. USA* **97**, 5083
- 148. Dunathan, H.C. (1966). Proc. Natl Acad. Sci. USA 55, 712
- 149. Shi, H. and Moore, P.B. (2000). RNA 6, 1091
- 150. Jovine, L., Djordjevic, S. and Rhodes, D. (2000). J. Mol. Biol. 301, 401
- 151. Dong, J., Drohat, A.C., Stivers, J.T., Pankiewicz, K.W. and Carey, P.R. (2000). Biochemistry 39, 13241
- 152. Bennett, M.J., Choe, S. and Eisenberg, D. (1994). Protein Sci. 3, 1444
- 153. Weiss, M.S., Blanke, S.R., Collier, R.J. and Eisenberg, D. (1995). Biochemistry 34, 773
- 154. Bennett, M.J. and Eisenberg, D. (1994). Protein Sci. 3, 1464
- 155. Bell, C.E. and Eisenberg, D. (1997). Biochemistry 36, 481
- 156. Choe, S., Bennett, M.J., Fujii, G., Curmi, P.M., Kantardjieff, K.A., Collier, R.J. and Eisenberg, D. (1992). *Nature* **357**, 216
- 157. Oppenheimer, N.J. and Bodley, J.W. (1981). J. Biol. Chem. 256, 8579
- 158. Wilson, B.A., Reich, K.A., Weinstein, B.R. and Collier, R.J. (1990). *Biochemistry* 29, 8643
- 159. Koshland, D.E., Jr. (1953). Biol. Rev. 28, 416
- 160. Davies, G., Sinnott, M.L. and Withers, S.G. (1997). *Comprehensive Biological Catalysis*. Sinnott, M. (ed.), p. 120. Academic Press, London
- 161. Indurugalla, D. and Bennet, A.J. (2001). J. Am. Chem. Soc. 123, 10889
- 162. Zhang, Y., Bommuswamy, J. and Sinnott, M.L. (1994). J. Am. Chem. Soc. 116, 7557
- 163. Tanaka, Y., Tao, W., Blanchard, J.S. and Hehre, E.J. (1994). J. Biol. Chem. 269, 32306
- 164. Deslongchamps, P. (1993). Pure Appl. Chem. 65, 1161
- 165. Streitwieser, A., Jr., Jagow, R.H., Fahey, R.C. and Suzuki, S. (1958). J. Am. Chem. Soc. 80, 2326
- 166. Wong, C.-H., Halcomb, R.L., Ichikawa, Y. and Kajimoto, T. (1995). Angew. Chem. Int. Ed. Engl. 34, 412
- 167. Wong, C.-H., Halcomb, R.L., Ichikawa, Y. and Kajimoto, T. (1995). *Angew. Chem. Int. Ed. Engl.* **34**, 521
- Simanek, E.E., McGarvey, G.J., Jablonowski, J.A. and Wong, C.-H. (1998). *Chem. Rev.* 98, 833
- 169. Sears, P. and Wong, C.-H. (1999). Angew. Chem. Int. Ed. Engl. 38, 2300
- 170. Yang, J., Schenkman, S. and Horenstein, B.A. (2000). Biochemistry 39, 5902
- 171. Schenkman, S., Eichinger, D., Pereira, M.E.A. and Nussenzweig, V. (1994). Annu. Rev. Microbiol. 48, 499
- 172. Tarnus, C. and Schuber, F. (1987). Bioorg. Chem. 15, 31
- 173. Jencks, W.P. (1981). Chem. Soc. Rev. 10, 345
- 174. Jencks, W.P. (1980). Acc. Chem. Res. 13, 161
- 175. Oppenheimer, N.J. and Tashma, R. (1992). Biochemistry 31, 2195
- 176. Cherian, X.M., Van Arman, S.A. and Czarnick, A.W. (1990). J. Am. Chem. Soc. 112, 4490
- 177. Ritchie, C.D. (1986). Can. J. Chem. 64, 2239
- 178. Roberts, D.D., Lewis, S.D., Ballou, D.P., Olson, S.T. and Shafer, J.A. (1986). *Biochemistry* 25, 5595
- 179. Karsten, W.E., Lai, C.-J. and Cook, P.F. (1995). J. Am. Chem. Soc. 117, 5914
- Stein, P.E., Boodhoo, A., Armstrong, G.D., Cockle, S.A., Klein, M.H. and Read, R.J. (1994). *Structure* 2, 45
- 181. Antoine, R. and Locht, C. (1994). J. Biol. Chem. 269, 6450
- 182. Bull, H.G., Ferraz, J.P., Cordes, E.H., Ribbi, A. and Apitz-Castro, R. (1978). J. Biol. Chem. 253, 5186
- 183. Parkin, D.W., Horenstein, B.A., Abdulah, D.R., Estupinan, B. and Schramm, V.L. (1991). J. Biol. Chem. 266, 20658

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- 184. Horenstein, B.A. and Schramm, V.L. (1993). Biochemistry 32, 7089
- 185. Horenstein, B.A. and Schramm, V.L. (1993). Biochemistry 32, 9917
- 186. Horenstein, B.A., Zabinski, R.F. and Schramm, V.L. (1993). Tetrahedron Lett. 34, 7213
- Boutellier, M., Horenstein, B.A., Semenyaka, A., Schramm, V.L. and Ganem, B. (1994). Biochemistry 33, 3994
- 188. Deng, H., Chan, A.W.-Y., Bagdassarian, C.K., Estupinan, B., Ganem, B., Callender, R.H. and Schramm, V.L. (1996). *Biochemistry* 35, 6037
- Gopaul, D.N., Meyer, S.L., Degano, M., Sacchettini, J.C. and Schramm, V.L. (1996). Biochemistry 35, 5963
- Degano, M., Gopaul, D.N., Scapin, G., Schramm, V.L. and Sacchettini, J.C. (1996). Biochemistry 35, 5971
- 191. Bagdassarian, C.K., Schramm, V.L. and Schwartz, S.D. (1996). J. Am. Chem. Soc. 118, 8825
- 192. Braunheim, B.B. and Schwartz, S.D. (1999). Methods Enzymol. 308, 398
- 193. Braunheim, B.B. and Schwartz, S.D. (2000). J. Theor. Biol. 206, 27
- Braunheim, B.B., Miles, R.W., Schramm, V.L. and Schwartz, S.D. (1999). *Biochemistry* 38, 16076
- 195. Parkin, D.W. and Schramm, V.L. (1984). J. Biol. Chem. 259, 9418
- 196. Tao, W., Grubmeyer, C. and Blanchard, J.S. (1996). Biochemistry 35, 14
- 197. Goitein, R.K., Chelsky, D. and Parsons, S.M. (1978). J. Biol. Chem. 253, 2963
- 198. Scapin, G., Grubmeyer, C. and Sacchettini, J.C. (1994). Biochemistry 33, 1287
- 199. Scapin, G., Ozturk, D.H., Grubmeyer, C. and Sacchettini, J.C. (1995). *Biochemistry* 34, 10744
- 200. Focia, P.J., Craig, S.P., III and Eakin, A.E. (1998). Biochemistry 37, 17120
- 201. Werner, R.M., Jiang, Y.L., Gordley, R.G., Jagadeesh, G.J., Ladner, J.E., Xiao, G., Tordova, M., Gilliland, G.L. and Stivers, J.T. (2000). *Biochemistry* **39**, 12585
- 202. Stivers, J.T. (1998). Nucleic Acids Res. 26, 3837
- 203. Stivers, J.T., Pankiewicz, K.W. and Watanabe, K.A. (1999). Biochemistry 38, 952
- 204. Sekino, Y., Bruner, S.D. and Verdine, G.L. (2000). J. Biol. Chem. 275, 36506
- 205. Olsnes, S. and Pihl, A. (1982). *Molecular Action of Toxins and Viruses*, Cohen, P. and van Heyningeneds, S. (eds), p. 51. Elsevier Biomedical, New York
- 206. Endo, Y. and Tsurugi, K. (1987). J. Biol. Chem. 262, 8128
- 207. Endo, Y. and Tsurugi, K. (1988). J. Biol. Chem. 263, 8735
- 208. Gluck, A., Endo, Y. and Wool, I.G. (1992). J. Mol. Biol. 226, 411
- 209. Endo, Y., Gluck, A. and Wool, I.G. (1991). J. Mol. Biol. 221, 193
- Orlowski, M., Orlowski, R., Chang, J.C., Wilk, E. and Lesser, M. (1984). *Mol. Cell. Biochem.* 64, 155
- 211. Saenger, W. (1984). Principles of Nucleic Acid Structure. Springer, New York
- 212. Chen, X.Y., Link, T.M. and Schramm, V.L. (1998). Biochemistry 37, 11605
- 213. Venner, H. (1964). Z. Physiol. Chem. 339, 14
- 214. Wolfenden, R., Lu, X. and Young, G. (1998). J. Am. Chem. Soc. 120, 6814
- 215. Orita, M., Nishikawa, F., Kohno, T., Senda, T., Mitsui, Y., Yaeta, E., Kazunari, T. and Nishikawa, S. (1996). *Nucleic Acids Res.* 24, 611
- 216. Tanaka, K.S.E., Chen, X.-Y., Ichikawa, Y., Tyler, P.C., Furneaux, R.H. and Schramm, V.L. (2001). *Biochemistry* 40, 6845
- 217. Kati, W.M. and Wolfenden, R. (1989). Science 243, 1591
- 218. Wolfenden, R. (1999). Bioorg. Med. Chem. 7, 647
- 219. Deng, L., Scharer, O.D. and Verdine, G.L. (1997). J. Am. Chem. Soc. 119, 7865
- 220. Becke, A.D. (1988). Phys. Rev. A 38, 3098
- 221. Perdew, J.P. and Wang, Y. (1992). Phys. Rev. B 45, 13244
- 222. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, J. A.; Stratmann, R. E.; Burant, J. C.; Dapprich,

S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Gonzalez, C.; Challacombe, M.; Gill, P. M. W.; Johnson, B. G.; Chen, W.; Wong, M. W.; Andres, J. L.; Head-Gordon, M.; Replogle, E. S. and Pople, J. A. (1998). Gaussian-98, Gaussian, Inc., Pittsburgh, PA, 1998